

#### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

#### REGION 6 1445 ROSS AVENUE, SUITE 1200 DALLAS TX 75202-2733

October 20, 2015

Sarah Clem ADEQ Branch Manager Arkansas Department of Environmental Quality 5301 Northshore Drive Little Rock, AR 72118-5317

Re: Environmental Protection Agency comments on the Proposed 3<sup>rd</sup> party rule by the Southwestern Electric Power Company John W. Turk, Jr. Power Plant (SWEPCO facility)

Dear Ms. Clem:

The Arkansas Pollution Control and Ecology Commission (APC&EC) held a public hearing on November 17, 2014, to receive comments on a third-party proposal by the Southwestern Electric Power Company John W. Turk, Jr. Power Plant (Turk/SWEPCO) regarding water quality standards changes to APC&EC Regulation 2. The proposed amendments to Regulation 2 include a modification of the Total Dissolved Solids (TDS) water quality criterion and temperature criterion for the Little River from Millwood Lake to the mouth of the Little River, and modification of the TDS water quality criterion and removal of the designated, but not existing, domestic water supply use for the Red River from the mouth of the Little River to the Arkansas/Louisiana state line. The Environmental Protection Agency Region 6 (EPA) would like to offer the following general and enclosed more detailed comments for the Arkansas Department of Environmental Quality's (ADEQ) consideration.

We are concerned that the supporting use attainability analysis (UAA) did not provide a clear basis to support the contention that only naturally occurring sources of TDS are causing the impairment in the Little River. We acknowledge that there are predominately natural and anthropogenic sources of minerals/TDS introduced into the Red River through Texas and Oklahoma as described in the supporting UAA. These sources are some 200 miles upstream of confluence of the Little River. Further, the UAA did not provide information on additional sources of minerals or sources of dilution in Arkansas that may influence conditions in the Red River. Although there may be some basis to support a revised TDS criterion below the confluence with the Little River to some point downstream, given that the designated fishery use is being attained in the Red River, a revised criterion from the confluence of the Little River to the Arkansas/Louisiana line does not appear to be clearly supported.

EPA appreciates the opportunity to comment on the proposed 3<sup>rd</sup> party proposal. If you have any questions, please contact me at (214) 665-6646.

Sincerely,

Russell Nelson

Regional Standards Coordinator Watershed Management Section

Enclosure

#### **Technical Comments on:**

Exhibit F
Use Attainability Analysis
For Dissolved Minerals in Little and Red Rivers
Hempstead and Little River Counties, Arkansas (September 5, 2014)

Southwestern Electric Power Company, Inc, Dallas, TX NPDES No. AR0051I36

UAA Prepared by FTN Associates, Ltd., Little Rock, Arkansas

These comments are being provided to the Arkansas Department of Environmental Quality (ADEQ) for the document titled *Use Attainability Analysis for Dissolved Minerals in Little and Red Rivers, Hempstead and Little River Counties, Arkansas.* This use attainability analysis (UAA) is intended support the modification of Arkansas water quality standards (designated uses and water quality criteria), to remove the domestic water supply use in the Red River and establishment of site-specific criteria for total dissolved solids (TDS) in both the Red and Little Rivers, within the Gulf Coastal ecoregion.

#### **Technical Comments**

#### **Executive Summary**

#### ES.7 Proposed TDS Criteria Little River

#### Page xi:

The proposed TDS criterion for the Little River below Millwood Lake is based on a weight-of-evidence approach utilizing a mass balance model. The document states that a mass balance was prepared that would reflect naturally occurring elevated TDS concentrations in the Little River. The document refers to TDS data collected upstream of SWEPCO by FTN and SWEPCO during October 2010 through October 2013 that showed that the current site-specific TDS criterion 100 mg/L criterion was exceeded approximately 10% of the time. The authors conclude that a TDS criterion of 136.1 mg/L would be necessary in the Little River to reflect the naturally occurring mineral regime and to allow SWEPCO to operate efficiently.

It's unclear what the basis for presuming that only naturally occurring sources of TDS result in the 10% exceedance rate. Later, the document refers to input from the Little River itself above Millwood Lake and from Mine Creek, noting that inputs from the latter are significantly higher per volume than other tributaries to Millwood Lake. The name Mine Creek alone suggests that this creek is a likely source of anthropogenic. Given the mass balance budget described in Table J.1 describes the background in the Little River as 98 mg/L and the SWEPCO discharge as contributing 1620 mg/L, it is unclear how a presumption that the inputs to the Little River are

anthropogenic. This tends to undermine the validity of the mass balance and any conclusions that may be drawn from it.

Differentiating between natural background and anthropogenic sources causing exceedances is important. As articulated in Tudor Davies 1997 policy memo, the water quality standards regulation at 40 CFR 131.11 (b)( I) requires States and authorized Tribes to adopt numeric water quality criteria that are based on section 304(a) criteria, section 304(a) criteria modified to reflect site-specific conditions, or other scientifically defensible methods.

In summarizing the aquatic life evaluation, the document indicates that the aquatic life composition of the current fish community below Millwood dam is a subset of what would be expected given available habitat. What is described here represent a snapshot of what is likely impacted and/or degraded conditions in the Little River. While document current conditions is an important, the objective of a UAA should be determining what the highest attainable use that can be attained. The objective of a UAA is to determine what the highest attainable use the waters of interest can support. 40 CFR 131.10(d) requires at a minimum, that uses be deemed attainable if they can be achieved by the imposition of effluent limits required under sections 301(b) and 306 of the Act and cost-effective and reasonable best management practices for nonpoint source control.

#### 1.0 Introduction

#### 1.1 Background and Overview

#### Page 1-1:

The discussion here appears to be suggest that the (predominate) natural (and anthropogenic) mineral and TDS sources from TX and OK are not adequately accounted for and result in exceedances of AR's 500 mg/L criterion in the lower segment of the Red River below the confluence of the Little River to the AR-LA state line. Since there is certainly some dilution in the Red River as it moves to the confluence with the Little River and on downstream, it will be critical to provide clear support that these predominately natural upstream in TX and OK, and those in AR contribute to an inability to attain the current 500 mg/L criterion in the lower segment. The spatial inconsistence identified in and of themselves are not a basis for modifying criteria.

#### Page 1-1 and 1-3:

In describing this permit as not typical, the authors are referring to ADEQ's use of the 500 mg/L TDS criterion that applies to the Red River as an "end of pipe" value rather than the 100 mg/L TDS criterion that applies to the Little River. Based on the fact sheet, it appears that ADEQ based 500 mg/L TDS criterion in the SWEPCO permit on the applicable criterion in the Red River to ensure that the discharge would not contribute to the impairment of the Red River because it is on the state's most recent EPA approved 303(d) list for exceedances of chloride, sulfate and TDS.

Region 6 permitting staff have suggested that if the 100 mg/L TDS criterion applicable to the Little River were applied, when dilution and mixing are considered, a limit of 3800 mg/L would

be possible and would be protective of the designated fishery use in both the Little River and the Red River.

#### Page 1-4 Table 1.2:

The increase for both median and 95th percentile concentrations of TDS from the historical data the more recent data collected by SWEPCO suggest that the concentrations are the result of something other than "naturally occurring exceedences" as suggested in the next paragraph.

#### Page 1-5:

The authors refer to little to no assimilative capacity for TDS in the Little River at times (absent the SWEPCO facility's discharge) as a result of "naturally occurring exceedances" of the current site-specific 100 mg/L TDS criterion occurring "upstream" of the SWEPCO discharge, refers to more in depth discussed in Sections 3.0 and 5.0. However, there is very little discussion of possible upstream naturally occurring" sources for the Little River in these sections. The only sources for the Little River discussed in the document to any degree are tributaries to Millwood Lake, particularly Mine Creek. Although Mine Creek is identified as the most significant contributor of minerals to Millwood Lake and downstream to the Little River, there is no information concerning possible mining impacts to that creek. As a result, the basis for attributing a lack of assimilative capacity and a 10% exceedance rate for the current 100 mg/L TDS criterion in the Little River to naturally occurring sources has not been established. What are these natural sources of high TDS - Mine Creek? And are they natural or anthropogenic?

2.0 Background

2.3 Existing Water Quality and Impairments

2.3.1 Little River

2.3.1.1 TDS Sources to the Little River

This section includes an evaluation of existing data from tributaries upstream of Millwood Lake to identify potential recent sources of TDS that might explain the increased TDS values in samples collected from the Little River downstream of the dam on Millwood Lake. The major tributaries of Millwood Lake include the Little River, Cossatot River, Saline River, and Mine Creek (Figure 1.1). Table 2.2 identifies the major tributaries to Millwood Lake, including the Cossatot River, Saline River and Mine Creek and summaries of their TDS contribution. Although Mine Creek only contributes 2% of the inflow, the TDS concentrations are significantly higher than the other streams.

There is some inconsistency between Tables 1.1 and 1.2 presenting USGS data from 1967-1995 and SWEPCO data from 2010-2013 (although it's not clear what sites the SWEPCO data came from) that show an increase in TDS concentrations over time. The time-series plots of TDS data from these ADEQ monitoring stations on the tributaries upstream of Millwood Lake identified in Figures 2.1 through 2.4 that show no clear evidence of significant changes in TDS levels in any of the streams, yet the authors report an increase in TDS concentration downstream of Millwood Lake.

#### 2.3.1.2 Water Quality Impairments for Minerals in the Little River

The authors indicate that ADEQ does not considers the Little River downstream of Millwood Lake to be impaired and have not included it on any 303(d) list. Further, the document notes that this stream is classified by ADEQ as "unassessed" due to a lack of data for water quality (including minerals). It is important to note that "unassessed" simply means that there is no data and does not necessarily mean that the Little River is either impaired or unimpaired. That lack of data may be why neither ADEQ nor EPA have included the Little River on the state's 303(d) list.

2.3.2 Red River

2.3.2.1 TDS Sources to the Red River in Arkansas

2.3.2.2 TDS Sources to the Upper Red River (Oklahoma, Texas)

#### Pages 2-5 to 2-8:

Much of the literature cited in the Executive Summary and in these sections is dated, relying on several sources from a time period when oil production was high with minimal environmental regulation. Despite those issues, it is reasonable to say that there are significant natural as well as anthropogenic sources of minerals in the Red River and its tributaries.

While the more current information tends to show that the predominant mineral sources to the Red River are likely natural, despite remediation efforts by the US Army Corps of Engineers (USACE), it is likely that brine disposal and intrusions from oil production into the Little Wichita and other sites are sources that affect the Red River in Texas, Oklahoma and contribute to the loading to a lesser extent in Arkansas. Sources of information for oil and gas production and remediation should be available from the Texas Railroad Commission and the Oklahoma Corporation Commission.

Not having, or possibly not being able to provide any information to support historical oil extraction impacts and their contribution to minerals/TDS concentrations significantly weakens the claim that the "overwhelming portion" historical of the TDS inputs to the system are from natural sources.

#### 2.3.2.3 Water Quality Impairments for Minerals in the Red River

#### Page 2-9:

The authors note that upstream criteria for the Red River in Texas and Oklahoma reflect the elevated ambient mineral concentrations that are due to inputs from salt springs and seeps from the arid and semi-arid headwater regions. This is generally true, given salt springs are found as far west as the High Plains and Tableland Level II ecoregions. Although less predominate, there also anthropogenic and natural inputs from the Central Plains, Crosstimbers, Blackland Prairie and East and South Central Plains ecoregions.

It is important for Arkansas to recognize that an upstream state can establish uses and criteria in their waters as appropriate. 40 CFR 131.10(b) requires that those states take the downstream state's standards. Recent court decisions have affirmed EPA's ability to look at downstream waters to ensure protection.

This section also discusses assessment procedures used by states that share jurisdiction over portions of the Red River. Referring to Table 2.6, which compares criteria for minerals and impairments identified on state 303(d) lists, which shows that the Red River is not considered to be impaired upstream in Oklahoma and Texas or downstream in Louisiana. The authors state that this is partly due to different assessment procedures that are used in other states, suggesting that these states use less protective exceedance rates. As an example, the authors note that Louisiana does not consider a stream to be impaired for dissolved minerals unless 30% of the observed data exceed the water quality standard compared to Arkansas' 10% exceedance rate. The authors also state that Oklahoma and Texas also use less-stringent assessment procedures for dissolved minerals than Arkansas, and also state that the criteria in their water quality standards are less stringent than the criteria in Arkansas's standards. However, the authors have not explained how these approaches are any less protective than those used by Arkansas.

It is important to understand that simply comparison of exceedance percentages used in assessing use attainment does not tell the entire story. Assessing attainment includes defining the water quality indicators it measures and the procedures for analyzing and interpreting data in order to decide whether standards are met or water quality is impaired. This should include collection and analysis of multiple types of data providing information relevant to assessing attainment with approved WQS. As a result, a comparison of Louisiana's current use of a 30% exceedance rate to Arkansas 10% is not necessarily appropriate, although it should be noted that Region 6 is working with Louisiana to revise its approach. Oklahoma compares the mean of all values to a yearly mean standard, then looks for greater than 10% exceedances against a sample standard. In Texas, minerals criteria represent annual averages of all values that were collected and compared to the criterion for each parameter. Use attainment is based on the average concentration that applies to the entire length or area of the segment. For TDS, a value is calculated by multiplying specific conductance measured at the surface by a factor of 0.65. The chloride, sulfate, and TDS criteria are not supported if the average value exceeds the criteria.

The authors also state that Oklahoma and Texas water quality standards as less stringent than the criteria in Arkansas. Given the natural saline seeps and historical oil extraction activity along the Red River that this document discussed previously, higher criteria would be expected upstream in Texas and Oklahoma. The burden for SWEPCO is to show that predominately natural sources upstream in Oklahoma and Texas upstream as distant as Lake Texoma and/or that natural sources in Arkansas itself alter concentrations in the lower segment of the Red River to support the proposed criteria modifications.

#### Page 2-10:

In a discussion of Table 2 describing applicable criteria and 303(d) listings, the authors refer to ADEQ's practice in recent years of making specific permitting decisions and calculations based on the state's most recent 303(d) list that has been approved by EPA. This approach means that ADEQ is applying the 500 mg/L TDS criterion that applies to the Red River below its confluence with the Little River as an end-of-pipe limit rather than the 100 mg/L criterion that applies to the Little River which is the receiving water for the Turk/SWEPCO discharge.

Based on the NPDES permit fact sheet, applying the 100 mg/L TDS criterion that applies to the Little River, when dilution and mixing is considered, a limit of up to 3800 mg/l would be possible and still meet the in-stream criterion of 100 mg/L which would protect beneficial uses in the Little River and downstream. However, applying the 500 mg/L TDS criterion as an end-of-pipe limit results in a more restrictive permit limit. Without any information to the contrary, the effect of ADEQ's approach does not appears to have a significant environmental benefit for either the Little River or Red River, but appears to have the opposite effect. While applying 500 mg/L TDS criterion as an end-of-pipe limit may result in a small reduction in overall loads, the more restrictive limit reportedly requires the facility to operate at less than optimum efficiency. Thus ADEQ's approach appears to be the primary driver for Turk/SWEPCO to seek revised criteria for the Red River, which may be a less desirable environmental outcome.

# 2.4 Threatened and Endangered Species 2.4.1 Threatened and Endangered Aquatic Species

#### Page 2-12:

This section of the document discusses several endangered species listed by the US Fish and Wildlife Service (USFWS) including the Ouachita rock pocketbook (Arkansia wheeleri), which is federally listed throughout its range in Arkansas and Oklahoma. The document reports that there have been no live A. wheeleri collected from just above or below the Turk/SWEPCO discharge. Their absence indicates that the Turk/SWEPCO discharge is having an effect on water quality and the presence of A. wheeleri. Although the physical characteristics and inputs to Millwood Lake may affect the ability of the Little River to support A. wheeleri, an important aspect of a UAA is to not only to identify current conditions. The document does not provide any discussion of whether minerals/TDS or other stressors may be affecting or limiting the presence of A. wheeleri in this portion of the Little River.

The authors also note that the USFWS recently listed the rabbitsfoot mussel (*Quadrula cylindrica cylindrica*) as threatened (Federal Register 2013). That species is known from some streams in southwest Arkansas, including the Little River upstream of Millwood Lake. This species not been documented in the Little River downstream of Millwood Lake or the Red River. The rabbitsfoot is likely limited by the physical characteristics Millwood Lake and its effect on the Little River, as well as the Turk/SWEPCO discharge.

## 3.0 FIELD SURVEY 3.1 Overview

#### Page 3-1:

Rather than relying on biological evaluations in a upstream versus downstream comparison from a point source discharge and a comparison of communities in reference and receiving streams, the approach for field surveys is attempting to show that characteristics in Millwood Lake and at the Denison dam dominate the mineral regime in the Little and Red Rivers rather than the Turk/SWEPCO discharge.

There are sources of minerals above Millwood dam, primarily the Little River and Mine Creek upstream of Millwood Lake and there are certainly significant natural and anthropogenic sources of mineral in the upper reaches of the Red River in Texas and Oklahoma. The burden for Arkansas is to clearly show that the sources above Millwood dam are at least predominantly anthropogenic. Similarly, Arkansas must be able to make a reasonable showing that the predominately natural and some anthropogenic sources some 200 miles upstream have a significant effect on water quality in the Red River as far downstream as its confluence of the Little River and show that there are no natural or anthropogenic sources from the TX-AR line to the confluence.

If the Turk/SWEPCO discharge is not contributing to the potential exceedances in the Little River or potentially impacting the Red River, the important question as to whether the facility is seeking to revise the criteria in the Little River and the Red River because the criteria are inappropriate or because of ADEQ's policy decision to applying a criterion as an end-of-pipe limit rather than basing it on applicable criteria considering dilution and mixing in the receiving water.

#### 3.4.1.2 Macroinvertebrate Data Analysis

#### Page 3-10:

In discussions of field observations the authors indicated an attempt was made to assess the susceptibility of the existing benthic macro invertebrate community to a slight increase in TDS. Tolerance indicator values (TIVs) for specific conductance for various invertebrate taxa provided in Carlisle et al. 2007 were reviewed. However, the authors note that too few of the taxa listed in Carlisle et al. 2007 were present in the data set for this study to allow for a meaningful evaluation of ion tolerance. Accordingly, the benthic data presented herein provide a description of the presently attained fisheries use and a baseline for future studies.

It should be noted that small increases in TDS alone may have minimal effect on the existing benthic community however, TDS is rarely seen independent of other stressors. In a quick search, a number of sources were identified that discuss the effect TDS has on Appalachian streams. For example, Timpano et al. (2010), in discussing the effects of TDS in coalfield streams. As a way of isolating the effects, Timpano et al. used headwater streams to represent a gradient where confounding factors could be minimized. This and other similar papers done in Appalachian could have been used to inform this aspect of the analysis. One of the problems with the approach take here is a lack of reference stream comparisons.

#### 3.4.2.2 Fish Data Analysis

#### Page 3-11:

Fish data were first assessed to determine if there were impacts due to the Turk/SWEPCO discharge "under current conditions" by evaluating differences in total species richness, relative abundance, diversity, and similarity measures between upstream and downstream reaches.

Although unclear, it appears that "under current conditions" likely means under limits specified in the current NPDES, which as noted earlier, are based on a monthly average of 500 mg/l TDS

applying Red River Criteria end-of-pipe. This end of pipe value equates to an instream waste concentration of 43 mg/L. As noted earlier, if ADEQ were to base the Turk/SWEPCO permit on the Little River instream standard of 100 mg/L, once dilution and mixing is allowed, this would result in an allowable end-of-pipe discharge of up to 3800 mg/l to still meet the instream waste concentration of 100 mg/L. These conditions should be modeled and presented as a comparison rather than solely those based on ADEQ's current approach of basing permits on past 303(d) listings. Without this, there is some question to the validity to the findings here and in other aspects of this proposal.

#### 3.4.4 Discussion and Conclusions

#### Page 3-22:

The authors note that the evaluation of the fish community demonstrated similar fish communities upstream and downstream of the Turk/SWEPCO discharge in the Little River. The combined collections from their study and USGS sampling during 2009 and 2013 provide evidence that the designated fisheries use in the Little River is currently attained under the existing mineral conditions.

Further, the document reports that an evaluation of the mineral tolerances of the existing fish community indicate a fish community that can be expected to be relatively insensitive to moderate changes in TDS (as indicated by specific conductance). The UAA notes that fish species that were classified as intolerant to TDS or for which data in Meador and Carlisle (2007) were lacking can be shown to be present in the Red River, which has substantially higher TDS than will be experienced in the Little River due to the operation of the Turk/SWEPCO plant. The conclusion drawn is that the anticipated increase in TDS due to the operation of the Turk/SWEPCO plant will support the designated fisheries use in the Little River downstream of the Millwood Lake dam.

The question these conclusion raise is even if the designated fisheries use in the Little River would potentially be maintained with the proposed increase in TDS criterion to 138 mg/L, why is it necessary if that use is being attained at the current 100 mg/L criterion? The answer seems to go back to Turk/SWEPCO's desire to operate at maximum efficiency. Under current conditions, the facilities efficiency appears to only be affected by ADEQ's approach in using limits based on the 500 mg/L criterion for the Red River as an end-of-pipe value. As noted previously, given that the Turk/SWEPCO discharge appears to have a negligible effect on the Red River, if ADEQ were to use existing criteria and allow for mixing and dilution in the Little River, there would be no need for this proposal.

#### 3.5 Fisheries Evaluation in the Red River

#### Page 3-24:

The objective of this evaluation was to determine whether the existing mineral regime supports the designated fisheries use as indicated by fish populations. The approach relied on the historical record of mineral concentrations in the Red River from ADEQ monitoring stations RED0025, RED0046, and RED0045 and the use of a sampling survey and historical data review

of fish populations in the Arkansas portion of the Red River conducted by Buchanan et al. (2003). The intent was to characterize the existing fish communities and evaluate whether those existing communities are supported by the existing mineral regime, thus indicating attainment of the fisheries designated use.

Although the Buchanan et al. (2003) data suggest that mineral conditions during 1995 and 2001 supported a high fisheries use attainment, it is somewhat problematic to rely on data that represents a single snapshot in time from 14 years ago. The authors draw the conclusion that since the Red River was meeting designated uses during the Buchanan et al. study and that because TDS data from the post-study period (2002 through 2013) indicate that mineral concentrations where generally higher during the time frame of the Buchanan et al. study, that fisheries beneficial uses are likely currently being attained. While the post-study period (2002 through 2013) TDS data gives an indication that uses are currently being attained, it does not provide the same level of confidence as current field data would. Showing that broadly written designated aquatic life uses can be supported under the existing mineral regime isn't necessarily a high bar given how broadly beneficial use definitions tend to be written. Determining attainment tends to depend on the individual state's assessment methodologies rather than the use definition.

# 3.6 Biological Evaluation: Summary and Conclusions 3.6.1 Little River 3.6.2 Red River

#### Page 3-25:

The evaluation of the biological data demonstrated similar fish and communities upstream and downstream of the Turk/SWEPCO facility. The authors conclude that these finding reported suggest that the designated fisheries use is being attained that that the projected mineral concentration - presumably the proposed 138 mg/L TDS criterion will not be adversely affect attainment of the fisheries use.

Although the data indicate that the designated fisheries use is currently being attained, the document does not provide compelling information to support the contention that the proposed 138 mg/L TDS criterion will not adversely affect attainment. Further, concerns with impacts to benthic macroinvertebrate communities downstream of the discharge at the current criterion have been identified in previous comments.

#### 4.0 TOXICITY ANALYSIS

#### 4.1 Evaluation of Routine WET Testing

#### Page 4-1:

The recurring concern here is that the conditions simulated for toxicity analysis are reflective of current permit conditions, not what would likely be the conditions if the Red River was not listed.

More specifically, section 4.1 reports that a review of NPDES DMRs from what is likely the current permit (from November 1, 2012) indicate no episodes of sub-lethal toxicity for *C. dubia* and *P. promaleas*. Since the current permit is based on 500 mg/L as an end-of-pipe limit, sub-lethal effects would not be expected. The question is, how C. dubia and *P. promaleas* would respond under conditions that would be expected if a limit of up to 3800 mg/L was in place for the facility as it may be once the current 303(d) listing in the Red River is resolved.

Relying on DMRs that are not reflective of what conditions will likely be long-term in the Little River and downstream in the Red River could be considered misleading.

#### 4.2.3 Conclusions

#### Page 4-5:

The authors state that calculated IC25 values (1,744 mg/L for TDS; 232 mg/L for sulfate; and 600 mg/L for chloride) indicate that toxicity due to minerals in WET testing would not be expected at the critical dilution (26%) until effluent TDS values reached approximately 6,976 mg/L. And that these results indicate that the threshold for toxicity due to minerals (as TDS, sulfate, and chloride) is well above anticipated mineral concentrations at the critical dilution of 26%. Accordingly, their conclusion is that there is a large margin of safety associated with mineral discharges from the plant with respect to compliance with NPDES WET testing.

It is unclear what the basis for the statement that toxicity due to minerals would not be expected until TDS values reach approximately 6,976 mg/L. Although the IC25 may be considered functionally equivalent to a NOEC by definition, it represents a 25% reduction in the rate of survival, growth or reproduction among test organisms. Further, Table 4.2 indicates that average concentrations of 6,000 mg/L cause 100% lethality.

# 5.0 MASS BALANCE MODELING5.2 Critical Flow Conditions

#### Page 5-1:

Instream TDS concentrations in the Red River were modeled for two critical flow conditions. The authors note that calculations and model results for 7Q10 flow conditions were used to develop proposed criteria for the Little River, and for the Red River between the Little River and the AR-LA state line. The results for harmonic mean flow conditions were used to confirm that the proposed criteria in Arkansas will still allow the Louisiana criterion for TDS in the Red River (780 mg/L) to be maintained.

It's important that ADEQ be aware that the State of Louisiana is currently reviewing its mineral criteria. It is EPA's understanding that water quality data taken in the Red River near Shreveport, between 1991 and 2014 indicate that TDS concentrations of 780 mg/L are just above the 90th percentile. This may mean that it if the Arkansas' proposed criteria are adopted, there is a significant likelihood of TDS impairments in the downstream segment of the Red River in Louisiana.

It is important to note that 40 CFR 131.10(b) requires states to consider the water quality standards of downstream states when revising their criteria.

#### 5.5 Model Results for 7Q10 Conditions

#### Page 5-7:

Neither the RED0046 data nor the RED0009 data were considered to be a good representation of well-mixed concentrations in the Red River by the authors. Both the RED0046 station and the RED0009 station are located where the Red River does not appear to be laterally well-mixed because of significant tributary inflows a short distance upstream of each station.

The discussion explains that because of the location of the sampling sites for both RED0046 and RED0009, the data are likely measuring a disproportionate percentage of tributary water that has not mixed completely with the entire flow of the Red River. The authors contend that both tributaries (especially the Little River) contribute water that typically has TDS concentrations much lower than concentrations in the Red River. Because these two stations appear to be underestimating the TDS concentrations in the Red River and should not be used for comparison with predicted TDS values from the mass balance modeling, presumably because their inclusion could result in an underestimation the TDS concentrations in the Red River as compared to predicted TDS values from the mass balance modeling.

While it is possible that the inflow from the Little River at (RED0046) and the Sulphur River at (RED0009) may not be well-mixed, the problem with leaving them out is that it will likely mean an underestimation of TDS concentrations to the Red River, particularly in the lower reach near the LA state line.

It is unclear how the segment reaches in Table 5.4 compare spatially to the long-term sampling sites.

#### 5.6 Model Results for Harmonic Mean Conditions

#### Page 9:

The authors explain that the reason for simulating harmonic mean conditions was to confirm that proposed TDS criteria for the Red River in Arkansas would allow the Louisiana criterion for TDS in the Red River to be maintained under critical conditions. The harmonic mean simulation presented predicts that the TDS concentration at the AR-LA state line will be 661 mg/L, which is well below the Louisiana criterion for TDS in the Red River (780 mg/L). The conclusion is that the proposed criteria based on the mass balance modeling will be protective of the Louisiana criterion for TDS.

However, the harmonic mean simulation used the same input TDS concentrations as the 7Q10 simulation. As noted in the previous section, the exclusion of inflow from the Little River at (RED0046) and the Sulphur River at (RED0009) my mean an underestimation of TDS concentrations in the Red River, resulting in potential impairment of the current 780 mg/L TDS criterion that applies at the LA state line. As noted in comments on section 5.5, it is EPA's understanding that water quality data taken in the Red River near Shreveport, between 1991 and

2014 indicate that TDS concentrations of 780 mg/L are just above the 90th percentile. The 780 mg/L value is likely to go down.

It would be advisable to consider what potential effect lower criteria in the Red River at the AR/LA state line may have on the predicted TDS criterion of 661 mg/L if LA revises its criteria down.

# 6.0 DESIGNATED USES6.2 Red River6.2.2 Domestic Water Supply

#### Page 6-4:

The domestic water supply use for the reach of the Red River between the OK-AR state line and the Little River confluence has previously been removed. The authors provide data that indicate that the reach of the Red River from the Little River confluence to the AR-LA state line can also be expected to exceed secondary drinking water standards due to the same factors. Figure 6.1 shows that the Red River TDS concentrations at Fulton (downstream of the mouth of the Little River) frequently exceeded the secondary drinking water standard of 500 mg/L during the period of record. The authors also noted that both the Arkansas Department of Health and the Arkansas Natural Resource Commission to confirm that the stream is not used as an existing domestic water supply. Their conclusion is that the domestic water supply use is neither an existing nor attainable use in the Red River downstream of the mouth of the Little River.

Although it is within Arkansas' authority to remove the domestic water supply use for the segment of the Red River below the confluence with the Little River to the LA state line, AR must consider the downstream state standards and ensure that its action would not affect domestic water supply/drinking water uses in Louisiana. See 40 CFR 131.10(b).

#### 7.0 ALTERNATIVES EVALUATION

#### Page 7-1:

Arkansas' UAA guidance and CPP require that a petition to increase dissolved minerals concentrations above existing conditions must include an evaluation of alternatives to the direct discharge of the water. Four alternatives that have been reviewed including:

- 1. Distillation treatment,
- 2. Treatment using a constructed wetland,
- 3. Pumping the wastewater to a larger stream that holds the potential for dilution of the minerals, and
- 4. Reverse osmosis (RO) treatment of the wastewater.

From the outset, the use of constructed wetlands was dismissed as an option for this facility. The authors based this exclusion on the argument that constructed wetlands can only be used to reduce sulfate, which results in the production of bicarbonate in place of sulfate (Hedin et al.

1989). The authors do note that a constructed wetland could, in principle, reduce sulfate in the discharge from this facility, the resulting TDS concentration would not be decreased (due to the replacement of the sulfate ions with bicarbonate ions), but argue that no net benefit would be obtained, stressing that TDS is the only concern for this evaluation.

The premise in rejecting a constructed wetland appears to be based on the premise that a constructed wetlands would only reduce sulfate to bicarbonate, which would not provide a net reduction in TDS. Looking at the Hedin et al. paper, it is specific to the treatment of acid mine drainage, not the type of discharge that would result from blow down cooling water from a power plant. The paper does indicate that with an increase in sulfate reduction, bicarbonate alkalinity will be produced and pH will increase, but it does not say that sulfate is directly reduced to bicarbonate. In treatment of acid mine drainage, sulfate would primarily be reduced to sulfite and tend to react with available metal to form sulfide precipitates. The Hedin paper refers to bacterial production of bicarbonate formation, particularly in anoxic limestone drains, but does not refer to the reduction of sulfate to bicarbonate. Which is not unexpected since as noted earlier, the paper is looking at treatment of acid mine drainage, not sulfate or other TDS constituents in particular. Given this, EPA considers a constructed wetland to be a viable and the preferred alternative to altering the applicable TDS criterion.

The authors contend that there are two alternatives for achieving compliance with the existing TDS criteria for the Little River and Red River:

- 1. RO treatment to remove or reduce dissolved minerals, and
- 2. Pumping the effluent to a larger river that holds the potential for dilution of the minerals.

In considering these options, the authors state that there is not a river in the vicinity that could be used as dilution to completely avoid a change in the water quality standards. Although the Red River has sufficient volume, it has a TDS criterion of 500 mg/L, the expected concentration of TDS in the effluent from this facility would not meet the Red River TDS criterion and would still require the completion of a UAA or the installation of partial RO treatment.

EPA considers ADEQ's use of TDS criteria applicable to the Red River, particularly the way the criterion has been applied as the emerging central element here. As noted in previous comments, ADEQ issued permit Turk/SWEPCO is operating under is based on the existing 500 mg/L TDS criterion for the Red River as end-of-pipe limits. The permit carries those limits because the receiving Red River segment was on the state's 2008 303(d) list. From a numeric perspective, ADEQ has issued the permit with a monthly average of 500 mg/l TDS end-of-pipe. This end-of-pipe value equates to an instream waste concentration of 43 mg/L. If ADEQ based the permit on the Little River instream standard of 100 mg/L, once dilution and mixing is allowed, the result is an allowable end-of-pipe discharge of up to 3800 mg/L TDS. While the end-of-pipe concentration is higher, this approach would meet the instream waste concentration based on the ecoregion-based 100 mg/L TDS while allowing the Turk/SWEPCO facility to operate efficiently.

EPA strongly recommends that rather than revise TDS criteria particularly for the Little River, in what appears to be an effort to avoid this permitting scenario, that this approach be considered.

#### 7.1.2.2 Option 2: RO Treatment/Discharge to Little River at Future Effluent Limits

#### Page 7-9:

The authors anticipate that the TDS criterion in the Little River, and thus the effluent limitations in future NPDES permits for the SWEPCO plant, could be lowered to 100 mg/L. The conclusion drawn is that this would require that all of the effluent be treated through an RO system if it continues to be discharged to the Little River, in order to achieve an effluent limit of 100 mg/L For TDS.

However, as noted previously, the 100 mg/L criterion that applies to the Little River is not the same as a derived NPDES permit limit. The permit limit that would be derived from the 100 mg/L TDS criterion once dilution and mixing is allowed, would likely be an allowable end-of-pipe discharge of up to 3800 mg/L. Given that this concentration would allow the facility to meet the 100 mg/L TDS criterion in the Little River, there would be no need to consider RO.

#### 7.3 Summary of Costs

#### Page 7-10:

The authors summarize the estimated costs of various treatment scenarios compared to the cost of direct discharge based on the proposed site-specific criteria in Table 7.1. The cost described here appear to be the potential cost associated with RO treatment of the entire volume of the Turk/SWEPCO discharge. It is unclear what volume of the discharge would need to be treated.

#### **Technical Comments on:**

EXHIBIT G
TECHNICAL JUSTIFICATION FOR A SITE-SPECIFIC TEMPERATURE
CRITERION IN THE LITTLE RIVER, HEMPSTEAD & LITTLE RIVER COUNTIES,
ARKANSAS

#### **Technical Comments**

# 1.0 INTRODUCTION 1.1 Background & Overview

#### Page 1-1:

This document states that the Turk/SWEPCO facility's National Pollutant Discharge Elimination System (NPDES) permit (No. AR0051136) includes a daily maximum temperature permit limit of 35 °C (95 °F). Given that Arkansas Reg. 2 specifies and the authors acknowledge that the applicable temperature limits for streams is 30°C (86°F), the basis for the maximum temperature limit of 35°C (95°F) in SWEPCO's current permit is unclear.

The document also notes that the Little River downstream of Millwood Lake has a lower temperature criterion than Millwood Lake and the Red River immediately downstream of the Little River. Both Millwood Lake and the Red River in Arkansas have a temperature criterion of 32 °C (89.6 °F) (APCEC 2011). Figure 1.2 is presented to illustrate the spatial inconsistency of the temperature criteria.

The document notes that Millwood Lake has a temperature criterion of 32°C (89.6°F), the Little River, downstream of Millwood Lake has a 30°C (86°F), and the Red River immediately downstream of the Little River has a temperature criterion of 32°C (89.6°F) (APCEC 2011) to illustrate the spatial inconsistency of the temperature criteria downstream of Millwood Lake. Spatial inconsistency has no bearing on toxic or conventional criteria that are protective of aquatic life uses (ALU) and is not a basis for altering applicable criteria. It is not unusual for temperatures in a reservoir to be higher than upstream and typically downstream lotic waters given the retention time and particularly the shallow depth of Millwood Lake.

Although we acknowledge that it may be difficult to manage the flow through a lake as shallow as Millwood given its design, relatively high retention time and higher tailwater race temperatures, but it's unclear how that would affect the entire length of the Little River or why the riparian cover downstream along the Little River could not be enhanced or restored.

#### Page 1-3:

The authors note that if the temperature criterion is not changed and the impairment status continues, the facility's NPDES permit limit for temperature at Outfall 001 would be reduced to the Little River instream criterion of 30°C (86°F) without the usual allowance for a mixing zone (i.e., using typical end-of-pipe calculations to develop permit limits for a thermal discharge). As such, the authors note that facility's operation would be adversely impacted by a temperature impairment designation in the Little River that may limit or prevent discharges during warm weather and reduce the efficiency of the facility's operation during other times of the year.

Does this mean that the decision was keeping the existing criteria in place versus operating efficiency (i.e., cost)? If so, the basis for the proposed amendment 40 CFR 131.10(g)(6), which requires an economic analysis to determine if SWEPCO can support upgrades to its process/facilities w/out substantial and widespread impacts.

#### Page 1-4:

The authors indicate that adjusting the temperature criterion for the Little River downstream of Millwood Lake is necessary to reflect current ambient conditions during the critical summer conditions and would result in NPDES temperature limitations that are appropriate and protective of the designated uses of the Little River. An important question is if current ambient conditions reflect the highest attainable condition given the requirements outlined in 40 CFR 131.10?

### 2.0 BACKGROUND 2.1 NPDES Permit

#### Page 2-1:

The document again refers to the Turk/SWEPCO discharge to the Little River under NPDES Permit No. AR0051136 and the effluent limitation for temperature of 35°C (95°F). As noted previously, it's unclear how this facility can have an NPDES permit that allows a 35°C (95°F) when the applicable temperature for streams is The 32°C (89.6°F). Is there a specific implementation procedure or mixing zone allowance?

#### 2.2 Applicable Water Quality Standards

#### Page 2-2:

The authors suggest that the temperature regime of the Little River downstream of Millwood Lake has been altered as a result of physical hydrologic alteration of the system caused by the construction of Millwood Lake. The authors contend that Millwood Lake and its confluence with the Little River has a typical width between 200 and 400 feet with only a small portion of the channel being shaded by trees along the banks.

Based on images of the USACE dam, the Little River may be "200-400" wide" immediately below the dam as stated here, but appears to constrict below River Run Park and appears to be generally consistent with the average stream of reference streams in the Gulf Coastal Plain ecoregion. Without supporting information for multiple downstream locations on the Little River below the Millwood, this claim does not appear to be supported. Photographs, specifically aerial photos should be included in this proposal to support this claim.

#### 2.3 Water Temperature Monitoring

#### Page 2-4:

Tables 2.4 and 2.5 presents summary statistics for water temperature data collected by SWEPCO from April to October 2012. These tables indicate that the Turk/SWEPCO discharge increases instream temperatures. With the exception of a higher maximum temperature reading upstream of SWEPCO, which may be a single event, the 95th percentile, percent daily max  $> 30^{\circ}$ C and percent daily max  $> 32^{\circ}$ C indicate that the facility adversely impacts temperature in the Little River.

#### Page 2-8:

The authors note that temperatures in the Little River during 2013 (Figures 2.3 and 2.4) appear very similar to those measured in Millwood Lake near the dam (Figure 2.5). They note that exceedances of the Little River temperature criterion occur at all of the Little River sampling locations, both upstream and downstream of the SWEPCO outfall, during 2012 and 2013 (Figures 2.2 through 2.4).

These figures generally show that there is an increase in temperature associated with the time of the year sampled. There is a clear correlation with ambient air temperature and water temperature, which among other factors can have a significant influence instream temperatures. This correlation is not considered here. The comparison to temperatures in Millwood Lake near the dam are of limited value since this lake is relatively shallow and retention times tend to be fairly high in most lakes.

In addition, the data reported in Fig. 2.8 which show a decrease in temperature from LR0 to LR00 does not appear to be consistent with the summary statistics presented for the same site in Table 2.4, which reports an increase of all and daily maximum data  $> 30^{\circ}$ C.

#### Page 2-10:

Warming in a large shallow impoundment like Millwood is expected as it is in the immediate tail water area. However, it's unclear if this narrative is suggesting that temperatures in the Little River are the result of the Turk/SWEPCO discharge or continuing the claim that the river remains wide and unshaded further downstream. As noted earlier, a simple internet search produced maps and photographs that suggest that this is not the case downstream.

#### 2.4 Evaluation of Little River Water Temperatures Upstream of SWEPCO Outfall 001

#### Page 2-10:

The discussion indicates that Table 2.5 provides an inventory of water temperature data collected in the Little River downstream of Millwood Lake and upstream of SWEPCO Outfall 001 that were used to evaluate the temperature criterion for the Little River. However, Table 2.5 also includes data from LR00, which is reported as 1 mile below SWEPCO Outfall 001.

#### Page 2-15:

A comparison of temperature statistics between the two FTN monitoring locations near the dam below Millwood Lake and the two locations farther downstream (near SWEPCO) indicate that that locations farther downstream have slightly higher temperatures during July through August and more exceedances of the 30 °C (86 °F) criterion (Tables 2.3 and 2.4).

This paragraph touches on the potential source affecting ambient temperatures in Millwood Lake and the Little River, but provide no details. The summary data, statistics and statements presented here do not appear to have considered that air temperature is highly correlated with the significant positive heat flux components including solar radiation and with stream water temperature. The correlation is supported by the USGS' Stream Segment Temperature Model (SSTEMP), which asserts that air temperature will usually be the single most important factor in determining mean daily water temperature.

The authors also note that ADEQ's (303(d)) assessment for 2014 was conducted using the SWEPCO 2012 and 2013 data and FTN 2013 data from this stream reach and that ADEQ's draft 2014 303(d) list includes a temperature impairment for the Little River downstream of Millwood Lake. Did SWEPCO and FTN use ADEQ monitoring stations to collect these data? Is there information to support the reported listing? While the assessment methodology used to determine impairment appears to be consistent with ADEQ methodology, it is unclear if 3rd party (SWEPCO and FTN) data be was used by ADEQ to support a 303(d) listing?

In instances where the temperature exceeds the criteria, it would likely result in a reduction in overall species richness and/or abundance because of excessive heat. Discussions of the macroinvertebrate and fish communities do not appear to show such effects.

# 2.5 Threatened and Endangered Species 2.5.1 Threatened and Endangered Aquatic Species

#### Page 2-18:

The document reports that there have been no live *A. wheeleri* collected from just above or below the Turk/SWEPCO discharge. Their absence indicates that the Turk/SWEPCO discharge is having an effect on water quality and the presence of *A. wheeleri*. Although the physical characteristics and inputs to Millwood Lake may affect the ability of the Little River to support *A. wheeleri*, an important aspect of a UAA is to not only to identify current conditions. The document does not provide any discussion of whether temperature or other stressors may be affecting or limiting the presence of *A. wheeleri* in this portion of the Little River.

Although the rabbitsfoot mussel (Quadrula cylindrica cylindrica) is known to occur in some streams in southwest Arkansas, including the Little River upstream of Millwood Lake, it has not been documented in the Little River downstream of the Lake to the Red River. The rabbitsfoot is likely limited by the physical characteristics Millwood Lake and its effect on the Little River, as well as the Turk/SWEPCO discharge.

#### 3.0 BIOLOGICAL EVALUATION

3.1 Overview

3.4.3.2 Fish

Ecoregion Key and Indicator Species

The authors describe presence and absence of ecoregion key and indicator species per APCEC (20 11) which is summarized in Table 3.7. Of three key and indicator species expected to be present based on habitat preferences, one species was present in the pooled list. One species not expected to be present (redfin shiner) was present in the pooled list. The reported data is not specific as to site - is this a comparison of upstream/downstream of the SWEPCO discharge? The location of this sampling would be useful, as it would give some indication of whether the presence of only 25% of key and indicator species is the result of a generally degraded/impacted system, resulting from hydromodification and high ambient air temperatures driving water temperature, or exacerbated by temperature inputs from the SWEPCO facility.

#### 3.4.4 Discussion and Conclusions

#### Page 3-22:

The authors observed that the strongly similar fish and benthic communities upstream and downstream of the discharge. The suggestion is that this finding indicates that the current operation of the discharge has negligible effect on the existing biological communities. Although relative abundance upstream/downstream showed reasonably good comparison at around 71%, the Meador and Carlisle (2007) data only showed 61% of the expected species are present and only 25% of the expected species for the Little River present in the pooled sample. It is unclear if this is the result of a degraded system or that the discharge has more than a negligible effect.

#### Page 3-23:

The discussion notes that ecoregion key and indicator species are generally not expected to be present in the reach of the Little River due to the habitat preferences of those species and that the low numbers of key and indicator species present in the pooled list (two species) is a reflection of this expectation. This statement appears to contradict Table 3.7, comparing expected key/indicator species in the Little River to what was present in the pooled list (25%).



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January 15, 2016

#### VIA ELECTRONIC DELIVERY & FIRST CLASS MAIL

Mr. William K. Honker Director Water Quality Protection Division EPA Region 6, Mail Code 6WQ 1445 Ross Avenue, Suite 1200 Dallas, TX 75202

Re:

Southwestern Electric Power Company Third Party Rulemaking Petition,

APC&EC Docket No. 14-007-R (approved October 23, 2015); and

Domtar A.W. LLC Inc. Third Party Rulemaking Petition, APC&EC Docket No.

14-008-R (approved Oct. 23, 2015)

Dear Bill:

On October 23, 2015, the Arkansas Pollution Control & Ecology Commission approved two third-party rulemaking petitions, one presented by SWEPCO and the other presented by Domtar. Three days before the Commission was scheduled to vote on final approval of the petitions, EPA sent ADEQ a 19 page set of comments on the SWEPCO petition. One day before the Commission final vote, EPA sent ADEQ a two page letter commenting on Domtar petition. I am writing this letter to transmit the SWEPCO's responses to EPA's comments on the SWEPCO petition, and to respond on behalf of Domtar to EPA's letter comment on the Domtar petition.

As you will recall from previous discussions, the SWEPCO and Domtar rulemaking petitions are closely related to one another. In fact, the Domtar petition specifically relies on the data submitted by SWEPCO and is expressly conditioned on approval of the changes requested in the SWEPCO petition. The two petitions sought the following changes in site specific water quality criteria in the Red River and the Little River downstream of Millwood Dam:

#### Changes Proposed in Domtar & SWEPCO 3rd Party Rulemaking Petitions

	Red River From AR/OK Line To Little River	Red River From Little River To LA line	Little River From Millwood Dam To Red River
<b>Domtar Petition</b>	TDS 850→940 Sulfate 200→250	Sulfate 200→ <b>225</b>	
SWEPCO Petition		TDS 500→860 Remove Domestic Water Supply Use	TDS 100→138 Temp. 30°C→32°

Mr. William K. Honker, Division Director January 15, 2016 Page 2

A map of the area involved is attached as Exhibit 1. A schematic map depicting the existing water quality criteria and the proposed changes is attached as Exhibit 2. A graphic depiction of the proposed changes to the Red River TDS and sulfate criteria is attached as Exhibit 3.

A substantial portion of EPA's comments regarding SWEPCO's petition involved questions about ADEQ's calculation of permit limits for SWEPCO. It is our understanding that ADEQ has separately addressed those concerns with EPA. SWEPCO's responses to the remainder of EPA's comments on the SWEPCO petition are attached as Exhibits 4 and 5.

The letter commenting on Domtar's petition briefly reviewed the principal information supporting the petition and concluded with a recommendation that a new bioassessment be undertaken to determine if the proposed criteria changes will be protective of aquatic life. We are concerned about this recommendation because it appears inconsistent with earlier discussions between EPA, ADEQ, and representatives of Domtar.

You will recall that on October 21, 2014, ADEQ Acting Director Ryan Benefield, Water Division Chief Ellen Carpenter, and representatives of Domtar met with you and members of your staff to discuss how much additional information, beyond that developed for the SWEPCO petition, would be needed to support the closely related criteria changes proposed by Domtar. I attended that meeting. It is my recollection the discussion ended with general agreement that only limited additional information, as proposed by ADEQ and Domtar, would be needed so long as the SWEPCO petition was adequately supported.

The week following the October 21st meeting Ryan Benefield sent you a summary rationale for the Domtar change that was consistent with the apparent agreement reached on October 21st. As I recall, in response to ADEQ's summary rationale some members of your staff initially seemed to reverse field regarding the extent of additional information that would be needed. It is my understanding you discussed this matter further with Ryan Benefield, and by an e-mail dated November 6, 2014 you confirmed to Ryan that if the SWEPCO criteria changes in the Red River were approvable and the Domtar segment of the Red River was similar to the SWEPCO one, EPA did not think additional study, beyond that contemplated by ADEQ's summary rationale would be required. Based on those assurances, Domtar finalized and filed its petition and proceeded through the rulemaking process.

Domtar is concerned that the comment letter EPA submitted the day before the Commission's final vote once again appears to reverse field. The main point of the letter, namely that a new bioassessment should be undertaken, seems directly contrary to the consensus that appeared to be reached in the October 21<sup>st</sup> meeting and confirmed in your subsequent discussions with Ryan Benefield.

It is our hope that your review of SWEPCO's response to EPA's comments on the SWEPCO petition will address Agency questions adequately to allow approval of the changes involved in that petition. It is also our hope that if that is the case, you will also be persuaded

Mr. William K. Honker, Division Director January 15, 2016 Page 3

that the prior agreements upon which ADEQ and Domtar relied in proceeding with the Domtar petition, were appropriate and that no additional bioassessment is required to approve that petition.

For your convenience, I am attaching copies of EPA's comment letters on the SWEPCO and Domtar petitions. See Exhibit 6 (EPA comments on SWEPCO's petition) and Exhibit 7(EPA comment letter on Domtar petition).

After you have had a chance to review this letter, I would like to discuss with you whether any further response is necessary regarding the substance of EPA's October 22, 2015 comment letter on the Domtar petition. I would also like to discuss what more is needed for EPA to be able to review and approve these rulemaking changes.

Needless to say, if you have any questions or desire any further information, please do not hesitate to contact me.

With best regards, I am

Respectfully yours,

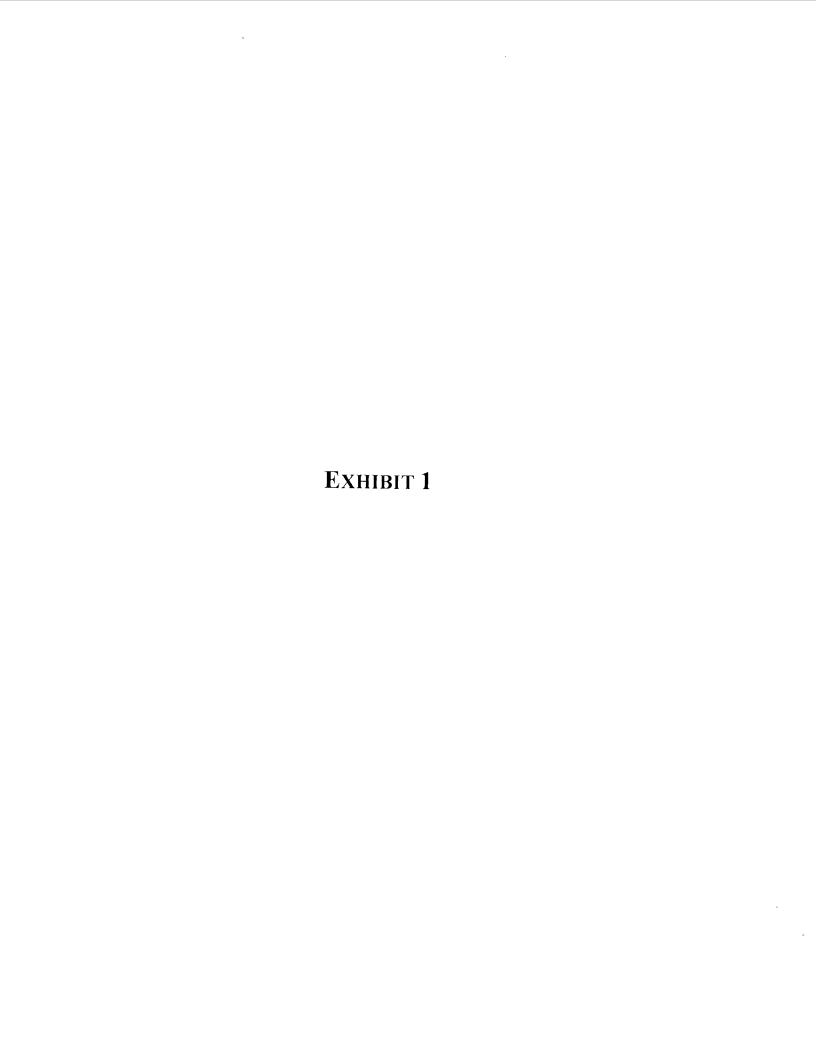
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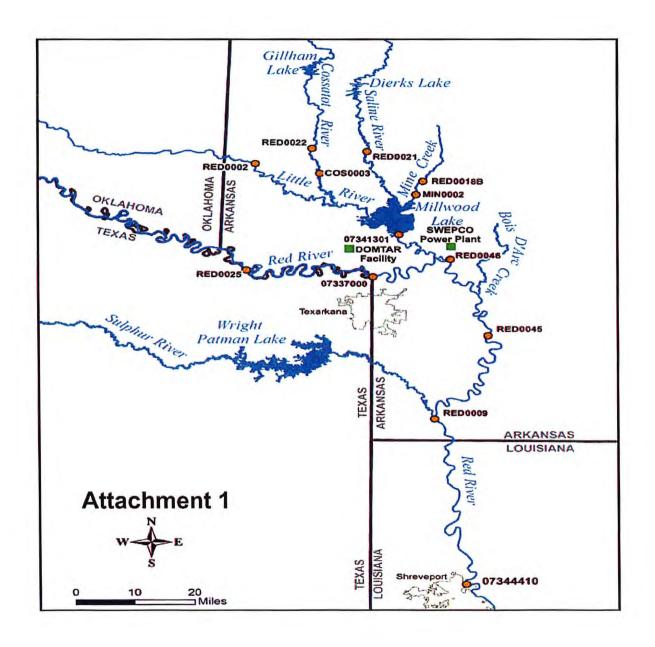
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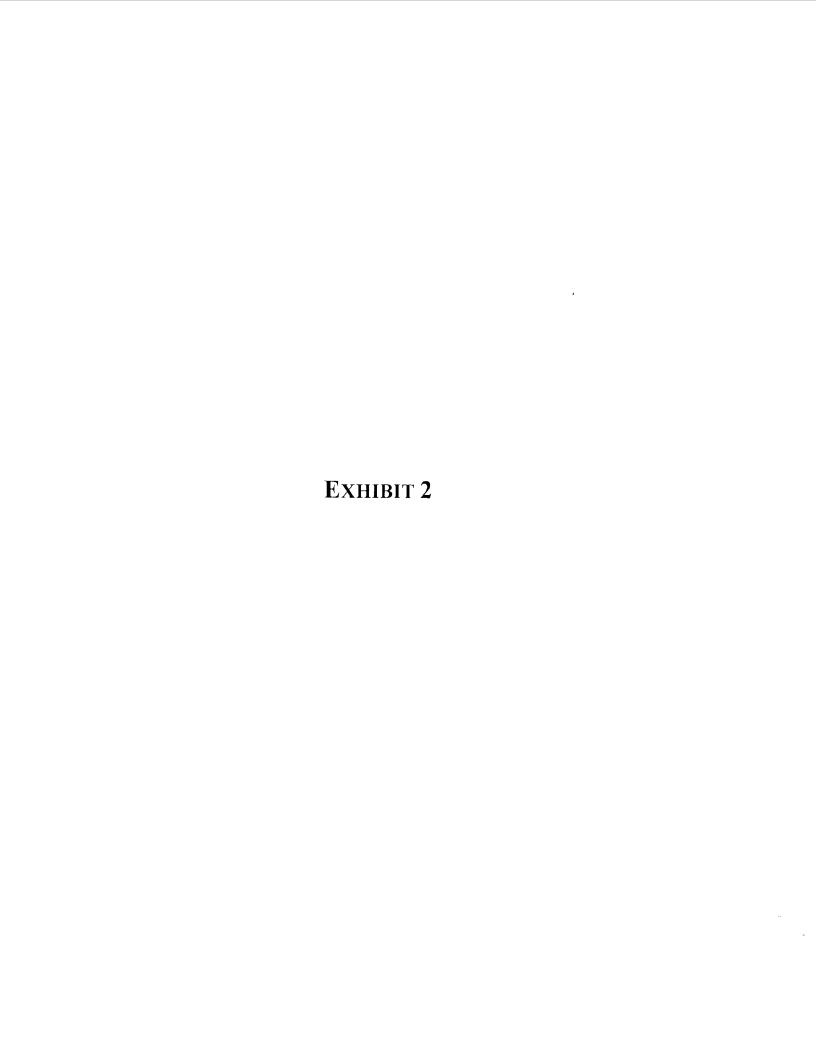
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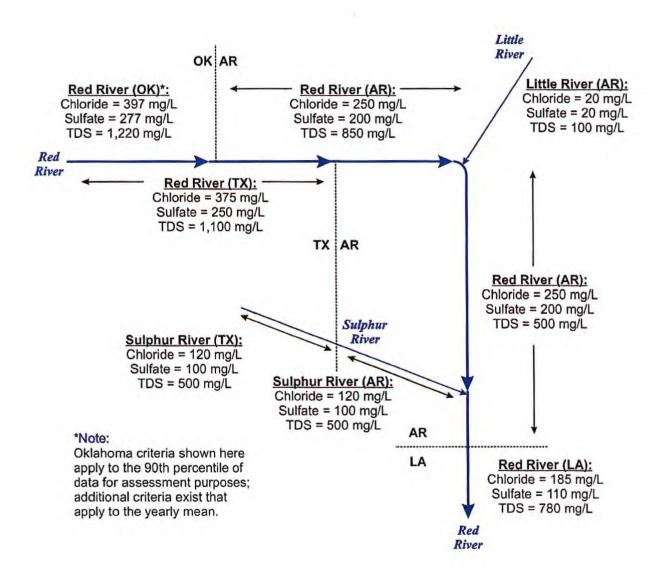
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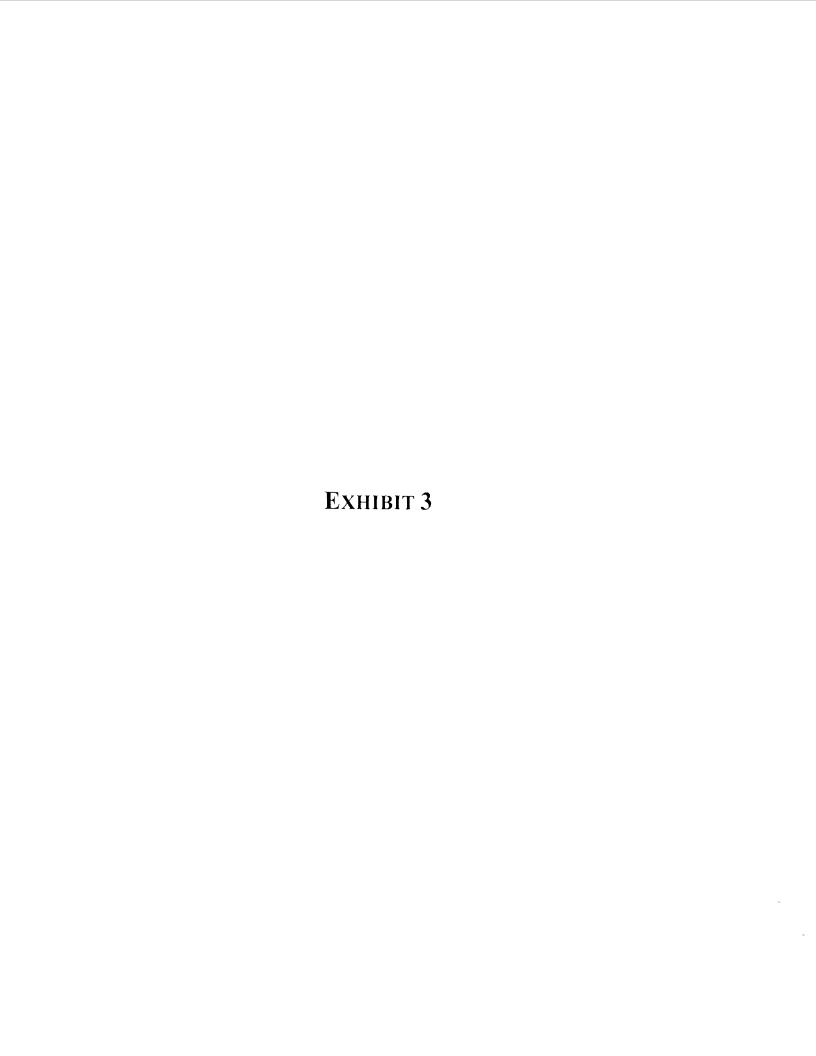
cc: Caleb Osborne, Esq., Deputy Director, ADEQ Ellen Carpenter, Esq., Water Division Manager, ADEQ

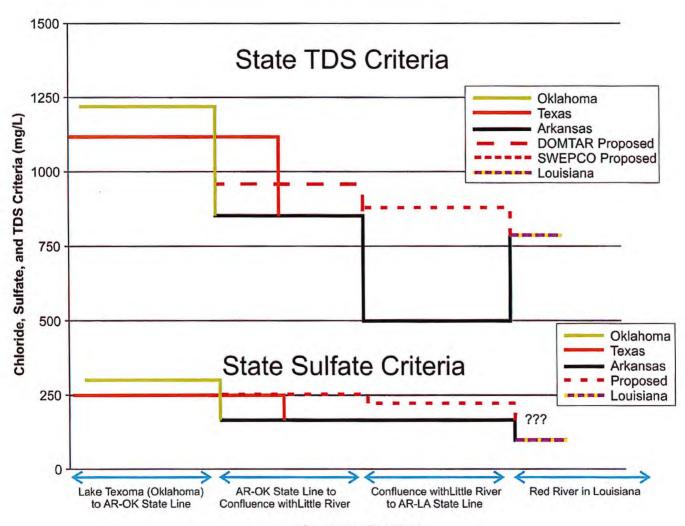




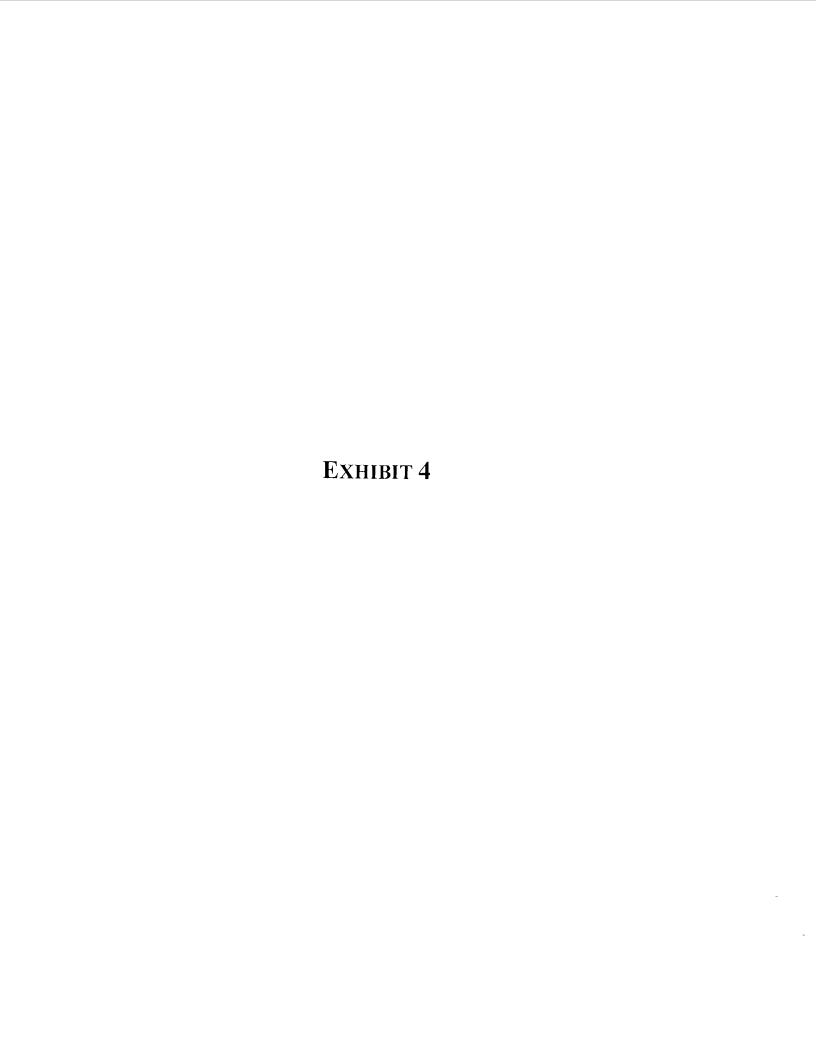








Reaches on Red River



### Response to EPA Technical Comments SWEPCO UAA for Dissolved Minerals in Little and Red Rivers

On October 20, 2015, Region 6 of the US Environmental Protection Agency (EPA) provided technical comments dated "September XX, 2015," to the Arkansas Department of Environmental Quality (ADEQ) on Southwestern Electric Power Company's (SWEPCO's) *Use Attainability Analysis for Dissolved Minerals in Little and Red Rivers, Hempstead and Little River Counties, Arkansas*, which was submitted to ADEQ on September 5, 2014.

EPA's comments have been reproduced below, followed by SWEPCO's responses in blue text. EPA's comments are given in italics for ease of distinguishing between the comment and SWEPCO's response, and the comments have been numbered to allow for referencing within the document.

Many comments throughout the document raised the same or similar questions. For three of those topics, SWEPCO has prepared general responses to address the questions raised. These general response sections are referenced throughout SWEPCO's responses to specific comments in order to avoid duplication.

#### 1.0 GENERAL RESPONSES

#### 1.1 Designated Uses and Sources of Total Dissolved Solids

A number of EPA's comments suggest that EPA understands the primary justification for the proposed site-specific criteria (SSC) to be elevated minerals concentrations due to "natural background," or that SSC must be based on natural background conditions. Although SSC can be based solely on natural background conditions, both state and federal regulations allow the development of SSC based on 304(a) guidance (i.e., water-effects ratios) or other scientifically defensible methods. This document represents development of SSC based on "other scientifically defensible methods." Although this document is referred to as a use attainability analysis (UAA), it is not a UAA in the sense of what is described in EPA's Water Quality Standards Handbook (2014 update), in which the UAA is intended to support changes to designated uses. The UAA term is used herein because the justification of the SSC still requires an evaluation of attainable uses and whether the uses can be supported with less-stringent SSC. However, the proposed SSC that are the subject of the SWEPCO study represent (a) existing conditions in the case of the Red River, and (b) a total dissolved solids (TDS) criterion that is comparable to an ecoregion least-disturbed stream in the case of the Little River.

Natural background conditions (in this case TDS and temperature) are *part* of the rationale for the SSC proposed herein, but they are not the sole basis, as considerable attention was given to evaluating the TDS conditions that support attainable designated uses. Therefore, the portion of the Tudor Davies 1997 policy memorandum that addresses criteria changes that are based *solely* on natural background has little relevance to this proposal. SWEPCO agrees, however, that in

some places, the draft report reads as if naturally occurring mineral loads are the only sources of minerals to the Red River and Millwood Lake. SWEPCO acknowledges that anthropogenic inputs are part of the mineral regime in both waterbodies, which is the reason for the focus on designated use attainment.

To justify less-stringent criteria, it is necessary to show that the proposed criteria protect the existing and attainable designated uses of the receiving streams (*Water Quality Standards Handbook*, EPA 2014). The study showed, and ADEQ has confirmed, that measured and predicted concentrations of dissolved minerals will not limit aquatic life or impair other existing or attainable uses.

In the case of the TDS SSC in the Red River, the report showed that the mineral regime at the Index, AR, gaging station is dominated by loading from Denison Dam, which is overwhelmingly due to natural inputs. SWEPCO has provided additional information to support that statement herein (see Sections 1.2 and 1.3 below). However, the reach of the river below Denison Dam undoubtedly sees some additional natural and anthropogenic input. Accordingly, since not *all* of the mineral loading to the station at Index is natural, the study focused on the attainment of designated uses in the reaches of the Red River in question. It is not appropriate for EPA to insist that "Arkansas must ...show that there are no natural or anthropogenic sources from the TX-AR line to the confluence." In addition to being a physical impossibility, this condition is not necessary for the full support of attainable designated uses (aquatic life/fisheries in particular). In EPA's comments, EPA has not disputed SWEPCO's conclusions that the existing mineral regime in the Red River supports designated uses. The report has proposed SSC based on existing conditions that are mainly (but not solely) due to known natural sources and that support attainable designated uses.

In contrast to the Red River, where the primary sources of upstream minerals are clear, the TDS concentrations below Millwood Lake dam cannot be attributed to a particular source. With the exception of Mine Creek, which has a negligible contribution, the major inflows to Millwood Lake all meet ecoregion minerals criteria. Accordingly, as with the Red River, the focus in the Little River below Millwood Lake dam is on attainment of designated uses. The report demonstrated that the existing conditions support the attainable designated fisheries use and that a slight increase in TDS due to plant operation would likely have little effect on the level of attainment. EPA did not dispute this conclusion in its comments.

# 1.2 Sources of TDS to the Red River at Denison Dam: Natural versus Oil-Field Brine Disposal

Information on TDS inputs to the Red River from oil-field brine disposal was available in the Red River Basin Chloride Control Project, Environmental Issues Summary Report (Red River Authority of Texas 1996), which is provided in Attachment 1. This report states the following:

"Pursuant to the agreements entered into between the states and the federal government, the states have been controlling approximately 85% of the man-made pollution to the river system

since 1980...Over 968 tons of chlorides are eliminated daily from entering the Red River watershed above Lake Texoma which were originating from oil field brine disposal operations."

This statement implies that the total chloride loading from oil-field brine disposal was 1,139 tons per day (calculated by dividing 968 tons per day of chloride eliminated daily by 85%). The statement further implies that the remaining 171 tons of chloride per day (1,139 T/d - 968 T/d) represents an ongoing source of chloride to the Red River due to oil-field brine. Table 1 below summarizes chloride and sulfate loading calculations based on the data provided in Appendix C of the report. Based on this summary, and assuming that the loading estimate from 1996 is representative of subsequent years, the loading due to the remaining oil field brine contributes 4.4% (171 T/d  $\div$  3,888\*100) of the total chloride loading from Denison Dam.

Table 1. Summary of selected parameters based on monitoring data from Denison Dam and Index.

Parameter	<b>Denison Dam</b>	Index
Average flow for period of record (POR) of 1997 to 2011 (cfs)	4,801	11,665
Average Cl (mg/L)	300	156
Average daily Cl load (kg/d)	3,527,321	4,440,700
Average daily Cl load (T/d)	3,888	4,895
Average SO <sub>4</sub> concentration (mg/L)	261	150
Average daily SO <sub>4</sub> load (kg/d)	3,064,566	4,265,497
Average daily SO <sub>4</sub> load (T/d)	3,378	4,705
Average Cl+SO <sub>4</sub> concentration (mg/L)	561	305
Average Cl+SO <sub>4</sub> load (kg/d)	6,591,887	8,705,151
Average Cl+SO <sub>4</sub> load (T/d)	7,266	9,595
Proportion of Index flow from Denison	10 Manual Committee of the State of the Stat	0.41
Proportion of Index Cl load from Denison		0.79
Proportion of Index SO <sub>4</sub> load from Denison		0.72
Proportion of Cl+SO <sub>4</sub> load from Denison		0.76
Average proportion of measured TDS as Cl+SO <sub>4</sub>	0.55	0.46
Average proportion of measured TDS as Cl+SO <sub>4</sub> based on POR of 1974 to 1986	0.86(1)	0.70
Estimated average TDS concentration <sup>(2)</sup> (mg/L)	1,020	649

Notes:

As shown in Table 1, the TDS of water from Denison Dam also contains a similar mass of sulfate derived from gypsum rocks in the natural springs that contribute the majority of salts to the Red River. Since oil-field brines are typically comprised predominantly of sodium and

<sup>1.</sup> Assuming all present as NaCl and CaSO4.

Based on average proportions of measured TDS as Cl+SO<sub>4</sub>.

chloride, the contribution to TDS due to brine disposal is therefore somewhat less than the 4.4% contribution indicated above.

This analysis demonstrates that oil-field brine disposal is a minor source of TDS to the Red River, as discharged from Denison Dam, compared to natural sources.

#### 1.3 Influence of Denison Dam Discharge on Flows and Minerals at Index

Although the monitoring station at Index, AR, is 200 miles downstream of Denison Dam, the discharge from Denison Dam dominates the flow and mineral regime at Index. While the flows from Denison Dam account for 41% of the average flow at Index (Table 1), they account for 76% (6,591,887 ÷ 8,705,151\*100) of the chloride and sulfate loading.

A straightforward calculation shows that the estimated average TDS concentration at Index would be 420 mg/L if there were no TDS source (natural or anthropogenic) downstream of Denison Dam. In other words, if all flow entering the Red River below Denison Dam was distilled water, the average TDS concentration at Index would be 420 mg/L. The same calculation can show, conversely, that the average TDS concentration at Index would be 229 mg/L without the loading from Denison Dam. Therefore, without the loading from the Denison Dam, there would be no exceedances of the existing criteria.

Since the TDS loading downstream of Denison Dam is clearly greater than zero, there are undoubtedly additional sources of TDS between Denison Dam and Index, some of which are likely point sources. However, this analysis demonstrates that TDS (primarily as chloride and sulfate) loading from Denison Dam, which is overwhelmingly due to natural sources, dominates the TDS loading at Index, 200 miles downstream from the Denison Dam.

#### 2.0 SPECIFIC RESPONSES

#### COMMENT 1

ES.7 Proposed TDS Criteria Little River, page xi:

The proposed TDS criterion for the Little River below Millwood Lake is based on a weight-of-evidence approach utilizing a mass balance model. The document states that a mass balance was prepared that would reflect naturally occurring elevated TDS concentrations in the Little River. The document refers to TDS data collected upstream of SWEPCO by FTN and SWEPCO during October 2010 through October 2013 that showed that the current site-specific TDS criterion 100 mg/L criterion was exceeded approximately 10% of the time. The authors conclude that a TDS criterion of 136.1 mg/L would be necessary in the Little River to reflect the naturally occurring mineral regime and to allow SWEPCO to operate efficiently.

It's unclear what the basis for presuming that only naturally occurring sources of TDS result in the 10% exceedance rate.

Please see Sections 1.1 and 1.2 of the general responses.

Later, the document refers to input from the Little River itself above Millwood Lake and from Mine Creek, noting that inputs from the latter are significantly higher per volume than other tributaries to Millwood Lake. The name Mine Creek alone suggests that this creek is a likely source of anthropogenic. Given the mass balance budget described in Table J.1 describes the background in the Little River as 98 mg/L and the SWEPCO discharge as contributing 1620 mg/L, it is unclear how a presumption that the inputs to the Little River are not anthropogenic.

Please see Section 1.1 of the general responses.

This tends to undermine the validity of the mass balance and any conclusions that may be drawn from it.

Whether the minerals sources to the Little River are or are not anthropogenic does not affect or invalidate the mass budget modeling prepared and submitted as part of the report. The mass budget modeling does not depend on the TDS source for predictions. However, the modeling does depend on data that represent current TDS concentrations in the river, as was demonstrated in the report.

Differentiating between natural background and anthropogenic sources causing exceedances is important. As articulated in Tudor Davies 1997 policy memo, the water quality standards regulation at 40 CFR 131.11(b)(I) requires States and authorized Tribes to adopt numeric water quality criteria that are based on section 304(a) criteria, section 304(a) criteria modified to reflect site-specific conditions, or other scientifically defensible methods.

#### Please see Section 1.1 of the general responses.

In summarizing the aquatic life evaluation, the document indicates that the aquatic life composition of the current fish community below Millwood dam is a subset of what would be expected given available habitat. What is described here represent a snapshot of what is likely impacted and/or degraded conditions in the Little River. While document current conditions is an important, the objective of a UAA should be determining what the highest attainable use that can be attained. The objective of a UAA is to determine what the highest attainable use the waters of interest can support. 40 CFR 131.10(d) requires at a minimum, that uses be deemed attainable if they can be achieved by the imposition of effluent limits required under sections 301(b) and 306 of the Act and cost-effective and reasonable best management practices for nonpoint source control.

Please see the first paragraph of Section 1.1. A major focus of the study was to document attainment of designated uses, particularly the aquatic life (fisheries use). EPA did not dispute the findings or conclusions regarding the aquatic life (fisheries) use attainment.

## **COMMENT 2**

1.1 Background and Overview, page 1-1:

The discussion here appears to be suggesting that the (predominate) natural (and anthropogenic) mineral and TDS sources from TX and OK are not adequately accounted for and result in exceedances of AR's 500 mg/L criterion in the lower segment of the Red River below the confluence of the Little River to the AR-LA state line. Since there is certainly some dilution in the Red River as it moves to the confluence with the Little River and on downstream, it will be critical to provide clear support that these predominately natural upstream in TX and OK, and those in AR contribute to an inability to attain the current 500 mg/L criterion in the lower segment.

Please see Section 1.1 of the general responses.

The spatial inconsistence identified in and of themselves are not a basis for modifying criteria.

SWEPCO agrees. The proposed SSC were not based on spatial inconsistencies of the existing criteria.

## **COMMENT 3**

1.1 Background and Overview, page 1-1 and 1-3:

In describing this permit as not typical, the authors are referring to ADEQ's use of the 500 mg/L TDS criterion that applies to the Red River as an "end of pipe" value rather than the 100 mg/L TDS criterion that applies to the Little River. Based on the fact sheet, it appears that ADEQ based 500 mg/L TDS criterion in the SWEPCO permit on the applicable criterion in the Red River to ensure that the discharge would not contribute to the impairment of the Red River because it is on the state's most recent EPA approved 303(d) list for exceedances of chloride, sulfate and TDS.

Comment acknowledged.

Region 6 permitting staff have suggested that if the 100 mg/L TDS criterion applicable to the Little River were applied, when dilution and mixing are considered, a limit of 3800 mg/L would be possible and would be protective of the designated fishery use in both the Little River and the Red River.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

1.1 Background and Overview, page 1-4, Table 1.2:

The increase for both median and 95th percentile concentrations of TDS from the historical data the more recent data collected by SWEPCO suggest that the concentrations are the result of something other than "naturally occurring exceedences" as suggested in the next paragraph.

Please see Section 1.1 of the general responses.

## **COMMENT 5**

1.1 Background and Overview, page 1-5:

The authors refer to little to no assimilative capacity for TDS in the Little River at times (absent the SWEPCO facility's discharge) as a result of "naturally occurring exceedances" of the current site-specific 100 mg/L TDS criterion occurring "upstream" of the SWEPCO discharge, refers to more in depth discussed in Sections 3.0 and 5.0. However, there is very little discussion of possible upstream naturally occurring" sources for the Little River in these sections. The only sources for the Little River discussed in the document to any degree are tributaries to Millwood Lake, particularly Mine Creek. Although Mine Creek is identified as the most significant contributor of minerals to Millwood Lake and downstream to the Little River, there is no information concerning possible mining impacts to that creek. As a result, the basis for attributing a lack of assimilative capacity and a 10% exceedance rate for the current 100 mg/L TDS criterion in the Little River to naturally occurring sources has not been established.

The data that support the statements are provided in Table 1.2 of the report.

What are these natural sources of high TDS - Mine Creek? And are they natural or anthropogenic?

Please see Section 1.1 of the general responses regarding natural versus anthropogenic sources of TDS. Please note that the report shows that although TDS concentrations in Mine Creek are significantly higher than concentrations in the other streams, the Mine Creek inflow provides only approximately 3 mg/L of TDS based on a flow-weighted mean inflow TDS concentration of 54.8 mg/L from the four major tributaries (54.8 mg/L including Mine Creek and 52.2 mg/L without Mine Creek).

## **COMMENT 6**

## 2.3.1.1 TDS Sources to the Little River

This section includes an evaluation of existing data from tributaries upstream of Millwood Lake to identify potential recent sources of TDS that might explain the increased TDS values in samples collected from the Little River downstream of the dam on Millwood Lake. The major

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tributaries of Millwood Lake include the Little River, Cossatot River, Saline River, and Mine Creek (Figure 1.1). Table 2.2 identifies the major tributaries to Millwood Lake, including the Cossatot River, Saline River and Mine Creek and summaries of their TDS contribution. Although Mine Creek only contributes 2% of the inflow, the TDS concentrations are significantly higher than the other streams.

Please see the last paragraph of the response to Comment 5.

There is some inconsistency between Tables 1.1 and 1.2 presenting USGS data from 1967-1995 and SWEPCO data from 2010-2013 (although it's not clear what sites the SWEPCO data came from) that show an increase in TDS concentrations over time.

The data presented in Table 1.2 were collected by SWEPCO and FTN at sampling station LR-0, which is shown on Figure 3.1 of the report. SWEPCO will clarify the source of the data in the table title for Table 1.2 and the corresponding section discussing the data in the report.

The two tables are not inconsistent but rather simply present summary information showing that TDS concentrations are different from two different time periods and that the more recent data show an increase in TDS concentrations.

The time-series plots of TDS data from these ADEQ monitoring stations on the tributaries upstream of Millwood Lake identified in Figures 2.1 through 2.4 that show no clear evidence of significant changes in TDS levels in any of the streams, yet the authors report an increase in TDS concentration downstream of Millwood Lake.

The second paragraph of Section 2.3.1.1 states, "Overall, there is no clear evidence of significant changes in TDS levels in any of the streams" [contributing to Millwood Lake]. SWEPCO agrees there is no obvious source or cause of the increase. With the exception of Mine Creek, which contributes a negligible loading, the major inflows to Millwood Lake all meet ecoregion minerals criteria.

## **COMMENT 7**

# 2.3.1.2 Water Quality Impairments for Minerals in the Little River

The authors indicate that ADEQ does not considers the Little River downstream of Millwood Lake to be impaired and have not included it on any 303(d) list. Further, the document notes that this stream is classified by ADEQ as "unassessed" due to a lack of data for water quality (including minerals). It is important to note that "unassessed" simply means that there is no data and does not necessarily mean that the Little River is either impaired or unimpaired. That lack of data may be why neither ADEQ nor EPA have included the Little River on the state's 303(d) list.

SWEPCO agrees.

2.3.2.2 TDS Sources to the Upper Red River (Oklahoma, Texas), pages 2-5 to 2-8:

Much of the literature cited in the Executive Summary and in these sections is dated, relying on several sources from a time period when oil production was high with minimal environmental regulation.

Information describing natural sources was from historical literature (Baldys et al. 1996, Keller et al. 1988, USGS 1961). Information describing current conditions is recent in our opinion (e.g., gage data with a period of record of 1997 to 2011; USACE publication from 2010; Red River Authority publication from 2013).

Despite those issues, it is reasonable to say that there are significant natural as well as anthropogenic sources of minerals in the Red River and its tributaries.

Please see Sections 1.1 and 1.2 of the general responses.

While the more current information tends to show that the predominant mineral sources to the Red River are likely natural, despite remediation efforts by the US Army Corps of Engineers (USACE), it is likely that brine disposal and intrusions from oil production into the Little Wichita and other sites are sources that affect the Red River in Texas, Oklahoma and contribute to the loading to a lesser extent in Arkansas. Sources of information for oil and gas production and remediation should be available from the Texas Railroad Commission and the Oklahoma Corporation Commission.

Not having, or possibly not being able to provide any information to support historical oil extraction impacts and their contribution to minerals/TDS concentrations significantly weakens the claim that the "overwhelming portion" historical of the TDS inputs to the system are from natural sources.

Please see Sections 1.1 through 1.3 of the general responses.

#### **COMMENT 9**

2.3.2.3 Water Quality Impairments for Minerals in the Red River, page 2-9:

The authors note that upstream criteria for the Red River in Texas and Oklahoma reflect the elevated ambient mineral concentrations that are due to inputs from salt springs and seeps from the arid and semi-arid headwater regions. This is generally true, given salt springs are found as far west as the High Plains and Tableland Level II ecoregions. Although less predominate, there also anthropogenic and natural inputs from the Central Plains, Crosstimbers, Blackland Prairie and East and South Central Plains ecoregions.

It is important for Arkansas to recognize that an upstream state can establish uses and criteria in their waters as appropriate. 40 CFR 131.10(b) requires that those states take the downstream state's standards. Recent court decisions have affirmed EPA's ability to look at downstream waters to ensure protection.

# Comment acknowledged.

This section also discusses assessment procedures used by states that share jurisdiction over portions of the Red River. Referring to Table 2.6, which compares criteria for minerals and impairments identified on state 303(d) lists, which shows that the Red River is not considered to be impaired upstream in Oklahoma and Texas or downstream in Louisiana. The authors state that this is partly due to different assessment procedures that are used in other states, suggesting that these states use less protective exceedance rates. As an example, the authors note that Louisiana does not consider a stream to be impaired for dissolved minerals unless 30% of the observed data exceed the water quality standard compared to Arkansas' 10% exceedance rate. The authors also state that Oklahoma and Texas also use less-stringent assessment procedures for dissolved minerals than Arkansas, and also state that the criteria in their water quality standards are less stringent than the criteria in Arkansas's standards. However, the authors have not explained how these approaches are any less protective than those used by Arkansas.

These proposed SSC considered downstream compliance with Louisiana's criteria. The report does not use the term "less protective"; that is the commenter's usage. The lack of impairment in upstream states is due to "less stringent" criteria or assessment methodology with no suggestion one way or the other regarding the protectiveness of those criteria. SWEPCO agrees that less-stringent criteria and assessment methodology can be protective.

It is important to understand that simply comparison of exceedance percentages used in assessing use attainment does not tell the entire story. Assessing attainment includes defining the water quality indicators it measures and the procedures for analyzing and interpreting data in order to decide whether standards are met or water quality is impaired. This should include collection and analysis of multiple types of data providing information relevant to assessing attainment with approved WQS. As a result, a comparison of Louisiana's current use of a 30% exceedance rate to Arkansas 10% is not necessarily appropriate, although it should be noted that Region 6 is working with Louisiana to revise its approach. Oklahoma compares the mean of all values to a yearly mean standard, then looks for greater than 10% exceedances against a sample standard. In Texas, minerals criteria represent annual averages of all values that were collected and compared to the criterion for each parameter. Use attainment is based on the average concentration that applies to the entire length or area of the segment. For TDS, a value is calculated by multiplying specific conductance measured at the surface by a factor of 0.65. The chloride, sulfate, and TDS criteria are not supported if the average value exceeds the criteria.

Comment acknowledged.

The authors also state that Oklahoma and Texas water quality standards as less stringent than the criteria in Arkansas. Given the natural saline seeps and historical oil extraction activity along the Red River that this document discussed previously, higher criteria would be expected upstream in Texas and Oklahoma. The burden for SWEPCO is to show that predominately natural sources upstream in Oklahoma and Texas upstream as distant as Lake Texoma and/or that natural sources in Arkansas itself alter concentrations in the lower segment of the Red River to support the proposed criteria modifications.

Please see Section 1.3 of the general responses, which clarifies report statements that the sources are predominantly upstream in Oklahoma and Texas.

## **COMMENT 10**

Page 2-10:

In a discussion of Table 2 describing applicable criteria and 303(d) listings, the authors refer to ADEQ's practice in recent years of making specific permitting decisions and calculations based on the state's most recent 303(d) list that has been approved by EPA. This approach means that ADEQ is applying the 500 mg/L TDS criterion that applies to the Red River below its confluence with the Little River as an end-of-pipe limit rather than the 100 mg/L criterion that applies to the Little River which is the receiving water for the Turk/SWEPCO discharge.

Based on the NPDES permit fact sheet, applying the 100 mg/L TDS criterion that applies to the Little River, when dilution and mixing is considered, a limit of up to 3800 mg/l would be possible and still meet the in-stream criterion of 100 mg/L which would protect beneficial uses in the Little River and downstream. However, applying the 500 mg/L TDS criterion as an end-of-pipe limit results in a more restrictive permit limit. Without any information to the contrary, the effect of ADEQ's approach does not appears to have a significant environmental benefit for either the Little River or Red River, but appears to have the opposite effect. While applying 500 mg/L TDS criterion as an end-of-pipe limit may result in a small reduction in overall loads, the more restrictive limit reportedly requires the facility to operate at less than optimum efficiency. Thus ADEQ's approach appears to be the primary driver for Turk/SWEPCO to seek revised criteria for the Red River, which may be a less desirable environmental outcome.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

#### **COMMENT 11**

2.4.1 Threatened and Endangered Aquatic Species, page 2-12:

This section of the document discusses several endangered species listed by the US Fish and Wildlife Service (USFWS) including the Ouachita rock pocketbook (Arkansia wheeleri), which is federally listed throughout its range in Arkansas and Oklahoma. The document reports that there have been no live A. wheeleri collected from just above or below the Turk/SWEPCO discharge.

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Their absence indicates that the Turk/SWEPCO discharge is having an effect on water quality and the presence of A. wheeleri. Although the physical characteristics and inputs to Millwood Lake may affect the ability of the Little River to support A. wheeleri, an important aspect of a UAA is to not only to identify current conditions. The document does not provide any discussion of whether minerals/TDS or other stressors may be affecting or limiting the presence of A. wheeleri in this portion of the Little River.

The report notes that the hydraulic modification (i.e., construction of Millwood Lake dam on the Little River) affected mussel distribution and abundance before construction of the SWEPCO facility. Studies performed after Millwood Lake was formed but prior to construction of the facility did not document the presence of A. wheeleri in the Little River in the vicinity of the future intake and outfall locations (see Section 2.1.4 of the report for further explanation).

The authors also note that the USFWS recently listed the rabbitsfoot mussel (Quadrula cylindrica cylindrica) as threatened (Federal Register 2013). That species is known from some streams in southwest Arkansas, including the Little River upstream of Millwood Lake. This species not been documented in the Little River downstream of Millwood Lake or the Red River. The rabbitsfoot is likely limited by the physical characteristics Millwood Lake and its effect on the Little River, as well as the Turk/SWEPCO discharge.

SWEPCO agrees that the rabbitsfoot is not found in the Little River downstream of Millwood Lake. The changes in the physical characteristics, and therefore habitat, of the Little River due to construction of the Millwood Lake dam occurred before construction of the SWEPCO facility.

## **COMMENT 12**

3.1 Overview, page 3-1:

Rather than relying on biological evaluations in a upstream versus downstream comparison from a point source discharge and a comparison of communities in reference and receiving streams, the approach for field surveys is attempting to show that characteristics in Millwood Lake and at the Denison dam dominate the mineral regime in the Little and Red Rivers rather than the Turk/SWEPCO discharge.

The field surveys addressed aquatic life use attainment. Sections 2.3.1, 2.3.2, and 5.0 of the report demonstrate that characteristics in Millwood Lake and at Denison Dam, rather than SWEPCO's discharge, dominate the mineral regime in the Little River and Red River. As the report notes, "This evaluation is unique because the instream criteria exceedances have little or nothing to do with a point source that is not able to meet existing criteria. In the case of the Red River, the criteria exceedances are primarily due to upstream background concentrations. In the case of the Little River, existing mineral concentrations are also at or near the existing site-specific criteria without influence from the plant." Please see Sections 1.2 and 1.3 of the general responses.

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There are sources of minerals above Millwood dam, primarily the Little River and Mine Creek upstream of Millwood Lake and there are certainly significant natural and anthropogenic sources of mineral in the upper reaches of the Red River in Texas and Oklahoma. The burden for Arkansas is to clearly show that the sources above Millwood dam are at least predominantly anthropogenic.

This important sentence, which states Arkansas' burden, seems to have been mistyped. Surely, in line with previous statements, the commenter meant to say "..at least predominantly *natural*"? As shown in Table 2.2 of the UAA report, TDS concentrations in the Little River, Cossatot River, and Saline River are well below the ecoregion criterion of 129 mg/L. It is not clear how upstream waterbodies that meet criteria by a comfortable margin (with the exception of Mine Creek) might represent some sort of unnatural or anthropogenic input. It is noted that although TDS concentrations in Mine Creek are significantly higher than concentrations in the other streams, the Mine Creek inflow contributes only approximately 3 mg/L of TDS based on a flow-weighted mean inflow TDS concentration of 54.8 mg/L for the four major tributaries (54.8 mg/L including Mine Creek and 52.2 mg/L without Mine Creek). Therefore, there is little support that anthropogenic inputs of TDS dominate Millwood Lake.

Similarly, Arkansas must be able to make a reasonable showing that the predominately natural and some anthropogenic sources some 200 miles upstream have a significant effect on water quality in the Red River as far downstream as its confluence of the Little River and show that there are no natural or anthropogenic sources from the TX-AR line to the confluence.

This must be another mistyped sentence. Surely EPA does not imagine the existence of any reach of any stream for which "...there are no natural or anthropogenic sources." Table 1 of this document notes that 75% of the chloride and sulfate loading at Index is from Denison Dam. Please see Sections 1.2 and 1.3 of the general responses.

If the Turk/SWEPCO discharge is not contributing to the potential exceedances in the Little River or potentially impacting the Red River, the important question as to whether the facility is seeking to revise the criteria in the Little River and the Red River because the criteria are inappropriate or because of ADEQ's policy decision to applying a criterion as an end-of-pipe limit rather than basing it on applicable criteria considering dilution and mixing in the receiving water.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

# **COMMENT 13**

3.4.1.2 Macroinvertebrate Data Analysis, page 3-10:

In discussions of field observations the authors indicated an attempt was made to assess the susceptibility of the existing benthic macro invertebrate community to a slight increase in TDS.

Tolerance indicator values (TIVs) for specific conductance for various invertebrate taxa provided in Carlisle et al. 2007 were reviewed. However, the authors note that too few of the taxa listed in Carlisle et al. 2007 were present in the data set for this study to allow for a meaningful evaluation of ion tolerance. Accordingly, the benthic data presented herein provide a description of the presently attained fisheries use and a baseline for future studies.

# SWEPCO agrees.

It should be noted that small increases in TDS alone may have minimal effect on the existing benthic community however, TDS is rarely seen independent of other stressors.

Comment acknowledged. It is not clear why other stressors are important. The relevant question is whether or not the existing levels of TDS support the attainable designated use.

In a quick search, a number of sources were identified that discuss the effect TDS has on Appalachian streams. For example, Timpano et al. (2010), in discussing the effects of TDS in coalfield streams. As a way of isolating the effects, Timpano et al. used headwater streams to represent a gradient where confounding factors could be minimized. This and other similar papers done in Appalachian could have been used to inform this aspect of the analysis.

The study by Timpano et al. has little in common with this study, because as EPA has noted elsewhere, there are no differences in benthic or fish communities upstream versus downstream of the discharge. We agree that there might be other unknown stressors present in the effluent. However, the best approach to address that possibility is through the normal permitting process, which includes tests such as priority pollutant scans and biomonitoring requirements.

One of the problems with the approach take here is a lack of reference stream comparisons.

The issue of reference condition comparison was addressed in the study plan for the UAA that was reviewed and approved by ADEQ. There is no appropriate reference data set or reference location for comparison with this reach of the Little River. The several similarly sized reservoirs in the region (e.g., Dierks, Gillam, DeQueen) are deep, have cold tailwaters, and are not suitable analogs to the Little River tailwater system. The study utilized literature-based tolerance values and professional judgment to evaluate field data to assess the susceptibility of the benthic community to the proposed increase in TDS.

## **COMMENT 14**

3.4.2.2 Fish Data Analysis, page 3-11:

Fish data were first assessed to determine if there were impacts due to the Turk/SWEPCO discharge "under current conditions" by evaluating differences in total species richness, relative abundance, diversity, and similarity measures between upstream and downstream reaches.

Although unclear, it appears that "under current conditions" likely means under limits specified in the current NPDES, which as noted earlier, are based on a monthly average of 500 mg/l TDS applying Red River Criteria end-of-pipe.

## EPA's statement is correct.

This end of pipe value equates to an instream waste concentration of 43 mg/L. As noted earlier, if ADEQ were to base the Turk/SWEPCO permit on the Little River instream standard of 100 mg/L, once dilution and mixing is allowed, this would result in an allowable end-of-pipe discharge of up to 3800 mg/l to still meet the instream waste concentration of 100 mg/L. These conditions should be modeled and presented as a comparison rather than solely those based on ADEQ's current approach of basing permits on past 303(d) listings. Without this, there is some question to the validity to the findings here and in other aspects of this proposal.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

# **COMMENT 15**

# 3.4.4 Discussion and Conclusions, page 3-22:

The authors note that the evaluation of the fish community demonstrated similar fish communities upstream and downstream of the Turk/SWEPCO discharge in the Little River. The combined collections from their study and USGS sampling during 2009 and 2013 provide evidence that the designated fisheries use in the Little River is currently attained under the existing mineral conditions.

Further, the document reports that an evaluation of the mineral tolerances of the existing fish community indicate a fish community that can be expected to be relatively insensitive to moderate changes in TDS (as indicated by specific conductance). The UAA notes that fish species that were classified as intolerant to TDS or for which data in Meador and Carlisle (2007) were lacking can be shown to be present in the Red River, which has substantially higher TDS than will be experienced in the Little River due to the operation of the Turk/SWEPCO plant. The conclusion drawn is that the anticipated increase in TDS due to the operation of the Turk/SWEPCO plant will support the designated fisheries use in the Little River downstream of the Millwood Lake dam.

The question these conclusion raise is even if the designated fisheries use in the Little River would potentially be maintained with the proposed increase in TDS criterion to 138 mg/L, why is it necessary if that use is being attained at the current 100 mg/L criterion? The answer seems to go back to Turk/SWEPCO's desire to operate at maximum efficiency. Under current conditions, the facilities efficiency appears to only be affected by ADEQ's approach in using limits based on the 500 mg/L criterion for the Red River as an end-of-pipe value. As noted previously, given that the Turk/SWEPCO discharge appears to have a negligible effect on the Red River, if ADEO were

to use existing criteria and allow for mixing and dilution in the Little River, there would be no need for this proposal.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

## **COMMENT 16**

3.5 Fisheries Evaluation in the Red River, page 3-24:

The objective of this evaluation was to determine whether the existing mineral regime supports the designated fisheries use as indicated by fish populations. The approach relied on the historical record of mineral concentrations in the Red River from ADEQ monitoring stations RED0025, RED0046, and RED0045 and the use of a sampling survey and historical data review of fish populations in the Arkansas portion of the Red River conducted by Buchanan et al. (2003). The intent was to characterize the existing fish communities and evaluate whether those existing communities are supported by the existing mineral regime, thus indicating attainment of the fisheries designated use.

This was not the intent of the evaluation. It seems clear that the existing mineral regime supports the existing fish community. However, this conclusion alone does not indicate use attainment. The study evaluated whether or not the existing *community* attains the designated use, and concluded that it does.

Although the Buchanan et al. (2003) data suggest that mineral conditions during 1995 and 2001 supported a high fisheries use attainment, it is somewhat problematic to rely on data that represents a single snapshot in time from 14 years ago. The authors draw the conclusion that since the Red River was meeting designated uses during the Buchanan et al. study and that because TDS data from the post-study period (2002 through 2013) indicate that mineral concentrations where generally higher during the time frame of the Buchanan et al. study, that fisheries beneficial uses are likely currently being attained. While the post-study period (2002 through 2013) TDS data gives an indication that uses are currently being attained, it does not provide the same level of confidence as current field data would.

In this case it is not problematic to rely on historical data. What is important is not how long ago the study was conducted but rather under what conditions—in this case the mineral concentrations—the study took place. The high level of use attainment reported by Buchannan et al. occurred concurrently with a particular minerals regime. Therefore, that particular minerals regime, whatever it was, supported that high level of use. The minerals concentrations occurring during the Buchanan et al. study were in general higher than during the post-survey period. If a certain range of minerals concentrations supports a high level of use attainment, then a slightly lower range of concentrations should also support attainment. The age of the Buchanan et al. study would be problematic if post-survey minerals concentrations were significantly lower than those present during the survey. In that case, SWEPCO would not be

able to say definitively whether the higher concentrations support the designated use, but the support (or lack thereof) would have nothing to do with the age of the study. Rather it would be because the conditions present during the study could not be applied to current conditions.

Showing that broadly written designated aquatic life uses can be supported under the existing mineral regime isn't necessarily a high bar given how broadly beneficial use definitions tend to be written. Determining attainment tends to depend on the individual state's assessment methodologies rather than the use definition.

Comment acknowledged.

## **COMMENT 17**

3.6.2 Red River, page 3-25:

The evaluation of the biological data demonstrated similar fish and communities upstream and downstream of the Turk/SWEPCO facility. The authors conclude that these finding reported suggest that the designated fisheries use is being attained that that the projected mineral concentration – presumably the proposed 138 mg/L TDS criterion will not be adversely affect attainment of the fisheries use.

Although the data indicate that the designated fisheries use is currently being attained, the document does not provide compelling information to support the contention that the proposed 138 mg/L TDS criterion will not adversely affect attainment. Further, concerns with impacts to benthic macroinvertebrate communities downstream of the discharge at the current criterion have been identified in previous comments.

It was asserted that the fish community is made up of tolerant species that would not likely be affected by a small increase in TDS. EPA did not dispute this conclusion. Regarding benthic macroinvertebrate communities and potential effects from the proposed SSC, EPA stated, "...it should be noted that small increases in TDS alone may have minimal effect on the existing benthic community..." [but EPA is concerned about other stressors that might also be present in the effluent]. SWEPCO's proposed SSC for TDS in the Little River is 138 mg/L, which reflects a TDS concentration in an undisturbed reference stream. A TDS criterion greater than 138 mg/L would be classified as a "significant modification" from the ecoregion reference stream value for TDS as described in Regulation No. 2.511(B).

## **COMMENT 18**

4.1 Evaluation of Routine WET Testing, page 4-1:

The recurring concern here is that the conditions simulated for toxicity analysis are reflective of current permit conditions, not what would likely be the conditions if the Red River was not listed.

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Removing the Red River from the 303(d) list or adding it to the list has no bearing on this discussion. The report addresses the potential for toxicity due to TDS at critical conditions.

More specifically, section 4.1 reports that a review of NPDES DMRs from what is likely the current permit (from November 1, 2012) indicate no episodes of sub-lethal toxicity for C. dubia and P. promaleas. Since the current permit is based on 500 mg/L as an end-of-pipe limit, sub-lethal effects would not be expected. The question is, how C. dubia and P. promaleas would respond under conditions that would be expected if a limit of up to 3800 mg/L was in place for the facility as it may be once the current 303(d) listing in the Red River is resolved.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

Relying on DMRs that are not reflective of what conditions will likely be long-term in the Little River and downstream in the Red River could be considered misleading.

The report does not state or suggest that a lack of toxicity under current conditions (i.e., TDS concentrations under the current operating regime) implies no toxicity under all future conditions. DMR data established that the effluent is not toxic under current conditions. A toxicity test to estimate the toxic threshold of minerals in a mixture of plant effluent and receiving stream showed that the toxic threshold for toxicity due to minerals (as TDS, sulfate, and chloride) is well above anticipated mineral concentrations at the critical dilution of 26%. Accordingly, there is a large margin of safety associated with mineral discharges from the plant with respect to compliance with NPDES WET testing.

#### **COMMENT 19**

## 4.2.3 Conclusions, page 4-5:

The authors state that calculated IC25 values (1,744 mg/L for TDS; 232 mg/L for sulfate; and 600 mg/L for chloride) indicate that toxicity due to minerals in WET testing would not be expected at the critical dilution (26%) until effluent TDS values reached approximately 6,976 mg/L. And that these results indicate that the threshold for toxicity due to minerals (as TDS, sulfate, and chloride) is well above anticipated mineral concentrations at the critical dilution of 26%. Accordingly, their conclusion is that there is a large margin of safety associated with mineral discharges from the plant with respect to compliance with NPDES WET testing.

It is unclear what the basis for the statement that toxicity due to minerals would not be expected until TDS values reach approximately 6,976 mg/L. Although the IC25 may be considered functionally equivalent to a NOEC by definition, it represents a 25% reduction in the rate of survival, growth or reproduction among test organisms.

If preferable, SWEPCO can use the same reasoning based on the NOEC, IC10, or any other endpoint of EPA's choosing. See the example using the NOEC below.

Further, Table 4.2 indicates that average concentrations of 6,000 mg/L cause 100% lethality.

Note that that report states that "... toxicity <u>due to minerals</u> in WET testing would not be expected <u>at the critical dilution (26%)</u> until <u>effluent TDS values</u> reached approximately 6,976 mg/L." The report does not state or imply that TDS concentrations greater than 6,000 mg/L are not toxic. Note that there is an error in Section 4.2.3 of the report. Based on an IC25 of 1,744 mg/L and a critical dilution of 26%, the report should have stated that toxicity at the critical dilution would not be expected until TDS in the effluent reached 6,708 mg/L  $(1,744 \div 0.26)$  instead of 6,976 mg/L (see page 4-5 of the report). A similar calculation based on the TDS NOEC of 1,260 mg/L predicts no toxicity due to minerals at the critical dilution at an effluent concentration of 4,848 mg/L  $(1,260 \div 0.26)$ . In either case, the threshold for toxicity due to TDS is well above anticipated permit limit of 1,620 mg/L (monthly average).

## **COMMENT 20**

5.2 Critical Flow Conditions, page 5-1:

Instream TDS concentrations in the Red River were modeled for two critical flow conditions. The authors note that calculations and model results for 7Q10 flow conditions were used to develop proposed criteria for the Little River, and for the Red River between the Little River and the AR-LA state line. The results for harmonic mean flow conditions were used to confirm that the proposed criteria in Arkansas will still allow the Louisiana criterion for TDS in the Red River (780 mg/L) to be maintained.

It's important that ADEQ be aware that the State of Louisiana is currently reviewing its mineral criteria. It is EPA's understanding that water quality data taken in the Red River near Shreveport, between 1991 and 2014 indicate that TDS concentrations of 780 mg/L are just above the 90th percentile. This may mean that it if the Arkansas' proposed criteria are adopted, there is a significant likelihood of TDS impairments in the downstream segment of the Red River in Louisiana.

Based on EPA's statement that "TDS concentrations of 780 mg/L are just above the 90th percentile," it can be concluded that TDS concentrations in the Red River near Shreveport exceed the criterion of 780 mg/L less than 10% of the time. The threshold for impairment using Louisiana's assessment methodology is 30%; therefore, the proposed criterion will not cause an impairment in the Red River in Louisiana.

It is important to note that 40 CFR 131.10(b) requires states to consider the water quality standards of downstream states when revising their criteria.

This report did consider attainment of downstream water quality standards in Louisiana. The proposed Red River criterion is reflective of current conditions, not an increase in mineral concentrations. Therefore, if current conditions are not causing an impairment in Louisiana, then the proposed criterion will not cause an impairment in Louisiana.

5.5 Model Results for 7Q10 Conditions, page 5-7:

Neither the RED0046 data nor the RED0009 data were considered to be a good representation of well-mixed concentrations in the Red River by the authors. Both the RED0046 station and the RED0009 station are located where the Red River does not appear to be laterally well-mixed because of significant tributary inflows a short distance upstream of each station.

The discussion explains that because of the location of the sampling sites for both RED0046 and RED0009, the data are likely measuring a disproportionate percentage of tributary water that has not mixed completely with the entire flow of the Red River. The authors contend that both tributaries (especially the Little River) contribute water that typically has TDS concentrations much lower than concentrations in the Red River. Because these two stations appear to be underestimating the TDS concentrations in the Red River and should not be used for comparison with predicted TDS values from the mass balance modeling, presumably because their inclusion could result in an underestimation the TDS concentrations in the Red River as compared to predicted TDS values from the mass balance modeling.

While it is possible that the inflow from the Little River at (RED0046) and the Sulphur River at (RED0009) may not be well-mixed, the problem with leaving them out is that it will likely mean an underestimation of TDS concentrations to the Red River, particularly in the lower reach near the LA state line.

It appears that the commenter believes that RED0046 and RED0009 are located in the tributaries rather than on the main stem. Inflows from the Little River and Sulphur River were not excluded from the mass balance model. If inflows from these rivers had been excluded, TDS concentrations in the Red River at the Louisiana state line would have been overestimated (not underestimated). The flow rates and TDS concentrations that were used for these two inflows are shown in Tables 5.1 through 5.3 of the report, Tables J.2 and J.3 in Appendix J, and in the model output in Appendix K (Data Types 20, 22, 24, and 26).

It is unclear how the segment reaches in Table 5.4 compare spatially to the long-term sampling sites.

Table 5.4 is a table of TDS concentrations for point sources. Table 5.5 shows TDS concentrations predicted by the model for each assessment reach, but it already shows the 90th and 95th percentiles of data at the long-term monitoring sites (third column in table). Figure J.1 in Appendix J provides a schematic diagram of the assessment reaches and point source inputs.

5.6 Model Results for Harmonic Mean Conditions, page 9:

The authors explain that the reason for simulating harmonic mean conditions was to confirm that proposed TDS criteria for the Red River in Arkansas would allow the Louisiana criterion for TDS in the Red River to be maintained under critical conditions. The harmonic mean simulation presented predicts that the TDS concentration at the AR-LA state line will be 661 mg/L, which is well below the Louisiana criterion for TDS in the Red River (780 mg/L). The conclusion is that the proposed criteria based on the mass balance modeling will be protective of the Louisiana criterion for TDS.

However, the harmonic mean simulation used the same input TDS concentrations as the 7Q10 simulation. As noted in the previous section, the exclusion of inflow from the Little River at (RED0046) and the Sulphur River at (RED0009) my mean an underestimation of TDS concentrations in the Red River, resulting in potential impairment of the current 780 mg/L TDS criterion that applies at the LA state line.

Please see SWEPCO's response to Comment 21 regarding inflows from Little River and Sulphur River. Section 5.0 of the report explains how the model works and gives the reasoning why it is inappropriate to use data from Red River stations (RED0046 and RED0009) to estimate water quality for tributary inflows.

As noted in comments on section 5.5, it is EPA's understanding that water quality data taken in the Red River near Shreveport, between 1991 and 2014 indicate that TDS concentrations of 780 mg/L are just above the 90th percentile. The 780 mg/L value is likely to go down.

Comment acknowledged, SWEPCO agrees that the Louisiana criterion is currently being met.

It would be advisable to consider what potential effect lower criteria in the Red River at the AR/LA state line may have on the predicted TDS criterion of 661 mg/L if LA revises its criteria down.

SWEPCO does not believe it is appropriate to speculate what future criteria may be proposed and potentially adopted for Louisiana and then perform analyses to see if the potential future criteria may be met under the proposed SSC in Arkansas.

## **COMMENT 23**

6.2.2 Domestic Water Supply, page 6-4:

The domestic water supply use for the reach of the Red River between the OK-AR state line and the Little River confluence has previously been removed. The authors provide data that indicate that the reach of the Red River from the Little River confluence to the AR-LA state line can also be expected to exceed secondary drinking water standards due to the same factors. Figure 6.1

shows that the Red River TDS concentrations at Fulton (downstream of the mouth of the Little River) frequently exceeded the secondary drinking water standard of 500 mg/L during the period of record. The authors also noted that both the Arkansas Department of Health and the Arkansas Natural Resource Commission to confirm that the stream is not used as an existing domestic water supply. Their conclusion is that the domestic water supply use is neither an existing nor attainable use in the Red River downstream of the mouth of the Little River.

Although it is within Arkansas' authority to remove the domestic water supply use for the segment of the Red River below the confluence with the Little River to the LA state line, AR must consider the downstream state standards and ensure that its action would not affect domestic water supply/drinking water uses in Louisiana. See 40 CFR 131.10(b).

For many years (since 1999 at least), the Red River in Louisiana has been designated for drinking water supply with a TDS criterion of 780 mg/L TDS; SWEPCO must assume that Louisiana, with EPA's approval, considers this TDS criterion to be protective for domestic water supply/drinking water use.

## **COMMENT 24**

7.0 Alternatives Evaluation, page 7-1:

Arkansas' UAA guidance and CPP require that a petition to increase dissolved minerals concentrations above existing conditions must include an evaluation of alternatives to the direct discharge of the water. Four alternatives that have been reviewed including:

- 1. Distillation treatment,
- Treatment using a constructed wetland,
- 3. Pumping the wastewater to a larger stream that holds the potential for dilution of the minerals, and
- 4. Reverse osmosis (RO) treatment of the wastewater.

From the outset, the use of constructed wetlands was dismissed as an option for this facility. The authors based this exclusion on the argument that constructed wetlands can only be used to reduce sulfate, which results in the production of bicarbonate in place of sulfate (Hedin et al. 1989). The authors do note that a constructed wetland could, in principle, reduce sulfate in the discharge from this facility, the resulting TDS concentration would not be decreased (due to the replacement of the sulfate ions with bicarbonate ions), but argue that no net benefit would be obtained, stressing that TDS is the only concern for this evaluation.

The premise in rejecting a constructed wetland appears to be based on the premise that a constructed wetlands would only reduce sulfate to bicarbonate, which would not provide a net reduction in TDS. Looking at the Hedin et al. paper, it is specific to the treatment of acid mine drainage, not the type of discharge that would result from blow down cooling water from a

power plant. The paper does indicate that with an increase in sulfate reduction, bicarbonate alkalinity will be produced and pH will increase, but it does not say that sulfate is directly reduced to bicarbonate.

Sulfate cannot be reduced to bicarbonate and the report provides the appropriate reference that explains the chemistry that is the basis for the conclusion of the evaluation (Hedin et al. 1989). Further discussion is provided below.

In treatment of acid mine drainage, sulfate would primarily be reduced to sulfite and tend to react with available metal to form sulfide precipitates. The Hedin paper refers to bacterial production of bicarbonate formation, particularly in anoxic limestone drains, but does not refer to the reduction of sulfate to bicarbonate. Which is not unexpected since as noted earlier, the paper is looking at treatment of acid mine drainage, not sulfate or other TDS constituents in particular. Given this, EPA considers a constructed wetland to be a viable and the preferred alternative to altering the applicable TDS criterion.

The sulfate reduction reactions are not specific to acid mine drainage. The chemistry of the sulfate ion does not depend on its source and sulfate is not reduced to bicarbonate in any context. To understand the reasons constructed wetlands cannot be used to reduce TDS, it is useful to review the typical chemical reactions that occur in sulfate reduction. For sulfate-reducing bacteria to carry out their function, a source of carbon must be available. In a constructed wetland, the source of the carbon can take many forms, so a generic formula for the reaction has been developed.

Using sodium as the primary cation for this water, the reaction for sulfate reduction can be expressed as follows:

As stated by the commenter, the sulfide ion is subsequently available to react with heavy metals (Me) as follows:

$$HS^- + Me^{2+} \rightarrow MeS + H^+$$

While it is true that the metal sulfide will likely precipitate, the generation of sodium bicarbonate more than offsets the mass of sodium sulfate that has been reduced. In fact, using the above molecular weights of the two compounds, one mole of sodium sulfate (molecular weight of 98 grams/mole) reacts with organic carbon to produce two moles of sodium bicarbonate (molecular weight of 84 grams/mole \* 2 moles = 168 grams).

While the specific pathways can be complex, the overall reaction, if the process is taken to completion, demonstrates the reasons that constructed wetlands are not used for TDS reduction.

The authors contend that there are two alternatives for achieving compliance with the existing TDS criteria for the Little River and Red River:

- 1. RO treatment to remove or reduce dissolved minerals, and
- 2. Pumping the effluent to a larger river that holds the potential for dilution of the minerals.

In considering these options, the authors state that there is not a river in the vicinity that could be used as dilution to completely avoid a change in the water quality standards. Although the Red River has sufficient volume, it has a TDS criterion of 500 mg/L, the expected concentration of TDS in the effluent from this facility would not meet the Red River TDS criterion and would still require the completion of a UAA or the installation of partial RO treatment.

EPA considers ADEQ's use of TDS criteria applicable to the Red River, particularly the way the criterion has been applied as the emerging central element here. As noted in previous comments, ADEQ issued permit Turk/SWEPCO is operating under is based on the existing 500 mg/L TDS criterion for the Red River as end-of-pipe limits. The permit carries those limits because the receiving Red River segment was on the state's 2008 303(d) list. From a numeric perspective, ADEQ has issued the permit with a monthly average of 500 mg/L TDS end-of-pipe. This end-of-pipe value equates to an instream waste concentration of 43 mg/L. If ADEQ based the permit on the Little River instream standard of 100 mg/L, once dilution and mixing is allowed, the result is an allowable end-of-pipe discharge of up to 3800 mg/L TDS. While the end-of-pipe concentration is higher, this approach would meet the instream waste concentration based on the ecoregion-based 100 mg/L TDS while allowing the Turk/SWEPCO facility to operate efficiently.

EPA strongly recommends that rather than revise TDS criteria particularly for the Little River, in what appears to be an effort to avoid this permitting scenario, that this approach be considered.

It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

## **COMMENT 25**

7.1.2.2 Option 2: RO Treatment/Discharge to Little River at Future Effluent Limits, page 7-9:

The authors anticipate that the TDS criterion in the Little River, and thus the effluent limitations in future NPDES permits for the SWEPCO plant, could be lowered to 100 mg/L. The conclusion drawn is that this would require that all of the effluent be treated through an RO system if it continues to be discharged to the Little River, in order to achieve an effluent limit of 100 mg/L for TDS.

However, as noted previously, the 100 mg/L criterion that applies to the Little River is not the same as a derived NPDES permit limit. The permit limit that would be derived from the

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100 mg/L TDS criterion once dilution and mixing is allowed, would likely be an allowable end-of-pipe discharge of up to 3800 mg/L. Given that this concentration would allow the facility to meet the 100 mg/L TDS criterion in the Little River, there would be no need to consider RO.

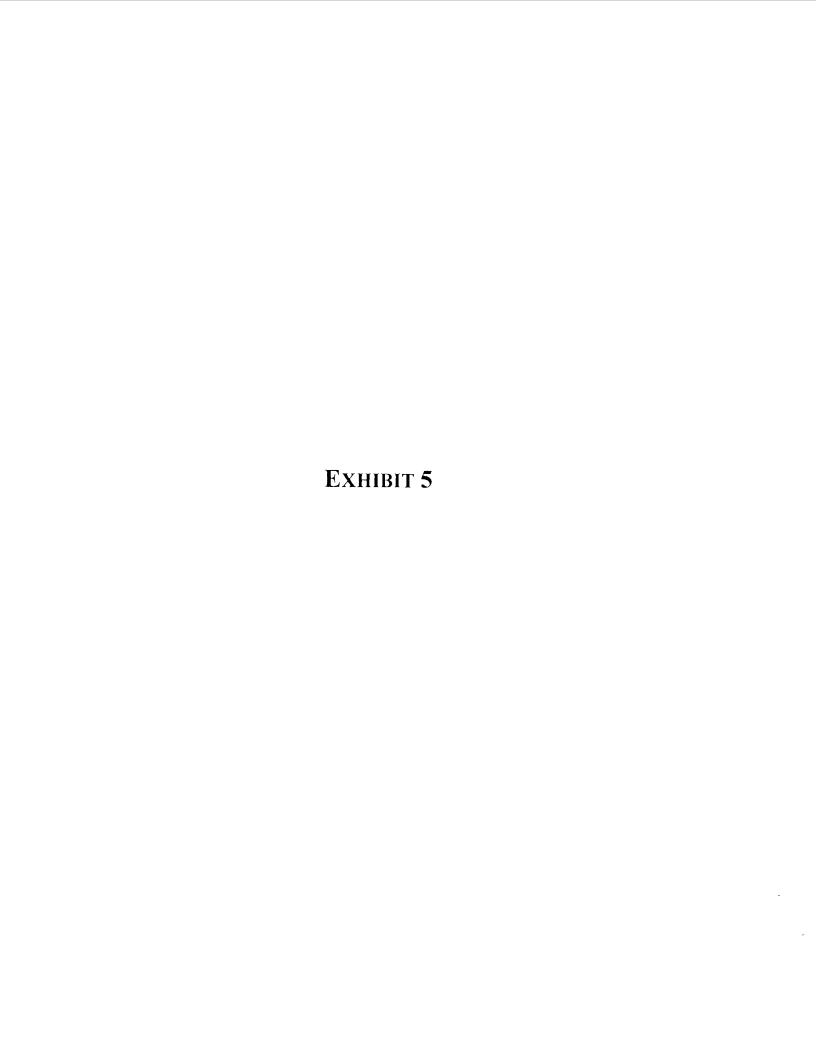
It is SWEPCO's understanding that discussions between ADEQ and EPA have addressed EPA's concern regarding this recurring comment.

## **COMMENT 26**

7.3 Summary of Costs, page 7-10:

The authors summarize the estimated costs of various treatment scenarios compared to the cost of direct discharge based on the proposed site-specific criteria in Table 7.1. The cost described here appear to be the potential cost associated with RO treatment of the entire volume of the Turk/SWEPCO discharge. It is unclear what volume of the discharge would need to be treated.

Please see Sections 7.1.2.1 through 7.1.2.3. Each of the three scenarios states how much of the discharge would be treated in order to meet the stated permit limitations.



# Response to EPA Technical Comments SWEPCO Technical Justification for a Site-Specific Criterion in the Little River

On October 20, 2015, Region 6 of the US Environmental Protection Agency (EPA) provided technical comments dated "September XX, 2015," to the Arkansas Department of Environmental Quality (ADEQ) on Southwestern Electric Power Company's (SWEPCO's) Technical Justification for a Site-Specific Temperature Criterion in the Little River, Hempstead and Little River Counties, Arkansas, which was submitted to ADEQ on September 10, 2014.

EPA's comments have been reproduced below, followed by SWEPCO's responses in blue text. EPA's comments are given in italics for ease of distinguishing between the comment and SWEPCO's response, and the comments have been numbered to allow for referencing within the document.

#### COMMENT 1

1.1 Background & Overview, page 1-1

This document states that the Turk/SWEPCO facility's National Pollutant Discharge Elimination System (NPDES) permit (No. AR0051136) includes a daily maximum temperature permit limit of 35°C (95°F). Given that Arkansas Reg. 2 specifies and the authors acknowledge that the applicable temperature limits for streams is 30°C (86°F), the basis for the maximum temperature limit of 35°C (95°F) in SWEPCO's current permit is unclear.

## Comment acknowledged.

The document also notes that the Little River downstream of Millwood Lake has a lower temperature criterion than Millwood Lake and the Red River immediately downstream of the Little River. Both Millwood Lake and the Red River in Arkansas have a temperature criterion of 32°C (89.6°F) (APCEC 2011). Figure 1.2 is presented to illustrate the spatial inconsistency of the temperature criteria.

EPA is correct. Comment acknowledged. As the report states in Section ES.1:

If the Little River is designated as impaired for temperature due to recent temperature measurements, the facility's NPDES permit limit for temperature at Outfall 001 would be reduced to the Little River instream criterion of 30°C (86°F) without the usual allowance for a mixing zone (i.e., using typical end-of-pipe calculations to develop permit limits for a thermal discharge). The facility's operation would be adversely impacted by a temperature impairment designation in the Little River that may limit or prevent discharges during warm weather and reduce the efficiency of the facility's operation during other times of the year. Adjusting the temperature criterion for the Little River downstream of Millwood Lake to reflect current ambient conditions during the critical summer conditions

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will (a) prevent the Little River from being inappropriately listed as impaired and (b) result in NPDES temperature limitations for the SWEPCO discharge that are appropriate and protective of the designated uses of the Little River.

Please note that the draft ADEQ 2014 303(d) list includes this reach of the Little River as impaired for temperature.

The document notes that Millwood Lake has a temperature criterion of 32°C (89.6°F), the Little River, downstream of Millwood Lake has a 30°C (86°F), and the Red River immediately downstream of the Little River has a temperature criterion of 32°C (89.6°F) (APCEC 2011) to illustrate the spatial inconsistency of the temperature criteria downstream of Millwood Lake. Spatial inconsistency has no bearing on toxic or conventional criteria that are protective of aquatic life uses (ALU) and is not a basis for altering applicable criteria.

The technical basis for the proposed SSC is identified and discussed in the report (see Sections ES-1, 3.4.4, and 4.0); the basis for the SSC for temperature in the Little River is not spatial inconsistency.

It is not unusual for temperatures in a reservoir to be higher than upstream and typically downstream lotic waters given the retention time and particularly the shallow depth of Millwood Lake.

SWEPCO agrees that it is not unusual for temperatures in a reservoir to be higher than downstream lotic water, but this fact is only true for deeper reservoirs with a cool- or cold-water discharge. Although Millwood Lake dam has an outlet near the bottom, Millwood Lake is shallow and generally unstratified such that the water temperature at the outlet reflects the temperature of the lake. Temperature measurements taken at the outlet as well as in-lake profiles confirm this statement (see Table 6.1 of the report). It is incorrect to state that the hydraulic residence time of Millwood Lake results in cooler downstream lotic waters.

Although we acknowledge that it may be difficult to manage the flow through a lake as shallow as Millwood given its design, relatively high retention time and higher tailwater race temperatures, but it's unclear how that would affect the entire length of the Little River or why the riparian cover downstream along the Little River could not be enhanced or restored.

The terms "high" or "low" retention times can be ambiguous; however, when compared to other reservoirs of similar surface areas, Millwood Lake has a relatively *low* retention time. Millwood Lake is a shallow lake with short hydraulic residence time (see page 6-3 of the report). The physical setting of a warm-water discharge from Millwood Lake primarily determines the downstream temperature regime of the Little River. Given the Little River's width in this reach (below Millwood Lake), restoring the riparian cover along this wide river would not significantly affect the river's temperature regime.

1.1 Background & Overview, page 1-3

The authors note that if the temperature criterion is not changed and the impairment status continues, the facility's NPDES permit limit for temperature at Outfall 001 would be reduced to the Little River instream criterion of 30°C (86°F) without the usual allowance for a mixing zone (i.e., using typical end-of-pipe calculations to develop permit limits for a thermal discharge). As such, the authors note that facility's operation would be adversely impacted by a temperature impairment designation in the Little River that may limit or prevent discharges during warm weather and reduce the efficiency of the facility's operation during other times of the year.

Does this mean that the decision was keeping the existing criteria in place versus operating efficiency (i.e., cost)? If so, the basis for the proposed amendment 40 CFR 131.10(g)(6), which requires an economic analysis to determine if SWEPCO can support upgrades to its process/facilities w/out substantial and widespread impacts.

The basis for the decision to develop the proposed SSC was based on the potential for this reach of the Little River to become listed as impaired on the 303(d) list, thus resulting in end-of-pipe permit limitations set at the current instream criterion for the facility. The SSC was proposed to reflect ambient conditions. The proposed criterion is based on data that are not impacted by SWEPCO's discharge. A correction to the existing criterion is needed even without the discharge.

If the described scenario comes to pass (which is already supported by the draft 2014 Arkansas 303(d) list), it will result in the economic impacts described in the report.

## **COMMENT 3**

1.1 Background & Overview, page 1-4:

The authors indicate that adjusting the temperature criterion for the Little River downstream of Millwood Lake is necessary to reflect current ambient conditions during the critical summer conditions and would result in NPDES temperature limitations that are appropriate and protective of the designated uses of the Little River. An important question is if current ambient conditions reflect the highest attainable condition given the requirements outlined in 40 CFR 131.10?

The temperature data provided in the report support this conclusion. Furthermore, there is nothing that SWEPCO, ADEQ, or the US Army Corps of Engineers (USACE) can do to "improve" water temperatures such that the existing criterion could be attained 90% of the time (to avoid being listed as impaired). The recommended SSC is based on data collected from stations *upstream* of the SWEPCO discharge.

2.1 NPDES Permit, page 2-1

The document again refers to the Turk/SWEPCO discharge to the Little River under NPDES Permit No. AR0051136 and the effluent limitation for temperature of 35°C (95°F). As noted previously, it's unclear how this facility can have an NPDES permit that allows a 35°C (95°F) when the applicable temperature for streams is the 32°C (89.6°F). Is there a specific implementation procedure or mixing zone allowance?

Please refer to ADEQ's Statement of Basis for Permit No. AR0051136.

## **COMMENT 5**

2.2 Applicable Water Quality Standards, page 2-2

The authors suggest that the temperature regime of the Little River downstream of Millwood Lake has been altered as a result of physical hydrologic alteration of the system caused by the construction of Millwood Lake. The authors contend that Millwood Lake and its confluence with the Little River has a typical width between 200 and 400 feet with only a small portion of the channel being shaded by trees along the banks.

Based on images of the USACE dam, the Little River may be "200-400" wide" immediately below the dam as stated here, but appears to constrict below River Run Park and appears to be generally consistent with the average stream of reference streams in the Gulf Coastal Plain ecoregion.

Without supporting information for multiple downstream locations on the Little River below the Millwood, this claim does not appear to be supported. Photographs, specifically aerial photos should be included in this proposal to support this claim.

The comment above is incorrect. As noted on page 2-2 of the report, the reference streams in the Gulf Coastal ecoregion had an average width of 23 ft (range was 6.4 ft to 35 ft) with an average canopy coverage of 84% (range was 47% to 100%). The Little River below Millwood Lake dam is about an order of magnitude wider than the ecoregion reference streams and, as a result, has significantly less instream cover. Figures 1 and 2 (below) show Google Earth imagery of the Little River from Millwood Lake dam to the mouth of the river on July 23, 2009, when flows were likely to be low. Visual inspection of the figures indicates somewhat uniform width throughout the entire reach. The figures also indicate the locations of 31 transects downstream of River Run Park for which wetted width was measured using the Google Earth measuring tool. The transects were selected to represent a range of wetted stream widths. Wetted width ranged from 174 to 479 ft and averaged 244 ft.



Figure 1. Google Earth image of upper portion of Little River downstream of Millwood Lake dam showing the locations of transects measured using the Google Earth measuring tool.



Figure 2. Google Earth image of lower portion of Little River downstream of Millwood Lake dam showing the locations of transects measured using the Google Earth measuring tool.

# 2.3 Water Temperature Monitoring, page 2-4

Tables 2.4 and 2.5 presents summary statistics for water temperature data collected by SWEPCO from April to October 2012. These tables indicate that the Turk/SWEPCO discharge increases instream temperatures. With the exception of a higher maximum temperature reading upstream of SWEPCO, which may be a single event, the 95th percentile, percent daily max > 30°C and percent daily max > 32°C indicate that the facility adversely impacts temperature in the Little River.

The data summary presented in Table 2.4 of the report was confusing and misleading. The summary statistics for each station represented the entire data set obtained at each station. Because the temperature recorders were not deployed at each station simultaneously, the summary statistics for some of the stations included data recorded earlier in the year, when water temperatures were cooler. Therefore, the data summarized in Table 2.4 contained a seasonal component that confounded differences among stations and was not apparent from the data as presented. The following revised Table 2.4 provides a summary of the data using the same period of record (July through August of 2013).

Table 2.4. Revised statistics for Little River temperature monitoring using a common period of record (July through August of 2013).

Summary Statistic	ML-0	LR-2	LR-1.5	LR-1	LR-0	LR-00
Number of days with measurements	62	62	62	62	62	62
Number of measurements	5,952	5,952	5,952	5,952	5,923	5,952
Minimum (°C)	25.5	24.2	24.9	25.0	26.3	26.3
Mean (°C)	29.2	28.6	28.7	29.5	29.5	29.3
Maximum (°C)	34.3	32.3	32.0	32.9	34.7	33.0
25 <sup>th</sup> percentile (°C)	28.2	27.9	28.0	28.8	28.8	28.5
Median (50 <sup>th</sup> percentile) (°C)	29.1	28.6	28.7	29.6	29.6	29.4
75 <sup>th</sup> percentile (°C)	30.2	29.3	29.5	30.2	30.2	30.1
95 <sup>th</sup> percentile (°C)	32.1	30.6	30.5	31.5	31.4	31.1
Percent of all data > 30°C	28.6%	10.4%	11.9%	33.9%	32.1%	27.1%
Percent of daily maximum data > 30°C	59.7%	30.6%	38.7%	69.4%	64.5%	56.5%
Percent of all data > 32°C	5.5%	0.1%	0.0%	1.2%	1.9%	1.6%
Percent of daily maximum data > 32°C	27.4%	1.6%	0.0%	14.5%	14.5%	4.8%

The summary in the revised Table 2.4 shows the following:

- 1. No evidence of warmer water temperature downstream of the discharge as compared to immediately upstream (LR-00 versus LR-0, respectively);
- 2. Daily maximum temperatures in Millwood Lake (ML-0) that exceed the criterion almost 60% of the time and daily maximum temperatures immediately below the dam (LR-2) and one mile downstream from the dam (LR-1.5) that exceed the criterion between 31% and 39% of the time; and
- 3. Frequent exceedances of the 30°C instream criterion as shown by 75<sup>th</sup> percentile values greater than 30°C at four out of six stations in the Little River.

## **COMMENT 7**

2.3 Water Temperature Monitoring, page 2-8

The authors note that temperatures in the Little River during 2013 (Figures 2.3 and 2.4) appear very similar to those measured in Millwood Lake near the dam (Figure 2.5). They note that exceedances of the Little River temperature criterion occur at all of the Little River sampling locations, both upstream and downstream of the SWEPCO outfall, during 2012 and 2013 (Figures 2.2 through 2.4).

These figures generally show that there is an increase in temperature associated with the time of the year sampled. There is a clear correlation with ambient air temperature and water temperature, which among other factors can have a significant influence instream temperatures. This correlation is not considered here.

A regression of air temperature and water temperature is not necessary for this section of the report. The monitoring data demonstrate that the reach of the Little River upstream of the SWEPCO discharge exceeds instream criteria and that the discharge does not have an appreciable effect on temperatures.

The comparison with temperatures in Millwood Lake near the dam are of limited value since this lake is relatively shallow and retention times tend to be fairly high in most lakes.

Again, please note that Millwood Lake has a relatively low retention time. The comparison to temperatures in Millwood Lake near the dam is highly relevant because it shows that the temperature of the Millwood Lake dam outflow is the same as the temperature of the lake.

In addition, the data reported in Fig. 2.8 which show a decrease in temperature from LR0 to LR00 does not appear to be consistent with the summary statistics presented for the same site in Table 2.4, which reports an increase of all and daily maximum data  $> 30^{\circ}$ C.

The revised Table 2.4 addresses this inconsistency.

2.3 Water Temperature Monitoring, page 2-10:

Warming in a large shallow impoundment like Millwood is expected as it is in the immediate tail water area. However, it's unclear if this narrative is suggesting that temperatures in the Little River are the result of the Turk/SWEPCO discharge or continuing the claim that the river remains wide and unshaded further downstream.

SWEPCO contends, and the data demonstrate, that the temperatures in the Little River are the result of the ambient conditions and are not due to the SWEPCO discharge.

As noted earlier, a simple internet search produced maps and photographs that suggest that this is not the case downstream.

Please see the response to Comment 5 (along with Figures 1 and 2 of this document) regarding the stream width of Little River downstream from Millwood Lake dam. Additionally, the revised Table 2.4 demonstrates that temperatures in the Little River consistently exceed the criterion upstream of the SWEPCO discharge. The temperatures in the Little River are the result of ambient conditions and are not due to the SWEPCO discharge.

## **COMMENT 9**

2.4 Evaluation of Little River Water Temperatures Upstream of SWEPCO Outfall 001, page 2-10

The discussion indicates that Table 2.5 provides an inventory of water temperature data collected in the Little River downstream of Millwood Lake and upstream of SWEPCO Outfall 001 that were used to evaluate the temperature criterion for the Little River. However, Table 2.5 also includes data from LR00, which is reported as 1 mile below SWEPCO Outfall 001.

That is correct. Although Table 2.5 includes data from LR-00, which is reported as one mile below SWEPCO's Outfall 001, the proposed criteria are based on the data collected upstream of Outfall 001.

## **COMMENT 10**

2.4 Evaluation of Little River Water Temperatures Upstream of SWEPCO Outfall 001, page 2-15

A comparison of temperature statistics between the two FTN monitoring locations near the dam below Millwood Lake and the two locations farther downstream (near SWEPCO) indicate that that locations farther downstream have slightly higher temperatures during July through August and more exceedances of the 30°C (86°F) criterion (Tables 2.3 and 2.4).

SWEPCO agrees.

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This paragraph touches on the potential source affecting ambient temperatures in Millwood Lake and the Little River, but provide no details. The summary data, statistics, and statements presented here do not appear to have considered that air temperature is highly correlated with the significant positive heat flux components including solar radiation and with stream water temperature. The correlation is supported by the USGS' Stream Segment Temperature Model (SSTEMP), which asserts that air temperature will usually be the single most important factor in determining mean daily water temperature.

A regression of air temperature and water temperature is not necessary for this section of the report. The monitoring data as summarized in the revised Table 2.4 and on Figures 2.9 through 2.11 of the report clearly indicate that reach of the Little River upstream of the SWEPCO discharge exceeds instream criteria and that the discharge does not have an appreciable effect on temperatures. In any case, meteorological influences that are beyond SWEPCO's control cause ambient water temperatures to exceed the existing criterion without any effects from a thermal discharge.

The authors also note that ADEQ's (303(d)) assessment for 2014 was conducted using the SWEPCO 2012 and 2013 data and FTN 2013 data from this stream reach and that ADEQ's draft 2014 303(d) list includes a temperature impairment for the Little River downstream of Millwood Lake. Did SWEPCO and FTN use ADEQ monitoring stations to collect these data?

No, ADEQ did not have any monitoring stations on the Little River.

Is there information to support the reported listing?

Please see ADEQ's draft 2014 303(d) list (http://www2.adeq.state.ar.us/water/branch\_planning/303d/pdfs/list\_of\_impaired\_waterbodies\_2\_0140401.pdf).

While the assessment methodology used to determine impairment appears to be consistent with ADEQ methodology, it is unclear if 3rd party (SWEPCO and FTN) data be was used by ADEQ to support a 303(d) listing?

It is SWEPCO's understanding that ADEQ used data collected by FTN and SWEPCO for the 2014 draft 303(d) listing.

In instances where the temperature exceeds the criteria, it would likely result in a reduction in overall species richness and/or abundance because of excessive heat. Discussions of the macroinvertebrate and fish communities do not appear to show such effects.

EPA is correct. The biological data show no effects due to the discharge.

2.5.1 Threatened and Endangered Aquatic Species, page 2-18

The document reports that there have been no live A. wheeleri collected from just above or below the Turk/SWEPCO discharge. Their absence indicates that the Turk/SWEPCO discharge is having an effect on water quality and the presence of A. wheeleri. Although the physical characteristics and inputs to Millwood Lake may affect the ability of the Little River to support A. wheeleri, an important aspect of a UAA is to not only to identify current conditions. The document does not provide any discussion of whether temperature or other stressors may be affecting or limiting the presence of A. wheeleri in this portion of the Little River.

The effects to A. wheeleri are due to the hydraulic modification (i.e., construction of Millwood Lake dam on the Little River). The reports note that construction of the dam affected mussel distribution and abundance before construction of the SWEPCO facility. Studies performed after Millwood Lake was formed but prior to construction of the facility did not document the presence of A. wheeleri in the Little River in the vicinity of the future intake and outfall locations.

Although the rabbitsfoot mussel (Quadrula cylindrica cylindrica) is known to occur in some streams in southwest Arkansas, including the Little River upstream of Millwood Lake, it has not been documented in the Little River downstream of the Lake to the Red River. The rabbitsfoot is likely limited by the physical characteristics Millwood Lake and its effect on the Little River, as well as the Turk/SWEPCO discharge.

SWEPCO agrees that the rabbitsfoot is not found in the Little River downstream of Millwood Lake. The changes in the physical characteristics, and therefore habitat, of the Little River due to construction of the Millwood Lake dam occurred before construction of the SWEPCO facility.

## **COMMENT 12**

3.4.3.2 Fish, Ecoregion Key and Indicator Species

The authors describe presence and absence of ecoregion key and indicator species per APCEC (2011) which is summarized in Table 3.7. Of three key and indicator species expected to be present based on habitat preferences, one species was present in the pooled list. One species not expected to be present (redfin shiner) was present in the pooled list. The reported data is not specific as to site - is this a comparison of upstream/downstream of the SWEPCO discharge?

Sampling locations are given in Section 3.4.2 of the report.

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The location of this sampling would be useful, as it would give some indication of whether the presence of only 25% of key and indicator species is the result of a generally degraded/impacted system, resulting from hydromodification and high ambient air temperatures driving water temperature, or exacerbated by temperature inputs from the SWEPCO facility.

Figure 3.1, which is referenced in the text, shows the sample reaches and the text in Section 3.4.2.1 describes how these reaches were sampled. SWEPCO noted the presence of one of the three expected key/indicator species and the presence of one key/indicator species that was not expected. SWEPCO is unsure how commenter arrived at the value of 25%; there is either one species present out of three expected or two species present out of four expected.

It is also misleading to use percentages to describe small numbers such as these. Since there are virtually no temperature impacts due to the plant, the presence of one out of three (or two out of four) key/indicator species, to the extent it needs explaining, must be due to factors other than temperature inputs from the plant.

## **COMMENT 13**

# 3.4.4 Discussion and Conclusions, page 3-22

The following is a preface to responses to comments on this section to clarify the approach and findings of this section.

The fish community data were analyzed two different ways. The first compared upstream versus downstream reaches (see Section 3.4.2 of the report for sampling locations); the methodology is described in Section 3.4.3.2 of the report ("Fish: Upstream versus Downstream"). This analysis demonstrated virtually no difference between the upstream and downstream samples (percent similarity between the two reaches was 81.3%; Morisita's similarity index value was 0.96). This analysis addressed the impact of the discharge on the fish community but did not address whether the fish community as a whole, under the elevated temperature regime (which is due to ambient warming), met the designated use.

The second analysis addressed use attainment of the overall community under the elevated temperature regime. Since there was no difference between upstream and downstream fish collections, the upstream and downstream data were combined with additional data collected by the US Geological Survey (USGS) in 2009 and 2013 within the immediate vicinity of the upstream sampling reach. The species list from this pooled data set was then evaluated as follows:

# A. Species Composition

The pooled data set was evaluated to assess whether or not the species present represented the expected assemblage of ecoregion fish species based on biogeography, habitat preferences,

seasonality and gear selectivity. The absence of species that "should" occur would indicate potential impacts due to water quality. The analysis was performed as follows:

- 1. Based on information in the literature, a list of "expected" species was generated based on geographic distribution and habitat preferences (Appendix D). This reference list provided a list of fish species that might be expected to be present in the Little River downstream of Millwood Lake dam based on geographical distribution and habitat preferences. Note that this list can only be a presence/absence list. It is not possible to generate expected relative abundance in the reference list.
- 2. This reference list of expected species was then compared to the pooled list.
- 3. Differences between the final reference and pooled lists were evaluated to determine if the differences could be attributed to factors such as habitat preferences, seasonality, rarity, or gear selectivity, as opposed to water quality.

This evaluation found that the pooled list contained 37 of the 52 (71%) expected species found in the reference list. SWEPCO evaluated the 15 "absent" species with respect to microhabitat preferences, seasonal distribution, and susceptibility to the sampling gear used and concluded that the pooled species list closely resembles expectations based on zoogeography, habitat preferences, and gear selectivity. The commenters did not dispute this finding.

# B. Temperature Tolerances

The tolerance of the existing community to thermal stress was assessed by compiling tolerance values from the literature and analyzing the distribution of tolerance values of fish in the pooled collection.

The result of this analysis indicated that the *expected* (reference) community was made up of tolerant to moderately tolerant species and should be relatively insensitive to moderate changes in water quality, including temperature. The distribution of tolerance classes among the fish species present in the pooled list was similarly skewed towards tolerant and moderately tolerant species and is a reflection of the expected fish community.

The analysis of the fish sampling data demonstrated that that the fish species composition of the Little River downstream of Millwood Lake dam (1) is not affected by the discharge, and (2) matches expectations based on biogeography, habitat, and tolerance.

The commenters did not dispute this finding.

Therefore, the existing temperature regime, which is the result of the combination of hydrologic modification and ambient climate, supports the designated fishery use.

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The authors observed that the strongly similar fish and benthic communities upstream and downstream of the discharge. The suggestion is that this finding indicates that the current operation of the discharge has negligible effect on the existing biological communities. Although relative abundance upstream/downstream showed reasonably good comparison at around 71%

The 71% value does not refer to the upstream/downstream comparison. The analysis of upstream versus downstream demonstrated virtually no difference between the upstream and downstream samples (percent similarity between the two reaches was 81.3%; Morisita's similarity index value was 0.96).

The Meador and Carlisle (2007) data only showed 61% of the expected species are present and only 25% of the expected species for the Little River present in the pooled sample. It is unclear if this is the result of a degraded system or that the discharge has more than a negligible effect.

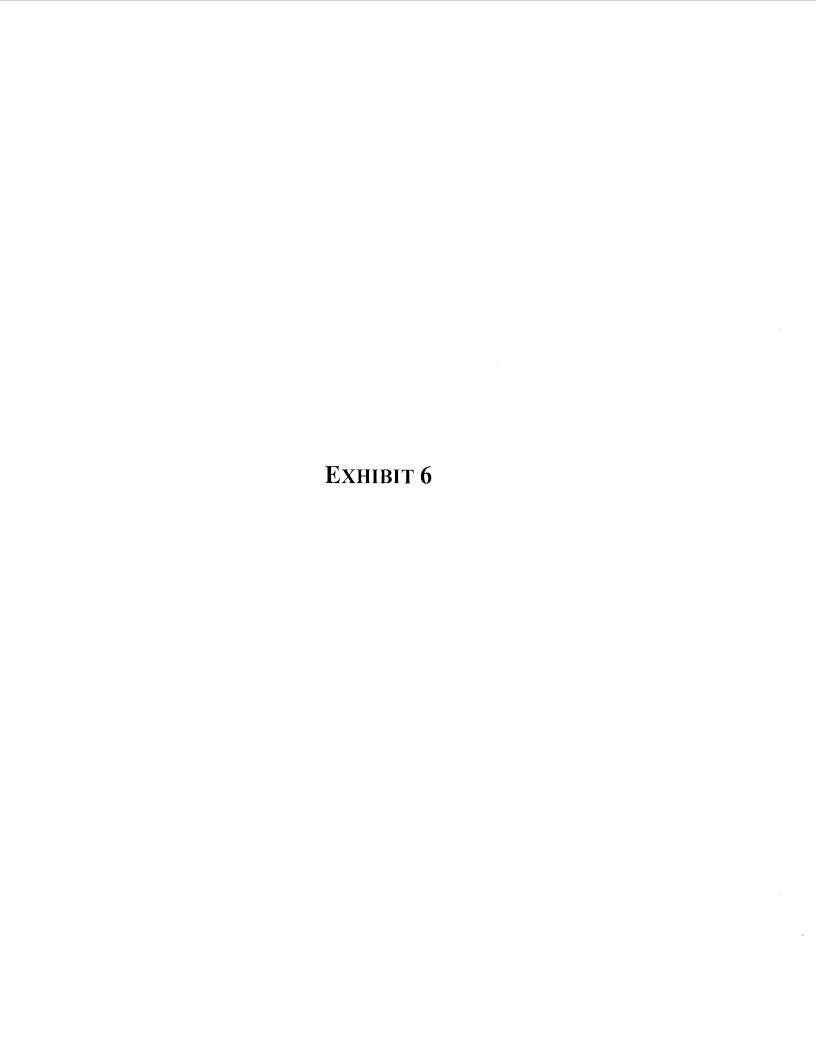
The report describes how the list of expected species was compiled in Section 3.4.2.2. The Meador and Carlisle data do not provide a list of expected species. They provide tolerance values for a list of North American fish species. The report stated that of the 52 expected species, Meador and Carlisle provide tolerance vales for 32 species (61%). This does not mean that only 61% of the expected species were present. The report does not state or imply "...only 25% of the expected species for the Little River were present in the pooled sample."

## **COMMENT 14**

3.4.4 Discussion and Conclusions, page 3-23

The discussion notes that ecoregion key and indicator species are generally not expected to be present in the reach of the Little River due to the habitat preferences of those species and that the low numbers of key and indicator species present in the pooled list (two species) is a reflection of this expectation. This statement appears to contradict Table 3.7, comparing expected key/indicator species in the Little River to what was present in the pooled list (25%).

Table 3.7 shows that only three (or four, depending on how they are counted; see response to Comment 12) of the 12 ecoregion key/indicator species are expected to be present based on habitat. Of these, we collected two. Noting that only three or four key/indicator species would generally be expected to be present does not contradict the observation that two key/indicator species were present.





#### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

REGION 6 1445 ROSS AVENUE, SUITE 1200 DALLAS TX 75202-2733

October 20, 2015

Sarah Clem ADEQ Branch Manager Arkansas Department of Environmental Quality 5301 Northshore Drive Little Rock, AR 72118-5317

Re: Environmental Protection Agency comments on the Proposed 3<sup>rd</sup> party rule by the Southwestern Electric Power Company John W. Turk, Jr. Power Plant (SWEPCO facility)

Dear Ms. Clem:

The Arkansas Pollution Control and Ecology Commission (APC&EC) held a public hearing on November 17, 2014, to receive comments on a third-party proposal by the Southwestern Electric Power Company John W. Turk, Jr. Power Plant (Turk/SWEPCO) regarding water quality standards changes to APC&EC Regulation 2. The proposed amendments to Regulation 2 include a modification of the Total Dissolved Solids (TDS) water quality criterion and temperature criterion for the Little River from Millwood Lake to the mouth of the Little River, and modification of the TDS water quality criterion and removal of the designated, but not existing, domestic water supply use for the Red River from the mouth of the Little River to the Arkansas/Louisiana state line. The Environmental Protection Agency Region 6 (EPA) would like to offer the following general and enclosed more detailed comments for the Arkansas Department of Environmental Quality's (ADEQ) consideration.

We are concerned that the supporting use attainability analysis (UAA) did not provide a clear basis to support the contention that only naturally occurring sources of TDS are causing the impairment in the Little River. We acknowledge that there are predominately natural and anthropogenic sources of minerals/TDS introduced into the Red River through Texas and Oklahoma as described in the supporting UAA. These sources are some 200 miles upstream of confluence of the Little River. Further, the UAA did not provide information on additional sources of minerals or sources of dilution in Arkansas that may influence conditions in the Red River. Although there may be some basis to support a revised TDS criterion below the confluence with the Little River to some point downstream, given that the designated fishery use is being attained in the Red River, a revised criterion from the confluence of the Little River to the Arkansas/Louisiana line does not appear to be clearly supported.

EPA appreciates the opportunity to comment on the proposed 3<sup>rd</sup> party proposal. If you have any questions, please contact me at (214) 665-6646.

Sincerely

Russell Nelson

Regional Standards Coordinator Watershed Management Section

Enclosure

#### **Technical Comments on:**

Exhibit F
Use Attainability Analysis
For Dissolved Minerals in Little and Red Rivers
Hempstead and Little River Counties, Arkansas (September 5, 2014)

Southwestern Electric Power Company, Inc, Dallas, TX NPDES No. AR0051I36

UAA Prepared by FTN Associates, Ltd., Little Rock, Arkansas

These comments are being provided to the Arkansas Department of Environmental Quality (ADEQ) for the document titled *Use Attainability Analysis for Dissolved Minerals in Little and Red Rivers, Hempstead and Little River Counties, Arkansas.* This use attainability analysis (UAA) is intended support the modification of Arkansas water quality standards (designated uses and water quality criteria), to remove the domestic water supply use in the Red River and establishment of site-specific criteria for total dissolved solids (TDS) in both the Red and Little Rivers, within the Gulf Coastal ecoregion.

#### **Technical Comments**

#### **Executive Summary**

#### ES.7 Proposed TDS Criteria Little River

# Page xi:

The proposed TDS criterion for the Little River below Millwood Lake is based on a weight-of-evidence approach utilizing a mass balance model. The document states that a mass balance was prepared that would reflect naturally occurring elevated TDS concentrations in the Little River. The document refers to TDS data collected upstream of SWEPCO by FTN and SWEPCO during October 2010 through October 2013 that showed that the current site-specific TDS criterion 100 mg/L criterion was exceeded approximately 10% of the time. The authors conclude that a TDS criterion of 136.1 mg/L would be necessary in the Little River to reflect the naturally occurring mineral regime and to allow SWEPCO to operate efficiently.

It's unclear what the basis for presuming that only naturally occurring sources of TDS result in the 10% exceedance rate. Later, the document refers to input from the Little River itself above Millwood Lake and from Mine Creek, noting that inputs from the latter are significantly higher per volume than other tributaries to Millwood Lake. The name Mine Creek alone suggests that this creek is a likely source of anthropogenic. Given the mass balance budget described in Table J.1 describes the background in the Little River as 98 mg/L and the SWEPCO discharge as contributing 1620 mg/L, it is unclear how a presumption that the inputs to the Little River are

Use Attainability Analysis for Dissolved Minerals In the Little and Red Rivers EPA Technical comments -September XX, 2015

anthropogenic. This tends to undermine the validity of the mass balance and any conclusions that may be drawn from it.

Differentiating between natural background and anthropogenic sources causing exceedances is important. As articulated in Tudor Davies 1997 policy memo, the water quality standards regulation at 40 CFR 131.11 (b)(1) requires States and authorized Tribes to adopt numeric water quality criteria that are based on section 304(a) criteria, section 304(a) criteria modified to reflect site-specific conditions, or other scientifically defensible methods.

In summarizing the aquatic life evaluation, the document indicates that the aquatic life composition of the current fish community below Millwood dam is a subset of what would be expected given available habitat. What is described here represent a snapshot of what is likely impacted and/or degraded conditions in the Little River. While document current conditions is an important, the objective of a UAA should be determining what the highest attainable use that can be attained. The objective of a UAA is to determine what the highest attainable use the waters of interest can support. 40 CFR 131.10(d) requires at a minimum, that uses be deemed attainable if they can be achieved by the imposition of effluent limits required under sections 301(b) and 306 of the Act and cost-effective and reasonable best management practices for nonpoint source control.

#### 1.0 Introduction

# 1.1 Background and Overview

#### Page 1-1:

The discussion here appears to be suggest that the (predominate) natural (and anthropogenic) mineral and TDS sources from TX and OK are not adequately accounted for and result in exceedances of AR's 500 mg/L criterion in the lower segment of the Red River below the confluence of the Little River to the AR-LA state line. Since there is certainly some dilution in the Red River as it moves to the confluence with the Little River and on downstream, it will be critical to provide clear support that these predominately natural upstream in TX and OK, and those in AR contribute to an inability to attain the current 500 mg/L criterion in the lower segment. The spatial inconsistence identified in and of themselves are not a basis for modifying criteria.

# Page 1-1 and 1-3:

In describing this permit as not typical, the authors are referring to ADEQ's use of the 500 mg/L TDS criterion that applies to the Red River as an "end of pipe" value rather than the 100 mg/L TDS criterion that applies to the Little River. Based on the fact sheet, it appears that ADEQ based 500 mg/L TDS criterion in the SWEPCO permit on the applicable criterion in the Red River to ensure that the discharge would not contribute to the impairment of the Red River because it is on the state's most recent EPA approved 303(d) list for exceedances of chloride, sulfate and TDS.

Region 6 permitting staff have suggested that if the 100 mg/L TDS criterion applicable to the Little River were applied, when dilution and mixing are considered, a limit of 3800 mg/L would

Use Attainability Analysis for Dissolved Minerals In the Little and Red Rivers EPA Technical comments –September XX, 2015

be possible and would be protective of the designated fishery use in both the Little River and the Red River.

# Page 1-4 Table 1.2:

The increase for both median and 95th percentile concentrations of TDS from the historical data the more recent data collected by SWEPCO suggest that the concentrations are the result of something other than "naturally occurring exceedences" as suggested in the next paragraph.

# Page 1-5:

The authors refer to little to no assimilative capacity for TDS in the Little River at times (absent the SWEPCO facility's discharge) as a result of "naturally occurring exceedances" of the current site-specific 100 mg/L TDS criterion occurring "upstream" of the SWEPCO discharge, refers to more in depth discussed in Sections 3.0 and 5.0. However, there is very little discussion of possible upstream naturally occurring" sources for the Little River in these sections. The only sources for the Little River discussed in the document to any degree are tributaries to Millwood Lake, particularly Mine Creek. Although Mine Creek is identified as the most significant contributor of minerals to Millwood Lake and downstream to the Little River, there is no information concerning possible mining impacts to that creek. As a result, the basis for attributing a lack of assimilative capacity and a 10% exceedance rate for the current 100 mg/L TDS criterion in the Little River to naturally occurring sources has not been established. What are these natural sources of high TDS - Mine Creek? And are they natural or anthropogenic?

# 2.0 Background

# 2.3 Existing Water Quality and Impairments

2.3.1 Little River

2.3.1.1 TDS Sources to the Little River

This section includes an evaluation of existing data from tributaries upstream of Millwood Lake to identify potential recent sources of TDS that might explain the increased TDS values in samples collected from the Little River downstream of the dam on Millwood Lake. The major tributaries of Millwood Lake include the Little River, Cossatot River, Saline River, and Mine Creek (Figure 1.1). Table 2.2 identifies the major tributaries to Millwood Lake, including the Cossatot River, Saline River and Mine Creek and summaries of their TDS contribution. Although Mine Creek only contributes 2% of the inflow, the TDS concentrations are significantly higher than the other streams.

There is some inconsistency between Tables 1.1 and 1.2 presenting USGS data from 1967-1995 and SWEPCO data from 2010-2013 (although it's not clear what sites the SWEPCO data came from) that show an increase in TDS concentrations over time. The time-series plots of TDS data from these ADEQ monitoring stations on the tributaries upstream of Millwood Lake identified in Figures 2.1 through 2.4 that show no clear evidence of significant changes in TDS levels in any of the streams, yet the authors report an increase in TDS concentration downstream of Millwood Lake.

#### 2.3.1.2 Water Quality Impairments for Minerals in the Little River

The authors indicate that ADEQ does not considers the Little River downstream of Millwood Lake to be impaired and have not included it on any 303(d) list. Further, the document notes that this stream is classified by ADEQ as "unassessed" due to a lack of data for water quality (including minerals). It is important to note that "unassessed" simply means that there is no data and does not necessarily mean that the Little River is either impaired or unimpaired. That lack of data may be why neither ADEQ nor EPA have included the Little River on the state's 303(d) list.

2.3.2 Red River

2.3.2.1 TDS Sources to the Red River in Arkansas

2.3.2.2 TDS Sources to the Upper Red River (Oklahoma, Texas)

#### Pages 2-5 to 2-8:

Much of the literature cited in the Executive Summary and in these sections is dated, relying on several sources from a time period when oil production was high with minimal environmental regulation. Despite those issues, it is reasonable to say that there are significant natural as well as anthropogenic sources of minerals in the Red River and its tributaries.

While the more current information tends to show that the predominant mineral sources to the Red River are likely natural, despite remediation efforts by the US Army Corps of Engineers (USACE), it is likely that brine disposal and intrusions from oil production into the Little Wichita and other sites are sources that affect the Red River in Texas. Oklahoma and contribute to the loading to a lesser extent in Arkansas. Sources of information for oil and gas production and remediation should be available from the Texas Railroad Commission and the Oklahoma Corporation Commission.

Not having, or possibly not being able to provide any information to support historical oil extraction impacts and their contribution to minerals/TDS concentrations significantly weakens the claim that the "overwhelming portion" historical of the TDS inputs to the system are from natural sources.

# 2.3.2.3 Water Quality Impairments for Minerals in the Red River

#### Page 2-9:

The authors note that upstream criteria for the Red River in Texas and Oklahoma reflect the elevated ambient mineral concentrations that are due to inputs from salt springs and seeps from the arid and semi-arid headwater regions. This is generally true, given salt springs are found as far west as the High Plains and Tableland Level II ecoregions. Although less predominate, there also anthropogenic and natural inputs from the Central Plains, Crosstimbers, Blackland Prairie and East and South Central Plains ecoregions.

It is important for Arkansas to recognize that an upstream state can establish uses and criteria in their waters as appropriate. 40 CFR 131.10(b) requires that those states take the downstream state's standards. Recent court decisions have affirmed EPA's ability to look at downstream waters to ensure protection.

This section also discusses assessment procedures used by states that share jurisdiction over portions of the Red River. Referring to Table 2.6, which compares criteria for minerals and impairments identified on state 303(d) lists, which shows that the Red River is not considered to be impaired upstream in Oklahoma and Texas or downstream in Louisiana. The authors state that this is partly due to different assessment procedures that are used in other states, suggesting that these states use less protective exceedance rates. As an example, the authors note that Louisiana does not consider a stream to be impaired for dissolved minerals unless 30% of the observed data exceed the water quality standard compared to Arkansas' 10% exceedance rate. The authors also state that Oklahoma and Texas also use less-stringent assessment procedures for dissolved minerals than Arkansas, and also state that the criteria in their water quality standards are less stringent than the criteria in Arkansas's standards. However, the authors have not explained how these approaches are any less protective than those used by Arkansas.

It is important to understand that simply comparison of exceedance percentages used in assessing use attainment does not tell the entire story. Assessing attainment includes defining the water quality indicators it measures and the procedures for analyzing and interpreting data in order to decide whether standards are met or water quality is impaired. This should include collection and analysis of multiple types of data providing information relevant to assessing attainment with approved WQS. As a result, a comparison of Louisiana's current use of a 30% exceedance rate to Arkansas 10% is not necessarily appropriate, although it should be noted that Region 6 is working with Louisiana to revise its approach. Oklahoma compares the mean of all values to a yearly mean standard, then looks for greater than 10% exceedances against a sample standard. In Texas, minerals criteria represent annual averages of all values that were collected and compared to the criterion for each parameter. Use attainment is based on the average concentration that applies to the entire length or area of the segment. For TDS, a value is calculated by multiplying specific conductance measured at the surface by a factor of 0.65. The chloride, sulfate, and TDS criteria are not supported if the average value exceeds the criteria.

The authors also state that Oklahoma and Texas water quality standards as less stringent than the criteria in Arkansas. Given the natural saline seeps and historical oil extraction activity along the Red River that this document discussed previously, higher criteria would be expected upstream in Texas and Oklahoma. The burden for SWEPCO is to show that predominately natural sources upstream in Oklahoma and Texas upstream as distant as Lake Texoma and/or that natural sources in Arkansas itself alter concentrations in the lower segment of the Red River to support the proposed criteria modifications.

# Page 2-10:

In a discussion of Table 2 describing applicable criteria and 303(d) listings, the authors refer to ADEQ's practice in recent years of making specific permitting decisions and calculations based on the state's most recent 303(d) list that has been approved by EPA. This approach means that ADEQ is applying the 500 mg/L TDS criterion that applies to the Red River below its confluence with the Little River as an end-of-pipe limit rather than the 100 mg/L criterion that applies to the Little River which is the receiving water for the Turk/SWEPCO discharge.

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Based on the NPDES permit fact sheet, applying the 100 mg/L TDS criterion that applies to the Little River, when dilution and mixing is considered, a limit of up to 3800 mg/l would be possible and still meet the in-stream criterion of 100 mg/L which would protect beneficial uses in the Little River and downstream. However, applying the 500 mg/L TDS criterion as an end-of-pipe limit results in a more restrictive permit limit. Without any information to the contrary, the effect of ADEQ's approach does not appears to have a significant environmental benefit for either the Little River or Red River, but appears to have the opposite effect. While applying 500 mg/L TDS criterion as an end-of-pipe limit may result in a small reduction in overall loads, the more restrictive limit reportedly requires the facility to operate at less than optimum efficiency. Thus ADEQ's approach appears to be the primary driver for Turk/SWEPCO to seek revised criteria for the Red River, which may be a less desirable environmental outcome.

# 2.4 Threatened and Endangered Species 2.4.1 Threatened and Endangered Aquatic Species

# Page 2-12:

This section of the document discusses several endangered species listed by the US Fish and Wildlife Service (USFWS) including the Ouachita rock pocketbook (Arkansia wheeleri), which is federally listed throughout its range in Arkansas and Oklahoma. The document reports that there have been no live A. wheeleri collected from just above or below the Turk/SWEPCO discharge. Their absence indicates that the Turk/SWEPCO discharge is having an effect on water quality and the presence of A. wheeleri. Although the physical characteristics and inputs to Millwood Lake may affect the ability of the Little River to support A. wheeleri, an important aspect of a UAA is to not only to identify current conditions. The document does not provide any discussion of whether minerals/TDS or other stressors may be affecting or limiting the presence of A. wheeleri in this portion of the Little River.

The authors also note that the USFWS recently listed the rabbits foot mussel (Quadrula cylindrica cylindrica) as threatened (Federal Register 2013). That species is known from some streams in southwest Arkansas, including the Little River upstream of Millwood Lake. This species not been documented in the Little River downstream of Millwood Lake or the Red River. The rabbits foot is likely limited by the physical characteristics Millwood Lake and its effect on the Little River, as well as the Turk/SWEPCO discharge.

# 3.0 FIELD SURVEY 3.1 Overview

#### Page 3-1:

Rather than relying on biological evaluations in a upstream versus downstream comparison from a point source discharge and a comparison of communities in reference and receiving streams, the approach for field surveys is attempting to show that characteristics in Millwood Lake and at the Denison dam dominate the mineral regime in the Little and Red Rivers rather than the Turk/SWEPCO discharge.

There are sources of minerals above Millwood dam, primarily the Little River and Mine Creek upstream of Millwood Lake and there are certainly significant natural and anthropogenic sources of mineral in the upper reaches of the Red River in Texas and Oklahoma. The burden for Arkansas is to clearly show that the sources above Millwood dam are at least predominantly anthropogenic. Similarly, Arkansas must be able to make a reasonable showing that the predominately natural and some anthropogenic sources some 200 miles upstream have a significant effect on water quality in the Red River as far downstream as its confluence of the Little River and show that there are no natural or anthropogenic sources from the TX-AR line to the confluence.

If the Turk/SWEPCO discharge is not contributing to the potential exceedances in the Little River or potentially impacting the Red River, the important question as to whether the facility is seeking to revise the criteria in the Little River and the Red River because the criteria are inappropriate or because of ADEQ's policy decision to applying a criterion as an end-of-pipe limit rather than basing it on applicable criteria considering dilution and mixing in the receiving water.

# 3.4.1.2 Macroinvertebrate Data Analysis

# Page 3-10:

In discussions of field observations the authors indicated an attempt was made to assess the susceptibility of the existing benthic macro invertebrate community to a slight increase in TDS. Tolerance indicator values (TIVs) for specific conductance for various invertebrate taxa provided in Carlisle et al. 2007 were reviewed. However, the authors note that too few of the taxa listed in Carlisle et al. 2007 were present in the data set for this study to allow for a meaningful evaluation of ion tolerance. Accordingly, the benthic data presented herein provide a description of the presently attained fisheries use and a baseline for future studies.

It should be noted that small increases in TDS alone may have minimal effect on the existing benthic community however. TDS is rarely seen independent of other stressors. In a quick search, a number of sources were identified that discuss the effect TDS has on Appalachian streams. For example, Timpano et al. (2010), in discussing the effects of TDS in coalfield streams. As a way of isolating the effects, Timpano et al. used headwater streams to represent a gradient where confounding factors could be minimized. This and other similar papers done in Appalachian could have been used to inform this aspect of the analysis. One of the problems with the approach take here is a lack of reference stream comparisons.

#### 3.4.2.2 Fish Data Analysis

#### Page 3-11:

Fish data were first assessed to determine if there were impacts due to the Turk/SWEPCO discharge "under current conditions" by evaluating differences in total species richness, relative abundance, diversity, and similarity measures between upstream and downstream reaches.

Although unclear, it appears that "under current conditions" likely means under limits specified in the current NPDES, which as noted earlier, are based on a monthly average of 500 mg/l TDS

applying Red River Criteria end-of-pipe. This end of pipe value equates to an instream waste concentration of 43 mg/L. As noted earlier, if ADEQ were to base the Turk/SWEPCO permit on the Little River instream standard of 100 mg/L, once dilution and mixing is allowed, this would result in an allowable end-of-pipe discharge of up to 3800 mg/l to still meet the instream waste concentration of 100 mg/L. These conditions should be modeled and presented as a comparison rather than solely those based on ADEQ's current approach of basing permits on past 303(d) listings. Without this, there is some question to the validity to the findings here and in other aspects of this proposal.

#### 3.4.4 Discussion and Conclusions

# Page 3-22:

The authors note that the evaluation of the fish community demonstrated similar fish communities upstream and downstream of the Turk/SWEPCO discharge in the Little River. The combined collections from their study and USGS sampling during 2009 and 2013 provide evidence that the designated fisheries use in the Little River is currently attained under the existing mineral conditions.

Further, the document reports that an evaluation of the mineral tolerances of the existing fish community indicate a fish community that can be expected to be relatively insensitive to moderate changes in TDS (as indicated by specific conductance). The UAA notes that fish species that were classified as intolerant to TDS or for which data in Meador and Carlisle (2007) were lacking can be shown to be present in the Red River, which has substantially higher TDS than will be experienced in the Little River due to the operation of the Turk/SWEPCO plant. The conclusion drawn is that the anticipated increase in TDS due to the operation of the Turk/SWEPCO plant will support the designated fisheries use in the Little River downstream of the Millwood Lake dam.

The question these conclusion raise is even if the designated fisheries use in the Little River would potentially be maintained with the proposed increase in TDS criterion to 138 mg/L, why is it necessary if that use is being attained at the current 100 mg/L criterion? The answer seems to go back to Turk/SWEPCO's desire to operate at maximum efficiency. Under current conditions, the facilities efficiency appears to only be affected by ADEQ's approach in using limits based on the 500 mg/L criterion for the Red River as an end-of-pipe value. As noted previously, given that the Turk/SWEPCO discharge appears to have a negligible effect on the Red River, if ADEQ were to use existing criteria and allow for mixing and dilution in the Little River, there would be no need for this proposal.

#### 3.5 Fisheries Evaluation in the Red River

#### Page 3-24:

The objective of this evaluation was to determine whether the existing mineral regime supports the designated fisheries use as indicated by fish populations. The approach relied on the historical record of mineral concentrations in the Red River from ADEQ monitoring stations RED0025, RED0046, and RED0045 and the use of a sampling survey and historical data review

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of fish populations in the Arkansas portion of the Red River conducted by Buchanan et al. (2003). The intent was to characterize the existing fish communities and evaluate whether those existing communities are supported by the existing mineral regime, thus indicating attainment of the fisheries designated use.

Although the Buchanan et al. (2003) data suggest that mineral conditions during 1995 and 2001 supported a high fisheries use attainment, it is somewhat problematic to rely on data that represents a single snapshot in time from 14 years ago. The authors draw the conclusion that since the Red River was meeting designated uses during the Buchanan et al. study and that because TDS data from the post-study period (2002 through 2013) indicate that mineral concentrations where generally higher during the time frame of the Buchanan et al. study, that fisheries beneficial uses are likely currently being attained. While the post-study period (2002 through 2013) TDS data gives an indication that uses are currently being attained, it does not provide the same level of confidence as current field data would. Showing that broadly written designated aquatic life uses can be supported under the existing mineral regime isn't necessarily a high bar given how broadly beneficial use definitions tend to be written. Determining attainment tends to depend on the individual state's assessment methodologies rather than the use definition.

3.6 Biological Evaluation: Summary and Conclusions 3.6.1 Little River 3.6.2 Red River

#### Page 3-25:

The evaluation of the biological data demonstrated similar fish and communities upstream and downstream of the Turk/SWEPCO facility. The authors conclude that these finding reported suggest that the designated fisheries use is being attained that that the projected mineral concentration - presumably the proposed 138 mg/L TDS criterion will not be adversely affect attainment of the fisheries use.

Although the data indicate that the designated fisheries use is currently being attained, the document does not provide compelling information to support the contention that the proposed 138 mg/L TDS criterion will not adversely affect attainment. Further, concerns with impacts to benthic macroinvertebrate communities downstream of the discharge at the current criterion have been identified in previous comments.

#### 4.0 TOXICITY ANALYSIS

# 4.1 Evaluation of Routine WET Testing

#### Page 4-1:

The recurring concern here is that the conditions simulated for toxicity analysis are reflective of current permit conditions, not what would likely be the conditions if the Red River was not listed.

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More specifically, section 4.1 reports that a review of NPDES DMRs from what is likely the current permit (from November 1, 2012) indicate no episodes of sub-lethal toxicity for *C. dubia* and *P. promaleas*. Since the current permit is based on 500 mg/L as an end-of-pipe limit, sub-lethal effects would not be expected. The question is, how *C.* dubia and *P. promaleas* would respond under conditions that would be expected if a limit of up to 3800 mg/L was in place for the facility as it may be once the current 303(d) listing in the Red River is resolved.

Relying on DMRs that are not reflective of what conditions will likely be long-term in the Little River and downstream in the Red River could be considered misleading.

#### 4.2.3 Conclusions

#### Page 4-5:

The authors state that calculated IC25 values (1,744 mg/L for TDS; 232 mg/L for sulfate; and 600 mg/L for chloride) indicate that toxicity due to minerals in WET testing would not be expected at the critical dilution (26%) until effluent TDS values reached approximately 6,976 mg/L. And that these results indicate that the threshold for toxicity due to minerals (as TDS, sulfate, and chloride) is well above anticipated mineral concentrations at the critical dilution of 26%. Accordingly, their conclusion is that there is a large margin of safety associated with mineral discharges from the plant with respect to compliance with NPDES WET testing.

It is unclear what the basis for the statement that toxicity due to minerals would not be expected until TDS values reach approximately 6,976 mg/L. Although the IC25 may be considered functionally equivalent to a NOEC by definition, it represents a 25% reduction in the rate of survival, growth or reproduction among test organisms. Further, Table 4.2 indicates that average concentrations of 6,000 mg/L cause 100% lethality.

# 5.0 MASS BALANCE MODELING 5.2 Critical Flow Conditions

#### Page 5-1:

Instream TDS concentrations in the Red River were modeled for two critical flow conditions. The authors note that calculations and model results for 7Q10 flow conditions were used to develop proposed criteria for the Little River, and for the Red River between the Little River and the AR-LA state line. The results for harmonic mean flow conditions were used to confirm that the proposed criteria in Arkansas will still allow the Louisiana criterion for TDS in the Red River (780 mg/L) to be maintained.

It's important that ADEQ be aware that the State of Louisiana is currently reviewing its mineral criteria. It is EPA's understanding that water quality data taken in the Red River near Shreveport, between 1991 and 2014 indicate that TDS concentrations of 780 mg/L are just above the 90th percentile. This may mean that it if the Arkansas' proposed criteria are adopted, there is a significant likelihood of TDS impairments in the downstream segment of the Red River in Louisiana.

It is important to note that 40 CFR 131.10(b) requires states to consider the water quality standards of downstream states when revising their criteria.

#### 5.5 Model Results for 7Q10 Conditions

#### Page 5-7:

Neither the RED0046 data nor the RED0009 data were considered to be a good representation of well-mixed concentrations in the Red River by the authors. Both the RED0046 station and the RED0009 station are located where the Red River does not appear to be laterally well-mixed because of significant tributary inflows a short distance upstream of each station.

The discussion explains that because of the location of the sampling sites for both RED0046 and RED0009, the data are likely measuring a disproportionate percentage of tributary water that has not mixed completely with the entire flow of the Red River. The authors contend that both tributaries (especially the Little River) contribute water that typically has TDS concentrations much lower than concentrations in the Red River. Because these two stations appear to be underestimating the TDS concentrations in the Red River and should not be used for comparison with predicted TDS values from the mass balance modeling, presumably because their inclusion could result in an underestimation the TDS concentrations in the Red River as compared to predicted TDS values from the mass balance modeling.

While it is possible that the inflow from the Little River at (RED0046) and the Sulphur River at (RED0009) may not be well-mixed, the problem with leaving them out is that it will likely mean an underestimation of TDS concentrations to the Red River, particularly in the lower reach near the LA state line.

It is unclear how the segment reaches in Table 5.4 compare spatially to the long-term sampling sites.

#### 5.6 Model Results for Harmonic Mean Conditions

### Page 9:

The authors explain that the reason for simulating harmonic mean conditions was to confirm that proposed TDS criteria for the Red River in Arkansas would allow the Louisiana criterion for TDS in the Red River to be maintained under critical conditions. The harmonic mean simulation presented predicts that the TDS concentration at the AR-LA state line will be 661 mg/L, which is well below the Louisiana criterion for TDS in the Red River (780 mg/L). The conclusion is that the proposed criteria based on the mass balance modeling will be protective of the Louisiana criterion for TDS.

However, the harmonic mean simulation used the same input TDS concentrations as the 7Q10 simulation. As noted in the previous section, the exclusion of inflow from the Little River at (RED0046) and the Sulphur River at (RED0009) my mean an underestimation of TDS concentrations in the Red River, resulting in potential impairment of the current 780 mg/L TDS criterion that applies at the LA state line. As noted in comments on section 5.5, it is EPA's understanding that water quality data taken in the Red River near Shreveport, between 1991 and

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2014 indicate that TDS concentrations of 780 mg/L are just above the 90th percentile. The 780 mg/L value is likely to go down.

It would be advisable to consider what potential effect lower criteria in the Red River at the AR/LA state line may have on the predicted TDS criterion of 661 mg/L if LA revises its criteria down.

# 6.0 DESIGNATED USES 6.2 Red River 6.2.2 Domestic Water Supply

#### Page 6-4:

The domestic water supply use for the reach of the Red River between the OK-AR state line and the Little River confluence has previously been removed. The authors provide data that indicate that the reach of the Red River from the Little River confluence to the AR-LA state line can also be expected to exceed secondary drinking water standards due to the same factors. Figure 6.1 shows that the Red River TDS concentrations at Fulton (downstream of the mouth of the Little River) frequently exceeded the secondary drinking water standard of 500 mg/L during the period of record. The authors also noted that both the Arkansas Department of Health and the Arkansas Natural Resource Commission to confirm that the stream is not used as an existing domestic water supply. Their conclusion is that the domestic water supply use is neither an existing nor attainable use in the Red River downstream of the mouth of the Little River.

Although it is within Arkansas' authority to remove the domestic water supply use for the segment of the Red River below the confluence with the Little River to the LA state line, AR must consider the downstream state standards and ensure that its action would not affect domestic water supply/drinking water uses in Louisiana. See 40 CFR 131.10(b).

#### 7.0 ALTERNATIVES EVALUATION

#### Page 7-1:

Arkansas' UAA guidance and CPP require that a petition to increase dissolved minerals concentrations above existing conditions must include an evaluation of alternatives to the direct discharge of the water. Four alternatives that have been reviewed including:

- 1. Distillation treatment.
- 2. Treatment using a constructed wetland,
- 3. Pumping the wastewater to a larger stream that holds the potential for dilution of the minerals, and
- 4. Reverse osmosis (RO) treatment of the wastewater.

From the outset, the use of constructed wetlands was dismissed as an option for this facility. The authors based this exclusion on the argument that constructed wetlands can only be used to reduce sulfate, which results in the production of bicarbonate in place of sulfate (Hedin et al.

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1989). The authors do note that a constructed wetland could, in principle, reduce sulfate in the discharge from this facility, the resulting TDS concentration would not be decreased (due to the replacement of the sulfate ions with bicarbonate ions), but argue that no net benefit would be obtained, stressing that TDS is the only concern for this evaluation.

The premise in rejecting a constructed wetland appears to be based on the premise that a constructed wetlands would only reduce sulfate to bicarbonate, which would not provide a net reduction in TDS. Looking at the Hedin et al. paper, it is specific to the treatment of acid mine drainage, not the type of discharge that would result from blow down cooling water from a power plant. The paper does indicate that with an increase in sulfate reduction, bicarbonate alkalinity will be produced and pH will increase, but it does not say that sulfate is directly reduced to bicarbonate. In treatment of acid mine drainage, sulfate would primarily be reduced to sulfite and tend to react with available metal to form sulfide precipitates. The Hedin paper refers to bacterial production of bicarbonate formation, particularly in anoxic limestone drains, but does not refer to the reduction of sulfate to bicarbonate. Which is not unexpected since as noted earlier, the paper is looking at treatment of acid mine drainage, not sulfate or other TDS constituents in particular. Given this, EPA considers a constructed wetland to be a viable and the preferred alternative to altering the applicable TDS criterion.

The authors contend that there are two alternatives for achieving compliance with the existing TDS criteria for the Little River and Red River:

- 1. RO treatment to remove or reduce dissolved minerals, and
- 2. Pumping the effluent to a larger river that holds the potential for dilution of the minerals.

In considering these options, the authors state that there is not a river in the vicinity that could be used as dilution to completely avoid a change in the water quality standards. Although the Red River has sufficient volume, it has a TDS criterion of 500 mg/L, the expected concentration of TDS in the effluent from this facility would not meet the Red River TDS criterion and would still require the completion of a UAA or the installation of partial RO treatment.

EPA considers ADEQ's use of TDS criteria applicable to the Red River, particularly the way the criterion has been applied as the emerging central element here. As noted in previous comments, ADEQ issued permit Turk/SWEPCO is operating under is based on the existing 500 mg/L TDS criterion for the Red River as end-of-pipe limits. The permit carries those limits because the receiving Red River segment was on the state's 2008 303(d) list. From a numeric perspective, ADEQ has issued the permit with a monthly average of 500 mg/l TDS end-of-pipe. This end-of-pipe value equates to an instream waste concentration of 43 mg/L. If ADEQ based the permit on the Little River instream standard of 100 mg/L, once dilution and mixing is allowed, the result is an allowable end-of-pipe discharge of up to 3800 mg/L TDS. While the end-of-pipe concentration is higher, this approach would meet the instream waste concentration based on the ecoregion-based 100 mg/L TDS while allowing the Turk/SWEPCO facility to operate efficiently.

EPA strongly recommends that rather than revise TDS criteria particularly for the Little River, in what appears to be an effort to avoid this permitting scenario, that this approach be considered.

# 7.1.2.2 Option 2: RO Treatment/Discharge to Little River at Future Effluent Limits

# Page 7-9:

The authors anticipate that the TDS criterion in the Little River, and thus the effluent limitations in future NPDES permits for the SWEPCO plant, could be lowered to 100 mg/L. The conclusion drawn is that this would require that all of the effluent be treated through an RO system if it continues to be discharged to the Little River, in order to achieve an effluent limit of 100 mg/L For TDS.

However, as noted previously, the 100 mg/L criterion that applies to the Little River is not the same as a derived NPDES permit limit. The permit limit that would be derived from the 100 mg/L TDS criterion once dilution and mixing is allowed, would likely be an allowable end-of-pipe discharge of up to 3800 mg/L. Given that this concentration would allow the facility to meet the 100 mg/L TDS criterion in the Little River, there would be no need to consider RO.

# 7.3 Summary of Costs

# Page 7-10:

The authors summarize the estimated costs of various treatment scenarios compared to the cost of direct discharge based on the proposed site-specific criteria in Table 7.1. The cost described here appear to be the potential cost associated with RO treatment of the entire volume of the Turk/SWEPCO discharge. It is unclear what volume of the discharge would need to be treated.

# Technical Comments on:

EXHIBIT G
TECHNICAL JUSTIFICATION FOR A SITE-SPECIFIC TEMPERATURE
CRITERION IN THE LITTLE RIVER, HEMPSTEAD & LITTLE RIVER COUNTIES,
ARKANSAS

#### **Technical Comments**

# 1.0 INTRODUCTION1.1 Background & Overview

# Page 1-1:

This document states that the Turk/SWEPCO facility's National Pollutant Discharge Elimination System (NPDES) permit (No. AR0051136) includes a daily maximum temperature permit limit of 35 °C (95 °F). Given that Arkansas Reg. 2 specifies and the authors acknowledge that the applicable temperature limits for streams is 30°C (86°F), the basis for the maximum temperature limit of 35°C (95°F) in SWEPCO's current permit is unclear.

The document also notes that the Little River downstream of Millwood Lake has a lower temperature criterion than Millwood Lake and the Red River immediately downstream of the Little River. Both Millwood Lake and the Red River in Arkansas have a temperature criterion of 32 °C (89.6 °F) (APCEC 2011). Figure 1.2 is presented to illustrate the spatial inconsistency of the temperature criteria.

The document notes that Millwood Lake has a temperature criterion of 32°C (89.6°F), the Little River, downstream of Millwood Lake has a 30°C (86°F), and the Red River immediately downstream of the Little River has a temperature criterion of 32°C (89.6°F) (APCEC 2011) to illustrate the spatial inconsistency of the temperature criteria downstream of Millwood Lake. Spatial inconsistency has no bearing on toxic or conventional criteria that are protective of aquatic life uses (ALU) and is not a basis for altering applicable criteria. It is not unusual for temperatures in a reservoir to be higher than upstream and typically downstream lotic waters given the retention time and particularly the shallow depth of Millwood Lake.

Although we acknowledge that it may be difficult to manage the flow through a lake as shallow as Millwood given its design, relatively high retention time and higher tailwater race temperatures, but it's unclear how that would affect the entire length of the Little River or why the riparian cover downstream along the Little River could not be enhanced or restored.

#### Page 1-3:

The authors note that if the temperature criterion is not changed and the impairment status continues, the facility's NPDES permit limit for temperature at Outfall 001 would be reduced to the Little River instream criterion of 30°C (86°F) without the usual allowance for a mixing zone (i.e., using typical end-of-pipe calculations to develop permit limits for a thermal discharge). As such, the authors note that facility's operation would be adversely impacted by a temperature impairment designation in the Little River that may limit or prevent discharges during warm weather and reduce the efficiency of the facility's operation during other times of the year.

Does this mean that the decision was keeping the existing criteria in place versus operating efficiency (i.e., cost)? If so, the basis for the proposed amendment 40 CFR 131.10(g)(6), which requires an economic analysis to determine if SWEPCO can support upgrades to its process/facilities w/out substantial and widespread impacts.

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# Page 1-4:

The authors indicate that adjusting the temperature criterion for the Little River downstream of Millwood Lake is necessary to reflect current ambient conditions during the critical summer conditions and would result in NPDES temperature limitations that are appropriate and protective of the designated uses of the Little River. An important question is if current ambient conditions reflect the highest attainable condition given the requirements outlined in 40 CFR 131.10?

# 2.0 BACKGROUND 2.1 NPDES Permit

## Page 2-1:

The document again refers to the Turk/SWEPCO discharge to the Little River under NPDES Permit No. AR0051136 and the effluent limitation for temperature of 35°C (95°F). As noted previously, it's unclear how this facility can have an NPDES permit that allows a 35°C (95°F) when the applicable temperature for streams is The 32°C (89.6°F). Is there a specific implementation procedure or mixing zone allowance?

#### 2.2 Applicable Water Quality Standards

# Page 2-2:

The authors suggest that the temperature regime of the Little River downstream of Millwood Lake has been altered as a result of physical hydrologic alteration of the system caused by the construction of Millwood Lake. The authors contend that Millwood Lake and its confluence with the Little River has a typical width between 200 and 400 feet with only a small portion of the channel being shaded by trees along the banks.

Based on images of the USACE dam, the Little River may be "200-400" wide" immediately below the dam as stated here, but appears to constrict below River Run Park and appears to be generally consistent with the average stream of reference streams in the Gulf Coastal Plain ecoregion. Without supporting information for multiple downstream locations on the Little River below the Millwood, this claim does not appear to be supported. Photographs, specifically aerial photos should be included in this proposal to support this claim.

# 2.3 Water Temperature Monitoring

#### Page 2-4:

Tables 2.4 and 2.5 presents summary statistics for water temperature data collected by SWEPCO from April to October 2012. These tables indicate that the Turk/SWEPCO discharge increases instream temperatures. With the exception of a higher maximum temperature reading upstream of SWEPCO, which may be a single event, the 95th percentile, percent daily max  $> 30^{\circ}$ C and percent daily max  $> 32^{\circ}$ C indicate that the facility adversely impacts temperature in the Little River.

#### Page 2-8:

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The authors note that temperatures in the Little River during 2013 (Figures 2.3 and 2.4) appear very similar to those measured in Millwood Lake near the dam (Figure 2.5). They note that exceedances of the Little River temperature criterion occur at all of the Little River sampling locations, both upstream and downstream of the SWEPCO outfall, during 2012 and 2013 (Figures 2.2 through 2.4).

These figures generally show that there is an increase in temperature associated with the time of the year sampled. There is a clear correlation with ambient air temperature and water temperature, which among other factors can have a significant influence instream temperatures. This correlation is not considered here. The comparison to temperatures in Millwood Lake near the dam are of limited value since this lake is relatively shallow and retention times tend to be fairly high in most lakes.

In addition, the data reported in Fig. 2.8 which show a decrease in temperature from LR0 to LR00 does not appear to be consistent with the summary statistics presented for the same site in Table 2.4, which reports an increase of all and daily maximum data  $> 30^{\circ}$ C.

# Page 2-10:

Warming in a large shallow impoundment like Millwood is expected as it is in the immediate tail water area. However, it's unclear if this narrative is suggesting that temperatures in the Little River are the result of the Turk/SWEPCO discharge or continuing the claim that the river remains wide and unshaded further downstream. As noted earlier, a simple internet search produced maps and photographs that suggest that this is not the case downstream.

# 2.4 Evaluation of Little River Water Temperatures Upstream of SWEPCO Outfall 001

#### Page 2-10:

The discussion indicates that Table 2.5 provides an inventory of water temperature data collected in the Little River downstream of Millwood Lake and upstream of SWEPCO Outfall 001 that were used to evaluate the temperature criterion for the Little River. However, Table 2.5 also includes data from LR00, which is reported as 1 mile below SWEPCO Outfall 001.

#### Page 2-15:

A comparison of temperature statistics between the two FTN monitoring locations near the dam below Millwood Lake and the two locations farther downstream (near SWEPCO) indicate that that locations farther downstream have slightly higher temperatures during July through August and more exceedances of the 30  $^{\circ}$ C (86  $^{\circ}$ F) criterion (Tables 2.3 and 2.4 ).

This paragraph touches on the potential source affecting ambient temperatures in Millwood Lake and the Little River, but provide no details. The summary data, statistics and statements presented here do not appear to have considered that air temperature is highly correlated with the significant positive heat flux components including solar radiation and with stream water temperature. The correlation is supported by the USGS' Stream Segment Temperature Model (SSTEMP), which asserts that air temperature will usually be the single most important factor in determining mean daily water temperature.

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The authors also note that ADEQ's (303(d)) assessment for 2014 was conducted using the SWEPCO 2012 and 2013 data and FTN 2013 data from this stream reach and that ADEQ's draft 2014 303(d) list includes a temperature impairment for the Little River downstream of Millwood Lake. Did SWEPCO and FTN use ADEQ monitoring stations to collect these data? Is there information to support the reported listing? While the assessment methodology used to determine impairment appears to be consistent with ADEQ methodology, it is unclear if 3rd party (SWEPCO and FTN) data be was used by ADEQ to support a 303(d) listing?

In instances where the temperature exceeds the criteria, it would likely result in a reduction in overall species richness and/or abundance because of excessive heat. Discussions of the macroinvertebrate and fish communities do not appear to show such effects.

# 2.5 Threatened and Endangered Species 2.5.1 Threatened and Endangered Aquatic Species

#### Page 2-18:

The document reports that there have been no live *A. wheeleri* collected from just above or below the Turk/SWEPCO discharge. Their absence indicates that the Turk/SWEPCO discharge is having an effect on water quality and the presence of *A. wheeleri*. Although the physical characteristics and inputs to Millwood Lake may affect the ability of the Little River to support *A. wheeleri*, an important aspect of a UAA is to not only to identify current conditions. The document does not provide any discussion of whether temperature or other stressors may be affecting or limiting the presence of *A. wheeleri* in this portion of the Little River.

Although the rabbitsfoot mussel (Quadrula cylindrica cylindrica) is known to occur in some streams in southwest Arkansas, including the Little River upstream of Millwood Lake, it has not been documented in the Little River downstream of the Lake to the Red River. The rabbitsfoot is likely limited by the physical characteristics Millwood Lake and its effect on the Little River, as well as the Turk/SWEPCO discharge.

#### 3.0 BIOLOGICAL EVALUATION

3.1 Overview

3.4.3.2 Fish

Ecoregion Key and Indicator Species

The authors describe presence and absence of ecoregion key and indicator species per APCEC (20 11) which is summarized in Table 3.7. Of three key and indicator species expected to be present based on habitat preferences, one species was present in the pooled list. One species not expected to be present (redfin shiner) was present in the pooled list. The reported data is not specific as to site - is this a comparison of upstream/downstream of the SWEPCO discharge? The location of this sampling would be useful, as it would give some indication of whether the presence of only 25% of key and indicator species is the result of a generally degraded/impacted system, resulting from hydromodification and high ambient air temperatures driving water temperature, or exacerbated by temperature inputs from the SWEPCO facility.

# 3.4.4 Discussion and Conclusions

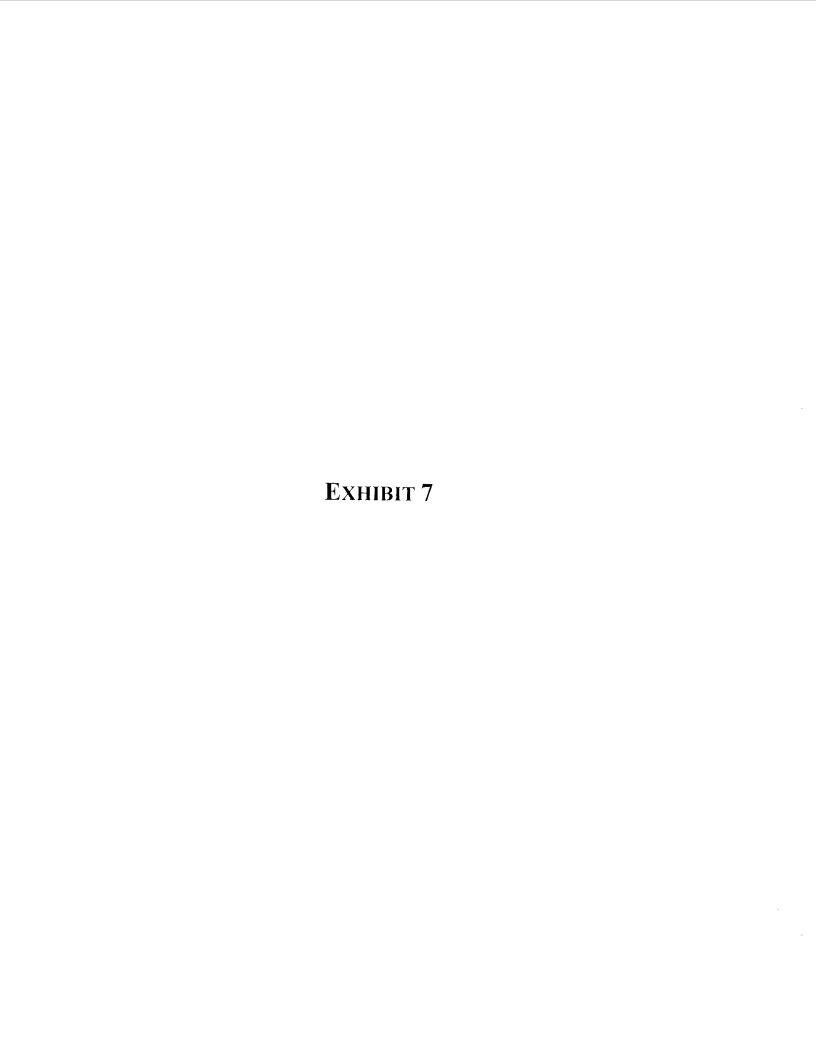
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# Page 3-22:

The authors observed that the strongly similar fish and benthic communities upstream and downstream of the discharge. The suggestion is that this finding indicates that the current operation of the discharge has negligible effect on the existing biological communities. Although relative abundance upstream/downstream showed reasonably good comparison at around 71%, the Meador and Carlisle (2007) data only showed 61% of the expected species are present and only 25% of the expected species for the Little River present in the pooled sample. It is unclear if this is the result of a degraded system or that the discharge has more than a negligible effect.

# Page 3-23:

The discussion notes that ecoregion key and indicator species are generally not expected to be present in the reach of the Little River due to the habitat preferences of those species and that the low numbers of key and indicator species present in the pooled list (two species) is a reflection of this expectation. This statement appears to contradict Table 3.7, comparing expected key/indicator species in the Little River to what was present in the pooled list (25%).





#### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

#### REGION 6 1445 ROSS AVENUE, SUITE 1200 DALLAS TX 75202-2733

October 22, 2015

Sarah Clem ADEQ Branch Manager Arkansas Department of Environmental Quality 5301 Northshore Drive Little Rock, AR 72118-5317

Re: Environmental Protection Agency comments on the Proposed 3<sup>rd</sup> party rule by Domtar A. W. LLC (DOMTAR)

Dear Ms. Clem:

The Arkansas Pollution Control and Ecology Commission (APC&EC) held a public hearing at Ashdown, AR on January 26, 2015, to receive comments on a third-party proposal by Domtar A.W. LLC (DOMTAR) to change APC&EC Regulation 2, the Arkansas Water Quality Standards, through a "technical adjustment," for the Red River from the Arkansas-Oklahoma state line to the Arkansas-Louisiana state line. The proposed amendments to Regulation 2 include:

- Modification of the Total Dissolved Solids (TDS) water quality criterion for the Red River from the Arkansas-Oklahoma state line to mouth of the Little River, from 850 milligrams per liter (mg/L) to 940 mg/L, and the sulfate (SO<sub>4</sub>) criterion from 200 mg/L to 250 mg/L; and
- Modification of the sulfate criterion for the Red River from the mouth of the Little River to the Arkansas-Louisiana state line from 200 mg/L to 225 mg/L.

The Environmental Protection Agency Region 6 would like to offer the following comments for the Arkansas Department of Environmental Quality's (ADEQ) consideration.

We are concerned that there is limited information supporting the proposed 3<sup>rd</sup> party rule. DOMTAR provided a "Summary Rationale" (Rationale) that indicates that the mineral concentrations in the Red River in Arkansas are primarily the result of naturally occurring sources in upstream states. We acknowledge that there are predominately natural as well as anthropogenic sources of minerals/TDS introduced into the Red River through Texas and Oklahoma. Although the proposal did not provide any information describing TDS or mineral levels, sources of minerals, or dilution in Arkansas that support the proposed TDS or sulfate criteria for the Red River, it does refer to a study initiated by Southwestern Electric Power Company John W. Turk, Jr. Power Plant (Turk/SWEPCO). Given that the DOMTAR proposal is specific to a revised TDS and sulfate criterion in the upper segment of the Red River and sulfate in the lower segment, whereas the SWEPCO proposal is specific to TDS criteria in the Little River and lower Red River, the study alone cannot be used to support the revisions proposed by

DOMTAR. It is worth noting that in referring to SWEPCO study, the Rationale notes that the Aquatic Life Designated Use is being attained within the Little and Red Rivers.

The DOMTAR proposal includes spiked toxicity test suggests there is no acute or chronic toxicity due to dissolved minerals levels. These tests relied on calculated ion concentrations of inorganic salts to mimic the ionic strength and makeup based on a single river sample. This test only provide an estimate of the toxic threshold for dissolved minerals for a single test species (*Ceriodaphnia dubia*) at a single point in the Red River. Similarly, we are concerned that the experimental design assumes that concentrations of TDS in the river, (i.e., ionic components S0<sub>4</sub>-2, Cl<sup>-</sup>, Na<sup>+</sup>, Mg<sup>+2</sup>, and Ca<sup>+2</sup>) do not vary downstream of the original sample site through the entire upper segment of the Red River. This approach does not appear to consider actual conditions where minor changes in ionic concentration in mineral salts and conductivity can have significant effects on aquatic species. It is unclear how a single sample location would be indicative of conditions throughout the segment of the Red River through Arkansas which would be covered by the proposed criteria modifications. Rather than establishing criteria to protect aquatic life based on an actual bioassessment of the Red River in Arkansas, the approach appears to be using spiked toxicity testing that should be used for permitting decisions to support proposed aquatic life criteria.

The proposal also cites Fishes of the Red River in Arkansas (Buchanan et al. 2003). Buchanan et al. (2003) demonstrates the designated fisheries use was being attained at the time of the survey (1995-2001) and suggests that fish species were more abundant and robust than either the Texas or Louisiana reaches of the Red River. For example, Buchanan et al. (2003) noted that two species have only been reported from the Arkansas segment, eleven species found in Arkansas have not been reported from the upstream segment, and seven species have not been reported from downstream segment in Louisiana. The study also reported that the Arkansas segment had 68% of the species reported from the upstream reaches, and 61% of those from the downstream segment.

Given that fish species are more abundant in the Arkansas reach, which has the lowest TDS and sulfate criteria, the limited information provided does not clearly show that the proposed criteria will not result in adverse effects on aquatic life. Although a Use Attainability Analysis (UAA) as defined in 40 CFR 131.3(f) is not required here, 40 CFR 131.6 requires that adequate information be provided to support the proposed revisions. We recommend that a bioassessment be developed to determine if the proposed criteria will be protective of aquatic life and designated uses in the Red River.

EPA appreciates the opportunity to comment on the proposed 3<sup>rd</sup> party proposal. If you have any questions, please contact me at (214) 665-6646.

Sincerely,
Russell Nelson

Regional Standards Coordinator Watershed Management Section



#### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

REGION 6 1445 ROSS AVENUE, SUITE 1200 DALLAS, TEXAS 75202 – 2733

FEB - 9 2016

Mr. Caleb Osborne
Associate Director – Water Division
Arkansas Department of Environmental Quality
5301 Northshore Drive
Little Rock, AR 72118-5317

Re: Final submittal of Regulation No. 2, as amended, Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas

Dear Mr. Osborne:

The Environmental Protection Agency (EPA) has completed its review of the amendments to Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas that were made in relation to the Third Party Rulemaking process initiated by Southwestern Electric Power Company (SWEPCO). The amended Regulation No. 2 was adopted by the Arkansas Pollution Control and Ecology Commission on October 23, 2015 and became effective on November 5, 2015. It was submitted to the EPA for approval on December 21, 2015.

At this time EPA is taking no action on the submitted amendment to Regulation No. 2 as the information provided in the submission was not sufficient to determine that the site specific criteria changes are protective of the aquatic life use in the Little River and the Red River. In addition, insufficient information was provided to demonstrate that the site specific criteria changes are protective of downstream uses in Louisiana.

A list of recommendations of additional information that can be provided to support these site specific criteria changes has been provided in the attached enclosure. EPA would be happy to meet with the Arkansas Department of the Environmental Quality, SWEPCO, and their consultant, FTN, at a technical level to discuss these recommendations in detail. If you have any questions or concerns, please contact me at 214-665-3187, or contact Russell Nelson at 214-665-6646 or Karen Kesler at 214-665-3185.

Sincerely,

William K. Honker, P.E.

Director

Water Division

Enclosure

#### Recommendations

- Identify the sources of TDS coming into Millwood Lake and calculate the loadings on the Little River below Millwood dam for each of the sources
- Present all data used to derive the TDS criterion for the Little River
- Provide water quality data for the Little River downstream of the SWEPCO outfall, particularly TDS, can either be newly or previously collected data from after the plant was operational
- Collect or provide existing temperature data downstream of SWEPCO outfall in winter months
- Provide numeric values for tolerance ranges of indigenous aquatic species in the Little River for temperature and TDS
- Use best fit regression model to determine temperature criterion
- Provide more complete data identifying the percent loading of minerals due to natural versus anthropogenic sources in the Red River; provide citations supporting the identification of the sources
- Revise the TDS mass balance with values that take into account changes in output from point source dischargers along the Red River if the Red River is delisted
- Demonstrate that the SSC changes in the Red River will be protective of the aquatic life use
  - o Numeric tolerance values from lab and field studies of species found in the Red River
  - o TDS values during the Buchanan et al. 2003 study only were ≥ 860 mg/L at Index (upstream of the reach of the SSC), therefore additional information needs to be presented to show that the aquatic life use will be met with the new proposed criteria

# **Heather Ferguson**

From: Kesler, Karen [Kesler.Karen@epa.gov]
Sent: Friday, February 26, 2016 2:16 PM

To: jim (jtm@ftn-ASSOC.COM)

Cc: clem@adeq.state.ar.us; Crocker, Philip; Nelson, Russell; Watson, Jane Subject: Request for additional information concerning the SWEPCO rulemaking

Dear Mr. Malcom,

During our discussion last week with ADEQ I expressed some concerns about the mass balance model from the SWEPCO UAA that was utilized to estimate whether the new site specific TDS criteria on the Red River from the confluence of the Little River to the Arkansas-Louisiana was protective of Louisiana's downstream criteria. As I have continued to think about the appropriateness of this mass balance this week, I believe I have come across another issue with the way it was executed. In the UAA for the SWEPCO rulemaking, the mass balance section states that:

Instream TDS concentrations in the Red River were modeled for two critical flow conditions: 7Q10 and harmonic mean flow. Regulation 2.106 of the Arkansas water quality standards (APCEC 2011; the latest version that is currently approved by EPA) specifies 7Q10 (referred to as "Q7 -10" in the standards) as the critical flow for meeting dissolved mineral criteria in streams such as the Little River and Red River that have site-specific criteria that are listed in Regulation No. 2.511 without an asterisk. Harmonic mean flow is the critical flow for meeting chloride, sulfate, and TDS criteria in Louisiana according to Section III5.C.8 of the Louisiana water quality standards (LDEQ 2012).

However, when I went back and reviewed Regulation 2 the definition for critical flows was that listed below:

Critical flows: The flow volume used as background dilution flows in calculating concentrations of pollutants from permitted discharges. These flows may be adjusted for mixing zones. The following critical flows are applicable:

For a seasonal fishery - 1 cfs minus the design flow of any point source discharge (may not be less than zero).

For human health criteria - harmonic mean flow or long term average flow.

For minerals criteria - harmonic mean flow or 4 cfs, except in those waters listed in Reg. 2.511. Those waters in Reg. 2.511 which are noted with an asterisk will have a critical flow of 4 cfs. (Also see minerals implementation procedure in CPP)

For all others - the critical flow will be Q7-10.

This indicates that a harmonic mean is more appropriate for the mass balance than the 7Q-10 is, which was used to set the value for the site specific criteria. The demonstration of downstream protection is contingent upon this difference in flow utilized to assess the levels of minerals in the Red River. The 860 mg/L value that was determined through the 7Q-10 model would be protective of Louisiana when it was assessed with a harmonic mean flow. However, minerals criteria in Arkansas are actually permitted using the harmonic mean flow, not the 7Q-10, meaning that the 860 mg/L criteria would actually violate Louisiana's criteria of 780 mg/L since they are both using the same flow regime. Based on this information, it does not seem that the 860 mg/L criteria is protective of Louisiana's 780 mg/L criteria. In addition, after reviewing the reference to Louisiana's standards, the reference to harmonic mean flow seems to apply to mixing zones:

- C. Mixing, Mixing Zone, and Flow Application
- 8. For chloride, sulfate, and total dissolved solids, criteria are to be met below the point of discharge after complete mixing. Because criteria are developed over a long-term period, harmonic mean flow will be applied for mixing.

When speaking to EPA staff in Assessments for Louisiana and speaking to LDEQ is appears the Louisiana criteria for minerals are determined by taking the arithmetic mean plus three standard deviations and that they do not consider flow. Based on this information, additional material would need to be provided to demonstrate downstream protection of Louisiana's criteria. I appreciate your help in providing all the necessary information to assess the appropriateness of the site specific criteria proposed by the SWEPCO rulemaking.

Sincerely, Karen Kesler

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

# **Heather Ferguson**

From: Jim Malcolm [jtm@ftn-assoc.com]
Sent: Monday, March 07, 2016 12:50 PM

To: 'Kesler, Karen'

Cc: 'clem@adeq.state.ar.us'; 'Crocker, Philip'; 'Nelson, Russell'; 'Watson, Jane'; James Malcolm

Subject: RE: Request for additional information concerning the SWEPCO rulemaking

Dear Karen,

Sorry for the delay in my reply but I wanted to discuss our reply with Sarah Clem before sending it out, we have completed that discussion.

This is written regarding your recent supplemental questions on the SWEPCO rulemaking. Thank you for reading this portion of the report very closely. Your question raises an important issue regarding implementation of the criteria we proposed which is a slightly different issue, to us, versus EPA's review and approval of the proposed criteria.

ADEQ has implemented AR minerals criteria at various flow conditions including 7Q-10 flow conditions based on the EPA approved version of Regulation No. 2 which you quoted below (e.g. see the existing TMDL for this watershed). This version was also the current version when we submitted the UAA report to the Arkansas Pollution Control & Ecology Commission. The SWEPCO and Domtar TDS and sulfate proposed SSC were developed anticipating implementation under 7Q-10 flow conditions to match the existing, previously approved (by EPA) TMDL. As such, once EPA has completed their review of the proposed criteria for minerals, SWEPCO will propose updates to the TMDL but we will not propose to modify the critical flow conditions – this will assure both continuation of the existing water quality conditions of the Red River (for minerals) as well as downstream compliance with LDEQ's TDS criteria. We agree with you that the LDEQ regulations (referenced below) require harmonic mean (HM) flow as critical flow for meeting dissolved minerals criteria in their state. As you stated, the HM flow conditions model was run and included to demonstrate our proposed TDS criterion will result in downstream compliance with the LA criteria. The HM flow simulation uses the same point source loads as in the 7Q-10 simulation.

If for some reason, both ADEQ and EPA desire to implement new SSC for TDS on the Red River based on a TMDL/permit that were HM flow conditions, we would need to recalculate the criterion. This would result in a much lower proposed TDS criterion in the range of 730 mg/L. We do not recommend changing the flow basis for our proposed criteria at this point but would defer to the agencies if they so desired.

Please do not hesitate to contact me or Phil Massirer with any further questions or comments. We will be providing ADEQ with our responses to your recent comments that were discussed in the ADEQ offices by later today. Hopefully, you will see the ADEQ review summary of the responses very soon.

Sincerely, Jim



Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Friday, February 26, 2016 2:16 PM

To: jim (jtm@ftn-ASSOC.COM)

**Cc:** clem@adeq.state.ar.us; Crocker, Philip; Nelson, Russell; Watson, Jane **Subject:** Request for additional information concerning the SWEPCO rulemaking

Dear Mr. Malcom,

During our discussion last week with ADEQ I expressed some concerns about the mass balance model from the SWEPCO UAA that was utilized to estimate whether the new site specific TDS criteria on the Red River from the confluence of the Little River to the Arkansas-Louisiana was protective of Louisiana's downstream criteria. As I have continued to think about the appropriateness of this mass balance this week, I believe I have come across another issue with the way it was executed. In the UAA for the SWEPCO rulemaking, the mass balance section states that:

Instream TDS concentrations in the Red River were modeled for two critical flow conditions: 7Q1O and harmonic mean flow. Regulation 2.106 of the Arkansas water quality standards (APCEC 2011; the latest version that is currently approved by EPA) specifies 7Q10 (referred to as "Q7 -10" in the standards) as the critical flow for meeting dissolved mineral criteria in streams such as the Little River and Red River that have site-specific criteria that are listed in Regulation No. 2.511 without an asterisk. Harmonic mean flow is the critical flow for meeting chloride, sulfate, and TDS criteria in Louisiana according to Section III5.C.8 of the Louisiana water quality standards (LDEQ 2012).

However, when I went back and reviewed Regulation 2 the definition for critical flows was that listed below:

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This indicates that a harmonic mean is more appropriate for the mass balance than the 7Q-10 is, which was used to set the value for the site specific criteria. The demonstration of downstream protection is contingent upon this difference in flow utilized to assess the levels of minerals in the Red River. The 860 mg/L value that was

determined through the 7Q-10 model would be protective of Louisiana when it was assessed with a harmonic mean flow. However, minerals criteria in Arkansas are actually permitted using the harmonic mean flow, not the 7Q-10, meaning that the 860 mg/L criteria would actually violate Louisiana's criteria of 780 mg/L since they are both using the same flow regime. Based on this information, it does not seem that the 860 mg/L criteria is protective of Louisiana's 780 mg/L criteria. In addition, after reviewing the reference to Louisiana's standards, the reference to harmonic mean flow seems to apply to mixing zones:

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Sincerely, Karen Kesler

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

# **Heather Ferguson**

From: Kesler, Karen [Kesler.Karen@epa.gov]
Sent: Monday, March 07, 2016 12:59 PM

To: Jim Malcolm

Cc: clem@adeq.state.ar.us; Crocker, Philip; Nelson, Russell; Watson, Jane Subject: RE: Request for additional information concerning the SWEPCO rulemaking

Hi Jim,

Thank you for your reply. Once we have a chance to discuss this issue further, I'll let you know if we need any additional information or if the criteria needs to be recalculated. I appreciate your help on clarifying this part of the supporting material for the rulemaking.

Thanks, Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Monday, March 07, 2016 12:50 PM
To: Kesler, Karen < Kesler. Karen@epa.gov>

Cc: clem@adeq.state.ar.us; Crocker, Philip <crocker.philip@epa.gov>; Nelson, Russell <nelson.russell@epa.gov>;

Watson, Jane <watson.jane@epa.gov>; James Malcolm <jtm@ftn-assoc.com> **Subject:** RE: Request for additional information concerning the SWEPCO rulemaking

Dear Karen,

Sorry for the delay in my reply but I wanted to discuss our reply with Sarah Clem before sending it out, we have completed that discussion.

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Sincerely, Jim



Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Friday, February 26, 2016 2:16 PM

To: jim (jtm@ftn-ASSOC.COM)

**Cc:** <u>clem@adeq.state.ar.us</u>; Crocker, Philip; Nelson, Russell; Watson, Jane **Subject:** Request for additional information concerning the SWEPCO rulemaking

Dear Mr. Malcom,

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Sincerely, Karen Kesler

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

# **Heather Ferguson**

From: Wentz, Tate [WENTZ@adeq.state.ar.us]
Sent: Wednesday, March 30, 2016 3:57 PM

To: kesler.karen@epa.gov; nelson.russell@epa.gov

Cc: Clem, Sarah; jtm@ftn-ASSOC.COM; agates@mwlaw.com

Subject: Domtar and SWEPCO Supplemental Information

Attachments: TM-W Honker 2016-03-07 Domtar.pdf; TM-W Honker 2016-03-08 SWEPCO.pdf;

Domtar\_signed.pdf; SWEPCO\_signed.pdf

#### Russell and Karen,

Please find attached digital copies of Domtar and SWEPCO supplemental information to complete documentation for the proposed site-specific criteria. We placed hard copies in the mail today.

Please let Sarah or I know if you have any additional questions or comments.

Tate Wentz
Aquatic Ecologist Coordinator
AFS Certified Fisheries Professional
Office of Water Quality-Planning Branch
Arkansas Department of Environmental Quality
North Little Rock, AR 72118
501-682-0661



CERTIFIED MAIL: 91 7199 9991 7035 3549 1694

March 30, 2016

Bill Honker, Director (6WQ)
Water Quality Protection Division
United States Environmental Protection Agency
Region 6
1445 Ross Avenue Suite 1200
Dallas, TX 75202-2733

Re: Supplemental Information to Address EPA Recommendations to Complete Documentation for Southwestern Electric Power Company's Proposed Site-Specific Criteria for Temperature in the Little River and Use Attainability Analysis For Dissolved Minerals in the Little River and Red River

Dear Mr. Honker,

ADEQ is pleased to provide you with Southwestern Electric Power Company's (SWEPCO) response to EPA's February 9, 2016 request for additional information. Office of Water Quality staff has reviewed SWEPCO's response to EPA and has no additional comments.

If you or your staff have any questions regarding, please feel free to contact me or Sarah Clem (clem@adeq.state.ar.us/501-682-0660).

Sincerely,

Caleb Osborne

Associate Director of the Office of Water Quality Arkansas Department of Environmental Quality

CC: Russell Nelson, Water Quality Protection Division, EPA Region 6

Karen Kesler, Water Quality Protection Division, EPA Region 6

Jim Malcom, FTN Associates, LTD

Allan Gates, Mitchell, William, Selig, Gates & Woodyard, P.L.L.C.



#### TECHNICAL MEMORANDUM

DATE:

March 8, 2016

TO:

William K. Honker

United States Environmental Protection Agency, Region VI

FROM:

Jim Malcolm, Patrick Downey, Philip Massirer Pall

FTN Associates, Ltd.

SUBJECT:

Supplemental Information to Address EPA Recommendations to Complete Documentation for Southwestern Electric Power Company's Proposed Site-Specific Criteria for Temperature in the Little River and Use Attainability Analysis for Dissolved

Minerals in the Little River and Red River

FTN No. R06510-0010-002

#### 1.0 BACKGROUND

On October 23, 2015, Region 6 of the US Environmental Protection Agency (EPA) provided technical comments to the Arkansas Department of Environmental Quality (ADEQ) on two reports¹ prepared for Southwestern Electric Power Company (SWEPCO) in support of a third-party rulemaking to modify dissolved minerals criteria in the Little River and Red River and the temperature criterion in the Little River. On January 15, 2016, SWEPCO provided EPA with responses to EPA's technical comments and additional information in support of the rulemaking. On February 9, 2016, ADEQ received additional comments on the responses and information provided by SWEPCO, along with requests for additional information. Representatives of EPA, ADEQ, SWEPCO, and FTN Associates, Ltd. (FTN) met at the ADEQ office on Friday, February 19, 2016, to clarify EPA's requests for additional information and discuss the projects. This memorandum addresses the comments received from EPA as clarified in the meeting at ADEQ.

## 2.0 ADDITIONAL INFORMATION IN SUPPORT OF THE PROPOSED RULEMAKING

EPA's comments as provided in correspondence dated February 9, 2016, are reproduced in italics below and have been numbered for reference purposes. SWEPCO's responses follow each comment.

<sup>&</sup>lt;sup>1</sup> Use Attainability Analysis for Dissolved Minerals in Little and Red Rivers, Hempstead and Little River Counties, Arkansas, submitted to ADEQ on September 5, 2014, and Technical Justification for a Site-Specific Temperature Criterion in the Little River, Hempstead and Little River Counties, Arkansas, submitted to ADEQ on September 10, 2014.

1. Identify the sources of TDS coming into Millwood Lake and calculate the loadings on the Little River below Millwood dam for each of the sources

Attachment 1 provides a discussion of TDS loads to Little River below Millwood Lake dam.

2. Present all data used to derive the TDS criterion for the Little River

The proposed TDS criterion for the Little River was derived from the sampling data collected by SWEPCO and FTN from October 2010 to October 2013 as part of the UAA effort. The data are provided as Attachment 2.

3. Provide water quality data for the Little River downstream of the SWEPCO outfall, particularly TDS, can either be newly or previously collected data from after the plant was operational

FTN and SWEPCO did not collect water quality data downstream of the SWEPCO outfall because current TDS measurements collected downstream of the SWEPCO outfall would not reflect anticipated future discharge conditions. SWEPCO's current permit limit for TDS is set equal to the Red River TDS criterion of 500 mg/L because the Red River is listed as impaired due to TDS on the Arkansas 2008 303(d) list (see Section 1.1 of the report). This "end-of-pipe" permit limit for TDS prevents the SWEPCO plant from operating as designed (i.e., re-circulation of cooling water up to ten cycles in the cooling tower).

A mass balance was prepared (utilizing anticipated operating conditions) to estimate effluent TDS concentrations. The model provided a proposed TDS criterion that would allow SWEPCO to operate the facility efficiently while still protecting the existing and attainable designated uses of the stream. The modeling was described in Section 5.0 of the report and additional documentation was provided in Appendices J through L. The model showed that an instream TDS criterion of 136.1 mg/L would be necessary in the Little River. This criterion would enable the SWEPCO plant to operate as designed once revised effluent limitations for TDS at Outfall 001 are calculated and incorporated into the facility's NPDES permit. The proposed TDS criterion for the Little River is 138 mg/L and is derived from the ecoregion reference stream values listed in Arkansas Pollution Control and Ecology Commission Regulation No. 2.511(B). The model results showed that TDS concentrations in the Little River under optimal plant operating conditions should remain below 138 mg/L under critical low-flow (7Q10) conditions.

4. Collect or provide existing temperature data downstream of SWEPCO outfall in winter months

Based on discussions held during the February 19 meeting at ADEQ, EPA withdrew its request for these data due to the  $\Delta 5^{\circ}$ F ( $\Delta 2.8^{\circ}$ C) criterion in the Arkansas water quality standards that is considered protective of aquatic life.

5. Provide numeric values for tolerance ranges of indigenous aquatic species in the Little River for temperature and TDS

Please see Attachment 3.



6. Use best fit regression model to determine temperature criterion

The regression model with the higher coefficient of determination (R<sup>2</sup>) was not the sole basis for the proposed criterion, because the high R<sup>2</sup> value was not indicative of the regression model's accuracy for predicting high water temperatures, which is important for developing a proposed temperature criterion. Attachment 4 provides a discussion of the modeling performed and the rationale for the proposed criterion.

7. Provide more complete data identifying the percent loading of minerals due to natural versus anthropogenic sources in the Red River; provide citations supporting identification of the sources

Sections 2.3.2.1 and 5.0 of the report provided information on sources of minerals to the Red River, which was then supplemented with additional information provided in the first set of responses to EPA comments (dated January 15, 2016). Appendices J and K to the report provided the model input and output. Additional data are provided in Attachment 5; a list of point sources to the Red River in Arkansas was extracted from the 2014 Arkansas 305(b) report and lists of point sources to the Red River in Texas and Oklahoma downstream of Lake Texoma were compiled by querying EPA's ECHO database for NPDES permits within the eight-digit hydrologic unit codes (HUCs) 111401-01 through -06 in Oklahoma and within HUCs 111401-01, 111401-05, and 111401-06 in Texas.

8. Revise the TDS mass balance with values that take into account changes in output from point source dischargers along the Red River if the Red River is delisted

Point sources will have to comply with the limits established in the total maximum daily load (TMDL) report. The proposed criteria have been demonstrated to be protective, and the TMDL and NPDES processes will ensure that the instream concentrations will be capped at the proposed criteria.

- 9. Demonstrate that the SSC changes in the Red River will be protective of the aquatic life use
  - a. Numeric tolerance values from lab and field studies of species found in the Red River
  - b. TDS values during the Buchanan et al. 2003 study only were > 860 mg/L at Index (upstream of the reach of the SSC), therefore additional information needs to be presented to show that the aquatic life use will be met with the new proposed criteria

Please see Attachment 6.

We appreciate the opportunity to work with you on this project. If you have questions or comments regarding this information, please do not hesitate to call Jim Malcolm, Pat Downey, or Philip Massirer, PE, at (501) 225-7779.

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TDS Sources and Loads to the Little River

## 1.0 POINT SOURCES AND TDS LOADS TO MILLWOOD LAKE

In the meeting at ADEQ's offices on February 19, 2016, it was understood that EPA's recommendation #1 on the SWEPCO proposed criteria (regarding sources of TDS to Millwood Lake and loadings on the Little River below Millwood Lake dam) could be addressed by identifying point sources in the Millwood Lake watershed and estimating their loadings of TDS.

Using EPA's Enforcement and Compliance History Online (ECHO) website, 25 facilities with individual NPDES permits were identified in the Millwood Lake watershed. The TDS load from each facility was estimated based on effluent flow rates and concentrations. The effluent flow rates were based on average (if available) or design flow rates in EPA's ICIS-NPDES database (accessed on EPA's ECHO web site). None of the facilities report TDS concentrations on discharge monitoring reports (DMRs); therefore, effluent TDS concentrations were assumed to be 400 mg/L for publicly owned treatment works (POTWs) and 1,000 mg/L for non-POTWs. These calculations yielded a total point source load of about 30 tons/day of TDS to Millwood Lake. Due to the lack of DMR data for TDS, no temporal trend could be identified for the point source loads. The point source facilities and their effluent flow rates and assumed TDS concentrations are listed in Table 1 below.

To provide perspective for the point source load, the total TDS load to Millwood Lake was estimated based on the long-term average release from Millwood Lake dam (3,642 million gallons per day [MGD] for October 1966 through September 2015) and the median TDS concentration for the Little River approximately 1 mile upstream of SWEPCO's outfall (63 mg/L based on 56 samples collected by FTN and SWEPCO between October 2010 and October 2013; see Attachment 2 to the memorandum). This yielded a total load of about 957 tons/day. Based on this result, the point source load is about 3% of the total load.

The nonpoint source load was estimated to be about 927 tons/day (the total load minus the point source load). A search was conducted to look for information concerning natural versus anthropogenic nonpoint source loads in the Millwood Lake watershed, but sufficient information was not found.

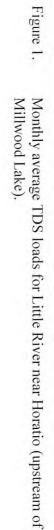
Table 1. Point source facilities, effluent flow rates, and assumed TDS concentrations.

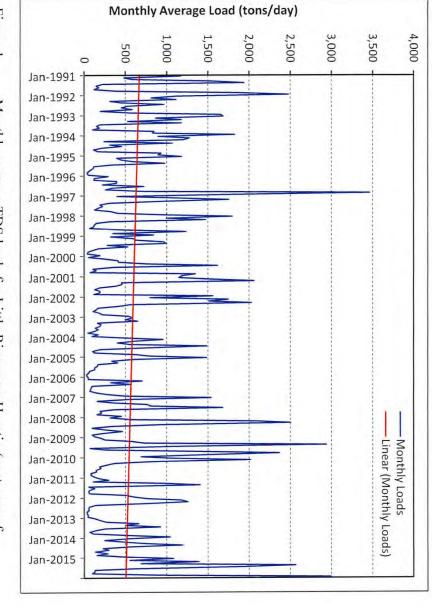
NPDES ID	Facility Name	Facility Type	TDS Concentration (mg/L)		TDS Load (lbs/day
AR0002909	WEYERHAEUSER NR CO DEQUEEN	NON-POTW	1,000	0.216	1,803
AR0002917	WEYERHAEUSER NR CO DIERKS MILL	NON-POTW	1,000	0.494	4,122
AR0003018	TYSON FOODS, INC GRANNIS	NON-POTW	1,000	0.864	7,210
AR0021261	MINERAL SPRINGS, CITY OF- WWTP	POTW	400	0.20	668
AR0021377	LOCKESBURG, CITY OF	POTW	400	0.15	501
AR0021709	DIERKS, CITY OF	POTW	400	0.23	768
AR0021733	DEQUEEN, CITY OF	POTW	400	4.00	13,352
AR0021776	NASHVILLE, CITY OF	POTW	400	2.30	7,677
AR0023817	FOREMAN, CITY OF	POTW	400	0.28	935
AR0035483	HATFIELD, CITY OF	POTW	400	0.07	234
AR0035785	HORATIO, CITY OF	POTW	400	0.15	501
AR0040886	WILTON, TOWN OF - WWTF	POTW	400	0.09	300
AR0041246	MILLWOOD WATER CORP	NON-POTW	1,000	0.03	250
AR0041734	TYSON FOODS, INC. NASHVILLE	NON-POTW	1,000	1.11	9,263
AR0045144	TOLLETTE, CITY OF	POTW	400	0.10	334
AR0047996	GILLHAM REGIONAL WASTEWATER	POTW	400	0.20	668
AR0048593	LOCKSBURG PLT #1544	NON-POTW	1,000	0.10	835
AR0049034	COSSATOT ROCK, LLC	NON-POTW	1,000	0.10	835
AR0049247	COVE, CITY OF	POTW	400	0.075	250
OK0000736	WEYERHAEUSER COMPANY	NON-POTW	1,000	0.10	835
OK0000795	TYSON FOODS, INC-BROKEN BOW	NON-POTW	1,000	0.10	835
OK0021521	BROKEN BOW PUBLIC WORKS AUTH.	POTW	400	0.761	2,540
OK0027677	IDABEL PUBLIC WORKS AUTHORITY	POTW	400	1.65	5,508
OK0032387	WRIGHT PUBLIC WORKS AUTHORITY	POTW	400	0.014	47
OK0044458	MIXON BROS. WOOD PRESERVING	NON-POTW	1,000	0.10	835
Total point so	ource load		-		61,102

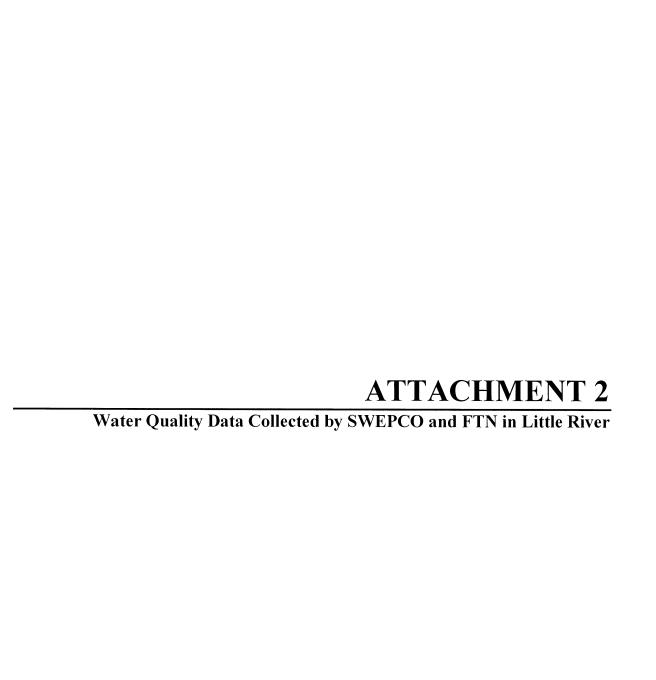
Land use data for the Millwood Lake watershed were downloaded from the USGS National Land Cover Database (NLCD). Based on 2011 data, the Millwood Lake watershed consists of approximately 61.6% forest/shrub, 25.9% pasture/hay/grass, 7.0% wetlands and water, 5.3% developed land, and 0.2% of other land (cultivated cropland and barren land). NLCD data from 2001 showed similar percentages: 67.8% forest/shrub, 19.9% pasture/hay/grass, 7.1% wetlands and water, 5.1% developed land, and 0.1% of other land. This shows that about 6% of the watershed changed from forest/shrub to pasture/hay/grass between 2001 and 2011. However, no information was found to estimate nonpoint source TDS loadings for different land uses. Therefore, it was not possible to evaluate what effect (if any) this land use change would have on the nonpoint source TDS load at Millwood Lake dam.

The TDS loads entering Millwood Lake were also analyzed temporally. Monthly TDS loads for the largest inflow to Millwood Lake (the Little River) were estimated for January 1991 through December 2015. These loads were estimated using USGS daily flow data and ADEQ routine samples (approximately monthly intervals) for the Little River at State Highway 41 near Horatio (USGS gage 07340000 and ADEQ station RED0002). These loads are shown on Figure 1 with a linear trend line that shows a decreasing trend over time.

Daily flow data were not available to calculate monthly TDS loads for other tributaries to Millwood Lake. However, TDS concentrations for other tributaries do not show significantly increasing trends over time (see Figures 2.2 through 2.4 in the UAA report). Therefore, it is expected that the TDS loads for those tributaries would not show increasing trends over time.







Dissolved mineral data in Little River upstream of Turk Outfall 001

Date	TDS (mg/L)	Chloride (mg/L)	Sulfate (mg/L)	Collected by
10/12/10	64	107	48	AEP
11/18/10	20	11	6	AEP
01/25/11	16	8	6	AEP
03/11/11	84	8	8	AEP
04/28/11	66	7	9	AEP
05/16/11	48	5	5	AEP
06/22/11	36	5	6	AEP
07/20/11	80	6	5	AEP
08/02/11	84	6	6	AEP
08/26/11	68			AEP
09/23/11	92			AEP
10/10/11	74	8	6	AEP
11/03/11	68	9	7	AEP
11/30/11	96	6	6	AEP
12/09/11	62	5	6	AEP
12/20/11	24	7	6	AEP
01/03/12	48	4	6	AEP
01/12/12	36	5	6	AEP
01/16/12	28	4	6	AEP
02/03/12	38			AEP
02/07/12	44	4	5	AEP
03/21/12	48	5	6	AEP
04/30/12	40	3	5	AEP
05/07/12	53	4	5	AEP
06/05/12	60	7	8	AEP
06/25/12	80	7	7	AEP
07/09/12	64	8	8	AEP
01/04/13	80	10	7	AEP
03/02/13	48	7	7	AEP
03/20/13	99	5.1	5.7	FTN
05/02/13		3.4	4.5	FTN
05/02/13	64	5	6	AEP
05/06/13	60	3	0	AEP
05/08/13	45	3.3	4.6	FTN
05/08/13		6	6	AEP
05/24/13		7	7	AEP
05/29/13		5	6	AEP
05/31/13		3.2	4.6	FTN
06/06/13		5	6	AEP
06/06/13		1.8	3.1	FTN
06/14/13		5	5	AEP
		5	5	AEP
06/20/13				
07/01/13		3.3	4.2	FTN
07/14/13		4	6	AEP
07/19/13		3.7	4.5	FTN
07/19/13		5	5	AEP
07/26/13		4	5	AEP
08/06/13	89	4.4	4.8	FTN

Date	TDS (mg/L)	Chloride (mg/L)	Sulfate (mg/L)	Collected by
08/07/13	72	6	6	AEP
08/12/13	96	4.5	4.9	FTN
08/15/13	47	4.5	4.3	FTN
08/16/13	94	6	6	AEP
08/22/13	88	6.2	5.7	FTN
08/29/13	100	4.8	4.6	FTN
09/19/13	71	4.9	4.3	FTN
10/17/13	150	5.5	4.7	FTN
ount:	56	51	. 51	
Ain:	10	1.9	3 1	

Count:	56	51	51
Min:	10	1.8	3.1
25th %tile:	48	4.2	5.0
Median:	63	5.0	6.0
75th %tile:	84	6.6	6.0
90th %tile:	98	8.0	7.0
95th %tile:	105	8.5	8.0
Max:	150	11.0	9.0
Start Date	10/12/10		

10/17/13

End Date

## **ATTACHMENT 3**

Discussion of TDS and Temperature Tolerance Ranges for Indigenous Aquatic Species in the Little River

## 1.0 ION TOLERANCE

### 1.1 Ion Tolerance in Fish

The use attainability analysis (UAA) report documented the presence of 40 species of fish collected by FTN in August 2013 and by the US Geological Survey (USGS) in June 2009 and August 2015 (referred to as the "pooled" list in the UAA report) in the Little River downstream of the Millwood Lake dam. The report concluded that the species makeup of these collections closely resembled expectations based on zoogeography (i.e., fish species known to be present in the Gulf Coastal Plain ecoregion), habitat preferences, and gear selectivity. EPA did not dispute this conclusion or ask for further documentation of this conclusion in any of its comments.

A comparison of the species present in the pooled list with the list of species present in the Red River (based on Buchanan et al. 2003) reveals that 37 of the 40 species from the pooled list (92%)<sup>1</sup> were present in the Red River based on the Buchanan et al. (2003) survey. As shown in Tables 1 and 2, the 10<sup>th</sup> percentile TDS values from the Garland and Index monitoring stations on the Red River during the Buchanan et al. study period are well above the proposed Little River TDS criterion of 138 mg/L. This information documents that virtually all of the fish species found in the Little River are also found in the Red River where TDS concentrations are substantially higher than the proposed TDS criterion. This finding proves that the proposed TDS criterion of 138 mg/L for the Little River is well within the TDS tolerance range of the fish present in the Little River. Therefore the proposed TDS criterion for the Little River will support the fisheries designated use based on the existing fish communities.

<sup>&</sup>lt;sup>1</sup> The fish species present in the Little River but not present in the Red River were bowfin (*Amia calva*), redfin shiner (*Lythrurus umbratilus*) and logperch (*Percina caprodes*).

Table 1. Summary of TDS concentrations in the Red River at Index (USGS 07337000) during and after the Buchanan et al. (2003) study.

		TDS at Index (USGS 07337000) (mg/L)			
Summary Statistic		Study Period	Post-Study Period		
	10	427	246.0		
	25	632	428.3		
Percentile	50	720	584.5		
Percentile	75	912	792.0		
	90	1,128	903.4		
	95	1,198	932.9		
Number of Values		37	72		
Period of Record		4/2/1996 – 12/11/2001	1/2/2002 - 10/22/2013		

Table 2. Summary of TDS concentrations in the Red River at Garland (RED0045) during and after the Buchanan et al. (2003) study.

		TDS at Garland (	RED0045) (mg/L)
Summary Statistic		Study Period	Post-Study Period
	10	157.2	156.0
	25	216.0	247.5
Percentile	50	464.0	438.5
Percennie	75	660.5	624.0
	90	801.8	750.6
	95	886.2	819.3
Number of Values		63	138
Period of Record		4/2/1996 — 12/11/2001	1/2/2002 — 10/22/2013

### 1.2 Ion Tolerance in Benthic Macroinvertebrates

## 1.2.1 Comparison with Field-Based Aquatic Life Benchmark for Conductivity

EPA (2011) used conductivity as a surrogate measure of TDS to develop a benchmark value of 300  $\mu$ S to protect aquatic life. The analysis is most applicable to high-gradient streams in the Appalachian Region (or comparable ecoregions) that are dominated by salts of Ca<sup>+2</sup>, Mg<sup>+2</sup>, SO<sub>4</sub><sup>2-</sup> and HCO<sub>3</sub><sup>-</sup>. Although the aforementioned salts dominate the Little River ionic matrix, the Little River is not located within a comparable ecoregion. The benthic macroinvertebrate communities that typify the high-gradient streams that were the focus of the EPA 2011 study

exhibit large numbers of sensitive EPT taxa. The protective benchmark conductivity value of  $300~\mu S$  is a reflection of the presence of those sensitive taxa. These sensitive taxa are not part of the existing benthic macroinvertebrate community of the Little River, which was shown in the UAA report to be dominated by Chironomidae and a single genus of Trichoptera. Therefore evaluating the Little River benthic community using the benchmark developed in EPA 2011 represents a conservative approach, because the benchmark is based on much more sensitive taxa than are found in the Little River.

Water quality data collected from the Little River during 2011 through 2013 indicated a poor correlation ( $R^2 = 0.20$ ) between TDS and conductivity in the Little River (Figure 1). However, a conservative estimate of the level of TDS that would correspond to 300  $\mu$ S was obtained by computing the upper 95<sup>th</sup> confidence interval of the ratio of conductivity to TDS and multiplying this factor by 138 mg/L. The data and statistics used for this computation are provided in Tables 3 and 4. Based on this computation, a conservative (i.e., high) estimate of the conductivity that would be associated with the proposed TDS criterion of 138 mg/L is 218  $\mu$ S, which is below the benchmark value of 300  $\mu$ S.

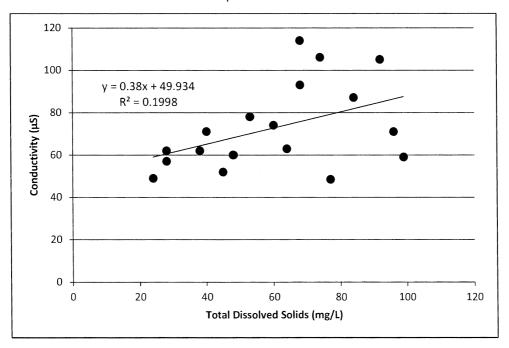


Figure 1. Scatter plot of relationship between TDS and conductivity measured in samples collected from the Little River downstream of Millwood Lake dam.

Table 3. Paired conductivity and TDS measurements from various sampling locations in the Little River downstream of the Millwood Lake dam.

	Conductivity	TDS	
Sampling Date	(µS)	(mg/L)	Conductivity/TDS
08/3/2011	87	84	1.036
08/26/2011	93	68	1.368
09/23/2011	105	92	1.141
10/11/2011	106	74	1.432
11/03/2011	114	68	1.676
11/30/2011	71	96	0.740
12/09/2011	338	62	Outlier; deleted from analysis
12/21/2011	49	24	2.042
01/03/2012	60	48	1.250
01/18/2012	57	28	2.036
02/03/2012	62	38	1.632
04/30/2012	71	40	1.775
05/07/2012	78	53	1.472
03/20/2013	59	99	0.596
05/02/2013	49	77	0.630
05/03/2013	63	64	0.984
05/06/2013	74	60	1.233
05/08/2013	52	45	1.156
05/13/2013	62	28	2.214

Table 4. Summary statistics from conductivity and TDS data provided in Table 3.

Summary Statistic	Value
Conversion factor (average conductivity/TDS)	1.3562
Standard deviation	0.4778
Number of values	18
Alpha	0.05
95% confidence interval (CI)	<u>+</u> 0.2207
Upper CI of conversion factor	1.5769
Estimated conductivity of 138 mg/L based on upper CI of conversion factor	138 x 1.5769 = 217.6188

## 1.2.2 Evaluation of the Tolerance of Indigenous Benthic Macroinvertebrates to Dissolved Ions

### 1.2.2.1 Non-Chironomid Taxa

The UAA study included sampling of benthic macroinvertebrates from woody debris, which is a dominant substrate type in the Little River below Millwood Lake dam. Communities were similar upstream versus downstream of the SWEPCO discharge point so that all samples were combined. Table 5 summarizes XC95 values ("95% Extirpation Concentration") for the taxa present in the Little River samples based on Appendices D and H of EPA 2011.

Table 5. Summary of relative abundance and XC95 values (from EPA 2011) for benthic macroinvertebrate taxa collected from woody debris substrates in the Little River.

			Relative	XC95 (μS)	
Order	Family	Genus	Abundance	Kentucky <sup>1</sup>	West Virginia <sup>2</sup>
Coleoptera	Elmidae	Stenelmis	< 0.01	> 1,520	>9,790
Diptera	Ceratopogonidae	Bezzia	0.03	ND	380
Diptera	Chironomidae		0.77	see Table 8	
Ephemeroptera	Caenidae	Caenis	0.02	>1,410	>3,923
Ephemeroptera	Heptageniidae	Stenacron	0.01	> 862	782
Megaloptera	Corydalidae	Nigronia	< 0.01	>1,197	>9,790
Odonata	Coenagrionidae	Argia	0.01	>1,410	ND
Odonata	Gomphidae	Erpetogomphus	< 0.01	ND	ND
Trichoptera	Polycentropodidae	Polycentropodidae	< 0.01	>570	>4,713
Trichoptera	Polycentropodidae	Cyrnellus	0.14	ND	ND

#### Notes:

- 1. Based on Appendix H of EPA 2011.
- 2. Based on Appendix D of EPA 2011.

XC95 values are statistically derived conductivity values associated with a 95% or greater probability that a particular genus will be absent at a site. Therefore an XC95 value near 218  $\mu$ S for a particular taxon would indicate that a TDS value of 138 mg/L could potentially limit the abundance of that taxon. XC95 values indicate a conductivity level associated with *absence* of a taxon. Therefore, for the purposes of this evaluation, any taxon with an XC95 value less than 900  $\mu$ S (three times EPA's field-based conductivity benchmark of 300  $\mu$ S) was considered to be potentially susceptible to adverse effects (i.e., decreased abundance) due to a TDS level corresponding to 281  $\mu$ S.

Table 5 shows that Chironomidae and a single genus of Trichoptera (*Cyrnellus*) comprised 91% of individuals collected. XC95 values were available for six of the remaining eight non-chironomid taxa, which comprised 9% of the population sampled. XC95 values of the remaining six taxa were generally greater than 900 μS (i.e., exceeding the 300-μS benchmark by three-fold or more) and ranged to up to nearly 10,000 μS; the exceptions were *Bezzia* and *Stenacron* (XC95 values of 380 and 782 μS, respectively), which comprised 4% of the individuals. This analysis indicates that TDS concentrations near 138 mg/L might potentially adversely affect the distribution and abundance of *Bezzia* and *Stenacron*, which comprise 4% of the benthic macroinvertebrate population in the Little River.

Potential effects on the trichopteran Cyrnellus, which made up 14% of the individuals in the Little River samples, could not be evaluated using EPA 2011 because this genus was not present in the EPA 2011 data set. There is only one North American species of Cyrnellus (C. fraternus), which is also found in South America<sup>2</sup>. Johnson et al. (1998) described the microdistribution of C. fraternus in the Elm Fork of the Trinity River in north Texas. The full text document for this study was not available, so any ancillary water quality data taken as part of the study are also unavailable. The USGS monitoring station located on Elm Fork (USGS 08050400<sup>3</sup>) does not include TDS or conductivity data. However, conductivity monitoring data are available from April 6, 2010, through March 1, 2016, at nearby Little Elm Creek (USGS 08052700<sup>4</sup>). Little Elm Creek is somewhat smaller than the Elm Fork (75.5 mi<sup>2</sup> versus 174 mi<sup>2</sup> at the respective monitoring locations). Conductivity data from Little Elm Creek can serve as a surrogate to represent conductivity in the Elm Fork because both waterbodies are located in the Cross Timbers ecoregion and should have similar geology. Table 6 summarizes available daily conductivity values from the Little Elm Creek monitoring station. If these values can be considered to be representative of Elm Fork, then they indicate that the Little River TDS criterion of 138 mg/L will support C. fraternus because the conductivity values greatly exceed the 218  $\mu$ S value that corresponds to 138 mg/L TDS.

<sup>4</sup> Located at latitude 33°17'00" and longitude 96°53'33" (NAD27).

http://www.waterbugkey.vcsu.edu/php/genusdetail.php?idnum=15&g=Cyrnellus&ls=larvae&f=Polycentropodidae

Located at latitude 33°37'27" and longitude 97°093'22" (North American Datum of 1927 [NAD27]).

Table 6. Summary of daily average conductivity monitoring from Little Elm Creek.<sup>5</sup>

	Daily Average Conductivity Values				
Summary Statistic	Median	80th Percentile			
Minimum	383	602			
Mean	673	861			
Maximum	989	1,420			
Number of values	351				
Period of record	April 16, 2010, to March 1, 2016				

This analysis indicates that TDS values exceeding the proposed site-specific criterion will support *C. fraternus*, which makes up 14% of the individuals in the Little River benthic macroinvertebrate samples.

### 1.2.2.2 Chironomid Taxa

Chironomidae, which made up 77% of the individuals in the Little River samples (Table 5), were not identified to genus, so the XC95 values for the 61 chironomid genera available in EPA 2011 cannot be directly applied to the Little River data set. However, 55 of the 61 XC95 values in EPA 2011 were reported as "greater than" values that were substantially greater than 218  $\mu$ S, with only two of those values (>681  $\mu$ S and >824  $\mu$ S) being less than 900  $\mu$ S (Table 7). Of the six definitive XC95 values for chironomid genera from EPA (2011), presented in Table 8, five are less than 900  $\mu$ S.

This information suggests that TDS concentrations at or near 138 mg/L could potentially adversely affect 7 of the 55 (9%) chironomid genera for which XC95 data are available. Available information on the geographic distribution of these genera was not sufficiently detailed to definitively assess whether the Little River is within the geographic range of these genera. However, Hudson et al. (1990) provide notes on biology, habitat, and distribution of chironomid taxa in Alabama, Florida, Georgia, Tennessee, South Carolina, and North Carolina. Table 8 provides this information for the genera having definitive XC95 values or XC95 values potentially less than 900 µS from EPA (2011). This information indicates that three of the seven

<sup>&</sup>lt;sup>5</sup> Conductivity data from USGS Station 08052700 were downloaded as percentile values for each day of the calendar year for the period of record. Table 6 summarizes the minimum, mean, and maximum values for the median and 80<sup>th</sup> percentile values for the entire period of record.

genera having (or potentially having) XC95 value less than 900  $\mu S$  are not likely to be found on woody debris habitat in the Little River.

Table 7. Summary of "greater than" XC95 values for chironomid general from EPA 2011.

Summ	ary Statistic	XC95 Value (μS)
	5 <sup>th</sup>	1,004
	25 <sup>th</sup>	1,616
Percentile	50 <sup>th</sup>	2,074
	75 <sup>th</sup>	4,971
	95 <sup>th</sup>	11,646
Minimum		686
Mean		4,349
Maximum		34,898
Number of value	ues	55

Table 8. Chironomid genera having definitive XC95 values or XC95 values potentially less than 1,000 μS from EPA 2011.

Genus	XC95 Value (μS)	Habitat/Distribution Notes from Hudson et al. 1990	Occurrence on Woody Debris in Little River?
Conchapelopia	546	More common in mountains than coastal plains; most often found in flowing water; some species are very tolerant of chemical and organic pollution.	Possible
Demicryptochironomus	322	Larvae characteristic of clean sandy areas of streams and rivers in the southeast.	Not likely
Micropsectra	462	Most abundant and diverse in cool mountain streams, rarely in coastal plains.	Not likely
Microtendipes	> 681	Widespread especially <i>M. caducus</i> and <i>M. pedellus</i> in clean streams. <i>M. pedellus</i> abundant in nutrient-enriched streams.	Possible
Rheopelopia	1,457	Found in small, fast-flowing stream in NC and large rivers in GA.	Possible
Stempellina	644	Most species are widespread.	Yes
Stenochironomus	> 824	Lotic and lentic habitats.	Yes
Zavrelia	413	Restricted to cool streams of mountains and sand hills.	Not likely

This analysis indicates that chironomid genera are in general tolerant of elevated conductivity levels and that only a small proportion of genera are likely to be sensitive to conductivity levels at or near 218  $\mu$ S (i.e., TDS concentrations at or near 138 mg/L). Therefore, TDS concentrations at or near the proposed TDS criterion of 138 mg/L should have negligible impacts on the existing chironomid population.

## 1.3 Summary and Conclusions

#### 1.3.1 Fish

Virtually all of the fish species found in the Little River are also found in the Red River where TDS concentrations are substantially higher than the proposed TDS criterion for the Little River of 138 mg/L. This finding proves that the proposed TDS criterion for the Little River is well within the TDS tolerance range of the fish present in the Little River. Therefore the proposed TDS criterion for the Little River will support the fisheries designated use based on the existing fish communities.

### 1.3.2 Benthic Macroinvertebrates

A conservative (i.e., high) estimate of the conductivity that would be associated with the proposed TDS criterion of 138 mg/L is 218 μS, which is below EPA's field-based benchmark value of 300 μS. The 300-μS benchmark developed in EPA 2011 represents a conservative value when applied to the Little River because the benchmark is based on much more sensitive taxa than are found in the Little River. This finding indicates that proposed TDS criterion of 138 mg/L should not impair the benthic macroinvertebrate community (including communities occupying habitats other than woody debris) in the Little River.

TDS concentrations near 138 mg/L might potentially adversely affect the distribution and abundance of *Bezzia* and *Stenacron*. However, this potential effect would affect only 4% of the benthic macroinvertebrates inhabiting woody debris in the Little River.

TDS values exceeding the proposed site-specific criterion will likely support *C. fraternus*, which makes up 14% of the individuals in the Little River benthic macroinvertebrate samples.

XC95 values for chironomid genera documented in EPA 2011 were all greater than 218  $\mu$ S and were overwhelmingly greater than 900  $\mu$ S (three times the 300- $\mu$ S benchmark developed in EPA 2011). This analysis shows that chironomid genera, which comprised 77% of the individuals found on woody debris substrates, are in general tolerant of elevated conductivity levels. Only a small proportion of genera are likely to be sensitive to conductivity levels at or near 218  $\mu$ S (i.e., TDS concentrations at or near 138 mg/L). Therefore, TDS levels at or near the proposed TDS criterion of 138 mg/L should have negligible impacts on the existing chironomid population.

### 2.0 TEMPERATURE TOLERANCE

## 2.1 Temperature Tolerance in Fish

As discussed in Section 1.1, the UAA report documented the presence of 40 species of fish collected by FTN in August 2013 and by USGS in June 2009 and August 2015 (referred to as the "pooled" list in the UAA report) in the Little River downstream of Millwood Lake dam. The report concluded that the species makeup of these collections closely resembled expectations based on zoogeography (i.e., fish species known to be present in the Gulf Coastal Plain ecoregion), habitat preferences, and gear selectivity. Therefore, the reach of the Little River below Millwood Lake dam currently attains its designated aquatic life (fishery) use based on the fish community. EPA did not dispute this conclusion or ask for further documentation of this conclusion in any of its comments.

A comparison of the species present in the pooled list (based on recent sampling) with the list of species present in the Red River (based on Buchanan et al. 2003) reveals that 37 of the 40 species from the pooled list (92%) were present in the Red River based on the Buchanan et al. (2003) survey. Available temperature data obtained from the USGS monitoring station on the Red River at Index, AR (USGS 07337000) (Table 9) indicate that temperatures exceeding 32°C are a common feature of the Red River. Table 9 shows that maximum *mean* temperatures exceed 32°C 11% of the time during the period of record. Since this summary refers to daily *mean* temperatures, maximum temperatures in excess of 32°C can be expected to occur even more

frequently. This characteristic of the Red River temperature regime is reflected in the temperature criterion of 32°C for the Red River (APCEC 2014). Therefore, since the large majority of the fish from the pooled list have also been documented in the Red River, temperatures in excess of 32°C will support the fisheries use in the Little River based on fish populations.

Summ	ary Statistic	Value (°C)	
	5 <sup>th</sup>	12.3	
	25 <sup>th</sup>	17.6	12.3 17.6 23.3 29.9 32.8 89.3 7.5
Percentile	50 <sup>th</sup>	23.3	
	75 <sup>th</sup>	29.9	
	95 <sup>th</sup>	32.8	,
Percentile Ra	ank of 32°C	89.3	
Minimum		7.5	
Mean		23.2	

33.5

October 1, 2003, to June 6, 2011

Table 9. Summary of maximum mean daily temperatures on each calendar day at Index, AR.

### 2.2 Benthic Macroinvertebrates

Maximum

Period of record

The proposed temperature criterion for the Little River below the Millwood Lake dam of 32°C is based on monitoring data collected during the UAA study and represents the conditions under which sampling of the indigenous macroinvertebrate community was performed. Therefore, it has already been shown that the indigenous macroinvertebrate community can tolerate a temperature regime that supports a 32°C criterion because the community was actually sampled under that regime. EPA's request to demonstrate that the indigenous biota can tolerate the conditions under which they were found represents circular logic.

No independent studies could be located that demonstrate that the indigenous macroinvertebrate taxa can tolerate the temperatures as high or higher than the indigenous taxa currently experience. Available field studies that addressed thermal tolerance occurred in northern locations and focused on cold-water species. The seasonal window for most field protocols for benthic macroinvertebrate sampling is generally early to mid-spring when maximum temperatures do not occur. Therefore benthic macroinvertebrate data sets with

accompanying temperature data typically do not address the warm temperatures shown during the late summer in the Little River.

## 2.3 Summary and Conclusions

Available temperature data obtained from the USGS monitoring station on the Red River at Index, AR (USGS 07337000) (Table 9) indicate that temperatures exceeding 32°C are a common feature of the Red River. Since the large majority of the fish collect from the Little River have also been documented in the Red River, temperatures in excess of 32°C will support the fisheries use in the Little River based in fish populations.

No applicable laboratory data or field data from other locations could be located showing that the indigenous invertebrate species present in the Little River can tolerate temperatures approaching or exceeding 32°C. However, the simultaneous occurrence of the indigenous biota and temperatures approaching and exceeding 32°C proves that a temperature criterion of 32°C will support indigenous biota.

### 3.0 REFERENCES

- APCEC. 2014. Regulation No. 2: Regulation establishing water quality standards for surface waters of the state of Arkansas. Little Rock, AR: Arkansas Pollution Control and Ecology Commission. Effective February 28, 2014.
- EPA. 2011. A Field-Based Aquatic Life Benchmark for Conductivity in Central Appalachian Streams, Final Report [EPA/600/R-10/023F]. Washington, DC: US Environmental Protection Agency.
- Hudson, P.L., D.R. Lenat, B.A. Caldwell, and D. Smith. 1990. *Chironomidae of the southeastern United States: a checklist of species and notes on biology, distribution and habitat* [Paper 173]. Washington, DC: United States Department of the Interior, Fish and Wildlife Service. ISSN 1040-2411.
- Johnson, Z.B., A.K. Riggs, and J.H. Kennedy. 1998. Microdistribution and secondary production of *Cyrnellus fraternus* (Trichoptera: Polycentropodidae) from snag habitats in the Elm Fork of the Trinity River, Texas. *Ann. Entomol. Soc. Am.* 91:641-646.

ATTACHMENT A
ATTACHMENT 4  Discussion of Best-Fit Regression Model Used for Temperature Criterion

# 1.0 DISCUSSION OF BEST FIT REGRESSION MODEL FOR TEMPERATURE CRITERION

The proposed criterion was developed using a weight-of-evidence approach with multiple sets of data and information, including the following:

- Observed water temperatures approximately 1 mile upstream of SWEPCO's outfall,
- Two regression models that predict daily maximum water temperatures, and
- Existing criteria for upstream and downstream waterbodies (Millwood Lake and the Red River).

The regression model with the higher coefficient of determination  $(R^2)$  was not the sole basis for the proposed criterion, because the high  $R^2$  value was not indicative of the regression model's accuracy for predicting high water temperatures, which is important for developing a proposed temperature criterion.

The second regression model presented in the report (described on pages 4-3 and 4-4 of the report) had a higher R<sup>2</sup> value than the first regression model (described on pages 4-1 and 4-2), but the higher R<sup>2</sup> does not necessarily mean that it is the best regression for developing a proposed criterion. The R<sup>2</sup> value represents the percentage of the observed data variation that is explained by the model. Comparisons of R<sup>2</sup> values for different regression models must consider differences in the data upon which each regression is based.

The first regression model was developed using only data for July and August, while the second regression model was developed using data for February through September (a longer period of observed data was needed to develop sine curves to represent normal temperatures). For the first regression model, the observed water temperatures during July and August exhibit a relatively small range of variation (27.6°C to 34.7°C). For the second regression model, though, the observed water temperatures for February through September exhibit a larger range of variation (8.6°C to 34.7°C). Therefore, a certain error in the predicted temperatures (say, 1.0°C for example) represents a much larger percentage of the observed data variation for the first regression model than for the second regression model. In other words, if both regression models

have the same error in predicted temperatures, the first regression model will inherently have a lower R<sup>2</sup> value than the second regression model.

A better metric for evaluating the accuracy of each regression model is the standard error of estimate, which was slightly lower for the first regression model (1.0°C) than for the second regression model (1.17°C). This shows that both regression models have similar accuracy over the range of observed temperatures used to develop each regression.

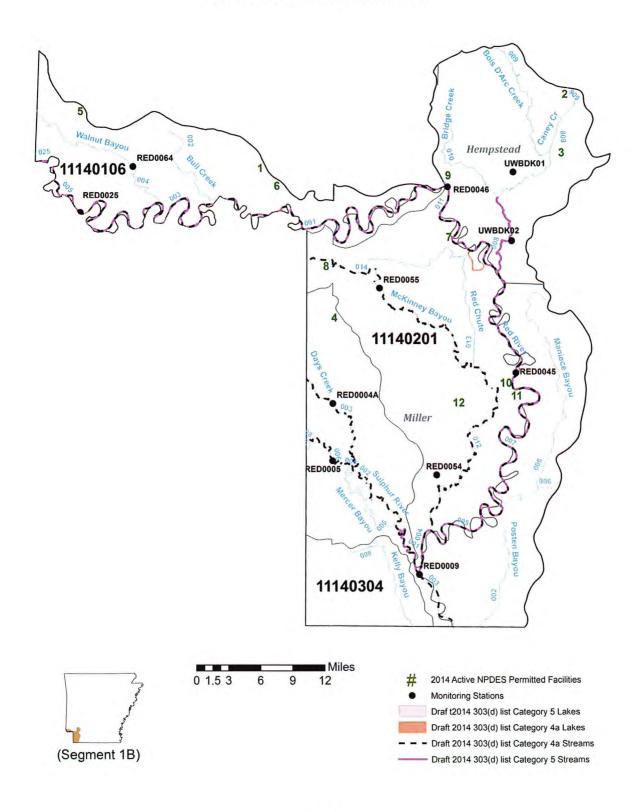
For development of temperature criteria, the accuracy of model predictions is important for days with high observed water temperatures. Unfortunately, both of these regression models tended to under-predict water temperatures on days with high observed water temperatures. The two regression models under-predicted water temperatures by an average of 1.4°C and 1.8°C, respectively, on days during 2013 when the observed maximum water temperature was 32°C or greater. Therefore, the 95<sup>th</sup> percentiles of daily temperatures predicted by these regression models are probably biased low.

The observed water temperature data only cover a 2-year period, but they provide valuable measurements that show numerous temperatures exceeding both the existing criterion (30°C) and the proposed criterion (32°C). Because these data are actual measurements (as opposed to model predictions), they must be given strong consideration in a weight-of-evidence evaluation.

## **ATTACHMENT 5**

**Point Sources to the Red River** 

Figure A-3: Planning Segment 1B



**Table A-4: Active NPDES permits for Planning Segment 1B** 

Permit Number	Facility Name	Receiving Waters	Reach	USGS H.U.C.	County	Map No.
AR0002968	DOMTAR A.W. LLC	PIPING & OPEN CANAL,RED R	001	11140106	Little River	1
AR0021326	TYSON FOODS, INC HOPE PROCESSING PLANT	UNNAMED TRIB,CANEY CR,BOIS D'ARC CR,RED R	909	11140201	Hempstead	2
AR0038466	HOPE, CITY OF-BOIS D'ARDC WWTP	BLACK BR,BOIS D'ARC CR,RED R	009	11140201	Hempstead	3
AR0038822	COOPER TIRE & RUBBER COMPANY	TRIB,NIX CR,DAYS CR,SULPHUR R,RED R	003	11140302	Miller	4
AR0042846	ASH GROVE CEMENT COMPANY	FRENCH CR,WALNUT BU,RED R	004	11140106	Little River	5
AR0042951	ASHDOWN WW TREATMENT PLANT	DOMTAR CANAL,RED R	001	11140106	Little River	6
AR0048356	TYSON FOODS, INCRIVER VALLEY ANIMAL FOODS	RED R	011	11140201	Miller	7
AR0048691	TEXARKANA, CITY OF- NORTH WWTP	MCKINNEY BU,RED R	014	11140302	Miller	8
AR0048810	FULTON, CITY OF-WWTP	RED R	011	11140201	Hempstead	9
AR0050857	GARLAND, CITY OF	RED R	007	11140201	Miller	10
AR0051942	CHIEFTAIN SAND AND PROPPANT, LLC	RED R	007	11140201	Miller	11
AR0052035	WOOTEN SAND AND GRAVEL, INC.	MCKINNEY BU,RED R	012	11140201	Miller	12

Facilities with NPDES Permits located in HUCs 111401-01, -02, -03, -04, -05, and -06 in Oklahoma, queried from ECHO.

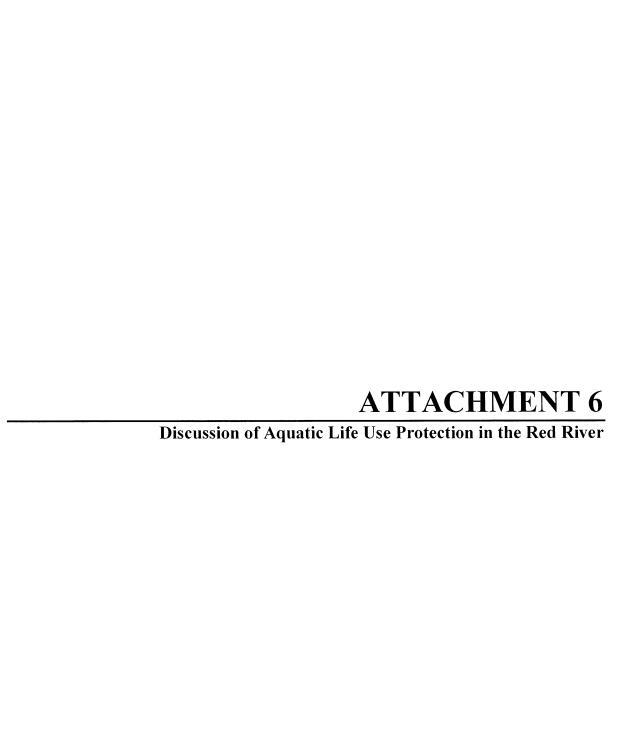
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110000608236	Western Farmers Electric Coop	Hwy. 70	Hugo	OK	Choctaw	34.016330	-95.321880 OK0035327	OK0035327
110000816528	Trinityrail Maintenance Services Plant	1806 West Garrett Street	Hugo	OK	Choctaw	34.017753	-95.540873 OKP003019	OKP003019
110001143361	T Bar M Ranch	Rt. 1 Box 233	Hendrix	OK	Bryan	33.815292	-96.258688 OKG010337	OKG010337
110002047384	Durant City Utilities Auth	1222 Davis Road	Durant	OK	Bryan	33.937083	-96,384000 OK0039063	OK0039063
110007384954	Lattimore Matl Corp / Stringtown Crusher No 67	1 mi NE of town off Hwy 69	Stringtown	OK	Atoka	34.481833	-96.048972 OK0040762	OK0040762
110007388291	Unimin Corp / Roff Plt	3rd and Walling St	Roff	OK	Pontotoc	34.636833	-96.814278 OK0040291	OK0040291
110009154654	Antlers PWA	100 SE 2nd St	Antlers	OK	Pushmataha	34.217972	-95.612139	-95.612139 OK0002615 OK0020346
110009154761	Millerton Pub Wrks Authority	P.O. Box 239	Millerton	OK	McCurtain	33.979382	-95.022535 OK0037028	OK0037028
110009154869	Ok St Dept of Corrections-Howa	1970 East Whippoorwill Lane	Atoka	OK	Atoka	34.162306	-95.822167	OK0033413 OKG580069
110010808395	Mack Alford Corr. Center	P.O. Box 220	Stringtown	OK	Atoka	34.522778	-96.030000 OK0044113	OK0044113
110011007622	Wapanucka Public Works Auth.	P.O. Box 163	Wapanucka	OK	Johnston	34.378750	-96.428417 OK0034011	OK0034011
110011007891	Calera PWA	110 West Main Street	Calera	OK	Bryan	33.924972	-96.440167	OK0031682
110011008024	Boswell, Town of	410 6th St	Boswell	OK	Choctaw	34.033583	-95.860806 OK0032255	OK0032255
110011008337	Clayton PWA	P.O. Box 279	Clayton	OK	Pushmataha	34.577600	-95.354800 OK0029157	OK0029157
110011008676	Talihina PWA	P.O. Box 457	Talihina	OK	Latimer	34.747779	-95.059297	OK0028380
110011008738	Atoka, City of	350 East North Hills Rd	Atoka	OK	Atoka	34.405201	-96.109161 OK0028576	OK0028576
110011008827	Bokchito, City of	Bryan County	Bokchito	OK	Bryan	34.012722	-96.136750 OK0027014	OK0027014
110011009791	Allen, Town of	P.O. Box 402	Allen	OK N	Pontotoc	34.876556	-96.423333 OK0020206	OK0020206
110011011591	Choctaw Manufacturing & Devlp	203 Choctaw Industrial Drive	Hugo	OK	Choctaw	34.016970	-95.534750 OKP003005	OKP003005
110011012812	Mansion Farm	P.O. Box 918	Holdenville	OK	Hughes	34.866944	-96.334444 OKG010292	OKG010292
110011012830	W-6 Swine Farms, Inc.	8159 E. 1515 Road	Calvin	OK	Hughes	34.816287	-96.234789 OKG010294	OKG010294
110011013358	Bolen Farms	Route 2, Box 328	Idabel	OK Yo	McCurtain	33.823611	-94.839444	OKG010230
110011013401	Roger Stinchcomb #2	Route 2, Box 25b	Stonewall	OK	Pontotoc	34.635556	-96.613889 OKG010243	OKG010243
110011013410	Skiles Farm	Route 1, Box 62	Holdenville	OK	Johnston	34.443333	-96.555278 OKG010244	OKG010244
110011013633	Phillips Farms	P.O. Box 35	Garvin	OK	McCurtain	33.956111	-94.960278 OKG010088	OKG010088
110011013786	Holts Egg Farm	Route 2, Box 67	Stonewall	OK	Johnston	34,448333	-96.609444 OKG010130	OKG010130
110011013820	Wood Farm, Larry	Route 1, Box 2	Milburn	OK	Johnston	34.266667	-96.541667 OKG010139	OKG010139
110011013875	Roger Stinchcomb #1	Route 2, Box 25b	Stonewall	OK	Pontotoc	34.636666	-96.610277 OKG010156	OKG010156
110011014302	Mahard Egg Farm, Inc Granite	Route 1, Box 25a	Milburn	OK	Johnston	34.349903	-96.582227	OKG010059
110012299789	Colbert WWT	% Colbert Water Department	Colbert	OK	Bryan	33.845556	-96.495389 OKG580017	OKG580017
110012864837	Achille	500 N 2nd Ave	Achille	OK	Bryan	33.831180	-96.390140 OK0027979	OK0027979
110012874354	Bryan County RWS & SWMD #2	Bryan County	Mead	OK	Bryan	34.049972	-96.380306 OKG380014	OKG380014
110014332775	Soper Public Works Authority	600 Main St	Soper	OK	Choctaw	34.034972	-95.700000 OK0044563	OK0044563
110017843565	Alan Ritchey Matl Co LC / Popes Point	6992 SR 74741	Hendrix	OK	Bryan	33.821944	-96.150278 OK0045519	OK0045519
110020151075	Sawyer Facility	6.5 mi N of Hwy 70 via SH147	Sawyer	OK	Choctaw	34.114611	-95.382028 OKG950017	OKG950017
110022402728	Dolese Brothers CoColeman	3 mi E & 1 mi S of Coleman	Coleman	OK	Atoka	34.258500	-96.366889 OKG950030	OKG950030
110022423028	Meridian Aggregates CoHugo	3 mi N of US Hwy 70	Hugo	OK	Choctaw	34.054826	-95.532990 OKG950021	OKG950021
110022590766	Choctaw Co. RW & SD #1	510 Everidge	Grant	OK	Choctaw	33.924917	-95.508278 OK0037826	OK0037826
110022616436	Pushmataha Co RWD #3	8 mi E of Antlers on SH 3	Antlers	OK	Pushmataha	34.201647	-95.487147	OK0045616
110022632150	Coalgate PWA	3 South Main	Coalgate	OK	Coal	34.508222	-96.214361 OKG580028	OKG580028
110024410653	Fittstone, Inc.	County Road 167, West of 99	Fittstown	OK	Pontotoc	34.593889	-96.646389 OKG950010	OKG950010
110037270823	Valliant PWA	111 N. Dalton	Valliant	OK	McCurtain	33.991861	-95.093278 OKG580065	OKG580065

Facilities with NPDES Permits located in HUCs 111401-01, -02, -03, -04, -05, and -06 in Oklahoma, queried from ECHO.

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110037271519	10037271519 Wapanucka Oolitic Limestone Quarry	10203mile Corner Rd	Bromide	OK	Johnston	34.404444	34.404444 -96.473056 OKG950047	OKG950047
110041245961	10041245961 Hale Scout Reservation- Camp Tom Hale	1 Camp Tom Hale Road	Talihina	OK	e Flore	34.735528	34.735528 -94.888028 OK0045764	OK0045764
110044270794	10044270794 Roos Resources, Inc.	7955 Cr 1670	Roff	OK	Pontotoc	34.569306	-96.824583 OKG950052	OKG950052
110044903174	.10044903174 Stringtown PWA	P.O. Box 98	Stringtown	OK	Atoka	34.459639	-96.063611 OK0030449	OK0030449
110046122995	10046122995 Weyerhaeuser Nr Co Idabel Sawmill	5500 NW Texas St	Idabel	OK	McCurtain	33.928477	-94.869819 OK0043885	OK0043885
110055639411	10055639411 Atoka Co. RW & SWMD #4	337 E. Hwy. 3	Atoka	OK	Atoka	34.351333	34.351333 -96.059417 OKG380059	OKG380059
110064606369	10064606369 Hugo Municipal Authority	201 South 2nd St.	Hugo	OK	Choctaw	33.988417	33.988417 -95.525778 OK0028487	OK0028487
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110064620548	10064620548 Kiowa Power Partner, LLC	4386 Old 69 Rd	Kiowa	OK	Pittsburg	34.683833	34.683833 -95.934722 OK0044083	OK0044083
110067190596	10067190596 E-Z Mart #108	802 N. Washington Ave	Durant	OK	Bryan	34.001387	-96.390410 OKG830048	OKG830048

Facilities with NPDES Permits located in HUCs 111401-01, -05, and -06 in Texas, queried from ECHO.

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registry_id	fac_name	fac_street	fac_city	fac_state	fac_state fac_county fac_lat	fac_lat	fac_long	npdes_ids	_
110000720970	City of Paris Water Treatment Plant	301 Lake Crook Road	Paris	X	Lamar	33.724468	-95.567306	TX0075931	1
110000758056	Wastewater Treatment Plant	1800 E. F.M. 1417	Sherman	TX	Grayson	33.601667	-96.574167	TX0024325	
110005103887	Grayson County College	6101 Grayson Dr	Denison	ΥL	Grayson	33.707972	-96.636611 TX0056235	TX0056235	
110006801924	Sherman Ready Mix - WWTP	6102 Teresa Drive	Sherman	Ϋ́	Grayson	33.693623		-96.581363 TXG110118 TXG111225	
110009115590	City of Honey Grove	2000f W FMR 100 & 3000 f N US	Honey Grove	XX	Fannin	33.613806	-95.918889 TX0117951	TX0117951	
110009747015	Reno WWTF	1 1/3m SW FMR 195 & NW 7th St	Reno	XL	Lamar	33.695856	-95.479678 TX0082309	TX0082309	
110009748023	Hooks Isd WWTP	E FMR 560 3 Mi N Ih-30	Hooks	XX	Bowie	33.517000	-94.285639 TX0118079	TX0118079	
110009774896	Camp Maxey WWTP	0.5 mi SE Intx US Hwy 271 and FMM 2648	Lamar County	TX	Lamar	33.778417	-95.540222 TX0101214	TX0101214	
110009778115	Town of Windom WWTP	0.25 mi SW Intx US Hwy 82 & FM	Windom	TX	Fannin	33.562583	-96.006889 TX0072711	TX0072711	
110009779187	City of Dodd City WWTP	2200 f SW Sthwy 897/82 2500 f SE	Dodd City	TX	Fannin	33.569639	-96.068889 TX0057169	TX0057169	
110009779613	City of Bells	US Hwy 69 and FM Hwy 1897, N	Bells	TX	Grayson	33.619806	-96.413028 TX0053368	TX0053368	
110009779640	Avery WWTP	On Mill Creek, 1.5 mi NE city	Avery	TX	Red River	33.561972	-94.776444 TX0053503	TX0053503	
110009780228	River Hills Owners Association	600' NW of FM 691 and FM 131	Denison	TX	Grayson	33.711167	-96.606444 TX0033154	TX0033154	
110009780246	City of Whitewright	1 BIk W FMR 898/Mk&T Railrd	Whitewright	TX	Grayson	33.520858	-96.392039 TX0033294	TX0033294	
110009781147	City of Ector	4700' N of US Hwy 82 & 800' W	Ector	TX	Fannin	33.589500	-96.278556 TX0025151	TX0025151	
110009781441	City of Hooks WWTP	449 Willow Oak Drive	Hooks	X	Bowie	33.477389	-94.252889	TX0022969	
110009781557	City of Savoy WWTF	601 N Cedar St	Savoy	TX	Fannin	33.606361	-96.367944 TX0023299	TX0023299	
110012702851	New Boston Lumber Mill	Hwy. 82 E.	New Boston	TX	Bowie	33.467499	-94,386940 TX0079723	TX0079723	
110015743622	Paris Energy Center	0.5 mi N Intx US Hwy 271 and US Hwy 82	Paris	TX	Lamar	33.696750	-95.557722 TX0103586	TX0103586	
110017693076	Coston & Son Ready Mix	155 East Oak Ave	Paris	TX	Lamar	33.677940	-95.554350 TXG110508	TXG110508	
110027363234	Randolph WWTP	FMR 896,1/2 m S Sthwy 121	Randolph	TX	Fannin	33.479695	-96.247995 TX0027928	TX0027928	
110031272469	Sherman Ready Mix	6202 Theresa St	Sherman	TX	Sherman	33.690992	-96.581447 TXG110755	TXG110755	
110033462731	Bonham Concrete	827 W Sam Rayburn Dr	Bonham	TX	Fannin	33.577259	-96.187356 TXG110317	TXG110317 TXG111177	
110037841681	Valley Steam Electric Station	1040 Cr 1225	Savoy	TX	Fannin	33.623890	-96.368126 TX0009067	TX0009067	
110038112271	City of Denison Ms4	Within the Denison city limits	Denison	TX	Grayson	33.755555	-96.562500 TXR040230	TXR040230	
110038179075	Texas Dept of Transportation	Paris Dist MS4	Paris	TX	Grayson	33.674139	-95.556036 TXR040166	TXR040166	
110039874053	Paw Paw WWTP	1600' E & 2200' N of Intx Cent	Denison	TX	Grayson	33.772798	-96.501864 TX0047228	TX0047228	
110042082572	Silgan Containers Paris Plant	500 NW Loop 286	Paris	TX	Lamar	33.718056	-95.568222 TX0008982	TX0008982	
110045506476	Hope Concrete	5815 N Travis St	Sherman	×⊥	Grayson	33.647340	-96.613050 TXG111178	TXG111178	
110059708400	Load Trail	220 FM 2216	Sumner	⊥X	Lamar	33.719020	-95.837720 TX0134431	TX0134431	
110064591419	Sherman WWTP	1800 E FM 1417	Sherman	X	Grayson	33.602434	-96.577777	TXL005020	
110064609847	City of Bonham WWTP	2501 Seven Oaks Road east of Bonham	Bonham		Fannin	33.587247	-96.160641	TX0021814	
110064648234	Bonham WWTP	2073 FM 273	Bonham	XT	Fannin	33.639143	-96.165859 TX0135569	TX0135569	



# 1.0 ASSESSMENT OF AVAILABLE DATA TO EVALUATE THE AQUATIC LIFE COMMUNITY & WATER QUALITY IN THE RED RIVER

The Buchanan et al. (2003) study and the availability of long-term monitoring data available from several long-term monitoring stations on the Red River provide a unique opportunity to link the results of an extensive and intensive survey of the fish communities with the TDS regime present during the study. This data set also has the distinct advantage of having been collected in the actual waterbody of interest. Therefore, conclusions based on this data set avoid the interpretational difficulties that inevitably arise when investigators are forced to rely on laboratory results (due to the usual difficulties encountered when applying laboratory studies to field situations) or other field studies (due to different taxa present).

We again wish to point out that EPA's concern that the age of the Buchanan et al. study (conducted from 1995 to 2001) undermines or weakens the conclusions based on the study is not valid because, as noted in responses submitted to EPA on January 15, 2016, the relevant issue regarding the use of the Buchanan et al. study is the water quality conditions present at the time of the study, not the calendar year(s) of the study. There is no reason to believe that the range of tolerance of any given species present during the study should be different from the range of tolerance that that species would show currently. Therefore, any species that tolerated the TDS concentrations present in the Red River from 1995 to 2001 should be able to tolerate the same or lower concentrations today.

The Buchanan et al. study included a comparison of the fish species present in the reach of the Red River upstream of the mouth of the Little River versus downstream of the mouth of the Little River ("upstream" and "downstream" reaches, respectively). The authors noted little difference in species richness between the upstream (67 species) and downstream (65 species) reaches. The upstream reach supported 67 of the total 72 species (93%) collected during the entire survey while the downstream reach supported 90% of the total species collected. Seven species were found only in the upstream segment and five species were found only in the downstream segment. The entire survey found nine species of darters (various species of *Amocrypta*, *Etheostoma*, *and Percina*), with all nine species found in the upstream reach and six

species found in the downstream reach. These results demonstrate that both the upstream and downstream reaches of the Red River show "high fish species richness."

The Red River monitoring station at Index, AR (USGS 07337000) provides a long-term record of TDS concentrations representative of the upstream reach of the Red River. Table 1 summarizes TDS concentrations present during the Buchanan et al. study and demonstrates that the 95th percentile TDS concentration during the study was 1,198 mg/L at the Index location. This result clearly demonstrates that TDS conditions that result in a 95<sup>th</sup> percentile concentration that exceeds the proposed criterion by almost 40% (1,198 mg/L versus 860 mg/L) will support high fish species richness and a high level of fisheries use support.

Table 1. Summary of TDS concentrations in the Red River at Index (USGS 07337000) during and after the Buchanan et al (2003) study.

		TDS at Index (U	JSGS 07337000) g/L)
Summary	<b>Statistic</b>	Study Period	Post-Study Period
	10	427	246.0
	25	632	428.3
Danaantila	50	720	584.5
Percentile	75	912	792.0
	90	1,128	903.4
	95	1,198	932.9
Number of Va	alues	37	72
Period of Rec	ord	4/2/1996 – 12/11/2001	1/2/2002 - 10/22/2013

Table 3.9 of the report suggested that TDS concentrations in the Red River downstream of its confluence with the Little River were lower than the proposed criterion for that reach during the Buchanan et al. study. However, Table 3.9, as presented, provided misleading results. The table was intended to show that TDS concentrations during the Buchanan et al. study were higher than the post-study period. However, it did not include the 95<sup>th</sup> percentile values from the monitoring stations, which formed part of the basis for the proposed 860 mg/L TDS criterion in the Red River downstream of the mouth of the Little River. The table also included data from the monitoring station at Fulton (RED0046), which was not used for the development of the criterion

because it is not representative of TDS conditions in the Red River at that location. This was explained in Section 5.5 of the report, which notes the following:

Neither the RED0046 data nor the RED0009 data were considered to be a good representation of well-mixed concentrations in the Red River. Both the RED0046 station and the RED0009 station are located in places where the Red River does not appear to be laterally well-mixed due to significant tributary inflows a short distance upstream of each station. Lateral gradients at these monitoring stations are likely created by inflows from the Little River (RED0046) and the Sulphur River (RED0009). In some instances, these lateral gradients can be observed from aerial photographs. According to a station description report from EPA's Legacy STORET web site, the RED0046 station is sampled at a railroad bridge pier on the east side of the river (i.e., the left bank), which is the same side of the river that the Little River enters. The RED0009 station is sampled at a boat ramp on the west side of the river, which is the same side of the river that the Sulphur River enters. Therefore, both the RED0046 data and the RED0009 data are likely measuring a disproportionate percentage of tributary water that has not mixed completely with the entire flow of the Red River. Both tributaries (especially the Little River) contribute water that typically has TDS concentrations much lower than concentrations in the Red River. Therefore, the data at these two stations appear to be underestimating the TDS concentrations in the Red River and should not be used for comparison with predicted TDS values from the mass balance modeling.

The monitoring station at Garland (RED0045) is not similarly affected and provides the most representative data set for assessing TDS concentrations in the reach of the Red River downstream of the mouth of the Little River. Accordingly, Table 3.9 should not have included data from the monitoring station at Fulton (RED0046) and should have included the 95<sup>th</sup> percentile values for the Garland station. Also, monitoring data from Index is not relevant for the purposes of assessing the reach of the Red River downstream of the Little River and should not have been included. The following Table 2 provides the percentile TDS values for the Garland station during and after the Buchanan et al. study and shows that the 95<sup>th</sup> percentile for TDS at Garland was 886 mg/L during the Buchanan et al. study period (from 1995 to 2001).

Table 2. Summary of TDS concentrations in the Red River at Garland (RED0045) during and after the Buchanan et al. (2003) study.

		TDS at Garland (RED0045) (mg/L)		
Summary Statistic		Study Period	Post-Study Period	
	10	157.2	156.0	
	25	216.0	247.5	
Percentile	50	464.0	438.5	
Percentile	75	660.5	624.0	
	90	801.8	750.6	
	95	886.2	819.3	
Number of Values Period of Record		63	138	
		4/2/1996 – 12/11/2001	1/2/2002 - 10/22/2013	

If a TDS regime characterized by a 95<sup>th</sup> percentile value of 886 mg/L supports a high level of fisheries use attainment ("high fish species richness" as characterized by Buchanan et al. 2003), then a TDS regime characterized by a 95<sup>th</sup> percentile value of 860 mg/L should support a similar level of use attainment. Therefore, the proposed site-specific criterion of 860 mg/L in the Red River downstream of the mouth of the Little River can be expected to support the attainable fisheries designated use (based on fish populations) in the Red River.

From: Allan Gates [AGates@mwlaw.com]
Sent: Wednesday, April 27, 2016 5:41 PM

To: Honker, William

Cc: osbornec@adeq.state.ar.us; Garcia, David; Watson, Jane; Crocker, Philip; Kesler, Karen

Subject: RE: Domtar & SWEPCO Third Party Rulemaking Proceedings

Dear Bill,

I am writing to inquire about the status of EPA's review of the Domtar and SWEPCO third party rulemaking changes to the Red River and Little River water quality criteria for minerals and temperature in Arkansas.

As I recall, the technical representatives for EPA, ADEQ, and the companies agreed in their February meeting on a timeline for review that targeted a decision by EPA in early to mid-April. It is my understanding that some of the documents to be delivered to EPA ran a week or two late. I am hoping that EPA will be able to make its decision in the same amount of time it originally proposed, and that a decision will soon be available.

We recognize the Domtar change to the sulfate criterion will probably be denied based on downstream impacts in Louisiana, but we hope the remainder of the changes can be approved soon. In particular, the temperature change in the SWEPCO petition is very time sensitive.

Would you please let me know when you think the Agency will make its decision on these rulemaking proceedings?

Thank you again for your assistance.

Best regards,

Allan



#### **Allan Gates**

T 501.688.8816 | F 501.918.7816

agates@mwlaw.com | MitchellWilliamsLaw.com

425 W. Capitol Ave. | Ste. 1800 | Little Rock, AR 72201

Mitchell, Williams, Selig, Gates & Woodyard, P.L.L.C.

From: Honker, William [mailto:honker.william@epa.gov]

Sent: Wednesday, March 02, 2016 8:32 AM

To: Allan Gates

Cc: osbornec@adeq.state.ar.us; Garcia, David; Watson, Jane; Crocker, Philip; Kesler, Karen

Subject: RE: Domtar & SWEPCO Third Party Rulemaking Proceedings

Allan,

Thanks for the feedback on the recent meeting. Sorry for the delay in responding to your question.

With regard to the portion of the Domtar petition that applies to the change in the sulfate criterion for the lower reach of the Red River, our proposed procedural path would be to deny this aspect of the petition with the clear rationale that it would not be protective of the downstream standard in LA. We could reconsider such a request if and when the LA standard were revised (with our approval) to a value consistent with the request.

Please feel free to contact me if you'd like to discuss further.

Bill

William K. Honker, P.E.
Director, Water Division (6WQ)
EPA Region 6
Dallas, TX
214-665-3187
http://www.epa.gov/region6/water/index.htm



From: Allan Gates [mailto:AGates@mwlaw.com]

Sent: Friday, February 19, 2016 6:09 PM

To: Honker, William

Cc: osbornec@adeq.state.ar.us

Subject: Domtar & SWEPCO Third Party Rulemaking Proceedings

Dear Bill,

This morning members of your staff met in Little Rock with ADEQ staff and consultants representing Domtar and SWEPCO. I am told the meeting was very productive. It is my understanding everyone involved reached agreement on how the questions raised by EPA would be answered. It is also my understanding the participants reached agreement on a specific schedule for bringing EPA's review of these rulemakings to a point where a decision can be made.

I want to thank you on behalf of Domtar and SWEPCO for making this matter a significant priority for your staff's attention.

I also want to compliment your staff for the cooperative and professional manner in which they helped bring the remaining questions into clear focus.

The discussions today did include one question on which I would like your thoughts. The computer modeling for the Domtar and SWEPCO rulemaking proceedings indicates that the change in the sulfate criterion in the lower reach of the Red River in Arkansas will result in a predicted exceedance of the sulfate standard immediately downstream in Louisiana. Domtar candidly acknowledged this issue in its papers, and the company plans to take the issue up with appropriate officials in Louisiana.

Here is my question for you: What is the best procedural path for dealing with the Arkansas part of the Domtar petition? Is it best to deny the petition with the understanding that it will be reopened after Louisiana takes action? Would it be better to postpone final action on the Domtar petition until Louisiana has acted? I imagine this type of question has come up in the past. We would welcome your thoughts on how things should proceed.

For our part, if the downstream impact predicted in Louisiana is the only reason the Domtar petition cannot be approved by EPA at the present time, we would like EPA's action to make that point clear. Then Domtar could proceed to address the downstream effects in Louisiana secure in the knowledge that everything else about the changes in Arkansas have been resolved.

I want to thank you and your staff once again for the prompt and professional manner in which the Domtar and SWEPCO rulemaking review has been handled by you and your staff.

Best regards,

Allan



#### **Allan Gates**

T 501.688.8816 | F 501.918.7816

agates@mwlaw.com | MitchellWilliamsLaw.com

425 W. Capitol Ave. | Ste. 1800 | Little Rock, AR 72201

Mitchell, Williams, Selig, Gates & Woodyard, P.L.L.C.

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From:

Kesler, Karen [Kesler.Karen@epa.gov]

Sent:

Thursday, April 28, 2016 8:29 AM

To: Cc: Jim Malcolm Crocker, Philip

Subject:

quick questions on the SWEPCO temperature regressions

Hi Jim,

I am currently reviewing the additional material on the SWEPCO/Domtar submissions and I had a few quick follow up questions. I was wondering if you could send me the p values and the results of the normality tests on the residuals for the regressions that were used for determining the temperature criteria. Also, were any data transformations performed?

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Jim Malcolm [jtm@ftn-assoc.com]
Sent: Thursday, April 28, 2016 9:06 AM

To: 'Kesler, Karen'

Cc: 'Crocker, Philip'; James Malcolm

Subject: RE: quick questions on the SWEPCO temperature regressions

Hi Karen.

Thanks for looking at this stuff! I hope you had a good vacay – we did the Hawaii cruise several years ago and loved it.

I forwarded your email to Phil who was our lead on that part of the work and he will reply very quickly. I asked him to also volunteer to call/discuss any details of this when he sends it if that is something you might find helpful or that would facilitate your review.

Again, thanks very much for your review and the contact!

Jim



#### Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Thursday, April 28, 2016 8:29 AM

**To:** Jim Malcolm **Cc:** Crocker, Philip

Subject: quick questions on the SWEPCO temperature regressions

Hi Jim,

I am currently reviewing the additional material on the SWEPCO/Domtar submissions and I had a few quick follow up questions. I was wondering if you could send me the p values and the results of the normality tests on the residuals for the regressions that were used for determining the temperature criteria. Also, were any data transformations performed?

Thanks,

# Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Kesler, Karen [Kesler.Karen@epa.gov]
Sent: Thursday, April 28, 2016 9:11 AM

To: Jim Malcolm

Subject: RE: quick questions on the SWEPCO temperature regressions

#### Thanks Jim,

I appreciate the quick response. Yes, the vacation was great, now back to reality. I don't think a follow up discussion is necessary at this time, but if something else comes up, I'll be sure to reach out.

#### - Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]

Sent: Thursday, April 28, 2016 9:06 AM
To: Kesler, Karen < Kesler. Karen@epa.gov >

Cc: Crocker, Philip < crocker.philip@epa.gov >; James Malcolm < jtm@ftn-assoc.com >

Subject: RE: quick questions on the SWEPCO temperature regressions

Hi Karen.

Thanks for looking at this stuff! I hope you had a good vacay – we did the Hawaii cruise several years ago and loved it.

I forwarded your email to Phil who was our lead on that part of the work and he will reply very quickly. I asked him to also volunteer to call/discuss any details of this when he sends it if that is something you might find helpful or that would facilitate your review.

Again, thanks very much for your review and the contact!

Jim



#### Jim Malcolm

jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Thursday, April 28, 2016 8:29 AM

**To:** Jim Malcolm **Cc:** Crocker, Philip

Subject: quick questions on the SWEPCO temperature regressions

Hi Jim,

I am currently reviewing the additional material on the SWEPCO/Domtar submissions and I had a few quick follow up questions. I was wondering if you could send me the p values and the results of the normality tests on the residuals for the regressions that were used for determining the temperature criteria. Also, were any data transformations performed?

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Philip Massirer [phm@ftn-assoc.com]
Sent: Philip Massirer [phm@ftn-assoc.com]
Friday, April 29, 2016 10:06 AM

To: 'Karen Kesler'

Cc: 'Phil Crocker'; 'Jim Malcolm'; 'Pat Downey'

Subject: RE: quick questions on the SWEPCO temperature regressions

Karen,

Thanks so much for reviewing our latest submittal for SWEPCO and for sending us questions right away. The information you requested is provided below.

As a quick reminder, the regression analysis was only one of the four lines of evidence used to develop the proposed temperature criterion:

- observed water temperatures in the Little River upstream of SWEPCO's discharge,
- the basis for the existing criterion (30°C) for the Little River (the ecoregion criterion was developed using data from small shaded streams),
- · existing criteria for upstream and downstream waterbodies (large unshaded waterbodies), and
- the temperature regression analysis.

The p values that Excel calculated for the first regression (the one shown at the top of page 4-2 in the September 2014 report) were:

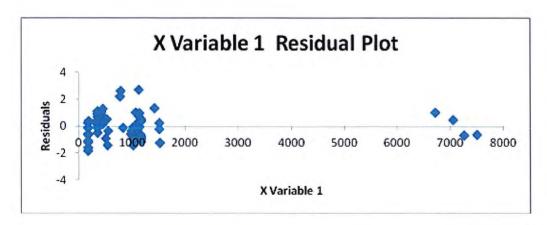
	P value
Intercept	0.00018
Coeff. for Q (X variable 1)	0.00026
Coefficient for $T_{A,0}$ (X variable 2)	0.00322
Coefficient for T <sub>A,1</sub> (X variable 3)	0.87515
Coefficient for T <sub>A,2</sub> (X variable 4)	0.03134

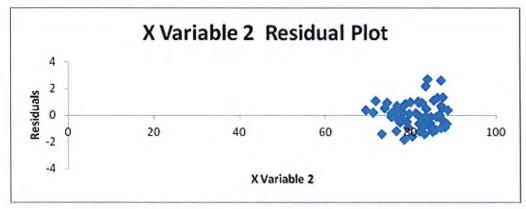
The p values that Systat calculated for the second regression (the one shown on pages 4-3 and 4-4 in the September 2014 report) were:

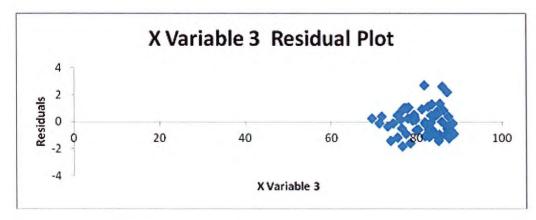
	P value
Intercept	<0.0005*
Coeff. for T <sub>SW</sub> (X variable 1)	<0.0005*
Coefficient for T <sub>AR,0</sub> (X variable 2)	<0.0005*
Coefficient for $T_{AR,1}$ (X variable 3)	0.068
Coefficient for T <sub>AR,2</sub> (X variable 4)	<0.0005*

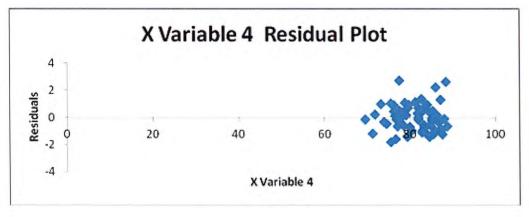
<sup>\*</sup>Systat only reported the p values to three decimal places; these were reported as "0.000".

We did not test the residuals for normality but we reviewed the four residual plots that Excel generated for the first regression (shown below). No data transformations were performed because that is not typically what we do for temperature analyses.









The weight of evidence approach that we used for the temperature criterion is similar to what we have done in other projects. A critically important requirement of the regressions is they need to accurately predict high water temperatures. As noted in Attachment 4 of our March 7 submittal to ADEQ, both regressions underpredicted high water temperatures. In fact, when observed water temperatures were 32.0°C or above, the two regressions under-predicted the temperature 83% and 100% of the time, respectively; therefore, these regressions are conservative. These under-predictions, when combined with the other lines of evidence mentioned above, provide a weight-of-evidence approach that supports the proposed 32.0°C criterion.

If you have any concerns about the temperature regressions, please let me know and we can discuss it over the phone. Thanks!

Philip Massirer 501-225-7779

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Thursday, April 28, 2016 8:29 AM

**To:** Jim Malcolm **Cc:** Crocker, Philip

**Subject:** quick questions on the SWEPCO temperature regressions

Hi Jim,

I am currently reviewing the additional material on the SWEPCO/Domtar submissions and I had a few quick follow up questions. I was wondering if you could send me the p values and the results of the normality tests on the residuals for the regressions that were used for determining the temperature criteria. Also, were any data transformations performed?

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From:

Philip Massirer [phm@ftn-assoc.com]

Sent:

Friday, April 29, 2016 11:19 AM

To: Cc: 'Jim Malcolm' 'Pat Downey'

Subject:

FW: quick questions on the SWEPCO temperature regressions

Jim - see Karen's response below.

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

**Sent:** Friday, April 29, 2016 10:12 AM

To: Philip Massirer

Subject: RE: quick questions on the SWEPCO temperature regressions

Hi Phil,

Thank you so much for getting back to me quickly. This is helpful in considering the accuracy of the regression. I am considering them, just as you pointed out, as one piece of the justification for the temperature criteria. I'll let you know if I have any additional questions.

Thanks, Karen

From: Philip Massirer [mailto:phm@ftn-assoc.com]

**Sent:** Friday, April 29, 2016 10:06 AM **To:** Kesler, Karen < Kesler. Karen@epa.gov >

Cc: Crocker, Philip < crocker.philip@epa.gov >; 'Jim Malcolm' < itm@ftn-assoc.com >; 'Pat Downey' < pjd@ftn-assoc.com >

**Subject:** RE: quick questions on the SWEPCO temperature regressions

Karen,

Thanks so much for reviewing our latest submittal for SWEPCO and for sending us questions right away. The information you requested is provided below.

As a quick reminder, the regression analysis was only one of the four lines of evidence used to develop the proposed temperature criterion:

- observed water temperatures in the Little River upstream of SWEPCO's discharge,
- the basis for the existing criterion (30°C) for the Little River (the ecoregion criterion was developed using data from small shaded streams),
- existing criteria for upstream and downstream waterbodies (large unshaded waterbodies), and
- the temperature regression analysis.

The p values that Excel calculated for the first regression (the one shown at the top of page 4-2 in the September 2014 report) were:

	P value
Intercept	0.00018

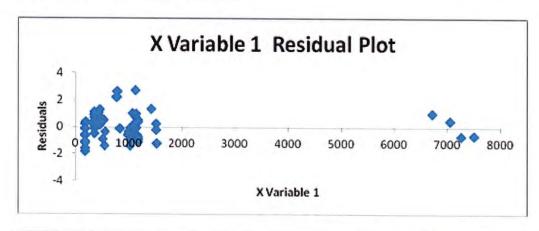
Coeff. for Q (X variable 1)	0.00026
Coefficient for $T_{A,0}$ (X variable 2)	0.00322
Coefficient for $T_{A,1}$ (X variable 3)	0.87515
Coefficient for T <sub>A,2</sub> (X variable 4)	0.03134

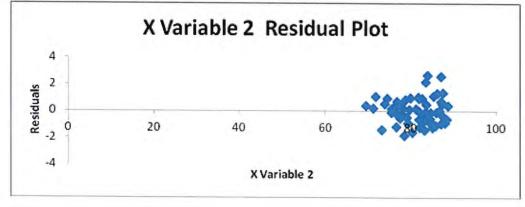
The p values that Systat calculated for the second regression (the one shown on pages 4-3 and 4-4 in the September 2014 report) were:

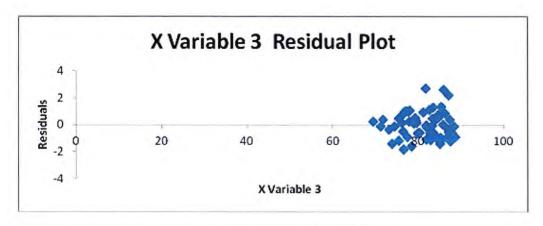
	P value
Intercept	<0.0005*
Coeff. for T <sub>SW</sub> (X variable 1)	<0.0005*
Coefficient for $T_{AR,0}$ (X variable 2)	<0.0005*
Coefficient for $T_{AR,1}$ (X variable 3)	0.068
Coefficient for T <sub>AR,2</sub> (X variable 4)	<0.0005*

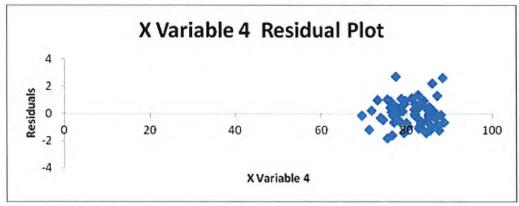
<sup>\*</sup>Systat only reported the p values to three decimal places; these were reported as "0.000".

We did not test the residuals for normality but we reviewed the four residual plots that Excel generated for the first regression (shown below). No data transformations were performed because that is not typically what we do for temperature analyses.









The weight of evidence approach that we used for the temperature criterion is similar to what we have done in other projects. A critically important requirement of the regressions is they need to accurately predict high water temperatures. As noted in Attachment 4 of our March 7 submittal to ADEQ, both regressions underpredicted high water temperatures. In fact, when observed water temperatures were 32.0°C or above, the two regressions under-predicted the temperature 83% and 100% of the time, respectively; therefore, these regressions are conservative. These under-predictions, when combined with the other lines of evidence mentioned above, provide a weight-of-evidence approach that supports the proposed 32.0°C criterion.

If you have any concerns about the temperature regressions, please let me know and we can discuss it over the phone. Thanks!

Philip Massirer 501-225-7779

From: Kesler, Karen [mailto:Kesler, Karen@epa.gov]

Sent: Thursday, April 28, 2016 8:29 AM

**To:** Jim Malcolm **Cc:** Crocker, Philip

**Subject:** quick questions on the SWEPCO temperature regressions

Hi Jim,

I am currently reviewing the additional material on the SWEPCO/Domtar submissions and I had a few quick follow up questions. I was wondering if you could send me the p values and the results of the normality tests on the residuals for the regressions that were used for determining the temperature criteria. Also, were any data transformations performed?

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Leann Kret [lkret@mwlaw.com] on behalf of Allan Gates [AGates@mwlaw.com]

Sent: Tuesday, May 24, 2016 11:28 AM

To: Brian Bond; Elizabeth Gunter; TimothyGross; Frank Mills; pjd@ftn-assoc.com; Jim Malcolm

Cc: Allan Gates; Marcy Taylor Subject: SWEPCO Letter and TSD

Attachments: SWEPCO TSD.pdf; SWEPCO Letter.pdf

All:

Allan is out of the office but asked that I forward the attached to you as quickly as possible for your review.

If there are any questions, please do not hesitate to let us know.

Thank you,

Leann Kret



Leann Kret | Assistant to Allan Gates, Chris Barrier, and Marcella Taylor T 501.370.4216 | F 501.918.7216 | Kret@mwlaw.com | MitchellWilliamsLaw.com 425 W. Capitol Ave. | Ste. 1800 | Little Rock, AR 72201 Mitchell, Williams, Selig, Gates & Woodyard, P.L.L.C.

From: Osborne, Caleb [mailto:osbornec@adeq.state.ar.us]

Sent: Tuesday, May 24, 2016 8:58 AM

To: Allan Gates

Cc: Blanz, Bob; Carpenter, Ellen; Clem, Sarah

Subject: SWEPCO Letter and TSD

Allan,

We have received EPA's decision on the SWEPCO Third Party Rulemaking. Attached is an electronic version of EPA's decision letter and TSD. Let me know if you have any questions. I know there is interest in moving ahead on the permitting front. Once you have had a chance to review, let me know how you all wish to proceed.

Thanks

Caleb

system without copying it, and notify the sender by reply e-mail or by calling (501) 688-8800 Little Rock, AR (479) 464-5650 Rogers, AR (512) 480-5100 Austin, TX (267) 757-8780 Newtown, PA or (870) 336-9292 Jonesboro, AR so that our address record can be corrected.



### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY **REGION 6** 1445 ROSS AVENUE, SUITE 1200

DALLAS, TEXAS 75202 - 2733

May 16, 2016

Mr. Caleb Osborne Associate Director – Water Division Arkansas Department of Environmental Quality 5301 Northshore Drive Little Rock, AR 72118-5317

Re: Final submittal of Regulation No. 2, as amended, Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas

Dear Mr. Osborne:

The Environmental Protection Agency (EPA) has completed its review of the amendments to Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas that were made in relation to the Third Party Rulemaking process initiated by Southwestern Electric Power Company (SWEPCO). The amended Regulation No. 2 was adopted by the Arkansas Pollution Control and Ecology Commission (APC&EC) on October 23, 2015 and became effective on November 5, 2015. It was submitted to the EPA for approval on December 21, 2015 by the Arkansas Department of Environmental Quality (ADEQ).

At this time EPA is approving the majority of the new and revised provisions to Regulation 2.511 and Appendix A of Regulation No. 2, including the site specific criteria changes for temperature and TDS in the Little River and the removal of the domestic water supply use in the Red River. These revisions are approved pursuant to the Clean Water Act (CWA) § 303(c) and its implementing regulations at 40 CFR Part 131. However, EPA is taking no action on the TDS criterion revision in the Red River. EPA has continuing concerns about appropriate demonstration of aquatic life use protection and protection of downstream uses in Louisiana. Consideration of these issues require further review by EPA management at this time. Once these issues have been sufficiently reviewed, EPA will issue an action on this portion of the SWEPCO submission. Please note that under 40 CFR § 313.21(c), new and revised standards do not go into effect for CWA purposes until approved by EPA. Therefore, previously approved language associated with the TDS criteria in the January 24, 2008 version of the Arkansas water quality standards (WQS) remains in effect for CWA purposes.

The approval of the new and revised water quality standards are subject to the results of consultation under section 7(a)(2) of the Endangered Species Act (ESA). Section 7(a)(2) requires that federal agencies consult with the U.S. Fish and Wildlife Service, as appropriate, to ensure that actions they take, fund, or authorize are not likely to jeopardize the continued existence of listed species or result in the adverse modification or destruction of habitat. As of today, this consultation has not been completed. By approving the WQS revisions "subject to the results of consultation under section 7(a)(2) of the Endangered Species Act," EPA retains the full range of options under CWA § 303(c) for ensuring that WQS are environmentally protective. EPA retains the discretion to revise its approval decision of these new and revised WQS if the consultation identifies deficiencies in the WQS that require remedial action.

I appreciate the APC&EC's and the ADEQ's effort in the review of these revised provisions of the State's standards and also appreciate ADEQ's assistance with coordinating meetings and correspondence with the third party. If you have any questions or concerns, please contact me at 214-665-3187, or contact Karen Kesler at 214-665-3185.

Sincerely,

William K. Honker, P.E.

Director

Water Division

Enclosure

cc: Sarah Clem, Branch Manager

Water Division ADEQ

## **TECHNICAL SUPPORT DOCUMENT:**

EPA REVIEW OF SITE-SPECIFIC CRITERIA AND USE REVISIONS TO REGULATION 2: REGULATION ESTABLISHING WATER QUALITY STANDARDS FOR SURFACE WATERS OF THE STATE OF ARKANSAS FOR THE LITTLE RIVER AND RED RIVER, ARKANSAS

Revisions Adopted by the Arkansas Pollution Control and Ecology Commission via Minute Order No. 15-21 for Southwestern Electric Power Company

> U.S. EPA REGION 6 WATER DIVISION May 2016

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#### I. Introduction

#### Background

As described in §303(c) of the Clean Water Act (CWA) and in the standards regulation within the Code of Federal Regulations (CFR) at 40 CFR §131.20, states and authorized tribes have primary responsibility to develop and adopt water quality standards to protect their waters. State and tribal water quality standards consist of three primary components: beneficial uses, criteria to support those uses, and an antidegradation policy. In addition, CWA §303(c)(1) and 40 CFR §131.20 require states to hold public hearings at least once every three years to review and, as appropriate, modify and adopt standards.

Under 40 CFR §131.21, EPA reviews new and revised surface water quality standards that have been adopted by states and authorized tribes. Authority to approve or disapprove new and/or revised standards submitted to EPA for review has been delegated to the Water Division Director in Region 6. Tribal or state water quality standards are not considered effective under the CWA until approved by EPA.

The purpose of this Technical Support Document (TSD) is to provide the basis for the Environmental Protection Agency's (EPA) actions on water quality standards revisions to Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas adopted by the Arkansas Pollution Control and Ecology Commission (APC&EC) in Minute Order 15-21 and further described in the subsection below titled "Summary of Revised Provisions."

## Chronology of Events

September 11, 2014	A third party, Southwestern Electric Power Company (SWEPCO), filed a petition with the APC&EC to amend Regulation No. 2
September 26, 2014	The APC&EC initiated the rulemaking proceedings via a Minute Order No. 14-33
October 1, 2014	Public notice of the proposed rule-making was published
November 17, 2014	Public hearing on the proposed rule-making was held in Hope, Arkansas
December 3, 2014	Public comment period ended on the proposed changes to Regulation No. 2
October 23, 2015	Becky Keogh, Director, Arkansas Department of Environmental Quality (ADEQ), signed Minute Order No. 15-21 adopting changes to Regulation No. 2

December 21, 2015

William K. Honker, Director, Water Division, EPA
Region 6, received letter from, Ellen Carpenter Water
Division Chief, ADEQ requesting EPA approval of the
adopted revisions and transmitting the water quality
standards submission package

February 9, 2016

EPA Region 6 issues a letter to ADEQ stating that no
action would be taken on the submission package to allow
the third party time to respond to EPA's comments

March 30, 2016

ADEQ submits additional information prepared by the
third party to EPA

#### Summary of Revised Provisions

By letter dated December 21, 2015, ADEQ submitted several water quality standards revisions adopted by the APC&EC via Minute Order No. 15-21 to EPA for review and approval. These revisions are located in Regulation 2.511 and Appendix A of Regulation No. 2. They include revisions of temperature (30°C to 32°C) and total dissolved solids (TDS) (100 mg/L to 138 mg/L) criteria for the Little River from Millwood Lake to its confluence with the Red River. They also include the removal of the domestic water supply use and the increase of the TDS criterion from 500 mg/L to 860 mg/L for the Red River from the mouth of the Little River to the Arkansas (AR)/Louisiana (LA) state line.

# II. Revised Provisions EPA is Approving

In accordance with the requirements found in Regulation No. 2.306 of the Arkansas Water Quality Standards, SWEPCO contracted with FTN Associates, Ltd., to complete a use attainability analysis (UAA) of the receiving waterbodies and a technical justification for the site specific criteria. The purpose of the UAA and technical justification were to provide scientific justification to support revised site-specific water quality criteria for temperature and TDS, as well as a domestic water supply use removal, for these waterbodies.

## Site-specific Water Quality Criterion for TDS in the Little River

**Table 1.** Site-specific water quality criterion revision for TDS in the Little River submitted by ADEQ to EPA for review and approval.

Reach Description	Current Criterion	Proposed Criterion
Little River from Millwood Lake to the Red River	100 mg/L	138 mg/L

#### Criterion Derivation

The TDS criterion for the Little River was developed by determining a value that would not be considered a "significant modification" from the ecoregion (ER) value by ADEQ, as defined in Regulation No.2. This value was the sum of 1/3 ER chloride criterion, 1/3 ER sulfate criterion, and the ER TDS criterion which totaled 138 mg/L. In addition, a mass balance model

was developed that both incorporated the ambient TDS concentrations in the Little River as well as the effluent TDS concentration from the SWEPCO plant. With these two input factors, the mass balance determined a site-specific criterion of 136.1 mg/L would be necessary for the plant to operate without causing impairments. Given that the mass balance value and the calculated value were very similar, the value of 138 mg/L was selected as the criterion that was proposed in this third party rulemaking.

### **Use Support Justification**

The following designated uses apply for the Little River from Millwood Lake to its confluence with the Red River: primary contact recreation, secondary contact recreation, domestic water supply, industrial water supply, agricultural water supply, and aquatic life. Of these uses, the Gulf Coastal Plain ecoregion aquatic life use is the most sensitive, and as such will be the focus of the following discussion of use support. According to the Arkansas Regulation No. 2, the aquatic life use "provides for the protection and propagation of fish, shellfish and other forms of aquatic biota."

A weight-of-evidence approach was taken to provide the justification for the support of the aquatic life use in the Little River, including field studies, a toxicity study, and a literature review of mineral impacts on aquatic life. Two groups of organisms, fish and benthic macroinvertebrates, were considered for the evaluation of protection of aquatic life use.

The benthic macroinvertebrate population of the Little River appeared to already be an impacted community. This impact is likely due to the presence of the dam that created Millwood Lake, upstream of this section of the Little River. When organisms were classified down to genus, only 11 unique taxa were found, with the predominant proportion consisting of Chironomidae (83%). Only 11.5 % of the community was made up of Ephemeroptera, Plecoptera, and Trichoptera (EPT) taxa. As most of this community is dominated by Chironomidae, a family that is generally pollution tolerant, it is not anticipated that these organisms will be very sensitive to changes in TDS concentration. Along with the assessment of the community in the field, a toxicity study was also completed with Ceriodaphnia dubia. These results found a no effect concentration of 1,260 for TDS (hardness in the range of 150-215 mg/L, chloride 171 mg/L, sulfate 432 mg/L) and an IC25 of 1,744 mg/L (hardness ~150-215 mg/L, chloride 232 mg/L, sulfate 600mg/L). While this indicates that a TDS of 138 mg/L will be supportive of the use, toxicity results need to be viewed only as a piece of the rationale, rather than the entire story. Toxicity studies are limited by their assessment of only one species and by the fact that they can't match the exact conditions of the field. C. dubia, while the typical species used for toxicity tests, is one of the less sensitive species when it comes to minerals toxicity. In addition, minerals toxicity is affected by a number of variables including the ionic ratio and the hardness of the water, making the determination of the toxicity of minerals challenging.

Given these limitations of laboratory studies, the protectiveness of a TDS of 138 mg/L for benthic macroinvertebrates for the Little River was also evaluated by utilizing the results from a field based study conducted by the EPA (2011). This study used conductivity as a measure to assess minerals and determined a benchmark conductivity value where 95 % of the genera in the community would be protected from extermination. This study was conducted in

high gradient streams in the Appalachian Region that were dominated by  $Ca^{2+}$ ,  $Mg^{2+}$ ,  $SO_4^{2-}$ ,  $HCO_3^-$  salts. These salts also dominate the Little River, but the Little River is located in a non-comparable ecoregion. The streams in the 2011 EPA study also had a greater number of EPT taxa than the Little River. Even though these streams varied geographically from each other, the results from the 2011 EPA study can still be used as a point of reference on toxicity. The 2011 EPA study found a benchmark value of 300  $\mu$ S. When 138 mg/L TDS was converted to a conductivity value for the Little River, it was estimated to be 218  $\mu$ S. This value falls below the 300  $\mu$ S benchmark found in the 2011 EPA study, adding support to the protectiveness of this criteria.

The fish community in the Little River was also considered when determining if the revised TDS criterion supported the aquatic life use. Field studies conducted in 2013 collected a total of 40 fish species in the Little River. Twenty-seven of those fish species could be assigned a tolerance classification according to Meador and Carlisle 2007 and 23 were classified as "tolerant" or "moderately tolerant." These classifications indicate that they are less likely to be sensitive to changes in TDS. Also, of the 40 fish species that were collected in the Little River, 37 of them were also found in the Red River in a study by Buchanan et al. 2003, which surveyed the fish population in the Arkansas portion of the Red River from 1996 to 2001. The 50<sup>th</sup> percentile of monitoring data collected at Garland, AR, was 464 mg/L during this study period, demonstrating that most of these fish species were living in a habitat with a much higher TDS than the criterion proposed for the Little River. This supports the rationale that the TDS criterion of 138 mg/L is protective of the aquatic life use.

#### Action

Based upon the information provided above that demonstrate that the TDS criterion revision will have minimal impact on aquatic life, EPA approves the revision of the TDS criterion from 100 mg/L to 138 mg/L for the reach of the Little River from Millwood Lake to the confluence with the Red River.

#### Site-specific Water Quality Criterion for Temperature in the Little River

**Table 2.** Site-specific water quality criterion revision for temperature in the Little River submitted by ADEQ to EPA for review and approval.

	Reach Description		Proposed Criterion
ĺ	Little River from Millwood Lake to the Red River	30 ° C	32 ° C

## **Criterion Derivation**

The Little River below Millwood Lake experiences temperatures near or higher than 32°C during summer months predominantly due to the geomorphic qualities of the lake and the river. Millwood Lake is a relatively shallow reservoir with an average depth of 1.5 m which leads to the entire water column of this lake increasing in temperature relatively quickly in summer months. The maximum depth measured near the old thalweg was 7 m. Therefore the

water that is released from this lake, even if it is from lower depths, is warm in summer months and can increase the temperature downstream in the Little River. In addition, the Little River is a wide river, with little overhead tree cover for the central section of the river. Its typical width is between 61 m and 122 m, resulting in minimal overhead cover for the center of the river. This exposure to direct sunlight leads to greater temperatures in the river. The ecoregion temperature value that is currently assigned to the river was based on background values of least disturbed streams in the ecoregion that had an average width of 7 m and average canopy cover of 84%. Given the difference in the geomorphology of the Little River from these reference streams and the impacts from the dam's presence, a higher temperature criterion value appears to be appropriate for this river. This is evidenced by daily maximum temperatures measured in the river from April to October 2012. The 95th percentile of temperature values upstream and downstream of SWEPCO were 32.11°C (89.8°F) and 32.5°C (90.5°F), respectively. In 2013 (Feb to Oct) the 95<sup>th</sup> percentile of the data ranged from 30.1°C (86.2°F) to 31°C (87.8°F) upstream of the SWEPCO effluent discharge. Both of these measurement periods demonstrate exceedances of the current criterion of 30°C. Twenty-six percent of both the data upstream and downstream of SWEPCO were found to exceed the current 30°C criteria from April to October 2012 and February to October 2013. A multiple linear regression was developed by FTN that used daily stream flow rates and daily average air temperatures in order to predict daily maximum water temperatures. Several variations of the parameters were tested to determine the best regression model and the model that had the lowest standard error of the estimate was selected. The regression was utilized to create a 5 year data set (April 2008 to March 2013) of daily maximum water temperatures. The 95<sup>th</sup> percentile of this data set was then determined to be 32.0°C and is the criteria value that is being proposed.

# **Use Support Justification**

Primary contact recreation, secondary contact recreation, domestic water supply, industrial water supply, agricultural water supply, and aquatic life uses apply to the Little River from Millwood Lake to its confluence with the Red River. The Gulf Coastal Plain ecoregion aquatic life use is the most sensitive, and as such will be the focus of the following discussion of use support. According to the Arkansas Regulation No. 2, the aquatic life use "provides for the protection and propagation of fish, shellfish and other forms of aquatic biota."

Assessment of benthic macroinvertebrates in the Little River showed a community that was likely already impacted by habitat factors. This river is wide with little overhead cover, where the predominant habitat for macroinvertebrates is woody debris. The river substrate is predominantly sand, with very few patches of gravel or cobble. In addition, the dam at the head of this reach of the Little River is also likely impacting flow and habitat qualities of this river which will in turn impact the benthic macroinvertebrates. The macroinvertebrate population appeared to already be an impacted community as when organisms were classified down to genus, only 11 unique taxa were found, with the predominant proportion consisting of Chironomidae (83%). Only 11.5 % of the community was made up of EPT taxa. As this community is dominated by Chironomidae, it is not anticipated that these organisms will be very sensitive to 2°C temperature change.

The fish community in the Little River was also assessed to determine whether it would be impacted by the increase in temperature criteria. Forty fish species were found in the Little River either by surveys conducted by FTN in 2013 or by the United States Geological Survey in 2009. Of the 40 fish species found, 28 of them had their temperature tolerance classified in Meador and Carlisle 2007. Of the 28 species, 2 were found to be intolerant, while the other 26 were "moderately tolerant" or "tolerant." In addition, 37 of the 40 fish found in the Little River were also found in the Red River, which has a temperature criterion of 32°C. The maximum mean temperatures exceeded 32°C in the Red River 11 % of the time from October 2003 to June 2011. The presence of these fish species in the Red River adds further support to the demonstration that these fish species can tolerate this higher temperature criteria with little impact.

#### Action

Based upon the information provided above, that demonstrates that the temperature criterion revision will have minimal impact on aquatic life and match background temperature conditions, EPA approves the revision of temperature criterion from 30°C to 32°C for the reach of the Little River from Millwood Lake to the confluence with the Red River.

## Removal of Domestic Water Supply Use in the Red River

This third party rulemaking is proposing the removal of the domestic water supply use from the Red River from its confluence with the Little River down to the Arkansas-Louisiana state line.

#### **Use Removal Justification**

In accordance with the requirements found in Regulation No. 2.306 of the Arkansas water quality standards, SWEPCO contracted with FTN Associates, Ltd. to complete a Use Attainability Analysis (UAA) for the Red River from the confluence of the Little River down to the Arkansas-Louisiana State Line. One of the things evaluated by the UAA was whether the domestic water supply designated use was an existing and/or attainable use for the Red River from the confluence with the Little River down to the Arkansas-Louisiana state line. The UAA study concluded that the domestic water supply designated use was neither an existing, nor an attainable, designated use and recommended its removal.

A letter dated December 8, 2013 from the Arkansas Natural Resources Commission provided along with the UAA study states that there are "no existing or planned public water supply uses documented for this reach of the Red River" and that "removal of the domestic water supply use designation does not conflict with the Arkansas Water Plan at this time." In addition, a letter dated November 4, 2013 from the Arkansas Department of Health which was included with the UAA stated that "[t]he stream segment in question has not been approved, nor is it under consideration, for use as a public water system source. Therefore, the Engineering Section has no objection to this stream segment being removed from the *Domestic Water Supply Use* designation."

The letters from the Arkansas Natural Resources Commission and the Arkansas Department of Health affirm that the domestic water supply designated use is not an existing use on this reach of the Red River and the reach is not currently being considered for a domestic water supply source. In addition, the high mineral content of this water body, partially due to natural salt seeps upstream, further justifies that the domestic water supply use is an unattainable use. The minerals criteria for water bodies with a domestic water supply use in Arkansas is 500 mg/L TDS, 250 mg/L sulfate, and 250 mg/L chloride. Water quality monitoring data from the Red River at Fulton, just downstream of the Little River, show frequent exceedances of the 500 mg/L TDS criterion starting from the time that monitoring began in 1997. In addition the 75<sup>th</sup> percentiles of ADEQ monitoring data at Garland (about the midpoint of this reach) were 660.5 mg/L from April 1996 to December 2001 and 624.0 mg/L from January 2002 to October 2013. Monitoring data from the ADEQ monitoring site at Doddridge also showed that the 90<sup>th</sup> and 95<sup>th</sup> percentiles for TDS at this location were 657 mg/L and 770 mg/L, respectively, for a period of record of September 1990 to October 2013. These data demonstrate that the minerals levels in this reach of the Red River are often above 500 mg/L.

#### Action

Based upon the current and historic data presented in the UAA for the TDS in this reach of the Red River, along with the letters from the Arkansas Natural Resources Commission and the Arkansas Department of Health, EPA determines that the domestic water supply use is neither an existing, nor an attainable use in the Red River from its confluence with the Little River down to the Arkansas-Louisiana state line. Therefore, EPA approves the removal of the domestic water supply use from the reach of the Red River from its confluence with the Little River down to the Arkansas-Louisiana state line.

# III. Revised Provision upon which EPA is Taking No Action

## Site-specific Water Quality Criterion for TDS in the Red River

**Table 3.** Site-specific water quality criterion revision for TDS in the Red River submitted by ADEO to EPA for review and approval.

Reach Description	Current Criterion	Proposed Criterion
Red River from the confluence with the Little River to the AR-LA state line	500 mg/L	860 mg/L

#### No Action Rationale

EPA is currently taking no action on the site-specific TDS criteria change for the Red River from its confluence with the Little River down to the Arkansas-Louisiana state line. EPA has continuing concerns about demonstration of appropriate aquatic life use protection and protection of downstream uses in Louisiana. Consideration of these issues require further review by EPA at this time. Once these issues have been sufficiently reviewed, EPA will issue an action on this revised provision.

As specified in 40 CFR § 131.21(c), these revised standards do not go into effect for CWA purposes until approved by EPA. Therefore, the previously approved criteria of 500 mg/L remains in effect for CWA purposes.

## IV. References

Buchanan, T.M., D. Wilson, L.G. Claybrook, and W.G. Layher. 2003. Fishes of the Red River in Arkansas. *J. Ark. Acad. Sci.* 57:18-26.

Meador, M.R., and D.M. Carlisle. 2007. Quantifying tolerance indicator values for common fish species of the United States. *Ecological Indicators* 7:329-338.

U.S. EPA (Environmental Protection Agency). 2011. A Field-Based Aquatic Life Benchmark for Conductivity in Central Appalachian Streams. Office of Research and Development, National Center for Environmental Assessment, Washington, DC. EPA/600/R-10/023F.

From: Jim Malcolm [jtm@ftn-assoc.com]

Sent: Wednesday, June 01, 2016 8:58 AM

To: 'Kesler Karen': 'Clem Sarah'

To: 'Kesler, Karen'; 'Clem, Sarah'

Cc: Patrick J. Downey; 'Philip Massirer'; James Malcolm

Subject: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

#### Hi Karen, Sarah;

Karen, thanks again for the May 2016 TSD and transmittal letter from Bill (May 16, 2016) discussing the referenced proposed site specific criteria (SSC) from SWEPCO. We sincerely appreciate the thorough review and discussions on the proposed SSC as well as the meeting with technical staff from both ADEQ and EPA that greatly facilitated the agency review and our understanding of agency needs!

In the TSD (Section III), EPA noted that it was taking no action on approval of the SWEPCO TDS proposed SSC for the lower Red River. The TSD referenced a focus on aquatic life protection in Louisiana as the reason for the 'no action' decision. On behalf of SWEPCO (and Domtar which also has a TDS proposed SSC at EPA for review for upper Red River in AR upstream of the Little River inflow), we request a conference call to better understand EPA's concern regarding the proposed TDS SSCs.

If this request is agreeable to you guys, please provide some possible days/times for a conference call in the next week or two and we will circulate a day/time for confirmation and set up a call in number.

Many thanks, Jim



#### Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Kesler, Karen [Kesler.Karen@epa.gov]
Sent: Thursday, June 02, 2016 8:22 AM

To: Jim Malcolm; 'Clem, Sarah'; Crocker, Philip

Cc: Patrick J. Downey; 'Philip Massirer'

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hi Jim,

The remaining action on the SWEPCO rulemaking and the actions on the Domtar rulemaking are currently under review by EPA management. Once those actions have taken place and you have had a chance to review the associated TSDs we would be happy to talk with you and ADEQ.

Thanks, Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Wednesday, June 01, 2016 8:58 AM

To: Kesler, Karen < Kesler.Karen@epa.gov>; 'Clem, Sarah' < CLEM@adeq.state.ar.us>

Cc: Patrick J. Downey <pid@ftn-assoc.com>; 'Philip Massirer' <phm@ftn-assoc.com>; James Malcolm <jtm@ftn-

assoc.com>

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If this request is agreeable to you guys, please provide some possible days/times for a conference call in the next week or two and we will circulate a day/time for confirmation and set up a call in number.

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Jim Malcolm jtm@ftn-assoc.com FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From:

Jim Malcolm [jtm@ftn-assoc.com]

Sent:

Thursday, June 02, 2016 10:54 AM

To: Cc: 'Kesler, Karen'; 'Clem, Sarah'; 'Crocker, Philip'
'Patrick J. Downey'; 'Philip Massirer'; James Malcolm

Subject:

RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hey Karen, thanks, do you have any schedule estimate on those TSDs? Ballpark?

Many thanks, Jim

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

**Sent:** Thursday, June 02, 2016 8:22 AM **To:** Jim Malcolm; 'Clem, Sarah'; Crocker, Philip **Cc:** Patrick J. Downey; 'Philip Massirer'

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

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Sent: Wednesday, June 01, 2016 8:58 AM

To: Kesler, Karen < Kesler. Karen@epa.gov >; 'Clem, Sarah' < CLEM@adeq.state.ar.us >

Cc: Patrick J. Downey <pid@ftn-assoc.com>; 'Philip Massirer' <phm@ftn-assoc.com>; James Malcolm <<u>itm@ftn-assoc.com</u>>; James Malcolm <<u>itm@ftn-assoc.com</u>>; James Malcolm <<u>itm@ftn-assoc.com</u>>; James Malcolm < <u>itm@ftn-assoc.com</u>>; James Malcolm < <u>itm@ftn-asso</u>

assoc.com>

Subject: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

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## Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

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From: Kesler, Karen [Kesler.Karen@epa.gov]
Sent: Thursday, June 02, 2016 11:15 AM
To: Jim Malcolm; 'Clem, Sarah'; Crocker, Philip
Cc: 'Patrick J. Downey'; 'Philip Massirer'

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hi Jim,

We are hoping to get them out in the next week or two, but part of that is dependent on management schedule.

- Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Thursday, June 02, 2016 10:54 AM

To: Kesler, Karen < Kesler.Karen@epa.gov >; 'Clem, Sarah' < CLEM@adeq.state.ar.us >; Crocker, Philip

<crocker.philip@epa.gov>

Cc: 'Patrick J. Downey' <pid@ftn-assoc.com'>; 'Philip Massirer' <phm@ftn-assoc.com'>; James Malcolm <itm@ftn-assoc.com'>; James Malcolm

assoc.com>

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hey Karen, thanks, do you have any schedule estimate on those TSDs? Ballpark?

Many thanks, Jim

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

**Sent:** Thursday, June 02, 2016 8:22 AM **To:** Jim Malcolm; 'Clem, Sarah'; Crocker, Philip **Cc:** Patrick J. Downey; 'Philip Massirer'

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hi Jim,

The remaining action on the SWEPCO rulemaking and the actions on the Domtar rulemaking are currently under review by EPA management. Once those actions have taken place and you have had a chance to review the associated TSDs we would be happy to talk with you and ADEQ.

Thanks, Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Wednesday, June 01, 2016 8:58 AM

To: Kesler, Karen < Kesler. Karen@epa.gov >; 'Clem, Sarah' < CLEM@adeq.state.ar.us >

**Cc:** Patrick J. Downey < pjd@ftn-assoc.com >; 'Philip Massirer' < phm@ftn-assoc.com >; James Malcolm < jtm@ftn-

assoc.com>

Subject: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hi Karen, Sarah;

Karen, thanks again for the May 2016 TSD and transmittal letter from Bill (May 16, 2016) discussing the referenced proposed site specific criteria (SSC) from SWEPCO. We sincerely appreciate the thorough review and discussions on the proposed SSC as well as the meeting with technical staff from both ADEQ and EPA that greatly facilitated the agency review and our understanding of agency needs!

In the TSD (Section III), EPA noted that it was taking no action on approval of the SWEPCO TDS proposed SSC for the lower Red River. The TSD referenced a focus on aquatic life protection in Louisiana as the reason for the 'no action' decision. On behalf of SWEPCO (and Domtar which also has a TDS proposed SSC at EPA for review for upper Red River in AR upstream of the Little River inflow), we request a conference call to better understand EPA's concern regarding the proposed TDS SSCs.

If this request is agreeable to you guys, please provide some possible days/times for a conference call in the next week or two and we will circulate a day/time for confirmation and set up a call in number.

Many thanks, Jim



Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Jim Malcolm [jtm@ftn-assoc.com]
Sent: Jim Malcolm [jtm@ftn-assoc.com]
Thursday, June 02, 2016 3:47 PM

To: 'Kesler, Karen'; 'Clem, Sarah'; 'Crocker, Philip'
Cc: 'Patrick J. Downey'; 'Philip Massirer'; James Malcolm

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Great, thanks again Karen.

Rgds, Jim

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

**Sent:** Thursday, June 02, 2016 11:15 AM **To:** Jim Malcolm; 'Clem, Sarah'; Crocker, Philip **Cc:** 'Patrick J. Downey'; 'Philip Massirer'

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

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Sent: Thursday, June 02, 2016 10:54 AM

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<<u>crocker.philip@epa.gov</u>>

Cc: 'Patrick J. Downey' pjd@ftn-assoc.com; 'Philip Massirer' phm@ftn-assoc.com; James Malcolm <<pre>jtm@ftn-

assoc.com>

Subject: RE: AR Rulemaking for proposed Red River TDS criteria (SWEPCO and Domtar proposals)

Hey Karen, thanks, do you have any schedule estimate on those TSDs? Ballpark?

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From: Kesler, Karen [mailto:Kesler, Karen@epa.gov]

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Cc: Patrick J. Downey; 'Philip Massirer'

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To: Kesler, Karen < Kesler.Karen@epa.gov>; 'Clem, Sarah' < CLEM@adeq.state.ar.us>

Cc: Patrick J. Downey <pid@ftn-assoc.com>; 'Philip Massirer' <phm@ftn-assoc.com>; James Malcolm <jtm@ftn-

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Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Sent: Kesler, Karen [Kesler.Karen@epa.gov] Wednesday, June 22, 2016 3:05 PM

To:

Jim Malcolm Crocker, Philip

Cc: Subject:

SWEPCO TDS supplemental information

Hi Jim,

I wanted to touch base and see when we should be expecting the additional information for the SWEPCO rulemaking in light of our meeting on June 10<sup>th</sup>. As this rulemaking has already been largely extended beyond the normal time frame, we would like to move forward with the final piece of rulemaking as soon as possible. Phil is pushing to get this action completed quickly, so we would prefer to receive the additional information by next Friday, July 1<sup>st</sup>.

Please let us know when you expect to send it along to ADEQ. I will be out of the office until June 30<sup>th</sup>, so please cc Phil with your response so he knows where things stand.

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From:

Jim Malcolm [jtm@ftn-assoc.com] Wednesday, June 29, 2016 3:40 PM

Sent: To:

'Kesler, Karen'

Cc:

Subject:

'Crocker, Philip'; James Malcolm RE: SWEPCO TDS supplemental information

Attachments:

TV summary from AQUIRE query.xlsx; tolerance values.docx; TM-K Kessler 2016-06-29.pdf

Karen, Phil,

Attached per your request are the following:

- 1. A technical memo describing macroinvertebrate data collected from the Red River upstream and downstream of the mouth of the Little River during October 2014;
- 2. A Word doc describing the results of a preliminary effort to develop numerical tolerance values for minerals based on laboratory toxicity data;
- 3. The spreadsheet referenced in the Word doc.

The development of the numerical tolerance values is preliminary. If you all think the information looks to be useful, we can refine the approach and provide more documentation of the methodology.

I am out of the office today, but I will be in tomorrow and Friday. Feel free to call me if you have any questions.

Thanks, Jim

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Wednesday, June 22, 2016 3:05 PM

To: Jim Malcolm Cc: Crocker, Philip

Subject: SWEPCO TDS supplemental information

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3 Innwood Circle, Suite 220 • Little Rock, AR 72211-2449 • (501) 225-7779 • Fax (501) 225-6738

#### TECHNICAL MEMORANDUM

DATE:

June 29, 2016

TO:

Ms. Karen Kessler

Water Division

US Environmental Protection Agency

FROM:

Jim Malcolm JTM

FTN Associates, Ltd.

SUBJECT:

FTN No. R06510-0010-002

#### 1.0 INTRODUCTION

Field observations in the reaches of the Red River upstream and downstream of the mouth of the Little River indicated that most of the available instream substrate suitable for invertebrate colonization was in the form of silt/sand and woody debris (WD). Accordingly, FTN Associates, Ltd. (FTN) and the Arkansas Department of Environmental Quality (ADEQ) planning division staff discussed and developed a protocol to sample WD habitat. All benthic macroinvertebrate sampling was conducted on areas of WD upstream and downstream of the mouth of the Little River to characterize the WD macroinvertebrate community.

#### 2.0 METHODS

WD was sampled in the upstream and downstream reaches (Figure 1) on October 17, 2014. A total of 20 submerged sticks with dimensions of 30 cm by 2.3 cm (approximately 0.49 square meter) were collected in each reach by cutting out sections of the WD below the water surface using limb loppers. The 20 WD pieces were placed into a single plastic bag and were immediately preserved with 70% ethanol.

Each WD sample was cleaned in the laboratory by emptying the plastic bag of its liquid contents into a sorting tray and gently rinsing each sample over the sorting tray using a rinsing bottle. Any loose bark was removed and rinsed over the sorting tray along with the uncovered area of the WD. Once all WD pieces were rinsed into the sorting tray, the plastic bag was rinsed into the sorting tray until all debris was removed. After all WD pieces and the plastic bag were thoroughly rinsed, the samples were returned to the plastic bag and preserved with 70% ethanol.

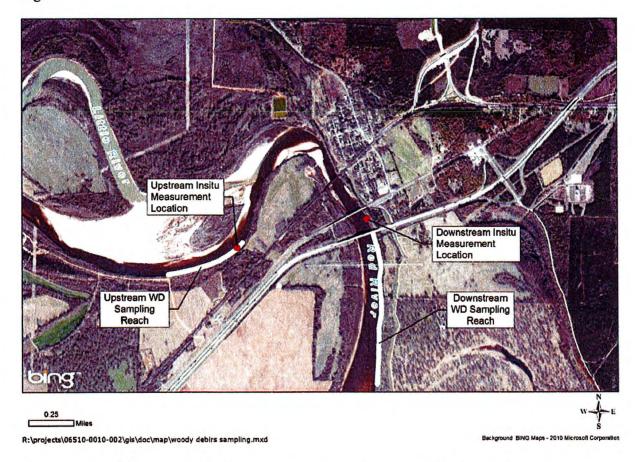


Figure 1. Woody debris and in situ sampling locations on the Red River, 2013.

WDs were sorted in the laboratory by rinsing the pooled sample in a 500-micron sieve and dispensing the entire sample onto a Caton grid. All organisms were sorted from randomly selected grids until a minimum of 270 organisms were collected. If the whole sample was sorted and the number of organisms fell short of 270, then the whole sample was used. Sorted organisms were transferred to 70% ethanol in glass vials. To assure thorough removal of specimens from the sample, the sorted residue was retained and examined by a second biological technician. If the second sorting produced fewer than 10% of the number of organisms found in the initial sorting, the sorting of that sample was considered complete. If the second sorting produced more than 10% of the number of organisms found in the initial sorting, the sample was resorted until the 10% goal was reached.



Ms. Karen Kessler June 29, 2016 Page 3

Taxonomic identifications were carried out to the lowest practical taxon according to Merritt and Cummins (1996), Thorp and Covich (2001), and Houston (1980). In general, macroinvertebrates were identified to genus except for bivalve mollusks, gastropods, dipteran larvae, and decaped shrimp, which were identified to family. A voucher collection of invertebrate taxa collected at the sites was retained for further reference. Taxonomic identifications in the voucher collection were verified by a second taxonomist and identification discrepancies were resolved.

Six in situ measurements of temperature (°C), specific conductance (μS/cm), pH (standard units) and dissolved oxygen (DO, mg/L) were taken between July 19 and October 17 at the locations indicated in Figure 1.

# 3.0 RESULTS AND DISCUSSION

Results of in situ measurements at the upstream and downstream locations are provided in Table 1. Values for temperature, pH and DO were within seasonal expectations. Specific conductance showed a general increase from mid-summer to early fall. This pattern is typical of conservative parameters under decreasing flow conditions. Downstream conductivity ranged from 4 to 27% (average = 16.8%) less than the upstream conductivity due to dilution from the Little River. The measured upstream and downstream conductivity values of 1192 and 1005  $\mu$ S, respectively, correspond to respective upstream and downstream total dissolved solids (TDS) concentrations of approximately 715 and 603 mg/L.

Taxa and raw counts from the WD samples are provided in Table 2. Selected benthic metrics are provided in Table 3. Both upstream and downstream WD communities showed low levels of total taxa and ephemeroptera, plecoptera, and trichoptera (EPT) richness; high proportions of chironomidae; and taxa that Carlisle et al. 2007 classify as moderately tolerant or tolerant to specific conductance.

There is no readily available data set to be used to develop a list of expected macroinvertebrate taxa for these types of substrates for the Red River. Accordingly, the benthic data presented herein characterize the fisheries use attained under the observed mineral conditions (i.e., TDS values up to  $600 - 700 \mu S$ ). This attained use can be described as a generally tolerant to moderately tolerant macro invertebrate community with low overall taxa richness dominated by chironomidae and certain trichoptera. This attained use likely represents the attainable use for these reaches of the Red River.



Table 1. In situ water quality measurements taken at the Red River location upstream and downstream of the mouth of the Little River during 2013.

Date	Parameter	Upstream	Downstream
7/19/2013		30.3	29.6
8/12/2013		34.3	34.1
8/15/2013	Temperature	29.6	29.3
8/29/2013	(°C)	32.2	31.5
9/13/2013		27.7	27.5
10/17/2013		21.4	21.6
7/19/2013		7.8	7.5
8/12/2013		8.5	8.4
8/15/2013	pH	8.6	8.5
8/29/2013	(S.U.)	7.8	7.7
9/13/2013		8.3	8.2
10/17/2013		7.7	7.7
7/19/2013		809	787
8/12/2013	Sandisa Cardantana	833	621
8/15/2013	Specific Conductance	1091	848
8/29/2013	— (μS/cm)	1607	1485
9/13/2013		1660	1590
10/17/2013		1192	1005
7/19/2013		9.1	8.3
8/12/2013		9.2	8.4
8/15/2013	DO	13.4	9.3
8/29/2013	(mg/L)	8.8	7.8
9/13/2013		7.1	6.6
10/17/2013		8.2	7.8

Table 2. Taxa and organism counts from woody debris samples collected from the Red River on October 17, 2013.

Class	Order	Family	Genus	TIV*	Upstream	Downstream
Am	phipoda		Hyalella	7/Moderate		4
Insecta	Diptera				8	
Insecta	Diptera	Chironomidae		5/Moderate	481	372
Insecta	Diptera	Simulidae	Simulium	6/Moderate		1
Insecta	Trichoptera					51
Insecta	Trichoptera	Hydropsychidae	Hydropshyche	8/Tolerant	103	30
Insecta	Trichoptera	Polycentropodidae		5/Moderate	31	

<sup>\*</sup> Tolerance indicator classification for specific conductance obtained from Carlisle et al. 2007.



Ms. Karen Kessler June 29, 2016 Page 5

Table 3. Selected community metrics from woody debris samples collected from the Red River on October 17, 2013.

Metric	Upstream	Downstream
Total organisms	623	458
Total unique taxa	3	4
Ratio of EPT to Chironomid Abundances	0.21	0.22
% Contribution of dominant taxa	77	81
Number of EPT Taxa	2	1
Proportion EPT	0.22	0.18
Proportion chiromonidae	0.77	0.81
Average tolerance value of non chironomidae	7.3	6.1

If you should have any questions or comments regarding this technical memorandum, please feel free to call me or Pat Downey at (501) 225-7779.

#### 4.0 LITERATURE CITED

Carlisle, D.M., M.R. Meador, S.R. Moulton, II, and P.M. Ruhl. 2007. Estimation and application of indicator values for common macroinvertebrate genera and families of the United States. *Ecological Indicators* 7:22-23.

Houston, J. 1980. Checklist of the Damselflies (Odonata: Zygoptera) of Arkansas. Arkansas Academy of Science, Arkansas Biota Survey Checklist No. 25. 1 p.

Merritt, R.W., and K.W. Cummins. 1996. An Introduction to the Aquatic Insects of North America. Dubuque, IA: Kendall/Hunt Publishing Company.

Thorpe, J.H., and A.P. Covich (eds). 2001. Ecology and Classification of North American Freshwater Invertebrates, second edition. Burlington, MA: Academic Press. 1,056 pp.

#### PJD/tas

R:\WP\_FILES\06510-0010-002\CORRESPONDENCE\2016-06-29 TO EPA BENTHIC MACROINVERTEBRATE SAMPLING RESULTS FROM RED RIVER UPSTREAM AND DOWNSTREAM OF LITTL RIVER\TM-K KESSLER 2016-06-29.DOCX



#### **Preliminary Development of Tolerance Values for Freshwater Species**

Preliminary tolerance values (TVs) for chloride and sulfate were developed from aquatic toxicity test data available through EPA's ECOTOX database. Test metrics (LCx, ECx, NOEC, LOEC and MATC) for Chloride and sulfate must be used as surrogates for TDS TVs because toxicity test results are seldom reported as TDS. The ECOTOX data base provides laboratory test results from a wide range of test conditions. Results of this compilation are provided in the "Fish summary", "Insects etc", and "Crustaceans" tabs in the attached accompanying spreadsheet. The tables for each species and chemical provide average, minimum and maximum values of all metrics at all test durations. Visual examination of these results indicated that maximum and minimum metric values are generally large in magnitude (almost all value well in excess of 1000mg/L) and vary with a factor of 2 or 3. Therefore average values for all test metrics (i.e. LCx, ECx, NOEC, LOEC and MATC) as mg/L chloride or sulfate for lethalityints were averaged across all test durations to provide a first approximation of TVs. These TVs are provided, as available from ECOTOX (Period of record 1954 through 2016), in Tables 1, and 3.

Table 1. Tolerance values for fish

		Tolerance Value (mg/L)	
Scientific name	Common name	Chloride	Sulfate
Pimephales promelas	Fathead minnow	4990	3461
Lepomis macrochirus	Bluegill sunfish	10378	
Gambusia affinis	Eastern mosquito fish	13400	
Anguilla rostrata	American eel	19665	
Cyprinella leedsi	Bannerfin shiner	6120	
Gambusia holbrooki	Western Mosquito fish	5820	
Carassius auratus	Goldfish	2734	
Morone saxatilis	Stripped Bass	3604	

Table 2. Tolerance values for insects and other invertebrates.

		Tolerance V	alue (mg/L)
Scientific name	Common name	Chloride	Sulfate
Culiseta incidens	Mosquito	7850	
Elliptio complanata	Eastern Elliptio	2450	
Epioblasma torulosa	Northern Riffleshell	244	
Erpobdella punctata	Red Leech	8125	
Gyraulus circumstriatus	FW mollusc	6020	
Gyraulus parvus	FW mussel	3043	
Hydroptila angusta	Caddisfly	6621	
Isonychia bicolor	Mayfly	2861	
Lampsilis cardium	Plain Pocketbook mussel	817	
Lampsilis fasciola	Wavy-Rayed Lampmussel	1133	
Lampsilis siliquoidea	Lamp-Mussel	1806	2882
Ligumia recta	Black Sandshell mussel	3130	
Limnodrilus hoffmeisteri	Oligochaete	6381	
Lumbriculus variegatus	Oligochaete, Worm	4626	
Maccaffertium sp.	Mayfly	380	
Musculium transversum	Long Fingernail Clam	1655	
Nais variabilis	Oligochaete	2569	
Nephelopsis obscura	Leech	4284	
Physa heterostropha	Pond Snail,	5067	
Planorbella campanulata	Snail, Bellmouth Rams-Horn	7190	
Pycnopsyche guttifera	Caddisfly	3526	
Pycnopsyche lepida	Northern Caddisfly	3526	
Sphaerium simile	Grooved Fingernail Clam	920	2290
Sphaerium sp.	Orb Cockle, Fingernail Clam	1612	
Stenonema modestum	Mayfly	4.7	
Tipula abdominalis	Crane Fly	1000	
Tubifex tubifex	Tubificid Worm	5311	
Villosa constricta	Notched Rainbow	3504	
Chironomus tentans	Midge		12908
Tricorythus sp.	Mayfly		660

Table 3. Tolerance values for insects and other invertebrates.

		Tolerance Value (mg/L)	
Scientific name	Common name	Chloride	Sulfate
Daphnia ambigua	Water flea	1097	
Daphnia hyalina	Water flea	3000	
Daphnia magna	Water flea	3813	
Daphnia pulex	Water flea		2573
Gammarus pseudolimnaeus	Aquatic Sowbug		10320
Hyalella azteca			1740
Lirceus fontinalis	Aquatic Sowbug	2970	
Pacifastacus leniusculus	Signal Crayfish	1000	

			Growth	
Chemical	Species		Endpoint Value	Test duration
CaCl	Pimephales promelas	Average	1410000	7
		Minimum	1410000	
		Maximum	1410000	
		N	1	
Na2SO4	Pimephales promelas	Average	1245972	11.2
		Minimum	101370	7
		Maximum	3650000	21
		N	10	
			Survival	
0.01		١.	Endpoint Value	
CaCl	Lepomis macrochirus	Average	10375000	4.166666667
		Minimum	9500000	4
		Maximum	11300000	5
CaCl	Dimonhalas	N A	11300000	2.75
CaCl	Pimephales promelas	Average	4990000	2.75
		Minimum Maximum	4630000 6560000	1 4
		N	13400000	4
CaCl	Gambusia affinis	Average	13400000	2.3
cuci	Guilibusia arriiriis	Minimum	13400000	2.3
		Maximum	13400000	4
		N	3	7
NaCl	Anguilla rostrata	Average	19665000	4
	American eel	Minimum	17880000	4
		Maximum	21450000	4
		N	2	
NaCl	Cyprinella leedsi	Average	6120115	4
	bannerfin shiner	Minimum	6069550	4
		Maximum	6069550	4
		N	4	
	Lepomis macrochirus	Average	8256650	. 4
	bluegill sunfish	Minimum	1294600	4
		Maximum	12946000	4
•		N	4	
	Gambusia holbrooki	Average	5820000	4.5
	Mosquito fish	Minimum	100000	4
		Maximum	11540000	5
	Corossius auretus	N 	2724200.62	2 527272727
	Carassius auratus Goldfish	Average	2734309.63	2.527272727
	Goldiisii	Minimum Maximum	4324000 11050000	1 10
		N	165	10
	Morone saxatilis	Average	3604166.667	2.5
	Striped bass	Minimum	1000000	1
	•	Maximum	7000000	4
		N	24	
	Gambusia affinis	Average	17916666.67	2.333333333
	Mosquito fish	Minimum	17916666.67	2.333333333
		Maximum	17916666.67	2.333333333
		N	1	
	Lepomis macrochirus	Average	10381428.57	4
	Bluegill sunfish	Minimum	3040000	4
		Maximum	13500000	4
N. 266.	[a:	N L	7	
Na2SO4	Pimephales promelas		3461232.759	
	Fathead minnow	Minimum	101370	1
		Maximum	15200000	21
		N	29	

Chemical NaCl

# Sublethal (Growth Reprod

Species	endpoint Value	Duration	
Brachionus calyciflorus	Average	1608200	2
Rotifer	Minimum	1120000	2
	Maximum	2230000	2
	N	5	
Chironomus dilutus	Average	2809200	20
	Minimum	2133000	20
	Maximum	3960000	20
	N	5	
Lumbriculus variegatus	Average	730750	28
	Minimum	366000	28
	Maximum	1366000	28
	N	4	
Polycelis felina	Average	108000	7
Tubellaria	Minimum	108000	7
	Maximum	108000	7
	N	1	
Stenonema modestum	Average	5650	14
	Minimum	2700	14
	Maximum	7000	14
	N	8	
Tubifex tubifex	Average	660600	28
Tubificid Worm	Minimum	462000	28
	Maximum	964000	28
	N	5	
Amerianna cumingi	Average	338000	4
snail	Minimum	338000	4
	Maximum	338000	4
	N	1	
Corbicula manilensis	Average	1500000	28
	Minimum	1500000	28
	Maximum	1500000	28
	N	4	

NaCl

# Lethal

Species		endpoint Value	Duration
Acroneuria abnormis	Average	10000000	4
Stonefly	Minimum	10000000	4
	Maximum	10000000	4
	Ν	1	
Agnetina capitata	Average	10000000	4
Stonefly	Minimum	10000000	4
	Maximum	10000000	4
	N	1	
Argia sp.	Average	27375000	2.375
Damdel fly	Minimum	23000000	1
	Maximum	32000000	4
	N	8	
Brachionus calyciflorus	Average	1645000	1
Rotifer	Minimum	1645000	
	Maximum		
	N	10 13000	
Caenorhabditis elegans	Average	18126250	
Nematode	Minimum	15460000	
remateuc	Maximum		
	N	21000000	
Chironomus dilutus	Average	6234856.667	
midge	Minimum	4162650	
mage	Maximum		
	N		2.111111111
Culiseta incidens			
Mosquito	Average Minimum	7850000	
Mosquito		4700000	
	Maximum		
	N A	2450000	
Elliptio complanata	Average	2450000	
	Minimum	2230000	
	Maximum		
New Picci I II	N	2	
Northern Riffleshell	Average	244000	
	Minimum	244000	
	Maximum	244000	1
	N	1	
Erpobdella punctata	Average	8125000	
Red Leech	Minimum	7500000	1
	Maximum	10000000	5
	N	4	
Gyraulus circumstriatus	Average	6020000	3
FW mollusc	Minimum	3200000	1
	Maximum	10000000	5
	Ν	5	

# Lethal

Species		endpoint Value	Duration
Gyraulus parvus	Average	3043500	4
FW mussel	Minimum	3009000	4
	Maximum	3078000	4
	N	2	
Hydroptila angusta	Average	6621000	2
Caddisfly	Minimum	6621000	2
	Maximum	6621000	2
	N	1	
Isonychia bicolor	Average	2861111.111	14
Mayfly	Minimum	500000	14
	Maximum	8000000	14
	N	18	
Lampsilis cardium	Average	817000	1
Plain Pocketbook	Minimum	817000	1
	Maximum	817000	1
	N	1	
Lampsilis fasciola	Average	1133461.538	1.307692308
Wavy-Rayed Lampmussel	Minimum	113000	1
	Maximum	3980000	4
	N	13	
Lampsilis siliquoidea	Average	1806300	1.8
Lamp-Mussel	Minimum	168000	1
	Maximum	4560000	4
	N	10	
Ligumia recta	Average	3130000	3
Black Sandshell	Minimum	2510000	2
	Maximum	3750000	4
Limnodrilus hoffmeisteri	N	2	
Tubificid Worm, Oligochaete	Average	6381250	4.859375
	Minimum	5800000	1
	Maximum	7500000	10.875
	N	8	
Lumbriculus variegatus	Average	4626592.5	4
Oligochaete, Worm	Minimum	3100000	4
	Maximum	5588680	4
	N	8	
Maccaffertium sp.	Average	3809953.478	5.239130435
Mayfly	Minimum	2700	1
	Maximum	14134000	14
	N	92	
Musculium transversum	Average	1655000	
Long Fingernail Clam	Minimum	1655000	
	Maximum	1655000	4
	N	1	

Summary			TV S
		Lethal	
Species		endpoint Value	Duration
Nais variabilis	Average	2569000	2
Oligochaete	Minimum	2569000	2
	Maximum	2569000	2
	N	1	
Nephelopsis obscura	Average	4284417.143	4
Leech	Minimum	3320000	4
	Maximum	5340000	4
	N	7	
Physa gyrina	Average	2540000	4
Pouch Snail	Minimum	2540000	4
	Maximum	2540000	
	N	1	7
Physa heterostropha	Average	5067647.059	2.647058824
Pond Snail, Pneumonate Snail	Minimum	3500000	
Tona Shan, Theamonate Shan	Maximum	7500000	
	N	17	5
Planorbella campanulata			2
	Average	7190000	3
Snail, Bellmouth Rams-Horn	Minimum	6150000	
	Maximum	10000000	5
Day of the same	N	5	_
Pycnopsyche guttifera	Average	3526000	
Caddisfly	Minimum	3526000	
	Maximum	3526000	4
	N	1	
Pycnopsyche lepida	Average	3526000	
Northern Caddisfly	Minimum	3526000	
	Maximum	3526000	4
	N	1	
Sphaerium simile	Average	920000	4
Grooved Fingernail Clam	Minimum	740000	4
	Maximum	1100000	4
	N	2	
Sphaerium sp.	Average	1612500	2.5
Orb Cockle, Fingernail Clam	Minimum	1100000	1
	Maximum	2400000	4
	N	8	
Stenonema modestum	Average	4700	14
Mayfly	Minimum	2700	14
	Maximum	7000	14
	N	4	
Tipula abdominalis	Average	1000000	4
Crane Fly	Minimum	1000000	4
	Maximum	1000000	4
	N	1	

Lethal
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			Letilai	
	Species		endpoint Value	Duration
	Tubifex tubifex	Average	5311333.333	3 4
	Tubificid Worm	Minimum	4278000	) 4
		Maximum	6008000	) 4
		N	3	3
	Villosa constricta	Average	3504000	) 2
	Notched Rainbow	Minimum	2590000	) 1
		Maximum	5230000	) 4
		N	Ç	5
Na2SO4	Chironomus tentans	Average	12908000	) 2
	Midge	Minimum	11682000	) 2
		Maximum	14134000	) 2
		N	2	2
	Lampsilis siliquoidea	Average	2882300	3
	Lamp-Mussel	Minimum	1702000	) 2
		Maximum	3729000	) 4
		N	10	10
	Sulfuric acid sodium salt (1:2)	Average	2290250	)
	Grooved Fingernail Clam	Minimum	1502000	)
		Maximum	2926000	)
		N	8	3
	Tricorythus sp.	Average	660000	) 4
	Mayfly	Minimum	660000	) 4
		Maximum	660000	) 4
		Ν	-	1

Crustacea	ins Summary			1 V Summary 1
			Sublethal (Growth Reprod	
	Species		endpoint Value	Duration
NaCl	Daphnia ambigua	Average	646666.6667	10
		Minimum	440000	10
		Maximum	850000	10
		N	3	
	Daphnia magna	Average	1041130.435	10.73913043
		Minimum	220000	2
		Maximum	2160000	21
		N	23	
Na2SO4	Daphnia pulex	Average	465083.3333	21
		Minimum	314000	21
		Maximum	1200000	21
		N	12	
	Eulimnogammarus toletanus	Average	108000	7
	Sand Shrimp	Minimum	108000	
		Maximum	108000	
		N	1	
	Hyalella azteca	Average	1876800	21
	Tryatena azteea	Minimum	380000	14
		Maximum	4237000	28
		N	10	20
		14	Lethal (survival)	
	Species		endpoint Value	Duration
CaCl2	Hyalella azteca	Average	2207555.556	3.925925926
CaCiz	Tryalella azteca	Minimum	512000	2
		Maximum	4345000	
		N	4343000	4
	Lireaus fantinalis			4
	Lirceus fontinalis	Average	2970000	4
	Aquatic Sowbug	Minimum	2970000	4
		Maximum	2970000	4
	5 16 1 1	N	1	
	Pacifastacus leniusculus	Average	1000000	71
	Signal Crayfish	Minimum	1000000	71
		Maximum	1000000	71
		N	1	
NaCl	Asellus communis	Average	7095454.545	
	Aquatic Sowbug	Minimum	5100000	1
		Maximum	10000000	7
		N	11	
	Daphnia ambigua	Average	1096666.667	7.333333333
		Minimum	440000	2
		Maximum	2000000	10
		N	3	
	Daphnia hyalina	Average	3000000	2
		Minimum	3000000	2
		Maximum	3000000	2
		N	1	

# Crustaceans Summary

			Lethal (survival)	
	Species		endpoint Value	Duration
	Daphnia magna	Average	3813532	2.725
		Minimum	467000	1
		Maximum	8600000	21
		N	40	)
Na2SO4	Daphnia pulex	Average	2573043.478	6.043478261
		Minimum	400000	1
		Maximum	5090000	21
		N	23	<b>,</b>
	Gammarus pseudolimnaeus	Average	10320000	3.142857143
		Minimum	7700000	3
		Maximum	12600000	4
		Ν	7	,
	Hyalella azteca	Average	1740373.942	
		Minimum	2185	;
		Maximum	5259000	)
		N	52	)

From: Sent: Kesler, Karen [Kesler.Karen@epa.gov] Wednesday, June 29, 2016 3:41 PM

To:

Jim Malcolm

Subject:

Automatic reply: SWEPCO TDS supplemental information

I will be out of the office between June 23rd and June 29th. I will respond to your email as quickly as possible when I return to the office on June 30th.

Thanks, Karen

From: Crocker, Philip [crocker.philip@epa.gov]
Sent: Wednesday, June 29, 2016 4:28 PM

To: Jim Malcolm Cc: Kesler, Karen

Subject: RE: SWEPCO TDS supplemental information

Hi Jim, thanks. I am getting ready to leave for several days. Karen should be back tomorrow so should be reviewing the info.

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To: Kesler, Karen < Kesler.Karen@epa.gov >

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Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

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Many thanks, Jim



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From: Sent: Jim Malcolm [jtm@ftn-assoc.com] Wednesday, July 06, 2016 8:02 AM

To:

'Jim Malcolm'; 'Crocker, Philip'
'Kesler, Karen'; James Malcolm; James Malcolm

Cc: Subject:

RE: SWEPCO TDS supplemental information

Hi Karen/Phil,

Circling back on this mainly to just make sure you guys got it and the previous email with the TLVs we dug up and the benthics data/memo. You have not had the information long but please let me know your thoughts when you guys have time to digest this? Thanks.

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From: Sent: Kesler, Karen [Kesler.Karen@epa.gov] Wednesday, July 06, 2016 8:08 AM

To:

Jim Malcolm; Crocker, Philip

Subject:

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To: Cc:

James Malcolm

Subject:

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Thanks Karen. Just an fyi, I've sent my two recent emails to you and Phil also to Sarah to keep her/ADEQ in all the loops.

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- A technical memo describing macroinvertebrate data collected from the Red River upstream and downstream
  of the mouth of the Little River during October 2014;
- 2. A Word doc describing the results of a preliminary effort to develop numerical tolerance values for minerals based on laboratory toxicity data;
- 3. The spreadsheet referenced in the Word doc.

The development of the numerical tolerance values is preliminary. If you all think the information looks to be useful, we can refine the approach and provide more documentation of the methodology.

I am out of the office today, but I will be in tomorrow and Friday. Feel free to call me if you have any questions.

Thanks, Jim

From: Kesler, Karen [mailto:Kesler,Karen@epa.gov]

Sent: Wednesday, June 22, 2016 3:05 PM

**To:** Jim Malcolm **Cc:** Crocker, Philip

Subject: SWEPCO TDS supplemental information

Hi Jim,

I wanted to touch base and see when we should be expecting the additional information for the SWEPCO rulemaking in light of our meeting on June  $10^{th}$ . As this rulemaking has already been largely extended beyond the normal time frame, we would like to move forward with the final piece of rulemaking as soon as possible. Phil is pushing to get this action completed quickly, so we would prefer to receive the additional information by next Friday, July  $1^{st}$ .

Please let us know when you expect to send it along to ADEQ. I will be out of the office until June 30<sup>th</sup>, so please cc Phil with your response so he knows where things stand.

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Kesler, Karen [Kesler.Karen@epa.gov]
Sent: Wednesday, July 06, 2016 9:49 AM

To: Jim Malcolm Crocker, Philip

Subject: RE: SWEPCO TDS supplemental information

Sounds good.

Thanks Jim.

- Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Wednesday, July 06, 2016 9:08 AM

To: Kesler, Karen < Kesler. Karen@epa.gov>; Crocker, Philip < crocker.philip@epa.gov>

Cc: James Malcolm <jtm@ftn-assoc.com>

Subject: RE: SWEPCO TDS supplemental information

Thanks Karen. Just an fyi, I've sent my two recent emails to you and Phil also to Sarah to keep her/ADEQ in all the loops.

Regards, Jim

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Wednesday, July 06, 2016 8:08 AM

To: Jim Malcolm; Crocker, Philip

Subject: RE: SWEPCO TDS supplemental information

Hi Jim,

Yes, we have received both the TLVs and the benthic data/memo. Thank you for sending both of those along so quickly. I have had a chance to go through them once. Once I have a chance to talk with everyone else about it, I will send you our thoughts.

Thanks, Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Wednesday, July 06, 2016 8:02 AM

To: 'Jim Malcolm' <jtm@ftn-assoc.com>; Crocker, Philip <crocker.philip@epa.gov>

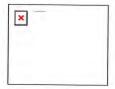
Cc: Kesler, Karen < Kesler.Karen@epa.gov >; James Malcolm < itm@ftn-assoc.com >; James Malcolm < itm@ftn-assoc.com >

Subject: RE: SWEPCO TDS supplemental information

Hi Karen/Phil,

Circling back on this mainly to just make sure you guys got it and the previous email with the TLVs we dug up and the benthics data/memo. You have not had the information long but please let me know your thoughts when you guys have time to digest this? Thanks.

### Regards, Jim



Jim Malcolm itm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Wednesday, June 29, 2016 5:38 PM

To: 'Crocker, Philip'

Cc: 'Kesler, Karen'; James Malcolm

Subject: RE: SWEPCO TDS supplemental information

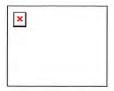
Hi Phil,

Thanks for the reply. Just so you and Karen know, from SWEPCO's perspective, the 780 mg/L TDS value is acceptable and they would be prepared to alter the Arkansas rulemaking to support the change if EPA is in agreement with this value on the Arkansas side of the AR-LA state line. The 780 mg/L value is the 90<sup>th</sup> percentile value for TDS in the lower Red River from our earlier analysis and therefore, FTN's concern was that the 780 mg/L might at some point end up triggering for impairment. However, this concern is mitigated somewhat because the Red River minerals levels appear a bit lower more recently and it is possible that EPA and ADEQ can still reach agreement on retaining 25% trigger for 303(d) assessment.

In looking back at the field work from the UAA, we had collected some benthics data from logs/woody debris in the lower Red River below the Little River as part of the SWEPCO work. We have compiled the data and they are attached in a technical memorandum along with other related information. We request that EPA review these data in the context of it completing the weight of evidence information needed by EPA relating to addressing AL characterization/use protection based on the revised 780 mg/L TDS criteria.

Please do not hesitate to contact me if there are any questions or if you wish to discuss this submittal.

Many thanks, Jim



#### Jim Malcolm

jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Crocker, Philip [mailto:crocker.philip@epa.gov]

Sent: Wednesday, June 29, 2016 4:28 PM

**To:** Jim Malcolm **Cc:** Kesler, Karen

Subject: RE: SWEPCO TDS supplemental information

Hi Jim, thanks. I am getting ready to leave for several days. Karen should be back tomorrow so should be reviewing the info.

Phil

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Wednesday, June 29, 2016 3:40 PM
To: Kesler, Karen < Kesler. Karen@epa.gov >

Cc: Crocker, Philip < crocker.philip@epa.gov >; James Malcolm < jtm@ftn-assoc.com >

Subject: RE: SWEPCO TDS supplemental information

Karen, Phil,

Attached per your request are the following:

- A technical memo describing macroinvertebrate data collected from the Red River upstream and downstream
  of the mouth of the Little River during October 2014;
- 2. A Word doc describing the results of a preliminary effort to develop numerical tolerance values for minerals based on laboratory toxicity data;
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The development of the numerical tolerance values is preliminary. If you all think the information looks to be useful, we can refine the approach and provide more documentation of the methodology.

I am out of the office today, but I will be in tomorrow and Friday. Feel free to call me if you have any questions.

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Sent: Wednesday, June 22, 2016 3:05 PM

**To:** Jim Malcolm **Cc:** Crocker, Philip

Subject: SWEPCO TDS supplemental information

Hi Jim,

I wanted to touch base and see when we should be expecting the additional information for the SWEPCO rulemaking in light of our meeting on June 10<sup>th</sup>. As this rulemaking has already been largely extended beyond the normal time frame, we would like to move forward with the final piece of rulemaking as soon as possible. Phil is pushing to get this action completed quickly, so we would prefer to receive the additional information by next Friday, July 1<sup>st</sup>.

Please let us know when you expect to send it along to ADEQ. I will be out of the office until June  $30^{th}$ , so please cc Phil with your response so he knows where things stand.

Thanks, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Clem, Sarah [CLEM@adeq.state.ar.us]

 Sent:
 Friday, July 22, 2016 1:35 PM

 To:
 jim (jtm@ftn-ASSOC.COM)

**Subject:** FW: SWEPCO Red River TDS site specific criterion **Attachments:** Final EPA Disapproval of TDS provision 7\_20\_16.pdf

Jim,

Received this morning. Caleb will be sending the attachment to Allan as well.

#### Sarah

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Friday, July 22, 2016 10:43 AM

To: Clem, Sarah

Cc: Crocker, Philip; Wentz, Tate; Barnett, Mary

Subject: SWEPCO Red River TDS site specific criterion

Hi Sarah,

Attached is the signed disapproval of the TDS site specific criterion for the lower Red River for the SWEPCO rulemaking. We know that FTN is working on collecting additional information about tolerance values, have provided additional benthic data for the Red River, and have stated that SWEPCO is ok with a 780 mg/L criterion. EPA believes these are all good steps forward towards determining an appropriate criterion that is both protective of downstream uses and demonstrates protection of Arkansas's uses. The attached letter and TSD reflects our issues with the initial submission and the 860 mg/L criterion that was proposed with this rulemaking. On Monday, I will follow up with an email to Jim Malcolm explaining how a preliminary assessment of the additional information submitted (since the last conference call) seems to indicate that this approach will add sufficient documentation to their weight of evidence demonstration of protection of aquatic life use and that the 780 mg/L value will address issues of downstream protection. If they decide to pursue another rulemaking with the 780 mg/L criteria value, we believe this approach will be useful for demonstrating protection of aquatic life use. Thank you for all your help with review of this rulemaking and coordination of communication between us and FTN.

Please let me know if you have any questions, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185



### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

# REGION 6 1445 ROSS AVENUE, SUITE 1200 DALLAS, TEXAS 75202 – 2733

July 20, 2016

Mr. Caleb Osborne
Associate Director – Water Division
Arkansas Department of Environmental Quality
5301 Northshore Drive
Little Rock, AR 72118-5317

Re: Final submittal of Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas, as amended by third party rulemaking initiated by the Southwestern Electric Power Company

Dear Mr. Osborne:

The Environmental Protection Agency (EPA) has completed its review of the revisions to Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas that were made in relation to the Third Party Rulemaking process initiated by Southwestern Electric Power Company. The revisions to Regulation No. 2 were adopted by the Arkansas Pollution Control and Ecology Commission (APC&EC) on October 23, 2015 and became effective on November 5, 2015. They were submitted to the EPA for approval on December 21, 2015 by the Arkansas Department of Environmental Quality (ADEQ).

In a letter, dated May 16, 2016, EPA approved the majority of the revised provisions to Regulation 2.511 and Appendix A of Regulation No. 2, including the site-specific criteria changes for temperature and total dissolved solids (TDS) in the Little River and the removal of the domestic water supply use in the Red River. These revisions were approved pursuant to the Clean Water Act (CWA) § 303(c) and its implementing regulations at 40 CFR Part 131. However, EPA did not take action on the site-specific TDS criterion revision in the Red River at that time. After further review, EPA is now disapproving this site-specific TDS criterion revision for the Red River from its confluence with the Little River to the Arkansas-Louisiana state line. Inadequate information was submitted to demonstrate protection of the aquatic life use and this criterion is not protective of the downstream use in Louisiana. Please note that under 40 CFR § 313.21(c), new and revised standards do not go into effect for CWA purposes until approved by EPA. Therefore, previously approved language associated with the TDS criterion in the January 24, 2008 version of the Arkansas water quality standards remains in effect for CWA purposes.

I appreciate the APC&EC's and the ADEQ's effort in the review of these revised provisions of the State's standards and also appreciate ADEQ's assistance with coordinating meetings and correspondence with the third party. If you have any questions or concerns, please contact me at 214-665-7101, or contact Karen Kesler at 214-665-3185.

Sincerely,

William K. Honker, P.E.

WK Houles

Director

Water Division

# Enclosure

cc: Sarah Clem, Branch Manager

Water Division ADEQ

# TECHNICAL SUPPORT DOCUMENT: EPA REVIEW OF SITE-SPECIFIC CRITERION REVISION TO REGULATION 2: REGULATION ESTABLISHING WATER QUALITY STANDARDS FOR SURFACE WATERS OF THE STATE OF ARKANSAS FOR THE RED RIVER, ARKANSAS

Revision Adopted by the Arkansas Pollution Control and Ecology Commission via Minute Order No. 15-21 for Southwestern Electric Power Company

> U.S. EPA REGION 6 WATER DIVISION July 2016

## I. Introduction

# Background

As described in § 303(c) of the Clean Water Act (CWA) and in the standards regulation within the Code of Federal Regulations (CFR) at 40 CFR § 131.20, states and authorized tribes have primary responsibility to develop and adopt water quality standards to protect their waters. State and tribal water quality standards consist of three primary components: beneficial uses, criteria to support those uses, and an antidegradation policy. In addition, CWA § 303(c)(1) and 40 CFR § 131.20 require states to hold public hearings at least once every three years to review and, as appropriate, modify and adopt standards.

Under 40 CFR § 131.21, the Environmental Protection Agency (EPA) reviews new and revised surface water quality standards that have been adopted by states and authorized tribes. Authority to approve or disapprove new and/or revised standards submitted to EPA for review has been delegated to the Water Division Director in Region 6. Tribal or state water quality standards are not considered effective under the CWA until approved by EPA.

The purpose of this Technical Support Document (TSD) is to provide the basis for the Environmental Protection Agency's disapproval of the site-specific total dissolved solids (TDS) water quality criterion revision for the Red River to Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas adopted by the Arkansas Pollution Control and Ecology Commission (APC&EC) in Minute Order 15-21. This revision is further described in the subsection below titled "Summary of Revised Provisions."

# Chronology of Events

September 11, 2014	A third party, Southwestern Electric Power Company (SWEPCO), filed a petition with the APC&EC to amend Regulation No. 2
September 26, 2014	The APC&EC initiated the rulemaking proceedings via Minute Order No. 14-33
October 1, 2014	Public notice of the proposed rule-making was published
November 17, 2014	Public hearing on the proposed rule-making was held in Hope, Arkansas
December 3, 2014	Public comment period ended on the proposed changes to Regulation No. 2
October 23, 2015	Becky Keogh, Director, Arkansas Department of Environmental Quality (ADEQ), signed Minute Order No. 15-21 adopting changes to Regulation No. 2

attainability analysis (UAA) of the receiving waterbodies affected by the revised site-specific criteria. The portions of the UAA that provided scientific justification for the appropriateness and protectiveness of the revised site-specific water quality criterion for TDS in the Red River from its confluence with the Little River to the AR/LA state line were considered for this action.

# Site-Specific Water Quality Criterion for TDS in the Red River

Table 1. Site-specific water quality criterion revision for TDS in the Red River submitted by ADEO to EPA for review and approval.

Reach Description	Current Criterion	Proposed Criterion
Red River from its confluence with the Little River to the AR/LA state line	500 mg/L	860 mg/L

# **Disapproval Justification**

In its review of the SWEPCO UAA and additional submitted material, EPA determined that the documents did not sufficiently demonstrate protection of aquatic life use which is required by 40 CFR § 131.6 and also did not demonstrate protection of downstream use which is required by 40 CFR § 131.10(b).

The predominant source of evidence that was submitted to show that the aquatic life use was protected was a study completed by Buchanan et al. in 2003. This study surveyed fish that were present in the Arkansas portion of the Red River from 1995 to 2001. While the Buchanan study provides a historic reference, it only provides us with an understanding of the fish community in the Red River in 2001, not the current fish community. In addition, the Buchanan et al. 2003 study assessed presence and determined broad classifications of abundance (rare, uncommon, common, abundant). This gives us information about acute tolerances, but does not give us information about chronic tolerances. The presence of the fishes does not indicate that they are growing and reproducing at unimpaired rates. In addition, there was no discussion of the protection of the macroinvertebrate community in the Red River. Without additional information about the fish and macroinvertebrate community currently present in the Red River, EPA could not determine whether those communities are protected with the new criterion. Based on these concerns, EPA requested that additional information be submitted that demonstrated numeric tolerances of species that are found in the Red River. However, no additional data or information, only a representation of the Buchanan et al. 2003 study, was provided with which to assess the proposed criterion. Without additional supporting documentation, EPA was unable to assess whether the aquatic life use was being protected.

Downstream protection was also not demonstrated for this revised criterion. The proposed criterion is 860 mg/L and the downstream criterion in Louisiana is 780 mg/L. In the UAA submitted, the contractor reported the results of a mass-balance they performed using a 7Q-10 flow rate. These results showed predicted TDS concentrations at various spots in the Red River, including at the AR/LA state line. Also included in the UAA were the results of a second mass-balance performed by the contractor that used the same input values and model, but used a harmonic mean flow in place of a 7Q-10 flow rate. This second mass-balance was performed to justify that the proposed criterion is protective of LA uses. But the use of the harmonic mean

From: Clem, Sarah [CLEM@adeq.state.ar.us]
Sent: Monday, July 25, 2016 9:57 AM

To: jim (jtm@ftn-ASSOC.COM)

Subject: FW: SWEPCO Red River TDS site specific criterion Attachments: Disapproval Letter\_SWEPCO TDS\_ 7\_20\_16.pdf

### Here is the rest of the pdf.

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Monday, July 25, 2016 8:21 AM

To: Clem, Sarah

Subject: RE: SWEPCO Red River TDS site specific criterion

Hi Sarah,

I'm sorry about that. The attached scan should be the whole thing.

### - Karen

From: Clem, Sarah [mailto:CLEM@adeq.state.ar.us]

Sent: Friday, July 22, 2016 3:48 PM

To: Kesler, Karen < Kesler. Karen@epa.gov>

Subject: RE: SWEPCO Red River TDS site specific criterion

Karen,

Looks like we only have up to page 3 of the SWEPCO TSD. Could you send the rest? Thanks.

#### Sarah

From: Kesler, Karen [mailto:Kesler, Karen@epa.gov]

Sent: Friday, July 22, 2016 10:43 AM

To: Clem, Sarah

Cc: Crocker, Philip; Wentz, Tate; Barnett, Mary

Subject: SWEPCO Red River TDS site specific criterion

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Please let me know if you have any questions, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185



### UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

## REGION 6 1445 ROSS AVENUE, SUITE 1200 DALLAS, TEXAS 75202 – 2733

July 20, 2016

Mr. Caleb Osborne
Associate Director – Water Division
Arkansas Department of Environmental Quality
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In a letter, dated May 16, 2016, EPA approved the majority of the revised provisions to Regulation 2.511 and Appendix A of Regulation No. 2, including the site-specific criteria changes for temperature and total dissolved solids (TDS) in the Little River and the removal of the domestic water supply use in the Red River. These revisions were approved pursuant to the Clean Water Act (CWA) § 303(c) and its implementing regulations at 40 CFR Part 131. However, EPA did not take action on the site-specific TDS criterion revision in the Red River at that time. After further review, EPA is now disapproving this site-specific TDS criterion revision for the Red River from its confluence with the Little River to the Arkansas-Louisiana state line. Inadequate information was submitted to demonstrate protection of the aquatic life use and this criterion is not protective of the downstream use in Louisiana. Please note that under 40 CFR § 313.21(c), new and revised standards do not go into effect for CWA purposes until approved by EPA. Therefore, previously approved language associated with the TDS criterion in the January 24, 2008 version of the Arkansas water quality standards remains in effect for CWA purposes.

I appreciate the APC&EC's and the ADEQ's effort in the review of these revised provisions of the State's standards and also appreciate ADEQ's assistance with coordinating meetings and correspondence with the third party. If you have any questions or concerns, please contact me at 214-665-7101, or contact Karen Kesler at 214-665-3185.

Sincerely,

William K. Honker, P.E.

WK Horlen

Director

Water Division

Enclosure

cc: Sarah Clem, Branch Manager

Water Division ADEQ

# TECHNICAL SUPPORT DOCUMENT:

EPA REVIEW OF SITE-SPECIFIC CRITERION REVISION TO REGULATION 2: REGULATION ESTABLISHING WATER QUALITY STANDARDS FOR SURFACE WATERS OF THE STATE OF ARKANSAS FOR THE RED RIVER, ARKANSAS

Revision Adopted by the Arkansas Pollution Control and Ecology Commission via Minute Order No. 15-21 for Southwestern Electric Power Company

> U.S. EPA REGION 6 WATER DIVISION July 2016

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# I. Introduction

# Background

As described in § 303(c) of the Clean Water Act (CWA) and in the standards regulation within the Code of Federal Regulations (CFR) at 40 CFR § 131.20, states and authorized tribes have primary responsibility to develop and adopt water quality standards to protect their waters. State and tribal water quality standards consist of three primary components: beneficial uses, criteria to support those uses, and an antidegradation policy. In addition, CWA § 303(c)(1) and 40 CFR § 131.20 require states to hold public hearings at least once every three years to review and, as appropriate, modify and adopt standards.

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The purpose of this Technical Support Document (TSD) is to provide the basis for the Environmental Protection Agency's disapproval of the site-specific total dissolved solids (TDS) water quality criterion revision for the Red River to Regulation No. 2: Regulation Establishing Water Quality Standards for Surface Waters of the State of Arkansas adopted by the Arkansas Pollution Control and Ecology Commission (APC&EC) in Minute Order 15-21. This revision is further described in the subsection below titled "Summary of Revised Provisions."

# Chronology of Events

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October 23, 2015	Becky Keogh, Director, Arkansas Department of Environmental Quality (ADEQ), signed Minute Order No. 15-21 adopting changes to Regulation No. 2

December 21, 2015

William K. Honker, Director, Water Division, EPA
Region 6, received letter from, Ellen Carpenter, Water
Division Chief, ADEQ, requesting EPA approval of the
adopted revisions and transmitting the water quality
standards submission package

February 9, 2016 EPA Region 6 issued a letter to ADEQ stating that no action would be taken on the submission package to allow the third party time to respond to EPA's comments

March 30, 2016 ADEQ submitted additional information prepared by the third party to EPA

May 16, 2016 EPA Region 6 approved site-specific criteria (SSC) revisions for temperature and TDS for the Little River and approved the removal of the domestic water supply use

from the Red River

# Summary of Revised Provisions

By letter dated December 21, 2015, ADEQ submitted several water quality standards revisions adopted by the APC&EC via Minute Order No. 15-21 to EPA for review and approval. These revisions are located in Regulation 2.511 and Appendix A. They include revisions of temperature (30°C to 32°C) and TDS (100 mg/L to 138 mg/L) criteria for the Little River from Millwood Lake to its confluence with the Red River. They also include the removal of the domestic water supply use and the increase of the TDS criterion from 500 mg/L to 860 mg/L for the Red River from its confluence with the Little River to the Arkansas (AR)/Louisiana (LA) state line.

# II. Revised Provisions EPA Previously Approved

By letter dated May 16, 2016, EPA acted on three of the four revised provisions that are described in the above paragraph. EPA approved the revision of temperature and TDS criteria for the Little River from Millwood Lake to its confluence with the Red River and approved the removal of the domestic water supply use for the Red River from its confluence with the Little River to the AR/LA state line. Details of that approval can be found in the May 16, 2016 letter and its enclosed TSD. This same letter stated that EPA was taking no action on the TDS provision for the Red River, as further consideration of protection of the aquatic life use and downstream uses were necessary. EPA has now completed its review of this final provision and this technical support document will detail the rationale of EPA's disapproval of this provision.

# III. Revised Provisions EPA is Disapproving

In accordance with the requirements found in Regulation No. 2.306 of the Arkansas Water Quality Standards, SWEPCO contracted with FTN Associates, Ltd., to complete a use

attainability analysis (UAA) of the receiving waterbodies affected by the revised site-specific criteria. The portions of the UAA that provided scientific justification for the appropriateness and protectiveness of the revised site-specific water quality criterion for TDS in the Red River from its confluence with the Little River to the AR/LA state line were considered for this action.

# Site-Specific Water Quality Criterion for TDS in the Red River

**Table 1.** Site-specific water quality criterion revision for TDS in the Red River submitted by ADEQ to EPA for review and approval.

Reach Description	Current Criterion	Proposed Criterion
Red River from its confluence with the Little River to the AR/LA state line	500 mg/L	860 mg/L

# **Disapproval Justification**

In its review of the SWEPCO UAA and additional submitted material, EPA determined that the documents did not sufficiently demonstrate protection of aquatic life use which is required by 40 CFR § 131.6 and also did not demonstrate protection of downstream use which is required by 40 CFR § 131.10(b).

The predominant source of evidence that was submitted to show that the aquatic life use was protected was a study completed by Buchanan et al. in 2003. This study surveyed fish that were present in the Arkansas portion of the Red River from 1995 to 2001. While the Buchanan study provides a historic reference, it only provides us with an understanding of the fish community in the Red River in 2001, not the current fish community. In addition, the Buchanan et al. 2003 study assessed presence and determined broad classifications of abundance (rare, uncommon, common, abundant). This gives us information about acute tolerances, but does not give us information about chronic tolerances. The presence of the fishes does not indicate that they are growing and reproducing at unimpaired rates. In addition, there was no discussion of the protection of the macroinvertebrate community in the Red River. Without additional information about the fish and macroinvertebrate community currently present in the Red River, EPA could not determine whether those communities are protected with the new criterion. Based on these concerns, EPA requested that additional information be submitted that demonstrated numeric tolerances of species that are found in the Red River. However, no additional data or information, only a representation of the Buchanan et al. 2003 study, was provided with which to assess the proposed criterion. Without additional supporting documentation, EPA was unable to assess whether the aquatic life use was being protected.

Downstream protection was also not demonstrated for this revised criterion. The proposed criterion is 860 mg/L and the downstream criterion in Louisiana is 780 mg/L. In the UAA submitted, the contractor reported the results of a mass-balance they performed using a 7Q-10 flow rate. These results showed predicted TDS concentrations at various spots in the Red River, including at the AR/LA state line. Also included in the UAA were the results of a second mass-balance performed by the contractor that used the same input values and model, but used a harmonic mean flow in place of a 7Q-10 flow rate. This second mass-balance was performed to justify that the proposed criterion is protective of LA uses. But the use of the harmonic mean

flow is not appropriate. Louisiana does not use flow when they are assessing their minerals criteria. Rather, when the State samples for minerals it determines whether the concentration is above the criteria in more than 30% of the samples collected. Given that this is how they perform their assessment, a criterion above the 780 mg/L could cause an impairment in LA waters and is not protective of downstream uses.

### Action

EPA is disapproving the site-specific TDS criterion change for the Red River from its confluence with the Little River to the AR/LA state line. Protection of aquatic life use was not demonstrated and downstream uses were not protected with this criterion.

As specified in 40 CFR § 131.21(c), this revised standard does not go into effect for CWA purposes until approved by EPA. Therefore, the previously approved criterion of 500 mg/L remains in effect for CWA purposes.

# IV. References

Buchanan, T.M., D. Wilson, L.G. Claybrook, and W.G. Layher. 2003. Fishes of the Red River in Arkansas. J. Ark. Acad. Sci. 57:18-26.

From:

Kesler, Karen [Kesler.Karen@epa.gov]

Sent:

Tuesday, July 26, 2016 10:46 AM

To: Cc: Jim Malcolm

Subject:

Clem, Sarah; Crocker, Philip Additional SWEPCO information

Hi Jim,

Thank you for the additional information that you have sent over about the SWEPCO rulemaking. EPA understands from your previous email that SWEPCO is willing to move forward with a 780 mg/L criteria for the lower Red River. The EPA believes this number will address issues of downstream protection. Unfortunately, since the previous 860 mg/L was adopted into Regulation No.2, a new rulemaking is going to have to occur for the 780 mg/L value to be considered. In order to allow this to happen, EPA is disapproving the previous 860 mg/L and the issues that we discussed during the previous conference call with downstream protection and protection of aquatic life use will be referenced in that technical support document. However with this email we wanted to recognize that we appreciate the additional information that you have put together for us and wanted to provide some feedback for the next rulemaking action.

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Please let me know if you have any questions.

Thank you, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Jim Malcolm [jtm@ftn-assoc.com]
Sent: Tuesday, July 26, 2016 11:13 AM

To: 'Kesler, Karen'

Cc: 'Clem, Sarah'; 'Crocker, Philip'; 'Philip Massirer'; Patrick J. Downey; James Malcolm

Subject: RE: Additional SWEPCO information

Hi Karen, this is really helpful and provides SWEPCO with the clear message from EPA that the revised proposed TDS criterion will address/resolve the downstream protection concern so that they can proceed with formalizing the revised criterion to the 780 mg/L value through the AR procedures in order to get it formally back in front of EPA for review/approval. As a further note, we will modify the tolerance value analysis as you have suggested below (i.e. only averaging alike metrics together) and circle back to that data to also search for hardness values to be included where we can find them. This revised information will be provided to EPA as soon as we prepare it and it will be part of the supplemental technical support/weight of evidence information that is submitted with the revised criterion also.

Thanks again, don't hesitate to let me or Sarah know if you, Phil, Russell or others have further questions or comments.

Sincerely, Jim



### Jim Malcolm jtm@ftn-assoc.com

FTN Associates 3 Innwood Circle, Suite 220 Little Rock, AR 72211

(501) 225-7779 work (501) 412-8185 mobile www.ftn-assoc.com webpage

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Tuesday, July 26, 2016 10:46 AM

To: Jim Malcolm

Cc: Clem, Sarah; Crocker, Philip

Subject: Additional SWEPCO information

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Please let me know if you have any questions.

Thank you, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From:

Kesler, Karen [Kesler.Karen@epa.gov]

Sent:

Tuesday, July 26, 2016 11:15 AM

To:

Jim Malcolm

Subject:

RE: Additional SWEPCO information

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I'll let you know if we have any further questions. Again, thank you for putting the additional information together.

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From: Jim Malcolm [mailto:jtm@ftn-assoc.com]

**Sent:** Tuesday, July 26, 2016 11:13 AM **To:** Kesler, Karen < Kesler. Karen@epa.gov >

Cc: 'Clem, Sarah' < CLEM@adeq.state.ar.us >; Crocker, Philip < crocker.philip@epa.gov >; 'Philip Massirer' < phm@ftn-

assoc.com>; Patrick J. Downey <pid@ftn-assoc.com>; James Malcolm <itm@ftn-assoc.com>

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Please let me know if you have any questions.

Thank you, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185

From: Allan Gates [AGates@mwlaw.com]
Sent: Thursday, August 11, 2016 11:32 AM
To: Honker, William (honker.william@epa.gov)

Cc: osbornec@adeq.state.ar.us; Kesler.karen@Epa.gov

Subject: SWEPCO 3rd Party Rulemaking -- TDS Site Specific Criterion in Lower Reach of the Red

River in Arkansas

Dear Bill,

I am writing to follow up on the conversation we had last Friday when you were in Little Rock.

As you know, your staff and FTN, the consultants working with SWEPCO in the above-styled rulemaking, have discussed the possibility of asking the Arkansas Pollution Control & Ecology Commission to reopen the SWEPCO rulemaking in order to reduce the new site specific TDS criterion in the lower reach of the Red River from 860 mg/L to 780 mg/L.

SWEPCO is willing to present such a request to the APC&E Commission, but only if it has confidence that such a change will ultimately be approved by EPA. The company does not want to ask the Commission to revisit this subject only to find out later that EPA will not approve the change.

For your convenience, I have reproduced below a recent e-mail exchange between Karen Kesler and Jim Malcolm. It is my understanding from our conversation that you are aware of this exchange and agree with the assessment it reflects.

To avoid any misunderstanding I would appreciate it if you would confirm whether my understanding on this subject is correct. I would also appreciate it if you would let me know if you are aware of any reason why you or your staff might have any problem approving a new TDS site specific criterion of 780 mg/L in the lower reach of the Red River in Arkansas using the data already generated and submitted.

Thank you in advance for your attention to this inquiry.

Needless to say, if you have any questions please do not hesitate to call or send me an e-mail.

Best regards,

Allan

# MITCHELL WILLIAMS

**Allan Gates** 

T 501.688.8816 | F 501.918.7816

<u>agates@mwlaw.com</u> | <u>MitchellWilliamsLaw.com</u>

425 W. Capitol Ave. | Ste. 1800 | Little Rock, AR 72201

Mitchell, Williams, Selig, Gates & Woodyard, P.L.L.C.

Karen Kesler – Jim Malcolm Email Exchange:

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Tuesday, July 26, 2016 11:15 AM

To: Jim Malcolm

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Thanks Jim,

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**Sent:** Tuesday, July 26, 2016 11:13 AM **To:** Kesler, Karen < Kesler.Karen@epa.gov>

Cc: 'Clem, Sarah' < CLEM@adeq.state.ar.us >; Crocker, Philip < crocker.philip@epa.gov >; 'Philip Massirer' < phm@ftn-

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agates@mwlaw.com | MitchellWilliamsLaw.com
425 W. Capitol Ave. | Ste. 1800 | Little Rock, AR 72201
Mitchell, Williams, Selig, Gates & Woodyard, P.L.L.C.

From: Allan Gates [AGates@mwlaw.com]
Sent: Monday, August 22, 2016 12:55 PM

To: 'jtm@ftn-assoc.com'

Subject: FW: SWEPCO 3rd Party Rulemaking -- TDS Site Specific Criterion inLower Reach of the Red

River in Arkansas [WARNING: SPF validation failed]

From: Honker, William [mailto:honker.william@epa.gov]

Sent: Friday, August 12, 2016 1:27 PM

To: Allan Gates

Cc: osbornec@adeq.state.ar.us; Kesler, Karen; Watson, Jane; Crocker, Philip

Subject: RE: SWEPCO 3rd Party Rulemaking -- TDS Site Specific Criterion in Lower Reach of the Red River in Arkansas

[WARNING: SPF validation failed]

Allan,

Thanks for your email concerning the SWEPCO rulemaking. I am aware of the email exchange below and concur with Karen Kesler's assessment of the situation. To clarify a couple of points regarding your inquiry:

- As you know, we disapproved the portion of the SWEPCO rulemaking that dealt with a TDS change for the lower Red River. It is our understanding that a new rulemaking would have to be undertaken if SWEPCO would like to propose a 780 mg/L criteria for TDS for the lower Red River.
- Regarding the Buchanan study, it can be used as supporting evidence, but it needs to be submitted in
  conjunction with tolerance values for species found in the Red River (or as closely related as they can
  find) and the benthic data that the applicant has for the Red River. All the pieces are needed for the
  justification, not solely the Buchanan study. Dr. Kesler's email below outlines recommendations for
  such data.

As we discussed last week, ADEQ will be the applicant's primary point of contact for any future proposal. However, I also encourage the applicant to solicit and incorporate input from my staff early in any subsequent rulemaking process, in order to promote early resolution of any issues. Of course, we cannot guarantee a specific outcome for a future rulemaking request. Our final decision on such a request must be based on the final submission and the quality of the information contained therein.

Bill

William K. Honker, P.E.
Director, Water Division (6WQ)
EPA Region 6
Dallas, TX
214-665-3187
<a href="http://www.epa.gov/region6/water/index.htm">http://www.epa.gov/region6/water/index.htm</a>



From: Allan Gates [mailto:AGates@mwlaw.com]
Sent: Thursday, August 11, 2016 11:32 AM
To: Honker, William < honker.william@epa.gov>

Cc: osbornec@adeq.state.ar.us; Kesler, Karen < Kesler.Karen@epa.gov>

Subject: SWEPCO 3rd Party Rulemaking -- TDS Site Specific Criterion in Lower Reach of the Red River in Arkansas

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Thank you in advance for your attention to this inquiry.

Needless to say, if you have any questions please do not hesitate to call or send me an e-mail.

Best regards,

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To: Jim Malcolm

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Cc: 'Clem, Sarah' < CLEM@adeq.state.ar.us >; Crocker, Philip < crocker.philip@epa.gov >; 'Philip Massirer' < phm@ftn-

assoc.com>; Patrick J. Downey <pid@ftn-assoc.com>; James Malcolm <jtm@ftn-assoc.com>

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**FTN Associates** 3 Innwood Circle, Suite 220 Little Rock, AR 72211

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Cc: Clem, Sarah; Crocker, Philip

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Thank you, Karen

Karen Kesler, Ph. D. Water Quality Standards Coordinator Water Division EPA Region 6 (214)665-3185



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Confidentiality Notice: This electronic mail transmission and any attachment may constitute an attorney-client communication that is privileged at law. It is not intended for transmission to, or receipt by, any unauthorized persons. If you have received this electronic mail transmission in error, please delete it from your system without copying it, and notify the sender by reply e-mail or by calling (501) 688-8800 Little Rock, AR (479) 464-5650 Rogers, AR (512) 480-5100 Austin, TX (267) 757-8780 Newtown, PA or (870) 336-9292 Jonesboro, AR so that our address record can be corrected.

From: Heather Ferguson [hlf@ftn-assoc.com]
Sent: Tuesday, September 27, 2016 2:19 PM

To: 'Kesler.Karen@epa.gov'; Philip Crocker; Russell Nelson

Cc: 'Allan Gates'; 'Frank Mills'; Patrick Downey; James Malcolm; 'Sarah Clem'

Subject: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the

Red River in Arkansas

Attachments: Dissolved minerals lethal and sublethal endpoints and hardness 2016-09-27.xlsx; 2016-09-27

M-K Kesler.pdf

Tracking: Recipient Read

'Kesler.Karen@epa.gov'

Philip Crocker
Russell Nelson
'Allan Gates'
'Frank Mills'
Patrick Downey
James Malcolm
'Sarah Clem'
PJD

Read: 9/27/2016 4:00 PM

### Good afternoon Karen,

On behalf of Southwestern Electric Power Company, please find attached the requested supplemental information for dissolved minerals tolerance values.

The attached information includes a memorandum that briefly describes our approach to obtaining the information you requested and a spreadsheet summarizing lethal and sublethal toxicity endpoints and associated hardness values. We believe this information satisfies your request and demonstrates through a weight-of-evidence approach the protection of aquatic life in the downstream reach of the Red River.

If you have any questions or comments regarding the attached information, please do not hesitate to call Jim Malcolm, Pat Downey, or me at (501) 225-7779.

Respectfully submitted,



Heather Ferguson FTN Associates, Ltd.

3 Innwood Circle, Suite 220 🗫 Little Rock, AR 72211

hlf@ftn-assoc.com

(501) 225-7779  $9\!\!o$  fax (501) 225-6738

http://www.ftn-assoc.com



### **MEMORANDUM**

DATE:

September 27, 2016

TO:

Karen Kesler, PhD

United States Environmental Protection Agency, Region VI

FROM:

Pat Downey, Jim Malcolm

FTN Associates, Ltd.

**SUBJECT:** 

Lethal and Sublethal Tolerance Values for Dissolved Minerals

FTN No. R06510-0010-002

On behalf of Southwestern Electric Power Company (SWEPCO), FTN Associates, Ltd. (FTN) appreciates the opportunity to offer supplemental information regarding lethal and sublethal tolerance values for dissolved minerals.

Per your request in an email to Jim Malcolm dated July 26, 2016, we reorganized the data set of tolerance values to summarize the values by lethal and sublethal endpoints. The attached spreadsheets summarize the lethal and sublethal endpoints. The spreadsheet tab titled "filter criteria" provides the list of steps taken to filter through the records and eliminate records that would not be useful in this comparison (such as studies that contained confounding factors or studies performed on species not found in North America).

You also requested hardness values associated with the toxicity values, which we were able to obtain for some studies in EPA's ECOTOX database or from the publication from which the tolerance values were derived. The tolerance values reported herein are from laboratory tests in which laboratory water was spiked with a particular salt of interest. When the experiment involves calcium- or magnesium-based salts of chloride or sulfate, the addition of chloride or sulfate automatically includes the addition of hardness. For example, a calcium chloride endpoint value (e.g., an LC50) of 1,000 mg/L automatically corresponds to a hardness of 1,330 mg/L. Therefore, all of the endpoints based on calcium or magnesium salts will be confounded with high levels of hardness. Endpoints based on sodium salts do not have this property. The attached spreadsheet includes two tabs that provide pairings of lethal and sublethal endpoints with hardness values. These would be the most appropriate data to use to evaluate hardness and toxicity.

Karen Kesler, PhD September 27, 2016 Page 2

If you have any questions or comments regarding this information, please do not hesitate to call Jim Malcolm, Pat Downey, or Heather Ferguson at (501) 225-7779.

CC: Philip Crocker, EPA Region 6 (with attachments)
Russell Nelson, EPA Region 6 (with attachments)
Sarah Clem, Arkansas Department of Environmental Quality (with attachments)
Frank Mills, American Electric Power (with attachments)
Allan Gates, Mitchell Williams Law (with attachments)

R:\WP\_FILES\06510-0010-002\CORRESPONDENCE\2016-09-27 M-K KESLER -- SUPPLEMENTAL TOLERANCE VALUES AND HARDNESS DATA TO EPA\2016-09-27 M-K KESLER.DOCX



Statistic         ICOO         LCSO           Minimum         1,000         1,250           Maximum         1,000         2,260           Minimum         401         1,250           Mean         1,000         2,260           Minimum         4,970         4,970           Maximum         4,970         1,188           Maximum         4,970         1,186           Maximum         1,196         3,000           Maximum         1,196         3,526           Ninimum         1,196         3,526           Ninimum         3,000         3,000           Mean         3,000         3,500           Mean         1,196         3,526           Nakimum         5         13           Maximum         1,196         3,526           Nakimum         1,000         3,526           Adrimum         1,000         3,526           Maximum         1,000         13,400           Adaximum         10,000         1,300           Adaximum         10,000         1,300           Adaximum         10,000         1,300           Inimum         10,000         1,300<					E	ENDPOINT (mg/L)	(1)	
Biomphalaria pjetiferi         Minimum         1,000         Incolor           Ceriodaphnia dubia         Namimum         1,000         1,599           Maximum         1,000         1,599         1,599           Maximum         2,160         1,599         1,599           Maximum         800         2,760         1,599           Maximum         8,970         1,599           Maximum         4,970         1,599           Maximum         1,250         3,136           Maximum         1,290         3,136           Daphnia nagna         1,136         3,136           Maximum         1,1300         1,1000           Maximum         1,1300         1,1000           Maximum         1,1300         1,1300           Maximum         1,0000         1,1300           Maximum         1,0000         1,1300           Maximum         1,0000         1,1300	Chemical	Species	Statistic	LC00 (NR-ZERO)		LC100 (NR-1 FTH)	1051/1051	orois, irois
Biomphalaria pjetiferi         Mean         1,000           Ceriodaphnia dubia         Maximuum         40.1           Ceriodaphnia dubia         Minimuum         40.1           Masimuum         800         2,260           Masimuum         800         2,260           Mean         4,970           Masimuum         4,970           Minimuum         4,970           Minimuum         3,000           Maximuum         1,266           Maximuum         3,000           Maximuum         1,196         3,500           Maximuum         1,3400           Maximuum         1,0000           Lepomis macrochirus         Maximuum         1,0000           Maximuum         10,000           Maximuum         10,000           Maximuum         10,000           Maximuum         10,000           Maximuum         2,573           Minimuum         10,000			Minimum	1,000		(	בטבר/ בטבר	NOEL/NOEC
Cyprinodon variegatus         Maximum         1,000           Cyprinodon variegatus         Minimum         4,970           Daphnia hyalina         Minimum         4,970           Daphnia hyalina         Minimum         1,256           Daphnia magna         Minimum         4,970           Maximum         3,000           Maximum         1,196         3,526           Maximum         1,13400           Maximum         1,13400           Maximum         5         13           Maximum         1,136         3,526         4           Maximum         5         13         10           Maximum         1,136         3,526         4           Maximum         1,136         3,526         4           Maximum         1,13400         13,400           Maximum         1,1300         13,400           Maximum         1,1300         1,3400           Maximum         1,0,000         1,300 <td></td> <td>Riomoholoria nfaiffari</td> <td>Mean</td> <td>1,000</td> <td></td> <td></td> <td></td> <td></td>		Riomoholoria nfaiffari	Mean	1,000				
Ceriodaphnia dubia         Immum         401         1,250           Mean         721         1,838           Maximum         800         2,260           N         A4970           Maximum         4,970           Maximum         4,970           Maximum         1,196         3,000           Maximum         1,196         3,526         4           Mean         Maximum         1,1300         1           Maximum         1,000         8,350         1           Maximum         1		naffiafd pupingidingia	Maximum	1,000				
Ceriodaphnia dubia         Minimum         401         1,250           Mean         721         1,898           Maximum         800         2,260           Minimum         4,970           Maximum         4,970           Maximum         3,000           Maximum         1,196         3,000           Maximum         1,196         3,000           Maximum         1,196         3,526           Maximum         1,196         3,526           Maximum         1,196         3,526           Maximum         1,196         3,526           Maximum         1,1300         3,000           Maximum         1,1300         3,000           Maximum         1,1340         13,400           Maximum         1,3400         13,400           Maximum         1,3400         13,400           Maximum         1,3400         10,000           Maximum         1,300         13,400           Maximum         1,300         13,300           Maximum         1,300         13,300           Maximum         1,000         1,300           Maximum         10,000         1,300			z	1				
Ceriodaphnia dubia         Mean         721         1,598           Maximum         800         2,260           Namimum         4,970           Maximum         4,970           Maximum         4,970           Maximum         4,970           Maximum         3,000           Maximum         1,196         3,500           Maximum         1,196         3,526         4           Maximum         1,3400         13,400           Maximum         1,3400         13,400           Maximum         1,3400         13,400           Maximum         1,3400         13,400           Maximum         1,000         8,350           Lepomis macrochirus         Maximum         10,000         11,300           N         N         A         A           N         Maximum         10,000         11,300           N         N         A			Minimum	401	1.250	1 599		
Cyprinodon variegatus         Maximum         800         2,550           Oxprinodon variegatus         Mean         4,970           Maximum         4,970           Maximum         3,000           Maximum         1,196         3,500           Maximum         1,196         3,526         4           Maximum         1,3400         13,400         13,400           Maximum         1,3400         13,400         13,400           Maximum         13,400         13,400         13,400           Maximum         1,000         8,356         1           Maximum         10,000         1,300         1           Lepomis macrochirus         Maximum         10,000         1,300           Mean         10,000         1,300         4           Maximum         10,000         1,300           Maximum         10,000         4           Maximum         10,000		Confordantantal	Mean	721	1.898	1 599		
Numinum         5         4           Cyprinodon variegatus         Minimum         4,970           Daphnia hyalina         Mean         4,970           Daphnia hyalina         Minimum         3,000           Daphnia hyalina         Maximum         1,136           Daphnia hyalina         Minimum         1,136           Daphnia hyalina         Minimum         1,136           Daphnia hyalina         Maximum         1,136           Daphnia hyalina         Minimum         1,136           Maximum         1,136         3,526           Maximum         13,400           Maximum         13,400           Maximum         13,400           Maximum         13,400           Maximum         10,000           Lepomis macrochirus         Minimum           Maximum         10,000           Maximum		cerioaapunia aubia	Maximum	800	2.260	1 500		
Opperition of on variegatus         Minimum magna         4,970           Daphnia hyalina polymorpha         Minimum magna         1,970           Daphnia hyalina magna polymorpha         Minimum magna         1,970           Gambusia affinis         Minimum maximum			z	2	4	1,333		
Cyprinodon variegatus         Mean         4,570           Dophnia hyalina         Minimum         4,570           Dophnia hyalina         Minimum         3,000           Daphnia magna         Minimum         1,196         3,526           Daphnia magna         Minimum         1,196         3,526           Naximum         1,196         3,526           Naximum         1,196         3,526           Naximum         1,196         3,526           Naximum         1,3400           Maximum         13,400           Naximum         13,400           Maximum         13,400           Maximum         13,400           Maximum         10,000         10,086           Maximum         10,000         10,086           Maximum         10,000         11,300           Maximum         10,000         11,300           Maximum         10,000         11,300           Minimum         10,000         11,300           Maximum         10,000         11,300           Maximum         10,000         11,300           Maximum         10,000         11,400           Maximum         10,000			Minimum		0707	7		
Cyprinodon variegatus Maximum 4,970			Mean		0/6,4			
Daphnia hyalina         Minimum         4,570           Daphnia hyalina         Maximum         3,000           Masimum         1,196         3,000           Masimum         1,196         3,526           Masimum         1,196         3,526           Maximum         1,196         3,526           Maximum         1,196         3,526           Maximum         1,3400           Maximum         13,400           Maximum         13,400           Maximum         13,400           Maximum         1,000           Maximum         10,000           Ictalurus punctatus         Maximum           Maximum         10,000           Maximum         4,485           Maximum         43           Maximu		Cyprinodon variegatus	Maximum		4,970			
Daphnia hyalina         Minimum         1           Daphnia hyalina         Minimum         3,000           Mean         3,000           Maximum         172         590           Maximum         1,196         3,526           Minimum         1,196         3,526           Minimum         1,196         3,526           Minimum         13,400         13,400           Maximum         10,000         8,350           Lymnaca offinis         Maximum         10,000         10,086           Minimum         10,000         11,300         Maximum           Maximum         10,000         10,000         Maximum           Maximum         10,000         Maximum         4,485           Maximum         4,38         Maximum         10,000           Maximum         43         Maximum         43			IVIDALITURII		4,970			
Daphnia hyalina         Minimum         3,000           Mean         3,000           Maximum         172         590           Daphnia magna         Maximum         1,196         3,526           Daphnia magna         Mean         1,196         3,526           N         N         5         13           Maximum         1,196         3,526         13           Maximum         1,196         3,526         13           Maximum         1,196         3,526         13           Maximum         13,400         13,400         13,400           Maximum         13,400         13,400         13,400           Mean         10,000         13,400         13,400           Mean         10,000         10,006         10,006           Lepomis macrochirus         Maximum         10,000         11,300           Minimum         10,000         11,300         10,000           Micropterus dolomieu         Maximum         10,000         4           Maximum         10,000         4         10,000           Maximum         43         10,000         10,000           Maximum         43         10,000			Z		1			
Daphnia hyalina         Mean         3,000           Daphnia magna         Minimum         172         590           Daphnia magna         Minimum         172         590           Daphnia magna         Maximum         1,196         3,526           Maximum         5         13         13,400           Maximum         13,400         13,400           Maximum         10,000         8,350           Maximum         10,000         11,300           Maximum         10,000         11,300           Maximum         10,000         11,300           Maximum         10,000         4           Minimum         10,000         4           Maximum         10,000         11,300           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum <t< td=""><td></td><td></td><td>Minimum</td><td></td><td>3,000</td><td></td><td></td><td></td></t<>			Minimum		3,000			
Opening in a control of the pomis macrocthirus         Maximum         3,000           Daphnia magna         Maximum         172         590           Mean         833         1,826           Maximum         5         13           Maximum         1,196         3,526           Maximum         1,196         3,526           Maximum         1,3400         13,400           Maximum         13,400         13,400           Maximum         13,400         13,400           Maximum         13,400         13,400           Maximum         10,000         8,350           Maximum         10,000         11,300           N         3         7           Minimum         10,000         4           Mean         10,000         4           Maximum         10,000         4           Mean         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         4,485           Maximum         43           Maximum         43           Ma		Danhaia hualina	Mean		3,000			
Minimum   172   590     Maximum   1,196   3,526     Maximum   1,196   3,500     Minimum   1,196   3,500     Maximum   1,000   1,300     Maximum   1,000   1,300     Maximum   1,000   4,485     Maximum   1,0,000   Maximum   1,0,000     Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000     Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maximum   1,0,000   Maxi			Maximum		3,000			
Daphnia magna         Minimum         172         590           Mean         833         1,826           Maximum         1,196         3,526           Minimum         1,196         3,526           Minimum         1,196         3,526           Maximum         13,400           Maximum         10,000         8,350           Maximum         10,000         11,300           Maximum         10,000         11,300           Minimum         3         7           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         Maximum           Maximum         4,485           Maximum         10,000         A           Maximum         4,33         A           Maximum         43         A           Maximum         43         <			Z		1			
Daphnia magna         Mean         833         1,826           Maximum         1,196         3,526           Minimum         3,196         3,526           Minimum         13,400         13,400           Maximum         13,400         13,400           Maximum         13,400         13,400           Minimum         13,400         13,400           Maximum         13,400         13,400           Maximum         13,400         13,400           Mean         13,400         13,400           Mean         10,000         10,006           Mean         10,000         10,086           Minimum         10,000         11,300           Micropterus dolonijeu         Minimum         4,485           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         4,485         Maximum           Maximum         10,000         4           Maximum         4,485         Maximum           Maximum         4,33         A           Maximum         43         A           Maximum <td></td> <td></td> <td>Minimum</td> <td>172</td> <td>290</td> <td>3,196</td> <td></td> <td></td>			Minimum	172	290	3,196		
Maximum 1,196 3,526     N		Danhaia maana	Mean	833	1,826	3.724		
Ninimum			Maximum	1,196	3,526	4,780		
Dreissena polymorpha  Mean  Gambusia affinis  Gambusia affinis  Gambusia affinis  Minimum  Lepomis macroctifirus  Micropterus dolomieu  Micropterus dolomieu  Morone saxatilis ssp. x  Maximum  Morone saxatilis ssp. x  Minimum  Morone saxatilis ssp. x  Minimum  Maximum  Maxi			z	2	13	6		
Dreissena polymorpha         Mean         Mean           Gambusia affinis         Minimum         13,400           Gambusia affinis         Mean         13,400           Maximum         13,400           N         3           Minimum         10,000           Lepomis macrochirus         Minimum           Lepomis macrochirus         Maximum           Minimum         10,000           N         3           Amean         10,000           Minimum         4,485           N         Amean           Minimum         10,000           Maximum         10,000           Maximum         10,000           Maximum         10,000           Maximum         43           Morone saxatilis ssp. x         Minimum           Maximum         43           Maximum         43           Maximum         43           Maximum         43           Maximum         43			Minimum			10.000		
Maximum   13,400   Maximum   13,400   Minimum   13,400   Maximum   13,400   Maximum   13,400   Maximum   13,400   Maximum   13,400   Maximum   13,400   Maximum   10,000   10,000   Minimum   10,000   11,300   Maximum   10,000   11,300   Maximum   10,000   11,300   Maximum   10,000   Maximum   10,		Orginal or propried	Mean			10,000		
Minimum   13,400     Mean   13,400     Mean   13,400     Maximum   13,400     Minimum   13,400     Minimum   10,000   10,086     Minimum   10,000   11,300     Minimum   10,000   11,300     Maximum   10,000   11,300     Maximum   10,000   11,300     Maximum   10,000   12,573     Minimum   10,000   4     Minimum   10,000   10,000     Maximum   10,000   10,000		orersseria polymorpha	Maximum			10,000		
Gambusia affinis         Minimum         13,400           Mean         13,400           Naximum         13,400           Minimum         3           Lepomis macrochirus         Minimum         10,000         8,350           Mean         10,000         10,086           Maximum         2,573           Minimum         3         7           Minimum         2,573           Maximum         4,485           Minimum         10,000         4           Minimum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         10,000         4           Maximum         4,38         1           Morone saxatilis ssp. x         Maximum         43           Maximum         43         Maximum           Maximum         43         Maximum           Maximum         43         Maximum	um chlorida		z			8		
io affinis         Mean         13,400           Maximum         13,400           Ninimum         3           Minimum         10,000         8,350           Maximum         10,000         10,086           Maximum         10,000         11,300           Maximum         3         7           Maximum         3,365         Minimum           Sp.         Maximum         4,485           N         Maximum         4,485           N         Maximum         4           Maximum         10,000         4           Maximum         10,000         N           N         2         Minimum           Amaximum         43         Maximum           Maximum         43         Maximum           Maximum         43         Maximum			Minimum		13,400			
Punctatus         Maximum         13,400           Punctatus         Naminum         3           Punctatus         Maximum         10,000         8,350           Maximum         10,000         10,086           Maximum         10,000         11,300           Maximum         3         7           Minimum         3,365           N         4485           N         Minimum         4485           Maximum         10,000         4           Maximum         10,000         4           Maximum         43         10,000           N         2         Minimum           Axatilis ssp. x         Mean         43           Maximum         43         10,000           Maximum         43         10,000		Gombineto affinis	Mean		13,400			
Punctatus         Minimum         3           Mean         10,000         8,350           Minimum         10,000         10,086           Maximum         10,000         11,300           N         3         7           Maximum         2,573           Maximum         4,485           N         4           Maximum         10,000           N         4           Maximum         10,000           N         2           Maximum         43           Maximum         43           Maximum         43           Maximum         43           Maximum         43		Sampasia allinis	Maximum		13,400			
punctatus         Mean         10,000         8,350           Maximum         10,000         8,350           Mean         10,000         11,300           Maximum         10,000         11,300           N         3         7           Minimum         3,365           Maximum         4,485           N         4           Minimum         10,000           N         2           Maximum         10,000           N         2           Maximum         43           Maximum         43           Maximum         43           Maximum         43			z		20,400			
punctatus         Mean         10,000         8,350           Maximum         10,000         8,350           Maximum         10,000         10,086           Maximum         3         7           Minimum         3,365           N         4,485           N         4           Minimum         10,000           Vadolonieu         Maximum           Maximum         10,000           N         2           Maximum         43           Maximum         43           Maximum         43           Maximum         43           Maximum         43			Minimim		)	000		
punctatus         Maximum         10,000         8,350           Maximum         10,000         10,086           Maximum         10,000         11,300           Maximum         3         7           Minimum         2,573           Maximum         10,000           vs dolomieu         Maximum         4,485           Maximum         10,000           Maximum         10,000           Maximum         43           Maximum         43           Maximum         43           Maximum         43           Maximum         43           Maximum         43			Mean			10,000		
Maximum         10,000         8,350           Maximum         10,000         10,086           Maximum         10,000         11,300           N         3         7           Minimum         2,573           Mean         4,485           N         4           Minimum         10,000           N         2           Maximum         43		Ictalurus punctatus	Maximim			10,000		
Moreorchirus         Minimum         10,000         8,350           Maximum         10,000         10,086           N         3         7           Minimum         2,573           Mean         3,365           Maximum         4,485           N         4           Minimum         10,000           N         2           N         2           N         2           Minimum         43           Minimum         43           Minimum         43           Maximum         43           Maximum         43			Z			10,000		
Mean			Minimim	10.000	0.350	7		
Maximum   10,000   Maximum   10,000   Maximum   10,000   Maximum   10,000   Maximum   10,000   Maximum   10,000   N   2   Minimum   43   Maximum   43   Ma		A Section Control of the Control of	Mean	10,000	10.086			
Sp.         Minimum         3           Mean         Mean         10,000           N         Minimum         10,000           Maximum         10,000           N         2           Minimum         43           Maximum         43           Maximum         43           Maximum         43           Maximum         43		repornis macrocnirus	Maximum	10.000	11 300			
Sp.         Minimum Maximum           Maximum         10,000           Winimum         10,000           Maximum         10,000           N         2           Minimum         43           Maximum         43           Maximum         43           Maximum         43			z	3	7			
Sp.         Mean           Maximum         10,000           Winimum         10,000           Maximum         10,000           N         2           Minimum         43           Maximum         43           Maximum         43			Minimum		2,573			
Sp.         Maximum         N           N         N         N           Minimum         10,000           Maximum         10,000           N         2           Minimum         43           Maximum         43           Maximum         43		Towns and the second	Mean		3 365			
N         N           Minimum         10,000           Mean         10,000           N         2           Minimum         43           Maximum         43           Maximum         43		туппией sp.	Maximum		4.485			
Minimum         10,000           Mean         10,000           Maximum         10,000           N         2           Minimum         43           Minimum         43           Maximum         43           Maximum         43			Z		4			
us dolomieu Maximum N Minimum Minimum Maximum Maximum Minimum Maximum Maximum Maximum			Minimum	10.000		Ī		
Maximum N Minimum Minimum Minimum Mean Maximum Maximum			Mean	10.000		1		
N Minimum Oxatilis ssp. x Mean Maximum			Maximim	10,000				
Minimum oxatilis ssp. x Mean Maximum			Z	20000				
axatilis ssp. x Mean Maximum			Minimim	7				
Maximum Maximum			Marinian	43				
Muaximum		axatılıs ssp. x	Mean	43				
			Maximum	43				71

# Toxicity Endpoint Definitions:

LC50/EC50: the concentration at which 50% lethality was observed LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival) LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically LC00 (NR-ZERO): the concentration at which no lethality was observed (100% survival)

significant effect was observed (compared to the control group)

NOEL/NOEC. the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

				Ġ	ENDPOINT (mg/L)	(1)	A COLUMN TO SERVICE STATE OF THE PERSON STATE
				1	10.00	,_	
Chemical	Species	Statistic	(NR-ZERO)	LC50/ EC50	(NR-LETH)	LOEL/LOEC	NOEL/NOEC
		Minimum		> 1,000			
	Operator latinos	Mean		> 1,000			
	Oryzius iutipes	Maximum		> 1,000			
		z		9			
		Minimum		2,110	2,640		
Calcium chloride	Dimenhales promelas	Mean		4,990	8,072		
(cont.)	rillepinales prometas	Maximum		> 6,660	20,000		
		Z		4	10		
		Minimum	10,000		20,000		
	Candarwitrans	Mean	10,000		20,000		
	Salinei vitieus	Maximum	10,000		20,000		
		z	co		17	3	
		Minimum		270			
	Austropotamobius	Mean		370			
	pallipes ssp. pallipes	Maximum		480			
		z		3			
		Minimum	467	880	933		
	Carindanhnia duhia	Mean	467	1,075	933		
	רבווסמחלווווח ממסום	Maximum	467	1,270	933		
		Z	2	2	2		
		Minimum	467	1,330	1,491		
	Danhair mann	Mean	548	2,693	1,861		
	המשנווום ווומשוום	Maximum	747	3,699	2,044		
Magnesium		z	4	5	5		
chloride		Minimum		16,500			
	of the state of the state of	Mean		17,667			
	Gambusia ajjinis	Maximum		18,750			
		z		33			
		Minimum		440			
		Mean		590			
	Orconectes limosus	Maximum		092			
		z		3			
		Minimum		2,120			
	Salar Salar Salar Salar	Mean		2,827			
	Pimephales promelas	Maximum		3,520			
		z		c			
		Minimum	616				
		Mean	616				
	Acipenser sinensis	Maximum	616				
		Z	1				
		Minimum	10,000	> 10,000			
Afterna of the same of the		Mean	10,000	> 10,000			
Sodium chioride	Acroneuria abnormis	Maximum	10,000	> 10,000			
		z	1	1			
		Minimum	10,000	> 10,000			
	Action contract	Mean	10,000	> 10,000			
	Agnetina capitata	Maximum	10,000	> 10,000			

Toxicity Endpoint Definitions:

LCOO (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LCSO/ECSO: the concentration at which 50% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

Lethal endpoints for calcium-, magnesium-, and sodium-based salts of chloride and sulfate.

				EN	ENDPOINT (mg/L)	()	
	i		0007	/C20/	LC100	3201/1201	Cacia) iacia
Chemical	Species	Statistic	(NR-ZEKO)	ECSU	(NK-LEIH)	LOEL/LOEC	NOEL/NOEC
		Minimum		17,880			
	Anguilla rostrata	Mean		19,665			
		Maximum		21,450			
		z		2			
		Minimum		23,000			
		Mean		27,375			
	Argia sp.	Maximum		32,000			
		z		80			
		Minimum		5,100			
	4 - 200 - 200 - 200 - 200	Mean		7,095			
	Asellus communis	Maximum		> 10,000			
		z		11			
		Minimum			3,700		
	Biomphalaria	Mean			4,120		
	alexandrina	Maximum			4,600		
		z			5		
		Minimum		1,645			
		Mean		1,645			
	Brachionus calyciflorus	Maximum		1,645			
		z		1			
		Minimum	10,000				
	Brachycentrus	Mean	10,000				
1	numerosus	Maximum	10,000				
Sodium chioride		Z	1				
(רסוורי)		Minimum			2,600	1	
	400	Mean			2,600		
	מחוווות בנותווכתות	Maximum			2,600		
		Z			1		
		Minimum		22,457		20,550	20,500
	Contraction of the Contraction o	Mean		24,524		20,775	20,725
	caenornabands elegans	Maximum		25,786		21,000	20,950
		z		9		2	2
		Minimum		008'9			
		Mean		8,154			
	Carassius auratus	Maximum		10,100			
		z		6			
		Minimum		13,750			
		Mean		13,750			
	cardssius cardssius	Maximum		13,750			
		z		1			
		Minimum	467	132	1,701	300	< 300
		Mean	881	1,453	2,442	2,306	1,182
	сеподарнија дирід	Maximum	1,250	3,380	4,500	5,000	2,500
		z	6	44	7	32	32
		Minimum		1,250			
		Mean		1,250			
	Ceriodaphnia reticulata	Maximum		1,250			
		z		1			

Toxicity Endpoint Definitions:

LC00 (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LC50/EC50: the concentration at which 50% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

Chironomus dilutus         Statistic         (NR-ZERO)         LCSO         (NR-ZERO)           Chironomus dilutus         Maximum         1,200         12,000         1,000           Chironomus dilutus         Maximum         1,200         12,000         1,200           Chironomus dilutus         Maximum         1,200         12,000         1,200           Maximum         1,200         12,000         1,200         1,200           Maximum         1,200         1,200         1,200         1,200           Maximum         1,200         2         4         1,200         1,200           Maximum         10,000         1         1,000         1,0	Channa punctata Minimum  Chironomus dilutus Mean  Chironomus tentans Maximum  Clarias gariepinus Mean  Carias gariepinus Mean  Calex sp.  Minimum  Culex sp.  Minimum  Mean  Culex sp.  Minimum  Mean  Culiseta incidens  Maximum  Mean  Maximum  Mean  Maximum  Mean  Maximum  Mean  Maximum				ENDPOINT (mg/L)	-)	
Chironopunctator         Mainimum         1,200         12,000           Chironopuus dilutus         Mainimum         1,200         12,000           Chironomus dilutus         Mainimum         4,394         5,7810         7,810           Chironomus dilutus         Minimum         4,580         5,781         7,810         7,810           Chironomus fentans         Minimum         10,000         3         1,1600         7,810           Chironomus fentans         Minimum         10,000         3         1,1600         7,810           Chironomus fentans         Minimum         1,000         3         1,1         1,1           Chironomus fentans         Minimum         1,000         3         1,1         1,1           Maximum         1,000         1,4,040         3         1,1         1,1           Corbicula manifensis         Minimum         4,850         2         1,0         1,0           Maximum         4,850         2         1,0         1,0         1,0         1,0           Culex sp.         Minimum         4,850         2,21         1,0         1,0         1,0           Culex sp.         Minimum         1,0,200         1,0,200         1,0,200	Chironomus dilutus Maximum Minimum Mean Minimum Mean Minimum Mean Maximum Minimum Mini			LC50/ EC50	LC100 (NR-LETH)	LOEL/LOEC	NOEL/NOEC
Channa punctata         Mean         1,200         12,000           Chironomus dilutus         Maximum         1,200         12,000           Chironomus dilutus         Minimum         4,940         5,793         7,810         7,810           Chironomus tentans         Maximum         7,810         6,280         11,600         7,810           Chironomus tentans         Maximum         10,000         4         3         1,11           Maximum         10,000         14,040         7,810         1,200           Maximum         10,000         14,040         7,810         1,810           Maximum         10,000         14,040         1,800         1,810           Corbicula manifensis         Maximum         4,850         10,800         1,800           Maximum         4,850         10,800         1,800         1,800           Colexicula manifensis         Maximum         4,850         10,800         1,900           Maximum         4,850         10,800         1,0800         1,0800           Colexicula manifensis         Maximum         4,850         10,800         1,0800           Maximum         4,850         10,800         1,0800         1,0800	Chironomus dilutus Mean Chironomus dilutus Mean Chironomus tentans Mean Clarias batrachus Maximum Mean Clarias batrachus Maximum Mean Clarias batrachus Maximum Mean Corbicula manilensis Maximum Mean Corbicula manilensis Maximum Mean Culex sp. Maximum Mean Maximum Mean Maximum Mean Maximum Maximum Mean Maximum Maximum Minimum Minimum Minimum Minimum Maximum Minimum Min	Minimun		12,000			
Chironomus tentons         Maximum         1,200         Name           Chironomus tentons         Minimum         4,340         5,793         7,810         7,810           Chironomus tentons         Minimum         1,200         4         3         1,810           Chironomus tentons         Minimum         10,000         4         3         1           Minimum         10,000         4         3         1           Mean         10,000         3         1           Minimum         14,040         1         1           Mean         14,040         1         1           Maximum         14,040         1         1           Minimum         4,850         10,800         1           Maximum         4,850         10,800         1           Minimum         4,850         10,800         1           Maximum         4,850         10,800         1           Maximum         10,500         10,800         1           Maximum         10,500         13,000         13,000           Culiex sp.         Minimum         4,850         10,800           Maximum         10,500         13,000	Chironomus dilutus  Chironomus tentans  Maximum  Maximum  Chironomus tentans  Maximum  Chironomus tentans  Maximum  Maximum  Chironomus tentans  Maximum  Maximu			12,000			
Chironomus dilutus         Minimum         4,940         5,781         7,810           Chironomus dilutus         Maximum         7,810         6,280         11,660         7,810           Maximum         7,810         6,280         11,600         7,810           Minimum         10,000         3         1,1           Chironomus tentans         Minimum         10,000         3         1,1           Maximum         10,000         1,440         1         1           Minimum         1,400         1,440         1         1           Maximum         1,400         1,080         1         1           Maximum         4,850         10,800         1         1           Maximum         4,850         10,800         1         1           Corbicula manijensis         Maximum         4,850         10,800         1           Maximum         4,850         10,800         1         1           Corbicula manijensis         Maximum         4,850         10,800         1           Maximum         4,850         10,800         1         1           Corbicula manijensis         Maximum         1,0,300         1,300         1,30	Chironomus dilutus Maximum Mean Chironomus tentans Maximum Mean Clarias batrachus Minimum Mean Clarias batrachus Minimum Mean Corbicula manilensis Maximum Mean Corbicula manilensis Maximum Mean Corbicula manilensis Maximum Mean Culex sp. Maximum Mean Culex sp. Maximum Mean Culiseta incidens Maximum Mean Culiseta incidens Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Maximum Mean Maximum			12,000			
Chironomus dilutus         Minimum         4,940         5,733         7,810         7,810           Chironomus dilutus         Naximum         7,810         6,280         3,013         7,810           Chironomus tentans         Maximum         10,000         4         3         1           Chironomus tentans         Mean         10,000         3         1           Mean         10,000         14,040         7           Mean         10,000         1         1           Minimum         14,040         1         1           Minimum         4,850         2         10,800           Maximum         4,850         10,800         1           Maximum         4,850         10,800         1           No         N         1         1         1           Naximum         4,850         10,800         10,800         10,800           Corbicula manifersis         Maximum         4,850         10,800         10,800           Naximum         1,050         10,800         N         10,800           Culex sp.         Maximum         4,700         13,000         N           Naximum         2,000         13,000	Chironomus dilutus Naximum Minimum Minimum Minimum Minimum Naximum Nax	Z	1	1			
Chironomus dilutus         Mean         6,375         5,993         9,073         7,810           Maximum         1,810         4         3         1           Chironomus tentans         Minimum         10,000         4         3         1           Chironomus tentans         Maximum         10,000         14,040         1         1           Maximum         1,000         14,040         1         1         1           Maximum         1,000         1,000         1         1         1           Clarias gariepinus         Maximum         4,850         10,800         1         1           Maximum         4,850         10,800         1         1         1         1           Corbicula manijensis         Maximum         4,850         10,800         1	Chironomus dilutus Maximum Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Mean Minimum Mean Minimum Mean Maximum Mean Maximum Mean Minimum Mean Maximum Mean Minimum Mean Minimum Mean Minimum Mean Minimum Mean Maximum Mean Maximum Mean Minimum Mean Minimum Mini	Minimum		5,793	7,810	7,810	4,940
Chironounus binitatos         Maximum         7,810         6,280         11,600         7,810           Chironomus tentans         Maximum         10,000         —         4         3         1           Chironomus tentans         Maximum         10,000         —         —         —         —           Chironomus tentans         Maximum         14,040         —         —         —         —           Maximum         10,000         —         14,040         —         —         —           Maximum         10,000         —         14,040         —         —         —           Maximum         4,850         —         10,800         —         —         —           Maximum         4,850         —         10,800         —         —         —           Corbicula manifensis         Maximum         4,850         —         10,800         —         —           Maximum         4,850         —         10,800         —         —         —         —         —         —         —         —         —         —         —         —         —         —         —         —         —         —         — <td< td=""><td>Clarias bartatas  Chironomus tentans  Maximum  Max</td><td></td><td>6,375</td><td>5,993</td><td>9,073</td><td>7,810</td><td>4,940</td></td<>	Clarias bartatas  Chironomus tentans  Maximum  Max		6,375	5,993	9,073	7,810	4,940
Corbicula maniferasis         Minimum         1,0,000         1,0,000           Corbicula maniferasis         Maximum         10,000         1           Corbicula maniferasis         Maximum         14,040         1           Corbicula maniferasis         Maximum         4,850         10,800           Culiseta incidens         Maximum         4,850         10,800           Culiseta incidens         Maximum         4,850         10,800           Culiseta incidens         Maximum         4,850         10,800           Maximum         4,850         10,800         10,800           Maximum         4,850         10,800         10,800           Maximum         4,850         10,800         10,800           Maximum         4,850         10,800         10,800           Maximum         4,850         10,200         10,800           Maximum         4,850         10,200         10,800           Maximum         4,700         13,000         10,800           Maximum         4,700         13,000         10,800           Maximum          4,700         13,000         10,800           Maximum         2,000         2,810         10,800	Clarias batrachus Minimum Minimum Minimum Maximum Maximum Maximum Minimum Minimum Minimum Minimum Minimum Minimum Minimum Minimum Mean Minimum Minimum Mean Minimum Mean Minimum Mean Minimum Minimum Mean Minimum Mean Minimum M			6,280	11,600	7,810	4,940
Chironomus tentons         Minimum         10,000         Minimum         10,000           Chironomus tentons         Mean         10,000         10,000           Maximum         10,000         14,040         10           Maximum         14,040         10         10           Maximum         14,040         10         10           Maximum         14,040         10         10           Maximum         10         10         10           Maximum         10         10,800         10           Maximum         10,200         10,800         10           Maximum         6,221         10,800         10           Maximum         6,221         1         10           Maximum         6,221         1         10           Maximum         6,221         1         1           Maximum         10,300         1         1         1           Maximum         10,300         1         1         1           Maximum         4,700         13,000         1         1           Maximum         4,700         13,000         1         1           Maximum         4,700         1	Carios batrachus Aminimum Aminimum Calarias batrachus Carios batrachus Carios gariepinus Maximum Mean Maximum Mean Maximum Max	Z	2	4	c	1	1
Chironomus tentons         Mean         10,000         Processor           Clarias batrachus         Maximum         14,040         Processor           Clarias batrachus         Mean         14,040         Processor           Clarias batrachus         Mean         14,040         Processor           Maximum         14,040         Processor         Processor           Maximum         4,850         10,800         Processor           Maximum         4,700         13,000         Processor	Carias batrachus Maximum Minimum Mean Minimum Minimum Mean Minimum	Minimum					
Corbicular spinarion         Maximum         10,000         Amonimum         Amonimum <td>Carias batrachus Maximum Mean  Carias gariepinus Maximum Mean  Carbicula manilensis Maximum Mean  Calex sp. Maximum Mean  Calex sp. Maximum Mean  Culiseta incidens Maximum Mean  Maximum Minimum Minimum Minimum Minimum Maximum Mean  Maximum Minimum Minimu</td> <td></td> <td>10,000</td> <td></td> <td></td> <td></td> <td></td>	Carias batrachus Maximum Mean  Carias gariepinus Maximum Mean  Carbicula manilensis Maximum Mean  Calex sp. Maximum Mean  Calex sp. Maximum Mean  Culiseta incidens Maximum Mean  Maximum Minimum Minimum Minimum Minimum Maximum Mean  Maximum Minimum Minimu		10,000				
Corbicula manifensis         N         1         4,040         N           Corbicula manifensis         Minimum         14,040         N	Carias batrachus Minimum Minimum Minimum Mean Maximum Mean Maximum Mean Minimum Minimu						
Clarias barrachus         Minimum         14,040         N           Clarias barrachus         Maximum         14,040         N           Clarias gariepinus         Minimum         610         N           Clarias gariepinus         Minimum         4,850         10,800           Maximum         4,850         10,800           Maximum         4,850         10,800           Maximum         4,850         10,800           Maximum         6,221         10,800           Maximum         1         1           Caricotopus trifasciatus         Maximum         6,221           Maximum         10,500         10,800           Maximum         10,500         10,800           Maximum         2         10,800           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,020         6,120           Maximum         2,000         850           Maximum         4,940         6,137           Maximum         2,000         850 <tr< td=""><td>Clarias batrachus Mean Mean Mean Mean Maximum Mean Minimum Mean Minimum Mean Minimum Mean Minimum Mean Minimum Mean Maximum Minimum Mean Maximum Mean Minimum Mean Minimum Min</td><td>Z</td><td>1</td><td></td><td></td><td></td><td></td></tr<>	Clarias batrachus Mean Mean Mean Mean Maximum Mean Minimum Mean Minimum Mean Minimum Mean Minimum Mean Minimum Mean Maximum Minimum Mean Maximum Mean Minimum Mean Minimum Min	Z	1				
Clarias batrachus         Mean         14,040         N           Clarias gariepinus         Minimum         610         N           Clarias gariepinus         Minimum         4,850         10,800         N           Corbicula manijensis         Mean         4,850         10,800         N           Corbicula manijensis         Maximum         4,850         10,800         N           Maximum         4,850         10,800         N           Maximum         4,850         10,800         N           Maximum         6,221         1         N           Maximum         1,850         N         N           Maximum         10,200         N         N           Maximum         4,700         13,000         N           Maximum         4,020         6,130         7,810           Maximum         4,020         6,130         7,810           Cyprinella leeds; <t< td=""><td>Clarias batrachus Mean  Maximum  Maximum  Minimum  Minimum  Mean  Minimum  Maximum  Minimum  Maximum  Maximum</td><td>Minimum</td><td></td><td>14,040</td><td></td><td></td><td></td></t<>	Clarias batrachus Mean  Maximum  Maximum  Minimum  Minimum  Mean  Minimum  Maximum  Minimum  Maximum	Minimum		14,040			
Carias partecinas         Maximum         14,040         1           Almimum         610         1         1           Almimum         655         10,800         1           Amimum         4,850         10,800         1           Amimum         4,850         10,800         1           Amimum         4,850         10,800         1           Maximum         4,850         10,800         1           Maximum         4,850         10,800         1           Maximum         6,221         1         1           Maximum         10,200         13,000         1           Maximum         10,200         13,000         1           Maximum         4,700         13,000         1           Maximum         4,700         13,000         1           Maximum         4,700         13,000         1           Maximum         4,020         6,120         7,810           Maximum         4,020         6,120         7,810           Cyprinella leeds i         Minimum         4,020         6,120         7,810           Mean         2,000         2,000         850         1      <	Clarias partachas  Minimum  Minimum  Corbicula manilensis  Corbicula manilensis  Minimum  Culex sp.  Culiseta incidens  Culiseta incidens  Maximum  Mean  Minimum  Mean  Minimum  Mean  Minimum  Mean  Minimum  Mean  Maximum  Mean  Minimum  Mean  Maximum  Mean  Minimum  Maximum  Minimum  Maximum			14,040			
Corbicula manifensis         N         1         0           Culex sp.         Minimum         4,850         10,800         10,800           Culex sp.         Maximum         4,850         10,800         10,800           Corbicula manifensis         Minimum         4,850         10,800         10,800           Maximum         4,850         10,800         10,800           Minimum         6,221         1         1           Maximum         10,200         1         1           Maximum         10,200         13,000         1           Culex sp.         Maximum         4,700         13,000         1           Maximum         4,700         13,000         1         1           Culex sp.         Maximum         4,700         13,000         1           Maximum         3,100         6,070         7,810         1           Maximum         4,020         6,130         7,810         1           Cyprinella leedsi         Maximum         2,000         850         1           Maan         2,000         13,000         850         1           Maan         3,000         2,000         850         1	Carias gariepinus Minimum Minimum Minimum Minimum Minimum Minimum Minimum Minimum Maximum Ninimum Mean Maximum Mean Minimum			14,040			
Clarias gariepinus         Minimum         610         Mes           Carbicula manilensis         Minimum         4,850         10,800         10,800           Carbicula manilensis         Minimum         4,850         10,800         10,800           Minimum         4,850         10,800         10,800         10,800           Minimum         4,850         10,800         10,800           Maximum         6,221         10,800         10,800           Maximum         6,221         10,800         10,800           Minimum         1,0,500         13,000         10,800           Minimum         4,700         13,000         13,000           Maximum         4,700         13,000         13,000           Culiseta incidens         Minimum         4,700         13,000         13,000           Maximum         4,700         6,120         7,810         10,800           Cyprinella leedsi         Maximum         4,940         6,120         7,810           Maximum         4,000         6,120         7,810         1           Masimum         4,000         6,120         7,810         1           Masimum         4,000         1,100	Carios gariepinus Minimum Mean  Corbicula manilensis Maximum Minimum Minimum Maximum N Minimum Mean  Culex sp. Maximum Mean  Culex sp. Maximum Mean  Culiseta incidens Maximum Mean  Culiseta incidens Maximum Mean  Culiseta incidens Maximum Minimum	Z		1			
Clarias gariepinus         Mean         655           Corbicula manilensis         Minimum         4,850         10,800           Corbicula manilensis         Minimum         4,850         10,800           Maximum         4,850         10,800         10,800           Maximum         4,850         10,800         10,800           Maximum         6,221         1         1           Maximum         6,221         1         1           Maximum         10,200         1         1           Minimum         4,700         13,000         1           Maximum         4,700         13,000         1           Maximum         4,700         13,000         1           Mean         4,700         13,000         1           Oxprinella leedsi         Mean         4,700         13,000           Maximum         2         4         2           Maximum         2,000         6,137         7,810           Maximum         2,000         6,137         7,810           Maximum         2,000         6,137         7,810           Maximum         2,000         4,020         1,1           Maximum <td>Carias gariepinus Mean  Corbicula manilensis Maximum  Corbicula manilensis Maximum  Minimum  Culex sp.  Culex sp.  Culiseta incidens  Culiseta incidens  Minimum  Minimum  Minimum  Minimum  Maximum  Max</td> <td>Minimum</td> <td></td> <td>610</td> <td></td> <td></td> <td></td>	Carias gariepinus Mean  Corbicula manilensis Maximum  Corbicula manilensis Maximum  Minimum  Culex sp.  Culex sp.  Culiseta incidens  Culiseta incidens  Minimum  Minimum  Minimum  Minimum  Maximum  Max	Minimum		610			
Carbicula manilensis         Maximum         700           Carbicula manilensis         Mean         4,850         10,800           Masimum         4,850         10,800         10,800           Maximum         4,850         10,800         10,800           Maximum         1,0,201         10,800         10,800           Maximum         6,221         1         1           Minimum         10,350         1         1           Maximum         10,500         13,000         1           Maximum         4,700         13,000         1           Culiseta incidens         Minimum         4,700         13,000         1           Maximum         4,700         13,000         1         1           Oxprinella leedsi         Minimum         4,020         6,120         7,810         1           Maximum         2         4         2         4         2         1           Oaphnia ambigua         Maximum         2,000         850         850         1           Maximum         2         4         2         4         1         1           Maximum         2,000         4,643         5,065         1 </td <td>Corbicula manilensis Maximum  Corbicula manilensis Maximum  Minimum  Minimum  Minimum  Maximum  Maximum  Maximum  Minimum  Minimum  Minimum  Minimum  Minimum  Maximum  Minimum  Minimum  Minimum  Minimum  Minimum  Minimum  Minimum  Minimum  Maximum  Maximu</td> <td></td> <td></td> <td>655</td> <td></td> <td></td> <td></td>	Corbicula manilensis Maximum  Corbicula manilensis Maximum  Minimum  Minimum  Minimum  Maximum  Maximum  Maximum  Minimum  Minimum  Minimum  Minimum  Minimum  Maximum  Minimum  Minimum  Minimum  Minimum  Minimum  Minimum  Minimum  Minimum  Maximum  Maximu			655			
On Minimum         4,850         2           Corbicula manilensis         Mean         4,850         10,800           Mean         4,850         10,800           Maximum         10,800         10,800           Minimum         6,221         1           Maximum         6,221         1           Minimum         10,350         1           Maximum         10,350         1           Maximum         4,700         13,000           Minimum         4,700         13,000           Minimum         4,700         13,000           Maximum         4,700         13,000           Minimum         4,700         13,000           Maximum         4,700         13,000           Opphnia ambigua         Minimum         4,920         6,120         7,810           Maximum         2         4         2         A	Corbicula manilensis Mean Minimum Mean Minimum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Minimum Min		,	700			
Corbicula manifensis         Minimum         4,850         10,800           Maximum         4,850         10,800         10,800           Maximum         1         1         1           Cricotopus trifasciatus         Minimum         6,221         1           Maximum         6,221         1         1           Maximum         10,200         1         1           Maximum         10,500         1         1           Maximum         4,700         13,000         1           Opphnia ambigua         Maximum         2         4         2           Maximum         2         4         2         850           Maximum         2         4         2         Minimum           Opphnia ambigua         Maximum         2,000         850         1           Maximum         2         4         2         850           Maximum         2	Corbicula manilensis  Mean  Maximum  Minimum  Mean  Culex sp.  Culiseta incidens  Culiseta incidens  Culiseta incidens  Minimum  Mean  Minimum  Mean  Maximum  Mean  Minimum  Mean  Maximum  Mean  Minimum  Mean  Maximum  Mean  Maximum  Mean  Maximum  Maximu	Z		2			
Corbicula manilensis         Mean         4,850         10,800           Maximum         4,850         10,800         1           Minimum         6,221         1         1           Maximum         6,221         1         1           Maximum         10,200         1         1           Maximum         10,500         1         1           Maximum         4,700         13,000         1           Culiseta incidens         Maximum         4,700         13,000         1           Minimum         4,700         13,000         1         1           Cyprinella leedsi         Minimum         4,700         13,000         1           Opphnia ambigua         Minimum         2         4         2           Maximum         2,000         6,120         7,810         1           Maximum         2,000         6,120         7,810         1           Maximum         2,000         6,120         7,810         850           Maximum         2,000         6,120         7,810         1           Maximum         2,000         8,500         1         1           Minimum         2,000	Corbicula manilensis Mean  Maximuum  Minimuum  Maximuum	Minimum			10,800		
Culexas inclusions         Maximum         4,850         10,800           Nanimum         6,221         1           Mean         6,221         8           Maximum         1         1           Maximum         10,200         8           Mean         10,500         8           Minimum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           N         1         1           Maximum         4,020         6,120         7,810           Cyprinella leedsi         Maximum         4,940         6,137         7,810           Maximum         2         4         2         850           Maximum         4,940         6,137         7,810         850           Maximum         2,000         8,500         850           Maximum         2,000         8,50         850           Maximum         2,000         8,50         850           Maximum         2,000         8,50         1           Maximum         2,000         8,50         8,50           Maximum         2,000         8,50	Culex sp.  Culex sp.  Culex sp.  Culiseta incidens  Culiseta incidens  Culiseta incidens  Culiseta incidens  Culiseta incidens  Maximum  Mean  Minimum  Mean  Minimum  Mean  Minimum  M				10,800		
Cricotopus trifasciatus         Minimum         1         1           Cricotopus trifasciatus         Mean         6,221         C           Mean         6,221         C         C           Maximum         10,200         C         C           Minimum         10,350         C         C           Maximum         4,700         13,000         C           Maximum         4,700         13,000         C           Maximum         4,700         13,000         C           Maximum         4,020         6,130         7,810         C           Oxprinello leedsi         Minimum         4,020         6,137         7,810         C           Oxprinello leedsi         Minimum         4,940         6,137         7,810         C           Maximum         2         4         2         Minimum           Maximum         2         4         2         Minimum           Maximum         2,000         8,500         8,50           Maximum         2,000         8,50         Minimum           Maximum         2,000         8,50         Minimum           Maximum         2,000         8,50         Mini	Culex sp.  Culex sp.  Culex sp.  Culiseta incidens  Culiseta incidens  Culiseta ancidens  Culiseta incidens  Minimum  Maximum  Ma				10,800		
Cricotopus trifasciatus         Mean         6,221         Mean         6,221           Mean         6,221         0 <td>Cricotopus trifasciatus Mean  Culex sp.  Culiseta incidens Maximum  Culiseta incidens Maximum  Culiseta incidens Maximum  Culiseta incidens Maximum  Minimum  Minimum  Minimum  Minimum  Minimum  Maximum  Maximum</td> <td>Z</td> <td></td> <td></td> <td>1.</td> <td></td> <td></td>	Cricotopus trifasciatus Mean  Culex sp.  Culiseta incidens Maximum  Culiseta incidens Maximum  Culiseta incidens Maximum  Culiseta incidens Maximum  Minimum  Minimum  Minimum  Minimum  Minimum  Maximum  Maximum	Z			1.		
Mean         6,221         Maximum         6,221           IN         1         1           Minimum         10,350         1           Maximum         10,500         13,000           Mean         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         7,810           Mean         4,700         5,130           Maximum         4,940         6,137         7,810           Maximum         2         4         2           Maximum         2         4         2           Minimum         2,000         850           Maximum         3,467         4,432           Maximum         3,466         7,500         8,000	Mean  Maximum  Mean  Maximum  Mean  Maximum  Maximum  Minimum  Minimum  Maximum  Max	Minimum		6,221			
Maximum         6,221           N         1           Minimum         10,350           Maximum         2           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         7,810           Maximum         4,020         6,120         7,810           Maximum         2         4         2	Maximum  Minimum  Maximum  Max			6,221			
Minimum         1           Mean         10,200           Maximum         10,500           Maximum         2           Minimum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         7,810           Maximum         4,020         6,120         7,810           Maximum         2         4         2           Maximum         2         4         2           Maximum         2,000         850           N         1         1           Maximum         2,000         8,000           Maximum         3,466         7,500           Maximum         3,466         7,500	Minimum Maximum N Minimum Maximum N Minimum Maximum N Minimum N Minimum N Minimum N Minimum N Minimum N Maximum N Maximum			6,221			
Minimum         10,200           Mean         10,350           Maximum         10,500           Minimum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Maximum         4,700         13,000           Mean         4,000         5,120           Maximum         4,920         6,120         7,810           Maximum         4,940         6,137         7,810           Maximum         2,000         850           Maximum         2,000         8,000           Maximum         3,466         7,500         8,000	Minimum Mean Maximum Minimum Mean Maximum Mean Maximum N Minimum N Minimum N Minimum N Minimum Maximum N Maximum	Z		1			
Mean         10,350           N         2           N         2           Minimum         4,700         13,000           Maximum         4,700         13,000           N         1         1           Minimum         3,100         6,070         7,810           Maximum         4,940         6,137         7,810           Maximum         2,000         850           Minimum         2,000         850           Maximum         2,000         850           Maximum         2,000         850           Maximum         2,000         850           Minimum         280         > 467         4,321           Maximum         3,466         7,500         8,000	Mean Naximum Naximum Mean Naximum Naximum Naximum Naximum N Maximum	Minimum		10,200			
Maximum         10,500           N         2           Minimum         4,700         13,000           Maximum         4,700         13,000           N         1         1           Minimum         3,100         6,070         7,810           Maximum         4,940         6,130         7,810           Maximum         2,000         850           Minimum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         > 467         4,321           Maximum         2,000         850           Maximum         2,000         850           Maximum         2,000         8,000           Maximum         3,466         7,500         8,000	Maximum Minimum Maximum Maximum Maximum Maximum N Minimum N Minimum N Minimum N Minimum Maximum N Maximum N Maximum	Mean		10,350			
N         2           Minimum         4,700         13,000           Mean         4,700         13,000           Maximum         4,700         13,000           N         1         1           Minimum         3,100         6,070         7,810           Maximum         4,940         6,137         7,810           Maximum         2,000         850           Minimum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         > 467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Minimum Mean Maximum Minimum Maximum N Minimum Mean Maximum Mean Maximum Mean Maximum	Maximum		10,500			
Minimum         4,700         13,000           Mean         4,700         13,000           Maximum         13,000         13,000           Minimum         3,100         6,070         7,810           Maximum         4,940         6,137         7,810           Minimum         2         4         2           Minimum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         2,600         850           N         1         1         1           Maximum         280         2,600         850           Maximum         3,466         7,500         8,000	Minimum Mean Maximum Mean Maximum Mean Maximum Mean Minimum Mean Maximum	Z		2			
Mean         4,700         13,000           Maximum         4,700         13,000           N         13,000         13,000           Minimum         3,100         6,070         7,810           Maximum         2         4         2           Minimum         2,000         850           Maximum         2,000         850           Maximum         2,000         850           N         1,000         850           N         467         4,321         1           Maximum         280         5,665         1           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Mean Maximum Minimum Mean Maximum Mean Mean Mean Maximum Maximum Maximum Maximum Maximum Maximum Maximum Maximum Maximum	Minimum		4,700	13,000		
Maximum         4,700         13,000           N         1         1           Mainimum         3,100         6,070         7,810           Maximum         4,020         6,137         7,810           N         2         4         2           Minimum         2,000         850           Maximum         2,000         850           N         2,000         850           N         1         1           Minimum         280         > 467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Maximum Ninimum Makimum Maximum Ninimum Ninimum Maximum Ninimum Naximum Naximum Naximum Naximum Naximum Naximum Naximum			4,700	13,000		
Minimum         3,100         6,070         7,810           Mean         4,020         6,120         7,810           Maximum         4,940         6,137         7,810           Minimum         2         4         2           Maximum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Minimum Maximum Naximum Ninimum Mean Maximum Naximum Maximum Maximum Maximum Maximum Maximum Maximum			4,700	13,000		
Minimum         3,100         6,070         7,810           Mean         4,020         6,120         7,810           Maximum         2         4         2           Minimum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Minimum Maximum N Minimum Maximum N Minimum N Maximum Maximum Maximum Maximum Maximum Maximum	Z		1	1		
Mean         4,020         6,120         7,810           Maximum         4,940         6,137         7,810           N         2         4         2           Minimum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         > 467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Mean Maximum Ninimum Mean Maximum N Minimum N Maximum Maximum Maximum Maximum	Minimum		6,070	7,810		
Maximum         4,940         6,137         7,810           N         2         4         2           Minimum         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         > 467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Maximum Minimum Mean Maximum N Minimum Maximum Maximum Maximum		4,020	6,120	7,810		
M         2         4         2           Minimum         2,000         850           Maximum         2,000         850           N         1         1         1           Minimum         280         467         4,321         1           Mean         1,902         4,643         5,065         8,000           Maximum         3,466         7,500         8,000         8,000	Minimum Mean Maximum N Minimum Maximum Maximum Maximum	Ĭ		6,137	7,810		
Minimum         2,000         850           Mean         2,000         850           Maximum         2,000         850           N         1         1           Minimum         280         > 467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Minimum Mean Maximum N Minimum Maximum Maximum Maximum	Z	2	4	2		
Mean         2,000         850           Maximum         2,000         850           N         1         1         1           Minimum         280         > 467         4,321         1           Mean         1,902         4,643         5,065         9           Maximum         3,466         7,500         8,000         8,000	Mean Maximum Ninimum Maximum Maximum	Minimum		2,000		820	440
Maximum         2,000         850           N         1         1         1           Minimum         280         > 467         4,321         1           Mean         1,902         4,643         5,065         9           Maximum         3,466         7,500         8,000         9	Maximum N Minimum Mean Maximum			2,000		850	440
N         1         1           Minimum         280         > 467         4,321           Mean         1,902         4,643         5,065           Maximum         3,466         7,500         8,000	Minimum Mean Maximim			2,000		850	440
Minimum         280         > 467           Mean         1,902         4,643           Maximum         3,466         7,500	Minimum Mean Maximim	Z		1		1	1
Mean 1,902 4,643 Maximum 3,466 7,500	Mean	Minimum			4,321		
Maximum 3,466 7,500	Maximim			4,643	5,065		
				7,500	8,000		

Toxicity Endpoint Definitions:

LCOO (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LCSO/ECSO: the concentration at which 50% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

OEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

VOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

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			E	IDPOINT (mg/	(1)	
		1000	1C50/	LC100		
Species	Statistic	(NR-ZERO)	ECS0	(NR-LETH)	LOEL/LOEC	NOEL/NOEC
	Minimum	441	1,250			
Danhaia aulay	Mean	441	1,883			
Capillia pales	Maximum	441	3,050			
	Z	1	4			
	Minimum			10,000		
Draiseana palamorpha	Mean			15,000		
Dieisseria polymorphia	Maximum			20,000		
	Z			2		
	Minimum		2,230			
Elliptic complanata	Mean		2,450			
בייוליום בסייוליום	Maximum		2,670			
	Z		2			
	Minimum		244			7
Epioblasma torulosa	Mean		244			
ssp. rangiana	Maximum		244			H
	z		1			
	Minimum		7,500			
Chapter of photons	Mean		8,000			
בו הספתבוות התעורותות	Maximum		> 10,000			
	z		. 5			7
	Minimum	16,000		24,000		
Fundulus cominglis	Mean	16,000		24,000		
	Maximum	16,000		24,000		
	z	1		1		
	Minimum		17,550			
Gambusia affinis	Mean		17,917			
cuit de presentation	Maximum		18,100			7
	z		3			
	Minimum		11,540			J.
Gambusia halbrooki	Mean		11,540			
	Maximum		11,540			
	z		1			
	Minimum	10,000	7,700			
Gammarus	Mean	10,000	7,700			
pseudolimnaeus	Maximum	10,000	7,700			
	z	1	1			
	Minimum		8,530	M		
Gammoriis roosell	Mean		10,757			
California de l'Oceani	Maximum		12,600			
	Z		9			
	Minimum		3,200			
Cutation of a contraction	Mean		6,020			
Gyrdaius circumstriatus	Maximum		> 10,000			
	z		5			
	Minimum		3,009			
Surger military	Mean		3,044			
dyladius parvas	Maximum		3,078			
	z		2			
	Chemical Species  Daphnia pulex  Daphnia pulex  Dreissena polymorpha  Elliptio complanata  Epioblasma torulosa ssp. rangiana  Erpobdella punctata  Erpobdella punctata  Gambusia affinis  Gambusia affinis  Gammarus  Gammarus	Species  Mananus Seminalis  Eundulus seminalis  Eundulus seminalis  Eundulus appropriate Mananus of precise and propriate and mananus of ambusia affinis  Mananus of ambusia affinis  Mananus of ambusia circumstriatus  Mananus of ambusia mananus  Mananus o	Species Statistic (Winimum Dreissena polymorpha Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Mean Maximum Mean Sp. rangiana torulosa Sp. rangiana feliptio complanta torulosa Sp. rangiana (Maximum Mean Maximum Mean Maximum M	Species         Statistic         (NRZERO)         LCSO/ ECSO           Dophnia pulex         Minimum         441         1,883           Dreissena polymorpha         Maximum         441         1,883           Eliptio complanata         Minimum         2,230           Eliptio complanata         Minimum         2,230           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         5,450           Maximum         16,000         5,450           Maximum         16,000         5,000           Fundulus seminalis         Minimum         1,000           Maximum         16,000         7,700           Maximum         10,000         7,700           Gambusia halbrooki         Maximum         1,0,000         7,700           Dopumarus         Maximum         10,000         7,700           Documarus         Maximum         10,000         7,700           Maximum         10,000         7,700           Maximum         10,000         7,700 </td <td>Species         Statistic         (NRZERO)         LCSO/ ECSO           Dophnia pulex         Minimum         441         1,883           Dreissena polymorpha         Maximum         441         1,883           Eliptio complanata         Minimum         2,230           Eliptio complanata         Minimum         2,230           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         5,450           Maximum         16,000         5,450           Maximum         16,000         5,000           Fundulus seminalis         Minimum         1,000           Maximum         16,000         7,700           Maximum         10,000         7,700           Gambusia halbrooki         Maximum         1,0,000         7,700           Dopumarus         Maximum         10,000         7,700           Documarus         Maximum         10,000         7,700           Maximum         10,000         7,700           Maximum         10,000         7,700     <!--</td--><td>Species         Statistic (NR.ZERO)         LC500 (NR.LETH)           Dophhila pulex         Manimum         441         1,250 (NR.LETH)           Mean         441         1,250 (NR.LETH)         NR.LETH           Maximum         41         1,250 (NR.LETH)         NR.LETH           Mean         441         1,283 (NR.LETH)         1,000           Mean         1         4         10,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,240 (NR.LETH)         2,000           Maximum         2,450 (NR.LETH)         2,000           Maximum         2,240 (NR.LETH)         2,000           Maximum         16,000 (NR.LETH)         2,400           Maximum         16,000 (NR.LETH)         2,400           Maximum         16,000 (NR.LETH)         2,400           Mean         11,540 (NR.LETH)           Maximum         10,000 (NR.LETH)           Maximum         10,000 (NR.LETH)           Mean         10,000 (NR.LETH)           Mean         10,000 (NR.LETH)           Maximum         10,000</td></td>	Species         Statistic         (NRZERO)         LCSO/ ECSO           Dophnia pulex         Minimum         441         1,883           Dreissena polymorpha         Maximum         441         1,883           Eliptio complanata         Minimum         2,230           Eliptio complanata         Minimum         2,230           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         2,450           Maximum         16,000         5,450           Maximum         16,000         5,450           Maximum         16,000         5,000           Fundulus seminalis         Minimum         1,000           Maximum         16,000         7,700           Maximum         10,000         7,700           Gambusia halbrooki         Maximum         1,0,000         7,700           Dopumarus         Maximum         10,000         7,700           Documarus         Maximum         10,000         7,700           Maximum         10,000         7,700           Maximum         10,000         7,700 </td <td>Species         Statistic (NR.ZERO)         LC500 (NR.LETH)           Dophhila pulex         Manimum         441         1,250 (NR.LETH)           Mean         441         1,250 (NR.LETH)         NR.LETH           Maximum         41         1,250 (NR.LETH)         NR.LETH           Mean         441         1,283 (NR.LETH)         1,000           Mean         1         4         10,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,240 (NR.LETH)         2,000           Maximum         2,450 (NR.LETH)         2,000           Maximum         2,240 (NR.LETH)         2,000           Maximum         16,000 (NR.LETH)         2,400           Maximum         16,000 (NR.LETH)         2,400           Maximum         16,000 (NR.LETH)         2,400           Mean         11,540 (NR.LETH)           Maximum         10,000 (NR.LETH)           Maximum         10,000 (NR.LETH)           Mean         10,000 (NR.LETH)           Mean         10,000 (NR.LETH)           Maximum         10,000</td>	Species         Statistic (NR.ZERO)         LC500 (NR.LETH)           Dophhila pulex         Manimum         441         1,250 (NR.LETH)           Mean         441         1,250 (NR.LETH)         NR.LETH           Maximum         41         1,250 (NR.LETH)         NR.LETH           Mean         441         1,283 (NR.LETH)         1,000           Mean         1         4         10,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,230 (NR.LETH)         2,000           Maximum         2,240 (NR.LETH)         2,000           Maximum         2,450 (NR.LETH)         2,000           Maximum         2,240 (NR.LETH)         2,000           Maximum         16,000 (NR.LETH)         2,400           Maximum         16,000 (NR.LETH)         2,400           Maximum         16,000 (NR.LETH)         2,400           Mean         11,540 (NR.LETH)           Maximum         10,000 (NR.LETH)           Maximum         10,000 (NR.LETH)           Mean         10,000 (NR.LETH)           Mean         10,000 (NR.LETH)           Maximum         10,000

Toxicity Endpoint Definitions:

LC00 (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LC50/ECS0: the concentration at which 50% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

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Chemical Species  Hyalella azteca  Hydroptila angusta  Ictalurus punctatus Isonychia bicolor Isonychia bicolor Isonychia bicolor Iampsilis fasciola  Lampsilis siliquoidea  Lampsilis siliquoidea	8	Statistic Minimum Mean Maximum NIN NIN Minimum Mean Naximum N Minimum Maximum N Minimum Mean Maximum Maximum Maximum N Maximum Maximum Maximum Maximum N N N N N N N N N N N N N N N N N N	LC00 (NR-ZERO)	LC50/ EC50	LC100 (NR-LETH)	TOEL/LOEC	0.000
	s	Minimum Mean Maximum N N Minimum Maximum N Maximum Maximum Maximum Minimum Mean Minimum Mean Minimum Minimum Minimum N N Minimum N N N N N N N N N N N N N N N N N N					NOEL/NOEC
	s	Mean Maximum Minimum Mean Maximum Maximum Minimum Minimum Minimum Mean Minimum Maximum Maximum N Minimum N Minimum N Minimum N Minimum Maximum Maximum		1,200			
	S	Maximum Minimum Mean Maximum N Minimum Maximum N Minimum Mean Maximum Maximum Maximum N Maximum N Minimum N Minimum N Minimum Maximum		1,322			
	s	Minimum Mean Maximum N Maximum Mean Maximum Minimum Maximum Maximum Minimum Minimum Minimum Maximum Maximum		1,510			
	s	Minimum Maximum Naximum Maximum Minimum Maximum Naximum Maximum Minimum Minimum Minimum Maximum Maximum		5			
	s	Mean Maximum Minimum Mean Maximum Maximum Maximum Minimum Minimum Minimum Minimum Minimum Minimum Minimum		6,621			
	5	Maximum Minimum Mean Maximum Minimum Minimum Minimum Minimum Maximum Maximum		6,621			
	S	N Minimum Mean Maximum N Minimum Mean Maximum		6,621			
	5	Minimum Mean Maximum N Minimum Mean Maximum Mean		1			
	3	Mean Maximum N Minimum Mean Maximum	10,000		20,000		
	5	Maximum Minimum Mean Maximum N	10,000		20,000		
		N Minimum Mean Maximum N	10,000		20,000		
		Minimum Mean Maximum N	2		1		
		Mean Maximum N				1,000	200
		Maximum				2,875	2,850
		z			Ī	4,000	8,000
						8	10
		Minimum		817			
	u	Mean		817			
		Maximum		817			
		z		1			
		Minimum		113			
		Mean		1,498			
		Maximum		3,980			
		z		6			
		Minimum		168			
rawbsiis siidaala		Mean		1,806			
		Maximum		4,560			
		z		10			
		Minimum	5,220	1,295	9,103		
		Mean	8,407	9,430	9,103		
Lepomis macrocuirus	snun	Maximum	10,000	14,125	9,103		
		z	3	5	1		
		Minimum		2,510			
of the state of th		Mean		3,130		Y	
Ligumia recta		Maximum		3,750			
		z		2			
		Minimum		5,800			
and the second s		Mean		6,381			
rimnoariids nojjineisten		Maximum		> 7,500			
		z		8			
		Minimum		2,970	4,580		
		Mean		2,970	4,580		
Lirceus Jontinalis		Maximum		2,970	4,580		
		Z		1	1		
		Minimum	3,100	3,100	7,810	4,940	3,100
Jumpriculus variegatus		Mean	4,480	5,012	7,810	4,940	3,100
1		Maximum	4,940	5,589	7,810	4,940	3,100

Toxicity Endpoint Definitions:

LC00 (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LC30/ECS0: the concentration at which 50% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

Chemical				EN	ENDPOINT (mg/L)	r)	
	Species	Statistic	LC00 (NR-ZERO)	LCS0/ ECS0	LC100 (NR-LETH)	LOEL/LOEC	NOEL/NOEC
		Minimum		3,388	2,600		
	0.000	Mean		3,400	2,600		
	Lynnueu sp.	Maximum		3,412	2,600		
		Z		2	1		
	Maccaffortium	Minimum	10,000				
	maccajjenani	Mean	10,000				
	mexicanum ssp.	Maximum	10,000				
	ıntegrum	z	1				
		Minimum				1,000	200
	A CONTRACTOR OF THE PARTY OF TH	Mean				4,714	2,786
	maccajjennum sp.	Maximum				10,000	8,000
		z				7	7
		Minimum	< 1,000	1,000	2,000		
		Mean	2,875	3,563	4,375		
	Morone saxatilis	Maximum	000'2	000'L <	> 7,000		
		z	8	8	8		
		Minimum		1,930			
		Mean		1,930			
	Musculium transversum	Maximum		1,930			
		z		1			
		Minimum		2.569			
		Mean		2.569			
	Nais variabilis	Maximim		2 569			
Sodium chloride		z		1			
(cont.)		Minimim	5 340	4 270	5 340	5 340	3 320
		Moon	5,340	A 280	6,133	5 340	3 320
	Nephelopsis obscura	Mariania	040,0	7 210	007.7	5,340	3 330
		Maximum	2,240	4,310	7,720	7,040	3,350
		2	n	4	0	T	4
		Minimum	10,000				
	Control for Control	Mean	10,000				
	בורח לותאבטרבווט	Maximum	10,000				
		z	2				
		Minimum		2,540	2,060		
		Mean		2,540	2,060		
	Physa gyrina	Maximum		2,540	2,060		
		z		1	1		
		Minimim		3.500			
		Mean		5.068			
1	Physa heterostropha	Maximim		7.500			
		Z		17			
		Minimi	3 035	0.070	6 060	201	253
		Mass	3003	7,005	11 176	2 010	2 100
4	Pimephales promelas	Medil	3,220	11,000	11,120	0766	4,000
		Maximum	10,000	11,400	20,000	8,000	4,000
		z	13	57	3	35	33
		Minimum		6,150	4		
1	Planorbella	Mean		7,190			
Ü	campanulata	Maximum		> 10,000			

Toxicity Endpoint Definitions:

LC00 (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LC100 (NR-ZERO): the concentration at which 100% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

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				E	ENDPOINT (mg/L)	(1)	
Chemical	Species	Statistic	LC00 (NR-ZERO)	LC50/ EC50	LC100 (NR-LETH)	LOEL/LOEC	NOEL/NOEC
		Minimum		16,595			
	Poecilia Intining	Mean		17,665			
		Maximum		18,735			
		Minimum	0677	11 700			
		Mean	9 710				
	Poecilia reticulata	Maximim	11 700				
		Z	2				
		Minimum	10,000	3,526			
		Mean	10,000	3,526			
	Pycnopsyche guttifera	Maximum	10,000	3,526			
		Z	2	1			
		Minimum	10,000	3,526			
	Durnonsuche lenida	Mean	10,000	3,526			
		Maximum	10,000	3,526			
		Minimim	4.000				
		Mean	4,000				
	Rocio octojasciata	Maximum	4,000				
		z	1				
		Minimum	10,000				
	Candarwitrans	Mean	10,000				
		Maximum	10,000				
Sodium chloride		z	2				
(cont.)		Minimum		740			
	Sahaerium simile	Mean		920			
		Maximum		1,100			
		Z		7			
		Minimum		1,100			
	Sphaerium sp.	Mean		1,613			
		Z		8			0
		Minimum	10,000				
	Stenacron	Mean	10,000				
	interpunctatum	Maximum	10,000				
		z	1				
		Minimum	10,000	> 10,000			
	Tipula abdominait	Mean	10,000	> 10,000			
	Similar page	Maximum	10,000	> 10,000			
		z	2	1			
		Minimum		4,278			
	T. hifen tuhifen	Mean		5,311			
	i daijex tabijex	Maximum		6,008			
		z		3			
		Minimum		2,590			
	Williams contributed	Mean		2,675			
	VIIIOSA CONSTITCTA	Maximum		2,760			
		N.1		2			

Toxicity Endpoint Definitions:

LC00 (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LC30/ECS0: the concentration at which 100% lethality was observed

LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

				EI	ENDPOINT (mg/L)	(1)	
Chemical	Species	Statistic	LC00 (NR-ZERO)	LCS0/ ECS0	LC100 (NR-LETH)	LOEL/LOEC	NOEL/NOEC
		Minimim		3 310			
Sodium chloride		Mean		4,057			
(cont.)	Villosa delumbis	Maximum		5,230			
		z		e			
		Minimum	765	> 1,910			
	Coriodophoia dubia	Mean	1,126	1,940			
	רבווסממלווווות ממסומ	Maximum	1,470	> 1,970			
		Z	9	3			
		Minimum	360	> 1,970			1,600
	Company of the Company	Mean	866	> 1,970			1,600
	Daphnia magna	Maximum	1,470	> 1,970			1,600
		Z	9	1			1
		Minimum		> 56,000			
alcium culfato	Gambucia affinis	Mean		> 56,000			
ורוחווו אחוופרב	cumpusion of firms	Maximum		> 56,000			
		z		3			
		Minimum		2,980			
	and the second second	Mean		2,980			
	דבלים ווומרו מרוווו מז	Maximum		2,980			
		z		2			
		Minimum	1,470	> 1,970			
	Dimontalor promolor	Mean	1,470	> 1,970			
	rimepriales prometus	Maximum	1,470	> 1,970			
		z	2	3			
		Minimum	1,000				
	Biomphalaria	Mean	1,000				
	alexandrina	Maximum	1,000				
		z	1		/11/		
		Minimum			4,000		
	Bulinus truncatus	Mean			4,000		
		Maximum			4,000		
		2	001	C C C C	7000		
		Minimum	580	1,770	2,076		
	Ceriodaphnia dubia	Mean	1,047	1,770	2,076		
		NIAXIMUM	1,394	1,770	6,0,0		
Aagnesium sulfate		Minimim	360	788	4 071		
		Mean	692	1 978	4 071		
	Daphnia magna	Maximum	1,079	4,300	4,071		
		z	5	8	2		
		Minimum		15,500			
		Mean		15,500			
	Gambusia affinis	Maximum		15,500			
		Z		3			
		Minimum					450
	Heterorhabditis	Mean		1			450
	bacteriophora	Maximum					450
		z					1

LCD0 (NR-ZERO): the concentration at which no lethality was observed (100% survival)
LC50/EC50: the concentration at which 50% lethality was observed
LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)
LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)
NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

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				EN	ENDPOINT (mg/L)	(1)	
Chemical	Species	Statistic	LC00 (NR-ZERO)	LC50/ EC50	LC100 (NR-LETH)	LOEL/LOEC	NOEL/NOEC
		Minimum		19,000			
		Mean		19,000			
	repomis macroculrus	Maximum		19,000			
		Z		1			
		Minimum		6,250			
	as papawit	Mean		7,401			
		Maximum		10,530			
Magnesium sulfate		Z		4			
(cont.)		Minimum		> 1,000			
	and anima	Mean		> 1,000			
	Oryzius iutipes	Maximum		> 1,000			
		Z		9			
		Minimum	2,076	2,820			2
	or losses of polaries of	Mean	2,076	4,715			
	rilliepitules profiteius	Maximum	2,076	7,900			
		Z	1	4			
		Minimum		880			
	poolidam	Mean		1,313			
	and dist	Maximum		2,380	TO THE PARTY OF TH		
		z		4			
		Minimum			4,800		
	Biomphalaria	Mean			4,800		
	alexandrina	Maximum			4,800		
		z			1		
		Minimum			900		
	Bulinus truncatus	INEGII			006		
		Maximum			900		
		2	CFF	0.14	0000	1 200	640
		Minimum	1,112	914	2,989	1,300	1 486
	Ceriodaphnia dubia	Maximim	2 178	3 716	3,463	3,557	2 264
		Z	6,1,2	69	6	6	11
Sodium sulfate		Minimim		14 134			
		Mean		14 134			
	Chironomus tentans	Maximum		14,134			
		z		1			
		Minimum		11,430			
		Mean		12,390			
	Culex sp.	Maximum		13,350			
		z		2			
		Minimum	927	930	5,492	7,460	3,650
	Danhaia maana	Mean	1,180	3,949	6,574	7,460	3,650
	ממשווים וווחמנים	Maximum	1,434	8,600	6,844	7,460	3,650
		z	2	23	5	H	1
		Minimum		10,000			
	Gambusia affinis	Mean		17,000			
	1	A A		0000			

<u>Ioxicity Endpoint Definitions:</u>
LCOO (NR-ZERO): the concentration at which no lethality was observed (100% survival)
LC50/EC50: the concentration at which 50% lethality was observed
LC100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)
LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically

gnificant effect was observed (compared to the control group)

OEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically gnificant effect was observed (compared to the control group)

T				EP	ENDPOINT (mg/L)	(1)	
Chemical	Species	Statistic	LC00 (NR-ZERO)	LC50/ EC50	LC100 (NR-LETH)	LOEL/LOEC	NOEL/NOEC
		Minimum	138	512	1,460	1,460	1,060
	Il and all a market	Mean	390	2,230	1,460	1,853	1,404
	Hyalella azteca	Maximum	642	5,259	1,460	3,650	1,637
		z	2	48	T	8	4
		Minimum		1,702			
	The second secon	Mean		2,882			
	Lampsilis siliquoided	Maximum		3,729			
		z		10			
		Minimum		3,040			
		Mean		11,271			
	repornis macrocuirus	Maximum		17,500			
		z		80			
		Minimum		4	1,000		
		Mean		2,702	1,000		
	Lynnaeu sp.	Maximum		5,401	1,000		
		Z		4	1		
		Minimum	100	99	1,000		
Sodium sulfate		Mean	1,350	009	3,125		
(cont.)	Mororie saxatilis	Maximum	2,500	1,100	4,500		
		z	80	16	8		
		Minimum	24	1,649		1,250	265
		Mean	300	6,699		3,713	1,721
	Firmephales promeias	Maximum	906	15,200		2,500	2,900
		z	9	8		4	9
		Minimum		15,996			
		Mean		18,018			
	Poecilia latipinna	Maximum		20,040			
		z		2			
		Minimum	1,502	2,078			
	Charles and the contract	Mean	1,502	2,403			
	Spriderium simile	Maximum	1,502	2,926			
		z	1	7			
		Minimum		300			
	Culex sp.	Mean		300			
	(Sodium sulfate 1:1)	Maximum		300			
		z		2			

Toxicity Endpoint Definitions:

LCOO (NR-ZERO): the concentration at which no lethality was observed (100% survival)

LCSO/ECSO: the concentration at which 50% lethality was observed

DELINEE: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically gnificant effect was observed (compared to the control group)

OEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically gnificant effect was observed (compared to the control group) :100 (NR-LETH): the concentration at which 100% lethality was observed (0% survival)

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

		Effect	N.		ENDPOI	ENDPOINT (mg/L)	A
Chemical	Species	Measurement	Statistic	EC50	1C50	LOEL/LOEC	NOEL/NOEC
			Minimum	750		1,106	638
			Mean	750		1,106	638
	Ceriodapunia aubia	Reproduction	Maximum	750		1,106	638
			z	1		1	1
			Minimum			305	< 305
			Mean	0		305	< 305
	Ceriodaphnia sp.	Keproduction	Maximum			305	
Calcium			z			1	1
chloride			Minimum	1,750		1,198	899
			Mean	1,750		1,198	899
	Барћпіа тадпа	Reproduction	Maximum	1,750		1,198	899
			z	1		1	1
			Minimum			> 1,410	1,410
			Mean			> 1,410	1,410
	Pimephales promelas	Growth	Maximum				1.410
			z				1
			Minimim		1 9/5	2 230	1120
			acond acond		1 945	2,230	1 130
	Brachionus calyciflorus	Reproduction	INICALI		C+C,1	2,230	1,120
			Maximum		1,945	2,230	1,120
			2	-	Т	T	1
			Minimum	> 467	161	250	130
	Ceriodaphnia dubia	Reproduction	Mean	1,579	829	1,193	653
	non number of the	in the second se	Maximum	2,750	1,900	2,500	1,500
			Z	4	39	46	59
			Minimum			1,650	1,155
			Mean			1,650	1,155
	cerioaaphnia sp.	Keproduction	Maximum			1,650	1,155
			z			1	1
			Minimum		3,047	3,960	2,133
	China and an annual and an annual and an	4	Mean		3,047	3,960	2,133
	Chironomus anutus	GLOWEI	Maximum		3,047	3,960	2,133
			z		1	1	1
			Minimum	650		850	440
Sodium			Mean	650		850	440
chloride	Daphnia ambigua	Keproduction	Maximum	650		850	440
			z	1		1	1
			Minimum	> 467	1,037	506	467
			Mean	2,359	1,037	1,583	808
	Daphnia magna	Reproduction	Maximum	4,250	1,037	2,160	1,296
			z	2	1	8	6
			Minimum			441	314
			Mean			441	314
		Growth	Maximum			441	314
			z			1	1
	Daphnia pulex		Minimim			441	314
			Mean			441	314
		Reproduction	Maximim			777	214
			MAXIIIIIIII			144	7
			2		2	7	7
			Minimum		2,298	4,237	2,210
	Hvalella azteca	Growth	Mean		2,298	4,237	2,210
		)	Maximum		2,298	4,237	2,210
			z		1	1	1

## Toxicity Endpoint Definitions:

EC50/IC50: the concentration at which a 50% inhibition or reduction of the associated effect measurement (e.g., growth, reproduction) is observed (compared to the control group)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant effect was observed (compared to the control group)

NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant effect was observed (compared to the control group)

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		Sodium sulf		Magnesium sulfate	Calcium sulfate		Sodium chloride (cont.)		Chemical
Pimephales promelas	Hyalella azteca	Ceriodaphnia sp. Sodium sulfate   Corbicula manilensis	Ceriodaphnia dubia	Daphnia magna	Daphnia magna	Tubifex tubifex	Pimephales promelas	Lumbriculus variegatus	Effect ENDPOINT (mg/L)  Species Measurement Statistic EC50 IC50 LOEL/LOEL
Growth	Reproduction	Reproduction Growth	Reproduction	Reproduction	Reproduction	Reproduction	Growth	Reproduction	Effect Measurement
Minimum Mean Maximum	Minimum Mean Maximum	Minimum Mean N Maximum N Minimum Mean Mean Mean N	Minimum Mean Maximum N	Minimum Mean Maximum N	Minimum Mean Maximum N	Minimum Mean Maximum N	Minimum Mean Maximum	Minimum Mean Maximum N	Statistic
	> 2,412 > 2,412 > 2,412 > 2,412 1		465 1,050 1,458 6						EC50
			715 1,199 2,061 4			752 752 752 1	958 958 958 1	1,366 1,366 1,366 1,366	ENDPOII IC50
220 220 220 1	2,412 2,412 2,412 1	2,220 2,220 2,220 1 1,500 1,500 1,500 2	150 1,150 3,650 14			964 964 964 1	1,000 3,041 10,000 28	366 366 366 1	ENDPOINT (mg/L)
< 220 760 1,301	1,637 1,637 1,637 1	< 2,220 < 2,220 < 2,220 < 2,220 1,500 1,500 1,500 2	< 150 710 1,301 10	360 360 360 2	360 980 1,600 4	462 462 462 1	210 1,673 5,000 47	< 366 < 366 < 366 1	NOEL/NOEC

Toxicity Endpoint Definitions:

EC50/IC50: the concentration at which a 50% inhibition or reduction of the associated effect measurement (e.g., growth, reproduction) is observed (compared to the control group)

LOEL/LOEC: the lowest-observed-effect level/concentration; the lowest concentration at which a statistically significant

Page 2 of 2

effect was observed (compared to the control group)

effect was observed (compared to the control group) NOEL/NOEC: the no-observed-effect level/concentration; the highest concentration at which no statistically significant

		Hardness val	ues associated wit	Hardness values associated with lethal toxicity endpoints	idpoints.			
Result				Concentration			Hardness	
Number	Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
160619 Ca	Calcium chloride (CaCl2)	Ceriodaphnia dubia	LC50	2,260	80		100	mg/L CaCO3
160620 Ca	Calcium chloride (CaCl2)	Ceriodaphnia dubia	LC50	1,830	80		100	mg/L CaCO3
768180 Ca	Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-LETH	1,599	80		100	mg/L CaCO3
768284 Ca	768284 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-LETH	1,599	80		100	mg/L CaCO3
768182 Ca	768182 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-LETH	1,599	80		100	mg/L CaCO3
767342 Ca	767342 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-ZERO	800	80		100	mg/L CaCO3
768283 Ca	768283 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-ZERO	800	80		100	mg/L CaCO3
768183 Ca	768183 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-ZERO	800	80		100	mg/L CaCO3
768181 Ca	768181 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-ZERO	800	80		100	mg/L CaCO3
768286 Ca	768286 Calcium chloride (CaCl2)	Ceriodaphnia dubia	NR-ZERO	401	80		100	mg/L CaCO3
160622 Ca	160622 Calcium chloride (CaCl2)	Daphnia magna	LC50	2,770	80		100	mg/L CaCO3
160621 Ca	160621 Calcium chloride (CaCl2)	Daphnia magna	LC50	3,250	80		100	mg/L CaCO3
768399 Ca	768399 Calcium chloride (CaCl2)	Daphnia magna	NR-LETH	3,196	08		100	mg/L CaCO3
768396 Ca	Calcium chloride (CaCl2)	Daphnia magna	NR-LETH	3,196	80		100	mg/L CaCO3
768457 Ca	Calcium chloride (CaCl2)	Daphnia magna	NR-LETH	4,780	08			mg/L CaCO3
768462 Ca	Calcium chloride (CaCl2)	Daphnia magna	NR-ZERO	1,196	08			mg/L CaCO3
768402 Ca	Calcium chloride (CaCl2)	Daphnia magna	NR-ZERO	800	80			mg/L CaCO3
768460 Cal	Calcium chloride (CaCl2)	Daphnia magna	NR-ZERO	1,196	80			mg/L CaCO3
768398 Cal	Calcium chloride (CaCl2)	Daphnia magna	NR-ZERO	800	80			mg/L CaCO3
2153763 Cal	2153763 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
2153676 Cal	2153676 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
740759 Cal	740759 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
740760 Cal	740760 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
740732 Cal	740732 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
2153700 Cal	2153700 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
2153697 Cal	2153697 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
740761 Cal	740761 Calcium chloride (CaCl2)	Dreissena polymorpha	NR-LETH	10,000		180		mg/L CaCO3
740765 Cal	740765 Calcium chloride (CaCl2)	Ictalurus punctatus	NR-LETH	10,000		140		mg/L CaCO3
740777 Cal	Calcium chloride (CaCl2)	Ictalurus punctatus	NR-LETH	10,000		140	1	mg/L CaCO3
196145 Cal	Calcium chloride (CaCl2)	Lepomis macrochirus	LC50	10,650		10	1	mg/L CaCO3
40507 Cal	Calcium chloride (CaCl2)	Lepomis macrochirus	LC50	10,650	6,314		11,900 r	mg/L CaCO3
-	Calcium chloride (CaCl2)	Lepomis macrochirus	NR-ZERO	10,000		140	_	mg/L CaCO3
741717 Cal	Calcium chloride (CaCl2)	Lepomis macrochirus	NR-ZERO	10,000		140	T	mg/L CaCO3

-	Hardness valu	ues associated wi	Hardness values associated with lethal toxicity endpoints.	ndpoints.			
Rossilt						Hardness	
Number Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
2153801 Calcium chloride (CaCl2)	Lepomis macrochirus	NR-ZERO	10.000		140		ma/I Cacoa
2153798 Calcium chloride (CaCl2)	Micropterus dolomieu	NR-ZERO	10,000		140		mg/l CaCO3
740767 Calcium chloride (CaCl2)	Micropterus dolomieu	NR-ZERO	10,000		140		mg/l CaCO3
160623 Calcium chloride (CaCl2)	Pimephales promelas	LC50	> 6,660	80		100	mg/L CaCO3
160624 Calcium chloride (CaCl2)	Pimephales promelas	LC50		80		100	mg/l CaCO3
160625 Calcium chloride (CaCl2)	Pimephales promelas	LC50	4,630	80		100	mg/L CaCO3
768541 Calcium chloride (CaCl2)	Pimephales promelas	NR-LETH	4,780	80		100	mg/l CaCO3
768539 Calcium chloride (CaCl2)	Pimephales promelas	NR-LETH	4,780	80		100	mg/L CaCO3
	Pimephales promelas	NR-LETH	10,000		140		mg/L CaCO3
	Pimephales promelas	NR-LETH	20,000		140		mg/L CaCO3
	Pimephales promelas	NR-LETH	20,000		140		mg/L CaCO3
740738 Calcium chloride (CaCl2)	Pimephales promelas	NR-LETH	10,000		140		mg/L CaCO3
	Sander vitreus	NR-LETH	20,000		140		mg/L CaCO3
	Sander vitreus	NR-ZERO	10,000		140		mg/L CaCO3
2153802   Calcium chloride (CaCl2)	Sander vitreus	NR-ZERO	10,000		140		mg/L CaCO3
	Sander vitreus	NR-ZERO	10,000		140		mg/L CaCO3
	Ceriodaphnia dubia	LC50	1,270	80		100	mg/L CaCO3
160514 Magnesium chloride	Ceriodaphnia dubia	LC50	880	80			mg/L CaCO3
/681/8 Magnesium chloride	Ceriodaphnia dubia	NR-LETH	933	80			mg/L CaCO3
/681/6 Magnesium chloride	Ceriodaphnia dubia	NR-LETH	933	80			mg/L CaCO3
768177 Magnesium chloride	Ceriodaphnia dubia	NR-ZERO	467	80		1	mg/L CaCO3
160516 Magnesium chloride	ibia	NR-ZERO	467	80			mg/L CaCO3
160515 Magnesium chloride		LC50	1,330	80		100 r	mg/L CaCO3
768454 Magnesium chloride		LC50	1,560	80			mg/L CaCO3
768393 Magnesium chloride	Daphnia magna	NR I ETU	1,044	80			mg/L CaCO3
768501 Magnesium chloride		NR-I ETH	1,004	000			mg/L CaCO3
768451 Magnesium chloride		NR-LETH	2 044	80 00			mg/L CaCO3
768390 Magnesium chloride		NR-LETH	1.864	80		100	mg/l Cacos
768392 Magnesium chloride		NR-ZERO	467	80			mg/l CaCO3
768395 Magnesium chloride	Daphnia magna	NR-ZERO	467	80			mg/L CaCO3
/68456 Magnesium chloride	Daphnia magna	NR-ZERO	512	80			mg/L CaCO3

187906	187901	187902	187903	187905	2152847	231564	231565	231570	231566	231563	231572	231573	231567	231571	231568	231569	231581	231579	231576	231575	231577	231574	231578	231580	2455	12210	2055980	160519	160518	160517	768503	Result Number	
187906 Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	231570 Sodium chloride (NaCl)	231566 Sodium chloride (NaCl)	231563 Sodium chloride (NaCl)	231572 Sodium chloride (NaCl)	231573 Sodium chloride (NaCl)	231567 Sodium chloride (NaCl)	231571 Sodium chloride (NaCl)	231568 Sodium chloride (NaCl)	231569 Sodium chloride (NaCl)	231581 Sodium chloride (NaCl)	231579 Sodium chloride (NaCl)	231576 Sodium chloride (NaCl)	231575 Sodium chloride (NaCl)	231577 Sodium chloride (NaCl)	231574 Sodium chloride (NaCl)	231578 Sodium chloride (NaCl)	231580 Sodium chloride (NaCl)	2455 Sodium chloride (NaCl)	12210 Sodium chloride (NaCl)	2055980 Sodium chloride (NaCl)	160519 Magnesium chloride	160518 Magnesium chloride	Magnesium chloride	Magnesium chloride	Chemical Name					
Caenorhabditis elegans	Caenorhabditis elegans	Caenorhabditis elegans	Caenorhabditis elegans	Caenorhabditis elegans	Brachionus calyciflorus	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Asellus communis	Argia sp.	Anguilla rostrata	Anguilla rostrata	Acipenser sinensis	Pimephales promelas	Pimephales promelas	Pimephales promelas	Daphnia magna	Species Scientific Name								
LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	LC50	NR-ZERO	LC50	LC50	LC50	NR-ZERO	Endpoint	
22,457	25,190	25,064	24,829	23,817	1,645	> 10,000	8,250	> 5,600	8,250	> 10,000	5,100	5,100	7,200	> 5,600	6,800	6,150	23,000	32,000	24,000	26,000	24,000	> 32,000	> 32,000	26,000	17,880	21,450	616	2,120	2,840	3,520	747	Concentration (mg/L)	
80	80	80	80	80	80																				40	40		80	80	80	80	Minimum	
						100	100	20	100	100	20	20	100	20	100	100	100	20	100	100	100	100	20	100			22					Mean	
90	90	90	90	90	100																				48	48		100	100	100	100	Maximum	Hardness
mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	Units	

						Hardness	
Result Chemical Name	Species Scientific Name	Endpoint	Concentration	Minimum	Mean	Maximum	Units
187904 Sodium chloride (NaCl)	Caenorhabditis elegans	LC50	25,786	80		_	mg/L CaCO3
742006 Sodium chloride (NaCl)	Caenorhabditis elegans	LOEL	20,550	80			mg/L CaCO3
742038 Sodium chloride (NaCl)	Caenorhabditis elegans	LOEL	21,000	80			mg/L CaCO3
742918 Sodium chloride (NaCl)	Caenorhabditis elegans	NOEL	20,500	80			mg/L CaCO3
742919 Sodium chloride (NaCl)	Caenorhabditis elegans	NOEL	20,950	80		ij,	mg/L CaCO3
167864 Sodium chloride (NaCl)	Carassius auratus	LC50	7,341		210		mg/L CaCO3
75451 Sodium chloride (NaCl)	Carassius auratus	LC50	4,324		149		mg/L CaCO3
167863 Sodium chloride (NaCl)	Carassius auratus	LETC	7,322		210		mg/L CaCO3
160319 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	3,380	80		100	mg/L CaCO3
160320 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,960	80			mg/L CaCO3
207716 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	2,250	94	102		mg/L CaCO3
207717 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	~ 2,000	94	102	E	mg/L CaCO3
741686 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,590	54		72	mg/L CaCO3
741547 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	> 467	100	101		mg/L CaCO3
2152724 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	2,750		98		mg/L CaCO3
2152723 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	2,750		98		mg/L CaCO3
2152825 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,068	80		100 1	mg/L CaCO3
2153271 Sodium chloride (NaCl)		LC50	540		40		mg/L CaCO3
2153268 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC20	316		20		mg/L CaCO3
2153274 Sodium chloride (NaCl)		LC50	1,134		80		mg/L CaCO3
2152874 Sodium chloride (NaCl)		LC50	132		10		mg/L CaCO3
2153277 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,240		160		mg/L CaCO3
2153280 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,303		320	7	mg/L CaCO3
2075467 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,489	276	278	280 r	mg/L CaCO3
2075442 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,836	792	796		mg/L CaCO3
2075434 Sodium chloride (NaCl)		LC50	1,779	560	565		mg/L CaCO3
2075468 Sodium chloride (NaCl)		LC50	1,317	280	282		mg/L CaCO3
2075410 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,249	95	96	Ļ	mg/L CaCO3
2075471 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,192	278	279		mg/L CaCO3
2075466 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,356	279	280		mg/L CaCO3
2075469 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,357	280	281	281 n	mg/L CaCO3
2075470 Sodium chloride (NaCI)	Ceriodaphnia dubia	LC50	1,154	280	285	290 n	mg/L CaCO3

	Hardness valu	ies associated wi	Hardness values associated with lethal toxicity endpoints	dpoints.			
Result			Concentration			Hardness	
Number Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
2074934 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	977	25	28	30	mg/L CaCO3
2075402 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	861	44	47	49	mg/L CaCO3
2075426 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,589	375	388	400	mg/L CaCO3
2075418 Sodium chloride (NaCl)	Ceriodaphnia dubia	LC50	1,402	180	187	194	mg/L CaCO3
742567 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,200	54		72	mg/L CaCO3
767903 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
766864 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	5,000	80		100	mg/L CaCO3
767897 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767961 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767879 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767895 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767872 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	08		100	mg/L CaCO3
767959 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	08		100	mg/L CaCO3
767870 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80			mg/L CaCO3
767967 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	08		100	mg/L CaCO3
766872 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80			mg/L CaCO3
766866 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80			mg/L CaCO3
767889 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	08			mg/L CaCO3
767856 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	08	Ì	100	mg/L CaCO3
767969 Sodium chloride (NaCl)		LOEC	2,500	80			mg/L CaCO3
767846 Sodium chloride (NaCl)	Ceriodaphnia dubia	DEC	2,500	80		100	mg/L CaCO3
767953 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767887 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767881 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767975 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80		100	mg/L CaCO3
767864 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	2,500	80			mg/L CaCO3
767862 Sodium chloride (NaCl)		LOEC	2,500	80		100	mg/L CaCO3
767848 Sodium chloride (NaCl)		LOEC	2,500	80			mg/L CaCO3
767854 Sodium chloride (NaCl)		LOEC	2,500	80		100	mg/L CaCO3
741687 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,700	54		72	mg/L CaCO3
767855 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767904 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3

						Hardness	
Result Chemical Name	Species Scientific Name	Endpoint	Concentration	Minimum	Mean	Maximum	Units
5 Sodium cl	Ceriodaphnia dubia	NOE	2,500	80		100	mg/L CaCO3
Sodium chloride	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767962 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767880 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767896 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767873 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767960 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767871 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767968 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
766873 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
766867 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767890 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767857 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	08		100	mg/L CaCO3
767970 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	08		100	mg/L CaCO3
767847 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	620	08		100	mg/L CaCO3
767954 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	08		100	mg/L CaCO3
767888 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767882 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767976 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767865 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767863 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
767849 Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	1,250	80		100	mg/L CaCO3
768272 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-LETH	1,701	80		100	mg/L CaCO3
768340 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-LETH	3,035	80		100	mg/L CaCO3
768268 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-LETH	1,701	80		100	mg/L CaCO3
768173 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-LETH	1,701	80		100	mg/L CaCO3
741695 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-LETH	2,260	54		72	mg/L CaCO3
741699 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-LETH	2,200	54		72	mg/L CaCO3
768266 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	760	80		100	mg/L CaCO3
768264 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	760	80			mg/L CaCO3
768175 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	872	80		100	mg/L CaCO3
207718 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	1,250	94	102	104	mg/L CaCO3

						Hardness	
Result Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
207719 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	1,250	94	102	104	mg/L CaCO3
207721 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	1,250	94	102		mg/L CaCO3
741660 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	850	54			mg/L CaCO3
742530 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	467	100	101		mg/L CaCO3
2152722 Sodium chloride (NaCl)	Ceriodaphnia dubia	NR-ZERO	467		98		mg/L CaCO3
741435 Sodium chloride (NaCl)	Ceriodaphnia reticulata	LC50	1,250		84		ppm
2060209 Sodium chloride (NaCl)	Chironomus dilutus	EC01	4,163		296		mg/L CaCO3
2060210 Sodium chloride (NaCl)	Chironomus dilutus	EC05	4,640		296		mg/L CaCO3
2060211 Sodium chloride (NaCl)	Chironomus dilutus	EC10	4,917		296		mg/L CaCO3
2060212 Sodium chloride (NaCl)	Chironomus dilutus	EC15	5,113		296		mg/L CaCO3
2060213 Sodium chloride (NaCl)	Chironomus dilutus	EC20	5,274		296		mg/L CaCO3
2060214 Sodium chloride (NaCl)	Chironomus dilutus	EC25	5,417		296		mg/L CaCO3
2060216 Sodium chloride (NaCl)	Chironomus dilutus	EC40	5,793		296		mg/L CaCO3
2060217 Sodium chloride (NaCl)	Chironomus dilutus	EC60	6,280		296		mg/L CaCO3
2060218 Sodium chloride (NaCl)	Chironomus dilutus	EC75	6,717		296		mg/L CaCO3
2060220 Sodium chloride (NaCl)	Chironomus dilutus	EC80	6,898		296		mg/L CaCO3
2060221 Sodium chloride (NaCl)	Chironomus dilutus	EC85	7,116		296		mg/L CaCO3
2060222 Sodium chloride (NaCl)	Chironomus dilutus	EC90	7,399		296		mg/L CaCO3
2060223 Sodium chloride (NaCl)	Chironomus dilutus	EC95	7,840		296		mg/L CaCO3
2060225 Sodium chloride (NaCl)	Chironomus dilutus	EC99	8,740		296		mg/L CaCO3
2060201 Sodium chloride (NaCl)	Chironomus dilutus	LC50	6,032		296		mg/L CaCO3
2152838 Sodium chloride (NaCl)	Chironomus dilutus	LC50	5,867	80		100	mg/L CaCO3
2060236 Sodium chloride (NaCl)	Chironomus dilutus	LOEC	7,810		296		mg/L CaCO3
2060237 Sodium chloride (NaCl)	Chironomus dilutus	MATC	6,211		296		mg/L CaCO3
2060228 Sodium chloride (NaCl)	Chironomus dilutus	NOEC	4,940		296		mg/L CaCO3
2060247 Sodium chloride (NaCl)	Chironomus dilutus	NR-LETH	11,600		296		mg/L CaCO3
2060275 Sodium chloride (NaCl)	Chironomus dilutus	NR-LETH	7,810		296		mg/L CaCO3
2060254 Sodium chloride (NaCl)		NR-LETH	7,810		296		mg/L CaCO3
2060259 Sodium chloride (NaCl)	Chironomus dilutus	NR-ZERO	7,810		296	-	mg/L CaCO3
2060270 Sodium chloride (NaCl)	Chironomus dilutus	NR-ZERO	4,940		296	T	mg/L CaCO3
2211037 Sodium chloride (NaCl)	Chironomus tentans	NR-ZERO	10,000	140		160 p	ppm
2211025 Sodium chloride (NaCl)	Corbicula manilensis	NR-LETH	10,800	140		160 p	mdd

						Hardness	
Result Number Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
2211024 Sodium chloride (NaCl)	Corbicula manilensis	NR-ZERO	4,850	140		160	mdd
chloride	Cricotopus trifasciatus	LC50	6,221		124		mg/L CaCO3
2060148 Sodium chloride (NaCl)	Cyprinella leedsi	LC50	6,070		296		mg/L CaCO3
2060157 Sodium chloride (NaCl)	Cyprinella leedsi	LC50	6,137		296		mg/L CaCO3
2060168 Sodium chloride (NaCl)	Cyprinella leedsi	LC50	6,137		296		mg/L CaCO3
2060161 Sodium chloride (NaCl)	Cyprinella leedsi	LC50	6,137		296		mg/L CaCO3
2060198 Sodium chloride (NaCl)	Cyprinella leedsi	NR-LETH	7,810		296		mg/L CaCO3
2060191 Sodium chloride (NaCl)	Cyprinella leedsi	NR-LETH	7,810		296		mg/L CaCO3
2060190 Sodium chloride (NaCl)	Cyprinella leedsi	NR-ZERO	3,100		296		mg/L CaCO3
2060189 Sodium chloride (NaCl)	Cyprinella leedsi	NR-ZERO	4,940		296		mg/L CaCO3
742566 Sodium chloride (NaCl)	Daphnia ambigua	LC50	2,000	54		72	mg/L CaCO3
741697 Sodium chloride (NaCl)	Daphnia ambigua	LOEC	850	54		72	mg/L CaCO3
741698 Sodium chloride (NaCl)	Daphnia ambigua	NOEC	440	54		72	mg/L CaCO3
2164727 Sodium chloride (NaCl)	Daphnia magna	EC50	4,040		150		ppm
144221 Sodium chloride (NaCl)	Daphnia magna	LC50	6,027		240		mg/L CaCO3
144220 Sodium chloride (NaCl)	Daphnia magna	LC50	6,027		240		mg/L CaCO3
144218 Sodium chloride (NaCl)	Daphnia magna	LC50	5,600		240		mg/L CaCO3
144219 Sodium chloride (NaCl)	Daphnia magna	LC50	5,020		240		mg/L CaCO3
144222 Sodium chloride (NaCl)	Daphnia magna	LC50	5,600		240		mg/L CaCO3
160322 Sodium chloride (NaCl)	Daphnia magna	LC50	4,770	80		100	mg/L CaCO3
160321 Sodium chloride (NaCl)	Daphnia magna	LC50	6,380	08		100	mg/L CaCO3
767302 Sodium chloride (NaCl)	Daphnia magna	LC50	3,222		106		mg/L CaCO3
772103 Sodium chloride (NaCl)	Daphnia magna	LC50	3,136		106		mg/L CaCO3
767308 Sodium chloride (NaCl)	Daphnia magna	LC50	3,137		106		mg/L CaCO3
742529 Sodium chloride (NaCl)	Daphnia magna	LC50	> 467	166	169	172	mg/L CaCO3
742019 Sodium chloride (NaCl)	Daphnia magna	LC50	5,480	160		180	mg/L CaCO3
759179 Sodium chloride (NaCl)	Daphnia magna	LC50	6,500	404	563	694	mg/L CaCO3
2152709 Sodium chloride (NaCl)	Daphnia magna	LC50	~ 6,000		170		mg/L CaCO3
2152708 Sodium chloride (NaCl)	Daphnia magna	LC50	7,500		170		mg/L CaCO3
2152826 Sodium chloride (NaCl)	Daphnia magna	LC50	3,630	08		100	mg/L CaCO3
768426 Sodium chloride (NaCl)	Daphnia magna	NR-LETH	6,069	80		100	mg/L CaCO3
768527 Sodium chloride (NaCl)	Daphnia magna	NR-LETH	4,321	80		100	mg/L CaCO3

mg/L CaCO3	192		170	2,670	EC50	Elliptio complanata	Sodium chloride (NaCl)	803856
mg/L CaCO3	192		170	2,230	EC50	Elliptio complanata	Sodium chloride (NaCl)	741563 9
mg/L CaCO3		180		20,000	NR-LETH	Dreissena polymorpha	Sodium chloride (NaCl)	740662 \$
mg/L CaCO3		180		10,000	NR-LETH	Dreissena polymorpha	Sodium chloride (NaCl)	740690 9
mg/L CaCO3		96		441	NR-ZERO	Daphnia pulex	Sodium chloride (NaCl)	201558 9
mg/L CaCO3	82		28	1,760	LC50	Daphnia pulex	Sodium chloride (NaCl)	2055812
ppm	100	84	08	1,250	LC50	Daphnia pulex	Sodium chloride (NaCl)	741434 9
mg/L CaCO3		93		1,470	LC50	Daphnia pulex	201545 Sodium chloride (NaCl)	201545
mdd		150		500	NR-ZERO	Daphnia magna	2164732 Sodium chloride (NaCl)	2164732
mg/L CaCO3		170		280	NR-ZERO	Daphnia magna	152707 Sodium chloride (NaCl)	2152707
mg/L CaCO3	172	169	166	467	NR-ZERO	Daphnia magna	741531 Sodium chloride (NaCl)	741531
mg/L CaCO3	100		80	2,213	NR-ZERO	Daphnia magna	768526 Sodium chloride (NaCl)	768526
mg/L CaCO3	100		80	1,519	NR-ZERO	Daphnia magna	768488 Sodium chloride (NaCl)	768488
mg/L CaCO3	100		80	2,766	NR-ZERO	Daphnia magna	768520 Sodium chloride (NaCl)	768520
mg/L CaCO3	100		80	2,766	NR-ZERO	Daphnia magna	768530 Sodium chloride (NaCl)	768530
mg/L CaCO3	100		80	3,466	NR-ZERO	Daphnia magna	768523 Sodium chloride (NaCl)	768523
mg/L CaCO3	100		80	2,766	NR-ZERO	Daphnia magna	768450 Sodium chloride (NaCl)	768450
mg/L CaCO3	100		80	2,766	NR-ZERO	Daphnia magna	768498 Sodium chloride (NaCl)	768498
mg/L CaCO3	100		80	1,519	NR-ZERO	Daphnia magna	768431 Sodium chloride (NaCl)	768431
mg/L CaCO3	100		80	3,457	NR-ZERO	Daphnia magna	768500 Sodium chloride (NaCl)	768500
mg/L CaCO3	100		80	2,766	NR-ZERO	Daphnia magna	768533 Sodium chloride (NaCl)	768533
mg/L CaCO3	100		80	1,519	NR-ZERO	Daphnia magna	768428 Sodium chloride (NaCl)	768428
ppm		150		8,000	NR-LETH	Daphnia magna	2164731 Sodium chloride (NaCl)	2164731
mg/L CaCO3	100		80	6,069	NR-LETH	Daphnia magna	768486 Sodium chloride (NaCl)	768486
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768521 Sodium chloride (NaCl)	768521
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768467 Sodium chloride (NaCl)	768467
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768499 Sodium chloride (NaCl)	768499
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768445 Sodium chloride (NaCl)	768445
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768496 Sodium chloride (NaCl)	768496
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768531 Sodium chloride (NaCl)	768531
mg/L CaCO3	100		80	4,321	NR-LETH	Daphnia magna	768475 Sodium chloride (NaCl)	768475
mg/L CaCO3	100		80	6,069	NR-LETH	Daphnia magna	768489 Sodium chloride (NaCl)	768489
Units	Maximum	Mean	Minimum	Concentration (mg/L)	Endpoint	Species Scientific Name	Chemical Name	Number
	Hardness							
			-					

Erpobdella punctata         LC50         7,500         100           Fundulus seminolis         NR-LETH         2,000         171         194         257           Fundulus circumstriatus         LC50         3,200         100         257           Gyraulus circumstriatus         LC50         3,700         100         100           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus circumstriatus         LC50         3,200         100         100         110         100         100         110 <th>mg/L CaCO3</th> <th></th> <th>700</th> <th></th> <th></th> <th></th> <th></th> <th></th> <th></th>	mg/L CaCO3		700						
Erpobdella punctata   LC50			100		113	EC50		2052773 Sodium chloride (NaCl)	2052773
Erpobdella punctata   LC50   7,500   100	mg/L CaCO3		322		1,391	EC50	A.	2052783 Sodium chloride (NaCl)	2052783
Probabella punctata   CCSO   7,500   100	mg/L CaCO3		292		1,559	EC50		2052781 Sodium chloride (NaCl)	2052781
Probabella punctata   CCSO   7,500   100	mg/L CaCO3	192		170	1,840	EC50		803857 Sodium chloride (NaCl)	803857
Erpobdella punctata         LC50         7,500         100         4           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Fundulus circumstriatus         LC50         3,200         171         194         257           Gyraulus circumstriatus         LC50         3,700         100         100         100           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus parvus         LC50         3,078         56         100         100         100           Gyraulus parvus         LC50         3,078         80         212         100         100         100         100         100         100         100         100         100         100         100         100         100         100         <	mg/L CaCO3	192		170	3,980	EC50		741584 Sodium chloride (NaCl)	741584
Erpobdella punctata         LC50         7,500         100         400           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus parvus         LC50         3,078         56         100         100         100           Gyraulus parvus         LC50         3,078         80         212         100         100         100         100         100         100         100         100         100         100         100         100         100         100         100	mg/L CaCO3	192		170	1,740	EC50		741564 Sodium chloride (NaCl)	741564
Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         110         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus circumstriatus         LC50         3,000         100         100         100           Gyraulus parvus         LC50         3,000         100         100         100           Gyraulus parvus         LC50         3,078         56         10         100         100           Hyalella azteca         LC50         3,009         212         10         10         10         10           Ictalurus punctatus         NR-ZERO         10,000	mg/L CaCO3		322		261	EC20		2052784 Sodium chloride (NaCl)	2052784
Erpobdella punctata         ICSO         7,500         100           Erpobdella punctata         ICSO         7,500         100         100           Erpobdella punctata         ICSO         7,500         100         100           Erpobdella punctata         ICSO         7,500         100         100           Erpobdella punctata         ICSO         7,500         171         194         257           Fundulus seminolis         NR-IETH         24,000         171         194         257           Fundulus circumstriatus         ICSO         3,200         171         194         257           Gyraulus circumstriatus         ICSO         3,700         100         100           Gyraulus circumstriatus         ICSO         3,200         100         100           Gyraulus circumstriatus         ICSO         3,000         100         100           Gyraulus circumstriatus         ICSO         3,078         56         100           Gyraulus circumstriatus         ICSO         3,078         56         212           Gyraulus parvus         ICSO         3,078         56         212           Justia         100         212         212         212 </td <td>mg/L CaCO3</td> <td></td> <td>306</td> <td></td> <td>153</td> <td>EC20</td> <td></td> <td>2052786 Sodium chloride (NaCl)</td> <td>2052786</td>	mg/L CaCO3		306		153	EC20		2052786 Sodium chloride (NaCl)	2052786
Erpobdella punctata         LC50         7,500         100         4           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus circumstriatus         LC50         3,200         100         100         100           Gyraulus circumstriatus         LC50         3,078         56         5         100         100         100         100         100         100         100         100         100         100         100         100         100         100         100         100	mg/L CaCO3		292		403	EC20		2052782 Sodium chloride (NaCl)	2052782
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100         100           Erpobdella punctata         LC50         7,500         100	mg/L CaCO3		278		432	EC20		2052780 Sodium chloride (NaCl)	2052780
Erpobdella punctata         IC50         7,500         100         100           Fundulus seminolis         NR-IETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         IC50         3,200         171         194         257           Gyraulus circumstriatus         IC50         3,700         100         100         100           Gyraulus circumstriatus         IC50         3,200         10	mg/L CaCO3		100		817	EC50		2052772 Sodium chloride (NaCl)	2052772
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus circumstriatus         LC50         3,200         171         194         257           Gyraulus circumstriatus         LC50         3,700         171         194         257           Gyraulus circumstriatus         LC50         3,700         100         100           Gyraulus circumstriatus         LC50         3,200         100         100           Gyraulus circumstriatus         LC50         3,200         100         100           Gyraulus parvus         LC50         3,000         100         100           Gyraulus parvus         LC50         3,078         56         100           Hydroptila angusta         LC50         1,382         80         124         100           Ictalurus punctatus         NR-ZERO         10,000         140	mg/L CaCO3		140		10,000	NR-ZERO	lctalurus punctatus	740689 Sodium chloride (NaCl)	740689
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         110         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus circumstriatus         LC50         3,200         171         194         257           Gyraulus circumstriatus         LC50         3,700         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100           Gyraulus parvus         LC50         > 10,000         100         100           Gyraulus parvus         LC50         3,078         56         100           Hydroptila angusta         LC50         1,382         80         100	mg/L CaCO3		140		10,000	NR-ZERO	Ictalurus punctatus	741710 Sodium chloride (NaCl)	741710
Erpobdella punctata         LC50         7,500         100         400           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         11         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus circumstriatus         LC50         3,200         171         194         257           Gyraulus circumstriatus         LC50         3,700         100         100         100           Gyraulus circumstriatus         LC50         3,200         100         100         100           Gyraulus parvus         LC50         3,000         100         100         100           Gyraulus parvus         LC50         3,078         56         100           Hyalella azteca         LC50         1,382         80         212         100           Hydroptila angusta         LC50         6,621         124         100	mg/L CaCO3		140		20,000	NR-LETH	lctalurus punctatus	740725 Sodium chloride (NaCl)	740725
Erpobdella punctata         LC50         7,500         100         4           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus circumstriatus         LC50         3,200         171         194         257           Gyraulus circumstriatus         LC50         3,700         100         100         257           Gyraulus circumstriatus         LC50         > 10,000         100         100         30           Gyraulus circumstriatus         LC50         > 10,000         100         30         100         30           Gyraulus parvus         LC50         > 10,000         56         30	mg/L CaCO3		124		6,621	LC50	Hydroptila angusta	16496 Sodium chloride (NaCl)	16496
Erpobdella punctata         LC50         7,500         100         400           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus circumstriatus         LC50         3,200         171         194         257           Gyraulus circumstriatus         LC50         3,700         100         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100         100           Gyraulus parvus         LC50         3,078         56         100         100         100           Gyraulus parvus         LC50         3,009         212         212         100	mg/L CaCO3	100		80	1,382	LC50	Hyalella azteca	2152842 Sodium chloride (NaCl)	2152842
Erpobdella punctata         LC50         7,500         100         400           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Fundulus circumstriatus         LC50         3,200         100         100         257           Gyraulus circumstriatus         LC50         3,700         100         100         30         100         100         30         100         100         30         100         100         30         30         100         30         30         100         30	mg/L CaCO3		212		3,009	LC50	Gyraulus parvus	2075475 Sodium chloride (NaCl)	2075475
Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100	mg/L CaCO3		56		3,078	LC50	Gyraulus parvus	2075474 Sodium chloride (NaCl)	2075474
Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100           Gyraulus circumstriatus         LC50         3,700         100         100           Gyraulus circumstriatus         LC50         > 10,000         100         100	ppm		100			LC50		231534 Sodium chloride (NaCl)	231534
Erpobdella punctata         LC50         7,500         100         4           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100           Gyraulus circumstriatus         LC50         3,700         100         100	ppm		100		3,200	LC50		231536 Sodium chloride (NaCl)	231536
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100	ppm		100			LC50		231533 Sodium chloride (NaCl)	231533
Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257           Gyraulus circumstriatus         LC50         3,200         100         100	ppm		100		3,700	LC50		231535 Sodium chloride (NaCl)	231535
Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Erpobdella punctata         LC50         7,500         100         100           Fundulus seminolis         NR-LETH         24,000         171         194         257           Fundulus seminolis         NR-ZERO         16,000         171         194         257	ppm		100		3,200	LC50	Gyraulus circumstriatus	231537 Sodium chloride (NaCl)	231537
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100           Fundulus seminolis         NR-LETH         24,000         171         194         257	mg/L CaCO3	257	194	171	16,000	NR-ZERO	Fundulus seminolis	2062416 Sodium chloride (NaCl)	2062416
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100	mg/L CaCO3	257	194	171	24,000	NR-LETH	Fundulus seminolis	2062417 Sodium chloride (NaCl)	2062417
Erpobdella punctata         LC50         7,500         100           Erpobdella punctata         LC50         7,500         100	ppm		100		7,500	LC50	Erpobdella punctata	231526 Sodium chloride (NaCl)	231526
Erpobdella punctata LC50 7,500 100	ppm		100		7,500	LC50	Erpobdella punctata	231525 Sodium chloride (NaCl)	231525
	ppm		100		7,500	LC50	Erpobdella punctata	231527 Sodium chloride (NaCl)	231527
	ppm		100			LC50	Erpobdella punctata	231523 Sodium chloride (NaCl)	231523
oride (NaCl)   Erpobdella punctata   LC50   7,500   100   ppm	ppm		100		7,500	LC50	Erpobdella punctata	231524 Sodium chloride (NaCl)	231524
oride (NaCl)   Epioblasma torulosa ssp.   EC50   244   100   mg/L CaCO3	mg/L CaCO3		100		244	EC50		2052775 Sodium chloride (NaCl)	2052775
Species Scientific Name Endpoint		Maximum	Mean	Minimum	(mg/L)	Endpoint	Species Scientific Name	Chemical Name	Number
Concentration Hardness		Hardness			Concentration			-	Recuil+

						Hardness	
Result  Number Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
5 Sodium c	Lampsilis fasciola	EC50	1,265		306		mg/L CaCO3
2052779 Sodium chloride (NaCl)	Lampsilis fasciola	EC50	1,313		278		mg/L CaCO3
741568 Sodium chloride (NaCl)	Lampsilis siliquoidea	EC50	550	170		192	mg/L CaCO3
803860 Sodium chloride (NaCl)	Lampsilis siliquoidea	EC50	560	170		192	mg/L CaCO3
741594 Sodium chloride (NaCl)	Lampsilis siliquoidea	EC50	4,560	170		192	mg/L CaCO3
2052777 Sodium chloride (NaCl)	Lampsilis siliquoidea	EC50	1,962		172		mg/L CaCO3
2052778 Sodium chloride (NaCl)	Lampsilis siliquoidea	EC50	1,870		322		mg/L CaCO3
2052770 Sodium chloride (NaCl)		EC50	168		100		mg/L CaCO3
2052776 Sodium chloride (NaCl)		EC50	763		47		mg/L CaCO3
2052771 Sodium chloride (NaCl)		EC50	1,430		99		mg/L CaCO3
2224815 Sodium chloride (NaCl)		EC50	2,630	160		180	mg/L CaCO3
2224806 Sodium chloride (NaCl)	Lampsilis siliquoidea	EC50	3,570	160		180	mg/L CaCO3
196203 Sodium chloride (NaCl)	Lepomis macrochirus	LC50	12,946		10		mg/L CaCO3
40512 Sodium chloride (NaCl)	0.0	LC50	1,295	39		52	mg/L CaCO3
201540 Sodium chloride (NaCl)	Lepomis macrochirus	LC50	5,840		102		mg/L CaCO3
220765 Sodium chloride (NaCl)	Lepomis macrochirus	NR-LETH	9,103		429		mg/L CaCO3
740684 Sodium chloride (NaCl)	Lepomis macrochirus	NR-ZERO	10,000		140		mg/L CaCO3
740724 Sodium chloride (NaCl)	Lepomis macrochirus	NR-ZERO	10,000		140		mg/L CaCO3
2210845 Sodium chloride (NaCl)	Lepomis macrochirus	NR-ZERO	5,220	140		160	ppm
2224820 Sodium chloride (NaCl)	Ligumia recta	EC50	3,750	160		180	mg/L CaCO3
2224821 Sodium chloride (NaCl)	Ligumia recta	EC50	2,510	160		180	mg/L CaCO3
231519 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	6,950		100		ppm
231583 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	5,800		100		ppm
231522 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	6,200		100		ppm
231584 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	5,800		100		ppm
231582 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	5,800		100		ppm
231521 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	6,200		100		ppm
231518 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	> 7,500		100		ppm
231520 Sodium chloride (NaCl)	Limnodrilus hoffmeisteri	LC50	6,800		100		mdd
203677 Sodium chloride (NaCl)	Lirceus fontinalis	LC50	2,970		101		mg/L CaCO3
203678 Sodium chloride (NaCl)	Lirceus fontinalis	NR-LETH	4,580		101		mg/L CaCO3
2060288 Sodium chloride (NaCl)	Lumbriculus variegatus	LC50	5,458		296		mg/L CaCO3

						Hardness	
			Concentration				
Number   Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Iviean	Maximum	Onits
2060289 Sodium chloride (NaCl)	Lumbriculus variegatus	LC50	5,506		296		mg/L CaCO3
2060290 Sodium chloride (NaCl)	Lumbriculus variegatus	LC50	5,589		296		mg/L CaCO3
2060276 Sodium chloride (NaCl)	Lumbriculus variegatus	LC50	5,407		296		mg/L CaCO3
2152829 Sodium chloride (NaCl)	Lumbriculus variegatus	LC50	3,100	80		100	mg/L CaCO3
2060292 Sodium chloride (NaCl)	Lumbriculus variegatus	LOEC	4,940		296		mg/L CaCO3
2060293 Sodium chloride (NaCl)	Lumbriculus variegatus	MATC	3,913		296		mg/L CaCO3
2060291 Sodium chloride (NaCl)	Lumbriculus variegatus	NOEC	3,100		296		mg/L CaCO3
2060317 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-LETH	7,810		296		mg/L CaCO3
2060318 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-LETH	7,810		296		mg/L CaCO3
2060307 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-LETH	7,810		296		mg/L CaCO3
2060306 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-LETH	7,810		296		mg/L CaCO3
2060315 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-ZERO	3,100		296		mg/L CaCO3
2060319 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-ZERO	4,940		296		mg/L CaCO3
2060302 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-ZERO	4,940		296		mg/L CaCO3
2060309 Sodium chloride (NaCl)	Lumbriculus variegatus	NR-ZERO	4,940		296		mg/L CaCO3
2075478 Sodium chloride (NaCl)	Musculium transversum	LC50	1,930		48		mg/L CaCO3
16494 Sodium chloride (NaCl)	Nais variabilis	LC50	2,569		124		mg/L CaCO3
2060327 Sodium chloride (NaCl)	Nephelopsis obscura	LC50	4,270	288	290	292	mg/L CaCO3
2060325 Sodium chloride (NaCl)	Nephelopsis obscura	LC50	4,310	288	290	292	mg/L CaCO3
2060326 Sodium chloride (NaCl)	Nephelopsis obscura	LC20	4,270	288	290	292	mg/L CaCO3
2060329 Sodium chloride (NaCl)	Nephelopsis obscura	LC50	4,270	288	290	292	mg/L CaCO3
2060335 Sodium chloride (NaCl)	Nephelopsis obscura	LOEC	5,340	288	290	292	mg/L CaCO3
2060336 Sodium chloride (NaCl)	Nephelopsis obscura	MATC	4,211	288	290	292	mg/L CaCO3
2060333 Sodium chloride (NaCl)	Nephelopsis obscura	NOEC	3,320	288	290	292	mg/L CaCO3
2060352 Sodium chloride (NaCl)	Nephelopsis obscura	NR-LETH	5,340	288	290	292	mg/L CaCO3
2060350 Sodium chloride (NaCl)	Nephelopsis obscura	NR-LETH	5,340	288	290	292	mg/L CaCO3
2060361 Sodium chloride (NaCl)		NR-LETH	7,720	288	290	292 r	mg/L CaCO3
2060360 Sodium chloride (NaCl)		NR-ZERO	5,340	288	290	292 r	mg/L CaCO3
2060342 Sodium chloride (NaCl)		NR-ZERO	5,340	288	290	292 r	mg/L CaCO3
2060354 Sodium chloride (NaCl)	Nephelopsis obscura	NR-ZERO	5,340	288	290	292 r	mg/L CaCO3
740693 Sodium chloride (NaCl)	Perca flavescens	NR-ZERO	10,000		140		mg/L CaCO3
741709 Sodium chloride (NaCl)	Perca flavescens	NR-ZERO	10,000		140	Ţ	mg/L CaCO3

						Hardness	
Result Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
201542 Sodium chloride (NaCl)	Physa gyrina	LC50	2,540		100		mg/L CaCO3
201543 Sodium chloride (NaCl)	Physa gyrina	NR-LETH	5,060		100		mg/L CaCO3
231546 Sodium chloride (NaCl)	Physa heterostropha	LC50	5,100		100		ppm
231545 Sodium chloride (NaCl)	Physa heterostropha	LC50	5,100		100		ppm
231550 Sodium chloride (NaCl)	Physa heterostropha	LC50	6,200		100		ppm
231549 Sodium chloride (NaCl)	Physa heterostropha	LC50	6,200		100		ppm
231554 Sodium chloride (NaCl)	Physa heterostropha	LC50	4,100		20		ppm
231544 Sodium chloride (NaCl)	Physa heterostropha	LC50	> 5,600		100		ppm
231553 Sodium chloride (NaCl)	Physa heterostropha	LC50	4,250		20		ppm
231547 Sodium chloride (NaCl)	Physa heterostropha	LC50	7,500		100		ppm
231542 Sodium chloride (NaCl)	Physa heterostropha	LC50	> 5,600		100		ppm
231551 Sodium chloride (NaCl)	Physa heterostropha	LC50	4,800		20		ppm
231548 Sodium chloride (NaCl)	Physa heterostropha	TC20	6,950		100		ppm
231540 Sodium chloride (NaCl)	Physa heterostropha	LC20	3,500		100		ppm
231543 Sodium chloride (NaCl)	Physa heterostropha	LC50	> 5,600		100		ppm
231539 Sodium chloride (NaCl)	Physa heterostropha	LC50	3,700		100		ppm
231538 Sodium chloride (NaCl)	Physa heterostropha	LC50	4,200		100		ppm
231552 Sodium chloride (NaCl)	Physa heterostropha	LC50	4,250		20		ppm
231541 Sodium chloride (NaCl)	Physa heterostropha	LC50	3,500		100		mdd
167861 Sodium chloride (NaCl)	Pimephales promelas	LC50	7,650		210		mg/L CaCO3
160325 Sodium chloride (NaCl)	Pimephales promelas	LC50	6,390	80		100 г	mg/L CaCO3
160324 Sodium chloride (NaCl)	Pimephales promelas	LC50	6,510	80		100 r	mg/L CaCO3
160323 Sodium chloride (NaCl)	Pimephales promelas	LC50	8,280	80			mg/L CaCO3
201537 Sodium chloride (NaCl)	Pimephales promelas	LC50	6,570		96	1	mg/L CaCO3
759180 Sodium chloride (NaCl)	Pimephales promelas	LC50	11,400	404	563	694 r	mg/L CaCO3
2055542 Sodium chloride (NaCl)	Pimephales promelas	LC50	8,360	52		96 r	mg/L CaCO3
2152828 Sodium chloride (NaCl)		LC50	4,079	80			mg/L CaCO3
167862 Sodium chloride (NaCl)		LETC	7,650		210	_	mg/L CaCO3
42866 Sodium chloride (NaCl)	Pimephales promelas	LOEC	8,000	86		94 r	mg/L CaCO3
96881 Sodium chloride (NaCl)	Pimephales promelas	LOEC	8,000	86		94 r	mg/L CaCO3
42423 Sodium chloride (NaCl)	Pimephales promelas	LOEC	8,000	86		94 n	mg/L CaCO3
96906 Sodium chloride (NaCl)	Pimephales promelas	LOEC	8,000	86		94 n	mg/L CaCO3

							Hardness	
Result				Concentration				
Number	Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
14389 So	14389 Sodium chloride (NaCl)	Pimephales promelas	LOEC	8,000	98		94	mg/L CaCO3
68598 So	68598 Sodium chloride (NaCl)	Pimephales promelas	LOEC	8,000	86		94	mg/L CaCO3
201561 So	201561 Sodium chloride (NaCl)	Pimephales promelas	LOEC	352		97		mg/L CaCO3
768104 So	768104 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,250	80		100	mg/L CaCO3
768072 So	768072 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	80		100	mg/L CaCO3
768157 So	768157 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	08		100	mg/L CaCO3
768066 So	768066 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	80			mg/L CaCO3
768058 So	768058 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	90		100	mg/L CaCO3
768096 So	768096 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	08		100	mg/L CaCO3
768127 So	768127 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80		100	mg/L CaCO3
768143 So	768143 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80			mg/L CaCO3
768090 So	768090 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80			mg/L CaCO3
768088 So	768088 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	80			mg/L CaCO3
768123 So	768123 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80			mg/L CaCO3
768163 So	768163 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	08			mg/L CaCO3
768153 So	768153 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80		100	mg/L CaCO3
768098 So	768098 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	08		100	mg/L CaCO3
768074 So	768074 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	08		100	mg/L CaCO3
768080 So	768080 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	08		100	mg/L CaCO3
768113 So	768113 Sodium chloride (NaCl)		LOEC	2,500	08		100	mg/L CaCO3
768107 So	768107 Sodium chloride (NaCl)		LOEC	2,500	08		100	mg/L CaCO3
768082 So	768082 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80		100	mg/L CaCO3
768137 So	768137 Sodium chloride (NaCl)		LOEC	2,500	08		100	mg/L CaCO3
768147 So	768147 Sodium chloride (NaCl)	Pimephales promelas	LOEC	2,500	80		100	mg/L CaCO3
768064 So	768064 Sodium chloride (NaCl)	Pimephales promelas	LOEC	5,000	80		100	mg/L CaCO3
768133 So	768133 Sodium chloride (NaCl)		LOEC	5,000	08		100	mg/L CaCO3
768117 So	768117 Sodium chloride (NaCl)		LOEC	2,500	80		100	mg/L CaCO3
741744 So	741744 Sodium chloride (NaCl)	Pimephales promelas	LOEL	1,140	90		110	ppm
742691 So	742691 Sodium chloride (NaCl)	Pimephales promelas	LOEL	210	90		110	ppm
741410 So	'41410 Sodium chloride (NaCl)	Pimephales promelas	LOEL	2,200	90		110	ppm
201562 Sov	201562 Sodium chloride (NaCl)		MATC	298		97		mg/L CaCO3
68599 So	68599 Sodium chloride (NaCl)	Pimephales promelas	NOEC	4,000	86		94	mg/L CaCO3

Result								
				Concentration				
42865 Sodium chloride (NaCl)	hloride (NaCl)	Pimenhales promelas	NOFC	4 000	86		_	mg/l CaCO3
	ide (NaCl)	Pimephales promelas	NOEC	4,000	86			mg/L CaCO3
96885 Sodium chloride	ide (NaCl)	Pimephales promelas	NOEC	4,000	86		94	mg/L CaCO3
69057 Sodium chloride	ide (NaCl)	Pimephales promelas	NOEC	4,000	86		94	mg/L CaCO3
96907 Sodium chloride	ide (NaCl)	Pimephales promelas	NOEC	4,000	86		94	mg/L CaCO3
201560 Sodium chloride	ide (NaCl)	Pimephales promelas	NOEC	252		97		mg/L CaCO3
768105 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	08		100	mg/L CaCO3
768073 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768158 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768067 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768059 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768097 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768128 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768144 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768091 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768089 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768124 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768164 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768154 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768099 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768075 Sodium chloride (NaCl)	ide (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768081 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768114 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768108 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768083 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768138 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	1,250	80			mg/L CaCO3
768148 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	1,250	80		100	mg/L CaCO3
768065 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	2,500	80			mg/L CaCO3
768134 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEC	2,500	80		100	mg/L CaCO3
768118 Sodium chloride (NaCl)	de (NaCl)		NOEC	1,250	80		100	mg/L CaCO3
741400 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEL	2,000	90		110	ppm
742666 Sodium chloride (NaCl)	de (NaCl)	Pimephales promelas	NOEL	860	90		110	ppm

						Hardness	
Result  Number Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
768558 Sodium chloride (NaCl)	Pimephales promelas	NR-LETH	6,069	80		100	mg/L CaCO3
201538 Sodium chloride (NaCl)	Pimephales promelas	NR-LETH	7,310		96		mg/L CaCO3
740697 Sodium chloride (NaCl)	Pimephales promelas	NR-LETH	20,000		140		mg/L CaCO3
768559 Sodium chloride (NaCl)	Pimephales promelas	NR-ZERO	3,035	80		100	mg/L CaCO3
740694 Sodium chloride (NaCl)	Pimephales promelas	NR-ZERO	10,000		140		mg/L CaCO3
740696 Sodium chloride (NaCl)	Pimephales promelas	NR-ZERO	10,000		140	.1	mg/L CaCO3
231532 Sodium chloride (NaCl)	lata	LC50	6,150		100		ppm
231528 Sodium chloride (NaCl)	Planorbella campanulata	LC50	> 10,000		100		ppm
231531 Sodium chloride (NaCl)	Planorbella campanulata LC50	LC50	6,150		100		ppm
231530 Sodium chloride (NaCl)	Planorbella campanulata LC50	LC50	6,150		100		ppm
231529 Sodium chloride (NaCl)	Planorbella campanulata LC50	LC50	7,500		100		ppm
2060321 Sodium chloride (NaCl)	Poecilia reticulata	LC50	> 11,700	288	290	292	mg/L CaCO3
2060324 Sodium chloride (NaCl)	Poecilia reticulata	NR-ZERO	11,700	288	290		mg/L CaCO3
2060323 Sodium chloride (NaCl)	Poecilia reticulata	NR-ZERO	7,720	288	290		mg/L CaCO3
2170552 Sodium chloride (NaCl)	Rocio octofasciata	NR-ZERO	4,000	132		205	ppm
740686 Sodium chloride (NaCl)	Sander vitreus	NR-ZERO	10,000		140		mg/L CaCO3
740685 Sodium chloride (NaCl)	Sander vitreus	NR-ZERO	10,000		140		mg/L CaCO3
2075473 Sodium chloride (NaCl)	Sphaerium simile	LC50	1,100		192		mg/L CaCO3
2075472 Sodium chloride (NaCl)	Sphaerium simile	LC50	740		51		mg/L CaCO3
231556 Sodium chloride (NaCl)	Sphaerium sp.	LC50	1,950		100		ppm
231560 Sodium chloride (NaCl)	Sphaerium sp.	LC50	1,550		20	_	ppm
231561 Sodium chloride (NaCl)		LC50	1,250		20		ppm
231559 Sodium chloride (NaCl)	Sphaerium sp.	LC50	2,400		20		ppm
231557 Sodium chloride (NaCl)		LC50	1,250		100		ppm
231558 Sodium chloride (NaCl)		LC50	1,100		100		ppm
231562 Sodium chloride (NaCl)		LC50	1,150		20		ppm
231555 Sodium chloride (NaCl)		LC50	2,250		100		ppm
2152833 Sodium chloride (NaCl)	Tubifex tubifex	LC50	5,648	80		100 r	mg/L CaCO3
2075476 Sodium chloride (NaCl)	Tubifex tubifex	LC50	4,278		52	_	mg/L CaCO3
2075477 Sodium chloride (NaCl)	Tubifex tubifex	LC50	6,008		220	_	mg/L CaCO3
803853 Sodium chloride (NaCl)	Villosa constricta	EC50	2,760	170		192 r	mg/L CaCO3
741562 Sodium chloride (NaCl)	Villosa constricta	EC50	2,590	170		192 r	mg/L CaCO3

	Hardness valu	ues associated wit	Hardness values associated with lethal toxicity end	dpoints.			
Result			Concentration			Hardness	
Number Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
741561 Sodium chloride (NaCl)	Villosa delumbis	EC50	3,310	170		192	mg/L CaCO3
	Villosa delumbis	EC50	5,230	170		192	mg/L CaCO3
Sodium chloride	Villosa delumbis	EC50	3,630	170		192	mg/L CaCO3
160288 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	LC50	1,770	80		100	mg/L CaCO3
Sulfuric	Ceriodaphnia dubia	LC50	1,770	80		100	mg/L CaCO3
767346 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-LETH	2,076	80		100	mg/L CaCO3
	Ceriodaphnia dubia	NR-LETH	2,076	80		100	mg/L CaCO3
	Ceriodaphnia dubia	NR-LETH	2,076	80		100	mg/L CaCO3
767402 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-LETH	2,076	80		100	mg/L CaCO3
767365 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-LETH	2,076	80		100	mg/L CaCO3
767407 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-LETH	2,076	80		100	mg/L CaCO3
767364 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,079	80		100	mg/L CaCO3
767366 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,079	80		100	mg/L CaCO3
767410 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	580	80		100	mg/L CaCO3
767347 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,079	80		100	mg/L CaCO3
767405 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,079	80		100	mg/L CaCO3
767461 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,036	80		100	mg/L CaCO3
767465 Sulfuric acid magnesium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,394	80		100	mg/L CaCO3
160289 Sulfuric acid magnesium salt (1:1)	Daphnia magna	LC50	2,360	80		100	mg/L CaCO3
160290 Sulfuric acid magnesium salt (1:1)	Daphnia magna	LC50	1,820	80		100	mg/L CaCO3
759182 Sulfuric acid magnesium salt (1:1)	Daphnia magna	LC50	4,300	404	563	694	mg/L CaCO3
767488 Sulfuric acid magnesium salt (1:1)	Daphnia magna	NR-LETH	4,071	80		100	mg/L CaCO3
767485 Sulfuric acid magnesium salt (1:1)	Daphnia magna	NR-LETH	4,071	80		100	mg/L CaCO3
767471 Sulfuric acid magnesium salt (1:1)		NR-ZERO	1,079	80		100	mg/L CaCO3
767487 Sulfuric acid magnesium salt (1:1)	Daphnia magna	NR-ZERO	1,079	80		100	mg/L CaCO3
767490 Sulfuric acid magnesium salt (1:1)	Daphnia magna	NR-ZERO	580	80		100	mg/L CaCO3
759473 Sulfuric acid magnesium salt (1:1)	Daphnia magna	NR-ZERO	360		170		mg/L CaCO3
759470 Sulfuric acid magnesium salt (1:1)	Daphnia magna	NR-ZERO	360		170		mg/L CaCO3
160291 Sulfuric acid magnesium salt (1:1)	Pimephales promelas	LC50	4,630	80		100	mg/L CaCO3
160292 Sulfuric acid magnesium salt (1:1)		LC50	3,510	80		100	mg/L CaCO3
160293 Sulfuric acid magnesium salt (1:1)   Pimephales promelas	Pimephales promelas	LC50	2,820	80		100	mg/L CaCO3
759181 Sulfuric acid magnesium salt (1:1)	Pimephales promelas	LC50	7,900	404	563	694	mg/L CaCO3

	Hardness valu	ies associated wit	Hardness values associated with lethal toxicity endpoints.	dpoints.			
						Hardness	
Result			Concentration				
Number Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
767519 Sulfuric acid magnesium salt (1:1)	Pimephales promelas	NR-ZERO	2,076	08		100	mg/L CaCO3
	Ceriodaphnia dubia	EC50	1,551		160		ppm
	Ceriodaphnia dubia	EC50	1,619		320		ppm
	Ceriodaphnia dubia	EC50	1,267		80		ppm
	Ceriodaphnia dubia	EC50	914		40		ppm
	Ceriodaphnia dubia	LC10	2,547		578		mg/L CaCO3
766427 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC10	2,173		194		mg/L CaCO3
	Ceriodaphnia dubia	LC10	2,793		484		mg/L CaCO3
	Ceriodaphnia dubia	LC10	2,216	80	107	100	mg/L CaCO3
766431 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC10	2,744		390		mg/L CaCO3
766429 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC10	2,389		288		mg/L CaCO3
	Ceriodaphnia dubia	LC10	1,759		94		mg/L CaCO3
	Ceriodaphnia dubia	LC50	3,590	80		100	mg/L CaCO3
160331 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,080	80		100	mg/L CaCO3
766604 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,853		288		mg/L CaCO3
766576 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,830		98		mg/L CaCO3
766609 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,716		484		mg/L CaCO3
	Ceriodaphnia dubia	LC50	2,526		102		mg/L CaCO3
766567 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,724		96		mg/L CaCO3
766580 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,328		96		mg/L CaCO3
766565 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,494		94		mg/L CaCO3
	Ceriodaphnia dubia	LC50	3,073		390		mg/L CaCO3
	Ceriodaphnia dubia	LC50	2,243		94		mg/L CaCO3
766574 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,757		96		mg/L CaCO3
766577 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,793		102		mg/L CaCO3
766560 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,402		94		mg/L CaCO3
766603 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,361		288		mg/L CaCO3
766575 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,184		104		mg/L CaCO3
766581 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,828		98		mg/L CaCO3
	Ceriodaphnia dubia	LC50	2,404		96		mg/L CaCO3
766578 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,095		102		mg/L CaCO3
766595 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,383		288		mg/L CaCO3

						Hardness	
Result  Chemical Name	Species Scientific Name	Endpoint	Concentration (mg/L)	Minimum	Mean	Maximum	Units
766586 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,868		94		mg/L CaCO3
	Ceriodaphnia dubia	LC50	3,506		484		mg/L CaCO3
766555 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,223		92		mg/L CaCO3
766566 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,198		100		mg/L CaCO3
766570 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,925		96		mg/L CaCO3
766605 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,297		288		mg/L CaCO3
766591 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,436		107		mg/L CaCO3
766572 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,481		96		mg/L CaCO3
766548 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,526		100		mg/L CaCO3
766606 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,369		390		mg/L CaCO3
766585 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,030		94		mg/L CaCO3
766588 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,538		107		mg/L CaCO3
766608 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,091		390		mg/L CaCO3
766551 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,357		100		mg/L CaCO3
766563 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,295		94		mg/L CaCO3
766592 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,059		194		mg/L CaCO3
766587 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,269		94		mg/L CaCO3
766612 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,338		484		mg/L CaCO3
766561 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,470		94		mg/L CaCO3
766590 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,607		107		mg/L CaCO3
766593 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,706		194		mg/L CaCO3
766594 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,265		194		mg/L CaCO3
766571 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,472		96		mg/L CaCO3
766584 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,116		94		mg/L CaCO3
766228 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,065		92		mg/L CaCO3
766227 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,446		92		mg/L CaCO3
766221 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,049		92	1	mg/L CaCO3
766226 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,442		92		mg/L CaCO3
766426 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,000		194	1	mg/L CaCO3
766432 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,516		484	1	mg/L CaCO3
766430 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,174		390	-	mg/L CaCO3
766418 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	TC20	2,526	80	107	100 r	mg/L CaCO3

						Hardness	
Result			Concentration				
Number Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
766428 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,946		288		mg/L CaCO3
Sulfuric	Ceriodaphnia dubia	LC50	2,050		94		mg/L CaCO3
766434 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,288		578		mg/L CaCO3
766720 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,419		100		mg/L CaCO3
766717 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,469		100		mg/L CaCO3
766722 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,272		100		mg/L CaCO3
766718 Sulfuric acid sodium salt (1:2)		LC50	2,289		100		mg/L CaCO3
766723 Sulfuric acid sodium salt (1:2)		LC50	2,417		100		mg/L CaCO3
766724 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,914		100		mg/L CaCO3
766726 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	1,496		100		mg/L CaCO3
766745 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	3,250	08		100	mg/L CaCO3
766748 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LC50	2,250	80		100	mg/L CaCO3
766224 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	3,000		92		mg/L CaCO3
766222 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	2,216		92		mg/L CaCO3
766739 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	3,000	80		100	mg/L CaCO3
766730 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	2,273	80		100	mg/L CaCO3
2017050 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	2,600		160		ppm
2017059 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	2,700		320		mdd
2017042 Sulfuric acid sodium salt (1:2)		LOEC	2,000		80		mdd
2017031 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	LOEC	1,300		40		ppm
766225 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	2,250		92		mg/L CaCO3
766223 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	1,750		92		mg/L CaCO3
766854 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	1,301	80		100	mg/L CaCO3
766740 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	2,264	80		100	mg/L CaCO3
766731 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	1,727	80		100	mg/L CaCO3
2017049 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	1,300		160		ppm
2017058 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	1,450		320		ppm
2017041 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	1,250		80		mdd
2017030 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NOEC	610		40		mdd
767457 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NR-LETH	2,989	80		100	mg/L CaCO3
767375 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NR-LETH	3,463	80		100	mg/L CaCO3
767380 Sulfuric acid sodium salt (1:2)	Ceriodaphnia dubia	NR-LETH	3,463	80		100	mg/L CaCO3

5		80		2,069	EC10	Hyalella azteca	2017113 Sulfuric acid sodium salt (1:2)	201711
mg/L CaCO3	100		80	1,434	NR-ZERO	Daphnia magna	Sulfuric acid sodium salt (1:2)	767507
mg/L CaCO3	100		80	927	NR-ZERO	Daphnia magna	67494 Sulfuric acid sodium salt (1:2)	76749
mg/L CaCO3	100		80	5,492	NR-LETH	Daphnia magna	767505 Sulfuric acid sodium salt (1:2)	76750
mg/L CaCO3	100		80	6,844	NR-LETH	Daphnia magna	767491 Sulfuric acid sodium salt (1:2)	76749
mg/L CaCO3	100		80	6,844	NR-LETH	Daphnia magna	767475 Sulfuric acid sodium salt (1:2)	76747
mg/L CaCO3	100		80	6,844	NR-LETH	Daphnia magna	767496 Sulfuric acid sodium salt (1:2)	76749
mg/L CaCO3	100		80	6,844	NR-LETH	Daphnia magna	767473 Sulfuric acid sodium salt (1:2)	76747
mg/L CaCO3	694	563	404	8,600	LC50	Daphnia magna	759175 Sulfuric acid sodium salt (1:2)	75917
mg/L CaCO3		106		3,203	LC50	Daphnia magna	767299 Sulfuric acid sodium salt (1:2)	76729
mg/L CaCO3		25		1,563	LC50	Daphnia magna	767305 Sulfuric acid sodium salt (1:2)	76730
mg/L CaCO3		100		3,203	LC50	Daphnia magna	767297 Sulfuric acid sodium salt (1:2)	76729
mg/L CaCO3		25		1,194	LC50	Daphnia magna	767298 Sulfuric acid sodium salt (1:2)	76729
mg/L CaCO3		50		1,551	LC50	Daphnia magna	767295 Sulfuric acid sodium salt (1:2)	76729
mg/L CaCO3		106		3,808	LC50	Daphnia magna	767303 Sulfuric acid sodium salt (1:2)	76730
mg/L CaCO3		75		3,342	LC50	Daphnia magna	767296 Sulfuric acid sodium salt (1:2)	76729
mg/L CaCO3		25		1,985	LC50	Daphnia magna	767306 Sulfuric acid sodium salt (1:2)	76730
mg/L CaCO3		106		4,395	LC50	Daphnia magna	767307 Sulfuric acid sodium salt (1:2)	76730
mg/L CaCO3		25		1,194	LC50	Daphnia magna	767294 Sulfuric acid sodium salt (1:2)	76729
mg/L CaCO3	100		80	4,580	LC50	Daphnia magna	160333 Sulfuric acid sodium salt (1:2)	16033
mg/L CaCO3	100		80	6,290	LC50	Daphnia magna	160332 Sulfuric acid sodium salt (1:2)	16033
mg/L CaCO3		94		14,134	LC50	Chironomus tentans	766217 Sulfuric acid sodium salt (1:2)	76621
mg/L CaCO3		94		11,682	LC10	Chironomus tentans	766218 Sulfuric acid sodium salt (1:2)	76621
mg/L CaCO3	100		80	1,772	NR-ZERO	Ceriodaphnia dubia	767376 Sulfuric acid sodium salt (1:2)	76737
mg/L CaCO3	100		80	1,772	NR-ZERO	Ceriodaphnia dubia	767353 Sulfuric acid sodium salt (1:2)	76735
mg/L CaCO3	100		80	1,772	NR-ZERO	Ceriodaphnia dubia	767412 Sulfuric acid sodium salt (1:2)	76741
mg/L CaCO3	100		80	2,178	NR-ZERO	Ceriodaphnia dubia	767456 Sulfuric acid sodium salt (1:2)	76745
mg/L CaCO3	100		80	2,178	NR-ZERO	Ceriodaphnia dubia	767459 Sulfuric acid sodium salt (1:2)	76745
mg/L CaCO3	100		80	1,772	NR-ZERO	Ceriodaphnia dubia	767351 Sulfuric acid sodium salt (1:2)	76735
mg/L CaCO3	100		80	2,989	NR-LETH	Ceriodaphnia dubia	767454 Sulfuric acid sodium salt (1:2)	76745
mg/L CaCO3	100		80	3,463	NR-LETH	Ceriodaphnia dubia	767411 Sulfuric acid sodium salt (1:2)	76741
mg/L CaCO3	100		80	3,463	NR-LETH	Ceriodaphnia dubia	767350 Sulfuric acid sodium salt (1:2)	76735
Units	Maximum	Mean	Minimum	Concentration (mg/L)	Endpoint	Species Scientific Name	r Chemical Name	Result Number
	Hardness							

fite Name         Endpoint         Concentration (mg/t)         Minimum         Mean         Maximum           1         EC25         2,246         80         Minimum         Mean         Maximum           1         EC50         2,461         80         80         40								Hardnace	
Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,246   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,461   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,461   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,461   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,461   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,461   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,461   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,741   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   2,741   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,336   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,336   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,336   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,336   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)   Hyalella azteca   ECS9   4,046   Sulfuric acid sodium salt (1.2)	Result				Concentration			naroness	
Hyalella azteca   EC25   2,246   80     Hyalella azteca   EC50   2,461   94     Hyalella azteca   LC50   2,840   94     Hyalella azteca   LC50   2,840   95     Hyalella azteca   LC50   2,724   392     Hyalella azteca   LC50   2,724   395     Hyalella azteca   LC50   1,779   96     Hyalella azteca   LC50   1,767   95     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,684   100     Hyalella azteca   LC50   1,684   100     Hyalella azteca   LC50   1,621   98     Hyalella azteca   LC50   1,621   98     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   2,002   1,901     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   2,955   2,955   2,955     Hyalella azteca   LC50   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,955   2,	Number	Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
Hyalella azteca   ECSO   2,461   80     Hyalella azteca   ICGO   262   94     Hyalella azteca   ICGSO   2,840   292     Hyalella azteca   ICGSO   4,145   486     Hyalella azteca   ICGSO   2,724   392     Hyalella azteca   ICGSO   1,779   96     Hyalella azteca   ICGSO   1,767   96     Hyalella azteca   ICGSO   1,634   100     Hyalella azteca   ICGSO   1,634   194     Hyalella azteca   ICGSO   1,621   98     Hyalella azteca   ICGSO   1,621   98     Hyalella azteca   ICGSO   1,840   194     Hyalella azteca   ICGSO   1,840   194     Hyalella azteca   ICGSO   1,840   194     Hyalella azteca   ICGSO   1,840   100     Hyalella azteca   ICGSO   1,840   100     Hyalella azteca   ICGSO   1,879   98     Hyalella azteca   ICGSO   2,000   100     Hyalella azteca   ICGSO   2,002   100     Hy	2017143 Sulfu	ric acid sodium salt (1:2)	Hyalella azteca	EC25	2,246		80		pm
Hyalella azteca   LC10   262   2,840   2,94     Hyalella azteca   LC50   2,840   2,92     Hyalella azteca   LC50   2,724   392     Hyalella azteca   LC50   2,724   395     Hyalella azteca   LC50   2,724   396     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,584   100     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   1,821   98     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   2,000   106     Hyalella azteca   LC50   2,000   100     Hyalella azteca   LC50   2,055   1,901   100     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,002   2,002   2,002     Hyalella azteca   LC50   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002	2017144 Sulfu	ric acid sodium salt (1:2)	Hyalella azteca	EC50	2,461		80		ppm
b) Hyalella azteca         LC50         2,840         292           c) Hyalella azteca         LC50         4,145         486           d) Hyalella azteca         LC50         4,145         486           e) Hyalella azteca         LC50         2,724         392           e) Hyalella azteca         LC50         1,767         96           e) Hyalella azteca         LC50         1,634         100           e) Hyalella azteca         LC50         1,634         100           e) Hyalella azteca         LC50         2,740         194           e) Hyalella azteca         LC50         1,621         98           e) Hyalella azteca         LC50         3,796         396           e) Hyalella azteca         LC50         1,480         100           e) Hyalella azteca         LC50         1,820         100           e) Hyalella azteca         LC50         1,879         98           e) Hyalella azteca         LC50         1,879         98           e) Hyalella azteca         LC50         1,879         98           e) Hyalella azteca         LC50         2,000         100           e) Hyalella azteca         LC50         2,002         100	766367 Sulfu	ric acid sodium salt (1:2)	Hyalella azteca	LC10	262		94	_	ng/L CaCO3
Hyalella azteca   LC50   4,145   392     Hyalella azteca   LC50   2,724   392     Hyalella azteca   LC50   1,779   96     Hyalella azteca   LC50   1,836   396     Hyalella azteca   LC50   1,884   100     Hyalella azteca   LC50   1,884   100     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   2,740   396     Hyalella azteca   LC50   2,740   396     Hyalella azteca   LC50   3,796   396     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,816   396     Hyalella azteca   LC50   1,829   398     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   2,000   100     Hyalella azteca   LC50   2,000   100     Hyalella azteca   LC50   2,000   100     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   293     Hyalella azteca   LC50   3,462   296     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2,002   2	766638 Sulfu	ric acid sodium salt (1:2)	Hyalella azteca	LC50	2,840		292		ng/L CaCO3
Hyalella azteca   LC50   2,724   392     Hyalella azteca   LC50   1,779   96     Hyalella azteca   LC50   4,336   396     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,684   100     Hyalella azteca   LC50   1,621   98     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,829   98     Hyalella azteca   LC50   2,000   106     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,003   1,438   10	766642 Sulfu	ric acid sodium salt (1:2)	Hyalella azteca	LC50	4,145		486		ng/L CaCO3
Hyalella azteca   LC50   1,779   96     Hyalella azteca   LC50   4,336   396     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   3,465   396     Hyalella azteca   LC50   3,766   398     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50	766639 Sulfu	ric acid sodium salt (1:2)	Hyalella azteca	LC50	2,724		392	1	ng/L CaCO3
Hyalella azteca	766618 Sulfu	ric acid sodium salt (1:2)		LC50	1,779		96	1	ng/L CaCO3
Hyalella azteca   LC50   1,767   96     Hyalella azteca   LC50   1,684   100     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   3,796   396     Hyalella azteca   LC50   3,796   396     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   2,000   196     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,121   296     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,830   100     Hyalella azteca   LC50   1,438   50     Hyalella azteca   LC50   1,438   50     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,448   50	766641 Sulfu	ric acid sodium salt (1:2)		LC50	4,336		396	7	ng/L CaCO3
Hyalella azteca   LC50   1,684   100     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   1,621   98     Hyalella azteca   LC50   4,046   396     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   2,002   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   1,438   100     Hyalella azteca   LC50   2,121   296     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,830   100     Hyalella azteca   LC50   1,830   100     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50   1,438   50     Hyalella azteca   LC50   1,448   50     Hyalella azteca   LC50   1,44	766616 Sulfu	ric acid sodium salt (1:2)		LC50	1,767		96	r	ng/L CaCO3
Hyalella azteca   LC50   2,740   194     Hyalella azteca   LC50   1,621   98     Hyalella azteca   LC50   4,046   396     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,579   98     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   2,000   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   1,438   100     Hyalella azteca   LC50   1,438   98     Hyalella azteca   LC50   2,002   100     H	766625 Sulfu	ric acid sodium salt (1:2)		LC50	1,684		100	r	ng/L CaCO3
Hyalella azteca   LC50   1,621   98     Hyalella azteca   LC50   4,046   396     Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,480   100     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   2,000   106     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   1,438   100     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,203   98     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   2,203   100     Hyalella azteca   LC50   1,438   100	766633 Sulfu	ric acid sodium salt (1:2)		LC50	2,740		194	r	ng/L CaCO3
Hyalella azteca   LC50   4,046   396   Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,480   100     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   2,000   106     Hyalella azteca   LC50   2,000   106     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   3,462   292     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,121   296     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   2,203   190     Hyalella azteca   LC50   1,830   100     Hyalella azteca   LC50   1,830   100	766631 Sulfu	ric acid sodium salt (1:2)		LC50	1,621		98	1	ng/L CaCO3
Hyalella azteca   LC50   3,796   482     Hyalella azteca   LC50   1,480   100     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,820   100     Hyalella azteca   LC50   1,879   98     Hyalella azteca   LC50   2,000   106     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,679   98     Hyalella azteca   LC50   1,901   100     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   2,955   196     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,002   100     Hyalella azteca   LC50   2,121   296     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   2,203   100     Hyalella azteca   LC50   2,203   100     Hyalella azteca   LC50   2,203   100     Hyalella azteca   LC50   2,203   192     Hyalella azteca   LC50   1,438   50     Hyalella azteca   LC50   1,438   50     Hyalella azteca   LC50   2,203   100     Hyalella azteca   LC50   1,438   50	766640 Sulfu	ric acid sodium salt (1:2)		LC50	4,046		396	r	ng/L CaCO3
Hyalella azteca         LC50         1,480         100           Hyalella azteca         LC50         1,820         100           Hyalella azteca         LC50         1,616         96           Hyalella azteca         LC50         1,879         98           Hyalella azteca         LC50         2,000         106           Hyalella azteca         LC50         4,345         482           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,202         102           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,203         192           Hyalella azteca <td>766644 Sulfui</td> <td>ric acid sodium salt (1:2)</td> <td></td> <td>LC50</td> <td>3,796</td> <td></td> <td>482</td> <td>1</td> <td>ng/L CaCO3</td>	766644 Sulfui	ric acid sodium salt (1:2)		LC50	3,796		482	1	ng/L CaCO3
Hyalella azteca         LC50         1,820         100           Hyalella azteca         LC50         1,616         96           Hyalella azteca         LC50         1,879         98           Hyalella azteca         LC50         2,000         106           Hyalella azteca         LC50         1,679         98           Hyalella azteca         LC50         1,679         98           Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,202         102           Hyalella azteca         LC50         2,203         193           Hyalella azteca         LC50         2,203         102           Hyalella azteca         LC50         2,203         100           Hyalella azteca	766627 Sulfui	ric acid sodium salt (1:2)		LC50	1,480		100	r	ng/L CaCO3
Hyalella azteca         LC50         1,616         96           Hyalella azteca         LC50         1,879         98           Hyalella azteca         LC50         2,000         106           Hyalella azteca         LC50         1,679         98           Hyalella azteca         LC50         4,345         482           Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,438         98           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         2,002         102           Hyalella azteca	766617 Sulfui	ric acid sodium salt (1:2)		LC50	1,820		100	r	ng/L CaCO3
Hyalella azteca         LC50         1,879         98           Hyalella azteca         LC50         2,000         106           Hyalella azteca         LC50         1,679         98           Hyalella azteca         LC50         4,345         482           Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196         292           Hyalella azteca         LC50         3,462         292         100           Hyalella azteca         LC50         2,002         100         100           Hyalella azteca         LC50         1,438         100         98           Hyalella azteca         LC50         2,002         100         100           Hyalella azteca         LC50         2,121         296         102           Hyalella azteca         LC50         2,203         192         100           Hyalella azteca         LC50         2,203         192         100           Hyalella azteca         LC50         1,830         100         100           Hyalella azteca         LC50         1,830         100         100	766624 Sulfui	ric acid sodium salt (1:2)		LC50	1,616		96	1	ng/L CaCO3
Hyalella azteca         LC50         2,000         106           Hyalella azteca         LC50         1,679         98           Hyalella azteca         LC50         4,345         482           Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196         196           Hyalella azteca         LC50         3,462         292         100           Hyalella azteca         LC50         1,438         100         100           Hyalella azteca         LC50         1,438         100         98           Hyalella azteca         LC50         2,121         296         102           Hyalella azteca         LC50         2,203         102         102           Hyalella azteca         LC50         2,121         296         102           Hyalella azteca         LC50         2,203         102         103           Hyalella azteca         LC50         1,830         100         103           Hyalella azteca         LC50         1,830         100         100           Hyalella azteca         LC50         1,448         50	766619 Sulfui	ric acid sodium salt (1:2)		LC50	1,879		98	ı	ng/L CaCO3
Hyalella azteca         LC50         1,679         98           Hyalella azteca         LC50         4,345         482           Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         2,002         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,433         98           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,002         102           Hyalella azteca         LC50         1,830         100           Hyalella azteca         LC50         1,448         50	766629 Sulfui	ric acid sodium salt (1:2)		LC50	2,000		106	1	ng/L CaCO3
Hyalella azteca         LC50         4,345         482           Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         2,002         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,202         102           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         1,830         192           Hyalella azteca         LC50         1,448         50	766623 Sulfui	ric acid sodium salt (1:2)		LC50	1,679		98	r	ng/L CaCO3
Hyalella azteca         LC50         1,901         100           Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         2,002         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,433         98           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,002         102           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         1,830         100           Hyalella azteca         LC50         1,448         50	766643 Sulfui	ric acid sodium salt (1:2)		LC50	4,345		482	ı	ng/L CaCO3
Hyalella azteca         LC50         2,955         196           Hyalella azteca         LC50         3,462         292           Hyalella azteca         LC50         2,002         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,433         98           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,203         102           Hyalella azteca         LC50         1,830         192           Hyalella azteca         LC50         1,448         50	766630 Sulfui	ric acid sodium salt (1:2)		LC50	1,901		100	r	ng/L CaCO3
Hyalella azteca       LC50       3,462       292         Hyalella azteca       LC50       2,002       100         Hyalella azteca       LC50       1,438       100         Hyalella azteca       LC50       1,433       98         Hyalella azteca       LC50       2,121       296         Hyalella azteca       LC50       2,002       102         Hyalella azteca       LC50       2,203       192         Hyalella azteca       LC50       1,830       100         Hyalella azteca       LC50       1,448       50	766635 Sulfur	ric acid sodium salt (1:2)		LC50	2,955		196	ı	ng/L CaCO3
Hyalella azteca         LC50         2,002         100           Hyalella azteca         LC50         1,438         100           Hyalella azteca         LC50         1,433         98           Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,002         102           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         1,830         100           Hyalella azteca         LC50         1,448         50	766637 Sulfur	ric acid sodium salt (1:2)		LC50	3,462		292	٦	ng/L CaCO3
Hyalella azteca       LC50       1,438       100         Hyalella azteca       LC50       1,433       98         Hyalella azteca       LC50       2,121       296         Hyalella azteca       LC50       2,002       102         Hyalella azteca       LC50       2,203       192         Hyalella azteca       LC50       1,830       100         Hyalella azteca       LC50       1,448       50	766622 Sulfur	ric acid sodium salt (1:2)		LC50	2,002		100	r	ng/L CaCO3
Hyalella azteca       LC50       1,433       98         Hyalella azteca       LC50       2,121       296         Hyalella azteca       LC50       2,002       102         Hyalella azteca       LC50       2,203       192         Hyalella azteca       LC50       1,830       100         Hyalella azteca       LC50       1,448       50	766628 Sulfur	ric acid sodium salt (1:2)		LC50	1,438		100	J	ng/L CaCO3
Hyalella azteca         LC50         2,121         296           Hyalella azteca         LC50         2,002         102           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         1,830         100           Hyalella azteca         LC50         1,448         50	766626 Sulfur	ric acid sodium salt (1:2)		LC50	1,433		98	n	ng/L CaCO3
Hyalella azteca         LC50         2,002         102           Hyalella azteca         LC50         2,203         192           Hyalella azteca         LC50         1,830         100           Hyalella azteca         LC50         1,448         50	766636 Sulfur	ric acid sodium salt (1:2)		LC50	2,121		296	n	ng/L CaCO3
Hyalella azteca       LC50       2,203       192         Hyalella azteca       LC50       1,830       100         Hyalella azteca       LC50       1,448       50	766632 Sulfur	ric acid sodium salt (1:2)		LC50	2,002		102	7	ng/L CaCO3
Hyalella azteca         LC50         1,830         100           Hyalella azteca         LC50         1,448         50	766634 Sulfur	ric acid sodium salt (1:2)		LC50	2,203		192	n	ng/L CaCO3
Hyalella azteca   LC50 1,448   50	766621 Sulfur	ric acid sodium salt (1:2)		LC50	1,830		100	п	ng/L CaCO3
	767290 Sulfur	ric acid sodium salt (1:2)	Hyalella azteca	LC50	1,448		50	u	mg/L CaCO3

						Hardness	
Result			Concentration				
Number Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
767310 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	2,725		102		mg/L CaCO3
767293 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	5,259		250		mg/L CaCO3
767291 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,580		75		mg/L CaCO3
767311 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	2,240		102		mg/L CaCO3
767312 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	2,101		102		mg/L CaCO3
767289 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	569		25		mg/L CaCO3
767292 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	3,144		123		mg/L CaCO3
766365 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	512		94		mg/L CaCO3
766415 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	2,855	08	107	100	mg/L CaCO3
766712 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,854		100		mg/L CaCO3
766716 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,470		100		mg/L CaCO3
766713 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,799		100		mg/L CaCO3
766714 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,938		100		mg/L CaCO3
766715 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,691		100	4	mg/L CaCO3
766710 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,563		100		mg/L CaCO3
766711 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,562		100		mg/L CaCO3
766709 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LC50	1,387		100		mg/L CaCO3
766446 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	1,460		300		mg/L CaCO3
766441 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	1,460		200		mg/L CaCO3
766438 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	1,460		100		mg/L CaCO3
766439 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	1,460		100		mg/L CaCO3
766443 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	1,460		200		mg/L CaCO3
766442 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	1,460		200		mg/L CaCO3
2017146 Sulfuric acid sodium salt (1:2)	Hyalella azteca	LOEC	2,412		80		ppm
766444 Sulfuric acid sodium salt (1:2)	Hyalella azteca	NOEC	1,460		300		mg/L CaCO3
766445 Sulfuric acid sodium salt (1:2)	Hyalella azteca	NOEC	1,460		300		mg/L CaCO3
2017145 Sulfuric acid sodium salt (1:2)	Hyalella azteca	NOEC	1,637	7	80		ppm
766440 Sulfuric acid sodium salt (1:2)	Hyalella azteca	NR-LETH	1,460		100		mg/L CaCO3
764945 Sulfuric acid sodium salt (1:2)	Hyalella azteca	NR-ZERO	138	132	137	140	mg/L CaCO3
764947 Sulfuric acid sodium salt (1:2)	Hyalella azteca	NR-ZERO	642	138	141	142	mg/L CaCO3
766659 Sulfuric acid sodium salt (1:2)	Lampsilis siliquoidea	LC50	3,525		300		mg/L CaCO3
766663 Sulfuric acid sodium salt (1:2)	Lampsilis siliquoidea	LC50	1,702		100		mg/L CaCO3

	Hardness valu	res associated wit	Hardness values associated with lethal toxicity end	dpoints.			
Result			Concentration			naroness	
Number Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
766664 Sulfuric acid sodium salt (1:2)	Lampsilis siliquoidea	LC50	1,727		100		mg/L CaCO3
766658 Sulfuric acid sodium salt (1:2)		LC50	3,523		300		mg/L CaCO3
Sulfuric acid sodium salt		LC50	1,822		100		mg/L CaCO3
766662 Sulfuric acid sodium salt (1:2)		LC50	3,729		500		mg/L CaCO3
766657 Sulfuric acid sodium salt (1:2)		LC50	3,377		100		mg/L CaCO3
766656 Sulfuric acid sodium salt (1:2)	Lampsilis siliquoidea	LC50	3,183		100		mg/L CaCO3
766665 Sulfuric acid sodium salt (1:2)		LC50	2,661		100		mg/L CaCO3
766661 Sulfuric acid sodium salt (1:2)	Lampsilis siliquoidea	LC50	3,574		500		mg/L CaCO3
200196 Sulfuric acid sodium salt (1:2)	Lepomis macrochirus	LC50	13,500		10		mg/L CaCO3
40509 Sulfuric acid sodium salt (1:2)	Lepomis macrochirus	LC50	3,040	35		62	mg/L CaCO3
40508 Sulfuric acid sodium salt (1:2)	Lepomis macrochirus	LC50	4,380	35		62	mg/L CaCO3
2017209 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC10	2,451		320		mdd
2017178 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC10	1,555		80		mdd
2017188 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC10	3,231		160		mdd
2017168 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC10	559		40		ppm
2017210 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC25	> 5,250		320		ppm
2017179 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC25	2,183		80		ppm
2017189 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC25	3,801		160		ppm
2017169 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC25	933		40		ppm
2017211 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC50	> 5,250		320		ppm
2017180 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC50	2,938		80		ppm
2017190 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC50	4,553		160	-	mdd
2017170 Sulfuric acid sodium salt (1:2)	Pimephales promelas	EC50	1,649		40	T	mdd
160335 Sulfuric acid sodium salt (1:2)	Pimephales promelas	LC50	> 7,960	80		100 r	mg/L CaCO3
160334 Sulfuric acid sodium salt (1:2)	Pimephales promelas	LC50	> 8,080	80		100 r	mg/L CaCO3
160336 Sulfuric acid sodium salt (1:2)		LC50	7,960	80			mg/L CaCO3
759176 Sulfuric acid sodium salt (1:2)		LC50	15,200	404	563		mg/L CaCO3
2017213 Sulfuric acid sodium salt (1:2)		COEC	5,250		320	7	ppm
2017182 Sulfuric acid sodium salt (1:2)		LOEC	2,850		80	T	ppm
2017172 Sulfuric acid sodium salt (1:2)	Pimephales promelas	LOEC	1,250		40	7	ppm
2017192 Sulfuric acid sodium salt (1:2)		LOEC	5,500		160	-	ppm
766856 Sulfuric acid sodium salt (1:2)		NOEC	1,301	80		100	mg/I CaCO3

Result         Chemical Name         Species Scientific Name         Endpoint         Concentration (mg/L)         Maintum (mg/L)         Hardness           2017/19/15/Sulfuric acid sodium salt (1/2)         Prinzephales promeios         NOEC         2,550         100         100         ppm           2017/19/Sulfuric acid sodium salt (1/2)         Prinzephales promeios         NOEC         2,900         320         ppm           2017/18/Sulfuric acid sodium salt (1/2)         Prinzephales promeios         NOEC         2,900         30         90         ppm           2017/18/Sulfuric acid sodium salt (1/2)         Prinzephales promeios         NOEC         2,900         30         40         ppm           764932/Sulfuric acid socium salt (1/2)         Prinzephales promeios         NR-ZERO         214         140         145         148         mg/L (aciO3)           764932/Sulfuric acid socium salt (1/2)         Prinzephales promeios         NR-ZERO         24         124         135         144         mg/L (aciO3)           764932/Sulfuric acid socium salt (1/2)         Prinzephales promeios         NR-ZERO         24         124         135         144         mg/L (aciO3)           764932/Sulfuric acid socium salt (1/2)         Prinzephales promeios         NR-ZERO         24         124 <t< th=""><th></th><th>Hardness valu</th><th>ues associated wit</th><th>Hardness values associated with lethal toxicity endpoints</th><th>dpoints.</th><th></th><th></th><th></th></t<>		Hardness valu	ues associated wit	Hardness values associated with lethal toxicity endpoints	dpoints.			
Sulfuric acid sodium salt (1.2)   Pimepholes promelos   NOEC   2,900   320   100   150   110   150   110   150   110   150   110   150				7/			Hardness	
Sulfuric acid sodium salt (1:2)   Pinnephales promelas   NNOEC   2,900   320   120   130   147   152   148   141   142   144   145   144   145   144   145   144   145   144   145	Result			Concentration				
Pimephales promeias   NOEC   2,850   2,900   2,900   2,900   2,900   330   2,900   2,900   330   330   2,900   2,900   330   330   2,900   2,900   330   330   2,900   330		Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
Pimephales promelas   NOEC   2,900   320   Pimephales promelas   NOEC   1,300   80   80   Pimephales promelas   NOEC   1,300   80   80   80   80   80   80   80	2017191 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NOEC	2,850		160		ppm
Pimephales promelas   NOEC   1,300   80     Pimephales promelas   NOEC   595   140   145   148     Pimephales promelas   NR-ZERO   214   140   147   152     Pimephales promelas   NR-ZERO   160   128   131   132     Pimephales promelas   NR-ZERO   24   124   136   144     Pimephales promelas   NR-ZERO   24   128   131   132     Pimephales promelas   NR-ZERO   2,078   94     Sphaerium simile   LC50   2,590   280   280     Sphaerium simile   LC50   2,926   269   293     Sphaerium simile   LC50   2,186   84     Sphaerium simile   LC50   2,186   84     Sphaerium simile   LC50   2,190   80   100     Sphaerium simile   LC50   2,197   80   100     Ceriodaphnia dubia   LC50   > 1,970   80   100     Ceriodaphnia dubia   NR-ZERO   765   80   100     Ceriodaphnia dubia   NR-ZERO   765   80   100     Ceriodaphnia dubia   NR-ZERO   765   80   100     Ceriodaphnia dubia   NR-ZERO   1,447   80   100     Ceriodaphnia magna   NR-ZERO   1,468   1	2017212 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NOEC	2,900		320		ppm
Pimephales promelas   NOEC   595   40   40     Pimephales promelas   NR-ZERO   214   140   145   148     Pimephales promelas   NR-ZERO   906   140   147   152     Pimephales promelas   NR-ZERO   906   140   147   152     Pimephales promelas   NR-ZERO   160   128   131   132     Pimephales promelas   NR-ZERO   24   124   136   144     Pimephales promelas   NR-ZERO   24   128   131   132     Pimephales promelas   NR-ZERO   24   128   131   132     Pimephales promelas   NR-ZERO   24   128   131   132     Pimephales promelas   LC50   2,978   94     Sphaerium simile   LC50   2,978   94     Sphaerium simile   LC50   2,926   280   280     Sphaerium simile   LC50   2,186   84     Sphaerium simile   LC50   2,186   84     Sphaerium simile   LC50   2,190   93   100     Sphaerium simile   LC50   2,190   80   100     Sphaerium simile   LC50   2,190   80   100     Sphaerium simile   LC50   > 1,970   80   100     Sphaerium simile   LC50   > 1,970   80   100     Ceriodaphnia dubia   LC50   > 1,970   80   100     Ceriodaphnia dubia   NR-ZERO   1,447   80   100     Ceriodaphnia dubia   NR-ZERO   1,447   80   100     Ceriodaphnia dubia   NR-ZERO   1,447   80   100     Ceriodaphnia dubia   NR-ZERO   1,470   80   100     Ceriodaphnia magna   NOEC   1,600   256   1,468     Daphnia magna   NOEC   1,600   80   100     Daphnia magna   NR-ZERO   1,470   80   100	2017181 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NOEC	1,300		80		ppm
sodium salt (1:2)         Pimephales promelas         NR-ZERO         214         140         145         148           sodium salt (1:2)         Pimephales promelas         NR-ZERO         906         140         147         152           sodium salt (1:2)         Pimephales promelas         NR-ZERO         160         128         131         132           sodium salt (1:2)         Pimephales promelas         NR-ZERO         24         124         136         144           sodium salt (1:2)         Pimephales promelas         NR-ZERO         474         128         131         132           sodium salt (1:2)         Sphaerium simile         LC50         2,078         94         131         136           sodium salt (1:2)         Sphaerium simile         LC50         2,956         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,956         289         280           sodium salt (1:2)         Sphaerium simile         LC50         2,950         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,960         290         280           sodium salt (1:2)         Sphaerium simile         LC50         2,190         80	2017171 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NOEC	595		40		ppm
sodium salt (1:2)         Piimephales promelas         NR-ZERO         906         140         147         152           sodium salt (1:2)         Piimephales promelas         NR-ZERO         160         128         131         132           sodium salt (1:2)         Piimephales promelas         NR-ZERO         474         128         131         132           sodium salt (1:2)         Piimephales promelas         NR-ZERO         474         128         131         132           sodium salt (1:2)         Sphaerium simile         LC10         2,078         24         128         131         132           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         280           sodium salt (1:2)         Sphaerium simile         LC50         2,280         299         293           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         269           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,186         84         34           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80	764930 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NR-ZERO	214	140	145		mg/L CaCO3
sodium salt (1:2)         Piimephales promelas         NR-ZERO         160         128         131         132           sodium salt (1:2)         Piimephales promelas         NR-ZERO         24         124         136         144           sodium salt (1:2)         Piimephales promelas         NR-ZERO         24         128         131         132           sodium salt (1:2)         Piimephales promelas         NR-ZERO         24         128         131         132           sodium salt (1:2)         Sphaerium simile         LC50         2,078         29         4           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,200         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,200         269         280           sodium salt (1:1)         Sphaerium simile         LC50         2,200         293         280           sodium salt (1:1)         Sphaerium simile         LC50         2,186         84         49           sodium salt (1:2)         Sphaerium simile         LC50         2,190         80         90           sodi	764935 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NR-ZERO	906	140	147		mg/L CaCO3
sodium salt (1:2)         Pimephales promelas         NR-ZERO         24         124         136         144           sodium salt (1:2)         Pimephales promelas         NR-ZERO         474         128         131         132           sodium salt (1:2)         Sphaerium simile         LC10         24         128         131         132           sodium salt (1:2)         Sphaerium simile         LC50         2,590         94         94           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         280           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         280           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         90           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,186         84         90           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,191         80         100           Calcium salt (1:1) <t< td=""><td>764929 Sulfuric acid sodium salt (1:2)</td><td>Pimephales promelas</td><td>NR-ZERO</td><td>160</td><td>128</td><td>131</td><td></td><td>mg/L CaCO3</td></t<>	764929 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NR-ZERO	160	128	131		mg/L CaCO3
sodium salt (1:2)         Pimephales promelas         NR-ZERO         474         128         131         132           sodium salt (1:2)         Pimephales promelas         NR-ZERO         24         128         131         136           sodium salt (1:2)         Sphaerium simile         LC50         2,078         94         94           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280         94           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280         269           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         29           sodium salt (1:1)         Sphaerium simile         LC50         2,186         84         90           sodium salt (1:1)         Sphaerium simile         LC50         2,190         93         90           calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia	764932 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NR-ZERO	24	124	136		mg/L CaCO3
sodium salt (1:2)         Pimephales promelas         NR-ZERO         24         128         131         136           sodium salt (1:2)         Sphaerium simile         LC10         1,502         94         94           sodium salt (1:2)         Sphaerium simile         LC50         2,078         94         94           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280         280           sodium salt (1:2)         Sphaerium simile         LC50         2,590         269         269           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         269           sodium salt (1:1)         Sphaerium simile         LC50         2,186         84         269           sodium salt (1:1)         Sphaerium simile         LC50         2,190         93         269           calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,186         84         4           calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         100           calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-	764934 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NR-ZERO	474	128	131		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC10         1,502         94           sodium salt (1:2)         Sphaerium simile         LC50         2,078         94           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280           sodium salt (1:2)         Sphaerium simile         LC50         2,590         269           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         269           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         93           sodium salt (1:1)         Sphaerium simile         LC50         2,190         93         90           calcium salt (1:1)         Sphaerium simile         LC50         2,190         80         90           calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         100           calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           c	764928 Sulfuric acid sodium salt (1:2)	Pimephales promelas	NR-ZERO	24	128	131		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,078         94           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280           sodium salt (1:2)         Sphaerium simile         LC50         2,590         280           sodium salt (1:2)         Sphaerium simile         LC50         2,926         269           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84           sodium salt (1:1)         Sphaerium simile         LC50         2,186         84           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         193           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,550         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,970         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia	766364 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC10	1,502		94		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,590         280           sodium salt (1:2)         Sphaerium simile         LC50         2,926         269           sodium salt (1:2)         Sphaerium simile         LC50         2,926         93           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84           sodium salt (1:1)         Sphaerium simile         LC50         2,190         80         193           sodium salt (1:1)         Ceriodaphnia dubia         LC50         2,190         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,970         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium sal	766363 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,078		94		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,926         269           sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         93           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         93           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         93           calcium salt (1:1)         Sphaerium simile         LC50         2,190         193         193           calcium salt (1:1)         Sphaerium simile         LC50         2,190         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,970         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,447	766706 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,590		280		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,200         93         4           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         84           sodium salt (1:2)         Sphaerium simile         LC50         2,186         84         84           sodium salt (1:2)         Sphaerium simile         LC50         2,190         80         193           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,650         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,940         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO	766705 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,926		269		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,186         84           sodium salt (1:2)         Sphaerium simile         LC50         2,190         193         193           sodium salt (1:2)         Sphaerium simile         LC50         2,190         193         193           sodium salt (1:1)         Sphaerium simile         LC50         2,650         90         193           calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,970         80         100           calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,940         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           calcium salt (1:1)         Daphnia magna         NC50         > 1,9	766707 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,200		93		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,190         193         49           sodium salt (1:2)         Sphaerium simile         LC50         2,650         90         193           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         2,650         90         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,970         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         NC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-Z	766703 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,186		84		mg/L CaCO3
sodium salt (1:2)         Sphaerium simile         LC50         2,650         90         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,970         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,940         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna <t< td=""><td>766704 Sulfuric acid sodium salt (1:2)</td><td>Sphaerium simile</td><td>LC50</td><td>2,190</td><td></td><td>193</td><td></td><td>mg/L CaCO3</td></t<>	766704 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,190		193		mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,970         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         NC50         1,447         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna <t< td=""><td>766708 Sulfuric acid sodium salt (1:2)</td><td>Sphaerium simile</td><td>LC50</td><td>2,650</td><td></td><td>90</td><td></td><td>mg/L CaCO3</td></t<>	766708 Sulfuric acid sodium salt (1:2)	Sphaerium simile	LC50	2,650		90		mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,910         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,940         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna <td< td=""><td>160500 Sulfuric acid, Calcium salt (1:1)</td><td>Ceriodaphnia dubia</td><td>LC50</td><td></td><td>08</td><td></td><td></td><td>mg/L CaCO3</td></td<>	160500 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	LC50		08			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         LC50         > 1,940         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna <td< td=""><td>160498 Sulfuric acid, Calcium salt (1:1)</td><td>Ceriodaphnia dubia</td><td>LC50</td><td></td><td>80</td><td></td><td></td><td>mg/L CaCO3</td></td<>	160498 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	LC50		80			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO<	160497 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	LC50		80			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Ceriodaphnia magna         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         NOEC         1,500         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100	767397 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,470	80			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         860         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,600         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100	767360 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,447	80			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,600         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100	767399 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	860	80			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         765         80         100           Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,600         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100	767463 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	765	80			mg/L CaCO3
Calcium salt (1:1)         Ceriodaphnia dubia         NR-ZERO         1,447         80         100           Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NOEC         1,600         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100	767462 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	765	80			mg/L CaCO3
Calcium salt (1:1)         Daphnia magna         LC50         > 1,970         80         100           Calcium salt (1:1)         Daphnia magna         NOEC         1,600         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         860         80         100	767317 Sulfuric acid, Calcium salt (1:1)	Ceriodaphnia dubia	NR-ZERO	1,447	80			mg/L CaCO3
Calcium salt (1:1)         Daphnia magna         NOEC         1,600         256         1,468           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         860         80         100	160499 Sulfuric acid, Calcium salt (1:1)	Daphnia magna	LC50		80			mg/L CaCO3
Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         860         80         100		Daphnia magna	NOEC	1,600	256		n	mg/L CaCO3
Calcium salt (1:1)         Daphnia magna         NR-ZERO         1,470         80         100           Calcium salt (1:1)         Daphnia magna         NR-ZERO         860         80         100	12.	Daphnia magna	NR-ZERO	1,470	80		1	mg/L CaCO3
Calcium salt (1:1) Daphnia magna NR-ZERO 860 80 100	Sulfuric acid,	Daphnia magna	NR-ZERO	1,470	80			mg/L CaCO3
	767469 Sulfuric acid, Calcium salt (1:1)	Daphnia magna	NR-ZERO	860	80			mg/L CaCO3

		Hardness valu	ies associated witl	Hardness values associated with lethal toxicity endpoints.	dpoints.			
			0.00				Hardness	
Result				Concentration				
Number	Chemical Name	Species Scientific Name	Endpoint	(mg/L)	Minimum	Mean	Maximum	Units
767484	767484 Sulfuric acid, Calcium salt (1:1)	Daphnia magna	NR-ZERO	1,470	80		100	mg/L CaCO3
759462	759462 Sulfuric acid, Calcium salt (1:1)	Daphnia magna	NR-ZERO	360		170		mg/L CaCO3
759465	759465 Sulfuric acid, Calcium salt (1:1)	Daphnia magna	NR-ZERO	360		170		mg/L CaCO3
196198	196198 Sulfuric acid, Calcium salt (1:1)	Lepomis macrochirus	LC50	2,980		10		mg/L CaCO3
160503	160503 Sulfuric acid, Calcium salt (1:1)	Pimephales promelas	LC50	> 1,970	80		100	mg/L CaCO3
160502	160502 Sulfuric acid, Calcium salt (1:1)	Pimephales promelas	LC50	> 1,970	80		100	mg/L CaCO3
160501	160501 Sulfuric acid, Calcium salt (1:1)	Pimephales promelas	LC50	> 1,970	80		100	mg/L CaCO3
767512	767512 Sulfuric acid, Calcium salt (1:1)	Pimephales promelas	NR-ZERO	1,470	80		100	mg/L CaCO3
767515	767515 Sulfuric acid, Calcium salt (1:1)	Pimephales promelas	NR-ZERO	1,470	80		100	mg/L CaCO3

1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fis	76054 Waller, D.L., S.W. Fisher,	741717
1996	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infes	76054 Waller, D.L., S.W. Fisher,	740778
1954	The Acute Toxicity of Some Common Salts of Sodium Proc. Acad. Nat. Sci. Philadelphia106:185	The Acute Toxicity of Some Comr	8037 Trama,F.B.	40507
1960	The Sensitivity of Aquatic Life to Certain Chemicals   Final Rep.No.RG-3965(C2R1), U.S.Public		5683 Academy of Natural Scie	196145
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa	76054 Waller, D.L., S.W. Fisher,	740777
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and	76054 Waller, D.L., S.W. Fisher,	740765
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa	76054 Waller, D.L., S.W. Fisher,	740761
1996	Dispersa	Prevention of Zebra Mussel Infestation and	76054 Waller, D.L., S.W. Fisher,	2153697
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and	76054 Waller, D.L., S.W. Fisher,	2153700
1996	Prog. Fis	Prevention of Zebra Mussel Infestation and Dispersa	76054 Waller, D.L., S.W. Fisher,	740732
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa	76054 Waller, D.L., S.W. Fisher,	740760
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog.	76054 Waller, D.L., S.W. Fisher,	740759
1996	station and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fis	76054 Waller, D.L., S.W. Fisher,	2153676
1996	76054 Waller, D.L., S.W. Fisher, Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infes	76054 Waller, D.L., S.W. Fisher,	2153763
1992	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pr Final Rep. GRI-92/0301, ENSR Contract No.	Development of a Salinity/Toxicit	14761 Mount, D.R., and D.D. Gu	768398
1992	ity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pu Final Rej	14761 Mount, D.R., and D.D. Gu	768460
1992	ity Relationship to PhFinal Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Py Final Rej	14761 Mount, D.R., and D.D. Gu	768402
1992	ity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Py Final Re	14761 Mount, D.R., and D.D. Gu	768462
1992	ity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Py Final Re	14761 Mount, D.R., and D.D. Gu	768457
1992	ity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No	Gu Development of a Salinity/Toxicity Relationship to Pr Final Re	14761 Mount, D.R., and D.D. Gu	768396
1992	ity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No	and D.D. Gu Development of a Salinity/Toxicity Relationship to PhFinal Re	14761 Mount, D.R., and D.D. Gu	768399
1997	Toxicity of Major Ion Environ. Toxicol. Chem.16(10): 2009-201	Statistical Models to Predict the Toxicity of Major Ion	D.D.	160621
1997	Toxicity of Major Ion Environ. Toxicol. Chem.16(10): 2009-201	Statistical Models to Predict the Toxicity of Major Ion	18272 Mount, D.R., D.D. Gulley,	160622
1992	ity Relationship to PrFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to Pr	14761 Mount, D.R., and D.D. Gu	768286
1992	Salinity/Toxicity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No.	Development of a	14761 Mount, D.R., and D.D. Gu	768181
1992	to PrFinal Rep.GRI-92/0301,	Development of a Salinity/Toxicit		768183
1992	ity Relationship to PrFinal Rep.GRI-92/0301, ENSR Contract No	Gu Development of a Salinity/Toxicity Relationship	14761 Mount, D.R., and D.D. Gu	768283
1992	Final Rep.GRI-92/0301,	Gu Development of a Salinity/Toxicity Relationship	14761 Mount, D.R., and D.D. Gu	767342
1992	ity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pr	14761 Mount, D.R., and D.D. Gu	768182
1992	ity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to PyFinal Re	14761 Mount, D.R., and D.D. Gu	768284
1992	ity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to PyFinal Re	14761 Mount, D.R., and D.D. Gu	768180
1997	Toxicity of Major Ion Environ. Toxicol. Chem.16(10): 2009-201	18272 Mount, D.R., D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ.	18272 Mount, D.R., D.D. Gulley,	160620
1997	Toxicity of Major Ion Environ. Toxicol. Chem.16(10): 2009-201	18272 Mount, D.R., D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ	18272 Mount, D.R., D.D. Gulley,	160619
Year		Title	Number Author	
Publication			Reference	Result Re
	Reference Information	Referenc		

1992	to PriFinal Rep.GRI-92/0301. ENSR Contract Nd	Gu  Development of a Salinity/Toxicity Relationship to PfFinal Rep.GRI-92/0301, ENSR Contract Nd	14761 Mount, D.R., and D.D. Gu	14761	/68456
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	and D.D. Gu	14761	768395
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	Mount, D.R., and D.D. Gu	14761	768392
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Gu Development of a Salinity/Toxicity Relationship to PyFinal Re	Mount, D.R., and D.D. Gu	14761	768390
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Gu Development of a Salinity/Toxicity Relationship to Pr Final Re	Mount, D.R., and D.D. Gu	14761	768451
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Gu Development of a Salinity/Toxicity Relationship to PyFinal Re	Mount, D.R., and D.D. Gu	14761	768501
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to PyFinal Re	Mount, D.R., and D.D. Gu	14761	768393
1992	73	Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to PyFinal Re	Mount, D.R., and D.D. Gu	14761	768454
1997	or lon Environ. Toxicol. Chem.16(10): 2009-2019	D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ.	Mount,D.R., D.D. Gulley,	18272	160515
1997	or lorEnviron. Toxicol. Chem.16(10): 2009-2019	D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ.	18272 Mount, D.R., D.D. Gulley,	18272	160516
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	Mount, D.R., and D.D. Gu	14761	768277
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Gu Development of a Salinity/Toxicity Relationship to Pr Final Rep.GRI-92/0301, ENSR Contract No	Mount, D.R., and D.D. Gu	14761	768177
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Py Final Rep.GRI-92/0301, ENSR Contract No	Mount, D.R., and D.D. Gu	14761	768176
1992	to PyFinal Rep.GRI-92/0301, ENSR Contract No	Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Py Final Rep. GRI-92/0301, ENSR Contract No	Mount, D.R., and D.D. Gu	14761	768178
1997	or lor Environ. Toxicol. Chem.16(10): 2009-201	D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ	Mount, D.R., D.D. Gulley,	18272	160514
1997	or IonEnviron. Toxicol. Chem.16(10): 2009-201	D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ	Mount, D.R., D.D. Gulley,	18272	160513
1996	persa Prog. Fish-Cult.58(2): 77-84	S.W. Fisher, Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	Waller, D.L.,	76054	740734
1996	persa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher,	76054	2153802
1996	persa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher,	76054	740744
1996	persa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher,	76054	740779
1996	persa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersa Prog. Fish-Cult.58(2): 77-84	1500	76054	740738
1996	persa Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Dispersal Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher,	76054	2153835
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Publication			Reference	Result
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1993	orated IPS Environmental and Analytical Services	Site Water Bioassays for Huff and Huff, Incorporated IPS Envi	116830 Sanders, D.F.	767896
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Reference  Number  Author  Title  Source  Public  Preserved  Author  Title  Author  Title  Author  Title  Source  Yes  Source  Yes  Source  Yes  Yes  Source  Yes  Yes  Tristi Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR Contract Nd. 14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pl Final Rep. GRI-92/0301. ENSR	2007		Acute and Chronic Toxicity of Techn	97 Bringolf,R.B., W.G. Cope	
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Author  Author  Title  Source  Yea  Pleckering,Q,H., J.M. Lazo Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pleckering,Q,H., J.M. Lazo Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pleckering,Q,H., J.M. Lazo Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pleckering,Q,H., J.M. Lazo Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pleckering,Q,H., J.M. Lazo Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pleckering,Q,H., J.M. Lazo Subchronic Sensitivity of One-, Four-, and Seven-Day Environmental and Analytical Services Subchronic Sensitivity of One-, Four-, and Seven-Day Environmental and Analytical Services Sunders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders,D.F.  Site Water Bioassays for Huff and Huff, Incorporated IPS Environmental and Analytical Services Sanders	10	IPS Environmental and Analytical Service	Site Water Bioassays for Huff and Huff, Incorporated		116830	768147
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Author  Title  Source  Yea  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environ. Toxicol. Chem.15(3): 353-359  Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environmental and Analytical Services    Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environmental and Analytical Services    Pickering, Q. H., J.M. Lazo   Subchronic Sensitivity of One-, Four-, and Seven-Day Environmental and Analytical Services    Pickering, Q. H., J.M. Lazo   Subchronic Services    Pickering, Q. H., J.M. Lazo   Subchronic Subchronic Subchronic Services    Pickering, Q. H., J.M. Lazo   Subchronic Subchronic Services    Pickering, Q. H., J.M. Lazo   Subchronic Subchronic Services    Pickering, Q. H., J.M. Lazo   Subchronic Subchronic Subchronic Services    Pickering, Q. H., J.M. Lazo   Subchronic	18	IPS Environmental and Analytical Services	Site Water Bioassays for Huff and Huff, Incorporate	0 Sanders, D.F.		768153
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2007	Environ. Toxicol. Chem.26(4): 773-779	Comparison of Hardness- and Chloride-Regulated Ad Environ.	Soucek,D.J.	96313	766560
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2007	Environ. Toxicol. Chem.26(4): 773-779	Comparison of Hardness- and Chloride-Regulated Ad Environ.	Soucek, D.J.		766574
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2007	dEnviron. Toxicol. Chem.26(4): 773-779	Comparison of Hardness- and Chloride-Regulated Ad Environ.	Soucek,D.J.	96313	766607
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2007		Comparison of Hardness- and Chloride-Regulated Ad Environ.	Soucek,D.J.	96313	766567
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Public   Author   Title   Source   Public   Public   Source   So	1992	PrFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to	Mount, D.R., and D.D.	14761	767462
Public   Author   Title   Source   Public   Public   Source   Public   Public   Author   Title   Source   Public   Source   Sou	1992	PrFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to	Mount, D.R., and D.D. Gu	14761	767463
Reference Information  Title  Author  Title  Source  Yes  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sul	1992	PrFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to	Mount, D.R., and D.D.	14761	767399
Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253	1992	PrFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to	Mount, D.R., and D.D.		767360
Public Author   Title   Source   Public   Publ	1992	PrFinal Rep.GRI-92/0301, ENSR Contract No	Development of a Salinity/Toxicity Relationship to	Mount, D.R., and D.D.		767397
Reference Information    Title	1997		Statistical Models to Predict the Toxicity of Major I	Mount, D.R.,	18272	160497
Reference Information  Title  Source  Yes  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Final Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Su	1997	7.	Statistical Models to Predict the Toxicity of Major I	Mount, D.R., D.D. Gulley,	18272	160498
Publick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253	1997	or Environ. Toxicol. Chem.16(10): 2009-2019	Statistical Models to Predict the Toxicity of Major I	Mount, D.R., D.D. Gulley,	18272	160500
Public Author   Title   Source   Public   Public   Source   Public   Publ	2005	ici Third Quarterly Report, Submitted to ∪.S	Effects of Water Quality on Acute and Chronic Tox		116831	766708
Reference Information    Public   Publi	2005	ici Third Quarterly Report, Submitted to ∪.S	Effects of Water Quality on Acute and Chronic Tox		116831	766704
Author         Title         Source         Public           Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation of Sulfate: The CEnviron. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, GAn Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute To	2005	ici Third Quarterly Report, Submitted to U.S	Effects of Water Quality on Acute and Chronic Tox		116831	766703
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Author       Title       Public         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P	2005		Effects of Hardness, Chloride, and Acclimation on		116417	766364
Reference Information         Author       Title       Source       Yei         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P         Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P         Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P	1998	of SFinal Report TR - 111878 to the Electric P	Effect of Sulfate Concentration on Acute Toxicity c	Brix, K.V.,	87256	764928
Reference Information         Author       Title       Source       Public         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric P       Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P         Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P         Brix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P	1998	of \$Final Report TR - 111878 to the Electric P	Effect of Sulfate Concentration on Acute Toxicity c	Brix, K.V., J.S. Volosin, W.	87256	764934
Reference InformationPublicAuthorTitleSourcePublicElphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation on Acute Toxicity of Sinal Report TR - 111878 to the Electric PBrix,K.V., J.S. Volosin, W. Effect of Sulfate Concentration on Acute Toxicity of Sinal Report TR - 111878 to the Electric P	1998	of SFinal Report TR - 111878 to the Electric P	Effect of Sulfate Concentration on Acute Toxicity c	Brix, K.V.,	87256	764932
Reference Information  Title  Source  Public Yea  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: T	1998	of \$Final Report TR - 111878 to the Electric P	Effect of Sulfate Concentration on Acute Toxicity c	Brix,K.V., J.S. Volosin, W.	87256	764929
Reference InformationAuthorTitleSourcePublicElphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253	1998	of \$Final Report TR - 111878 to the Electric P	Effect of Sulfate Concentration on Acute Toxicity c	Brix, K.V., J.S. Volosin, W.	87256	764935
Reference InformationPublicAuthorTitleSourcePublicElphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253	1998	port TR - 111878 to the Electric	Effect of Sulfate Concentration on Acute Toxicity of	Brix,K.V., J.S. Volosin, W.	87256	764930
Reference Information       Public         Author       Title       Source       Yea         Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253       Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate: The C Environ. Toxicol. Chem.30(1): 247-253       Environ. Toxicol. Chem.30(1): 247-253	2011		An Aquatic Toxicological Evaluation of Sulfate: The		153684	2017171
Reference Information    Publication   Publi	2011		An Aquatic Toxicological Evaluation of Sulfate: The			2017181
Reference Information  Reference Information  Public: Source Yea  Elphick,J.R., M. Davies, G An Aquatic Toxicological Evaluation of Sulfate; The C Environ. Toxicol. Chem.30(1): 247-253	2011	•	An Aquatic Toxicological Evaluation of Sulfate: The			2017212
Reference Information  Author Title Source	2011	Environ.	An Aquatic Toxicological Evaluation of Sulfate: The		153684	2017191
Reference Information	Year	Source	Title	Author	Number	Number
Reference Information	Publication				Reference	Result
			Reference Information			

			Reference Information		
Result	Reference				Publication
Number	Number	Author	Title	Source	Year
767484		Mount,D.R., and D.D. Gu	14761 Mount,D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to PyFinal Rep.GRI-92/0301, ENSR Contract No	Final Rep.GRI-92/0301, ENSR Contract No	1992
759462		Leblanc,G.A., and D.C. Su	107318 Leblanc, G.A., and D.C. Su The Influence of Mineral Salts on Fecundity of the W Hydrobiologia108:25-31	Hydrobiologia108:25-31	1984
759465		Leblanc,G.A., and D.C. Su	107318 Leblanc, G.A., and D.C. Su The Influence of Mineral Salts on Fecundity of the W Hydrobiologia108:25-31	Hydrobiologia108:25-31	1984
196198		Academy of Natural Sciel	5683 Academy of Natural Scie The Sensitivity of Aquatic Life to Certain Chemicals CFinal Rep.No.RG-3965(C2R1), U.S.Public	Final Rep.No.RG-3965(C2R1), U.S.Public H	1960
160503	1	Mount, D.R., D.D. Gulley,	18272 Mount, D.R., D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ.	Environ. Toxicol. Chem.16(10): 2009-2019	1997
160502		Mount, D.R., D.D. Gulley,	18272 Mount, D.R., D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ. Toxicol. Chem. 16(10): 2009-201	Environ. Toxicol. Chem.16(10): 2009-201	1997
160501	18272	Mount, D.R., D.D. Gulley,	18272 Mount, D.R., D.D. Gulley, Statistical Models to Predict the Toxicity of Major Ion Environ.	Environ. Toxicol. Chem.16(10): 2009-2019	1997
767512		Mount, D.R., and D.D. Gu	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pr Final Rep. GRI-92/0301, ENSR Contract No.	Final Rep.GRI-92/0301, ENSR Contract No	1992
767515		Mount, D.R., and D.D. Gu	14761 Mount, D.R., and D.D. Gu Development of a Salinity/Toxicity Relationship to Pr Final Rep. GRI-92/0301, ENSR Contract No.	Final Rep.GRI-92/0301, ENSR Contract No	1992

Result Number Chemical Name  2152811 Sodium chloride (NaCl)  2152812 Sodium chloride (NaCl)  2152813 Sodium chloride (NaCl)  2152815 Sodium chloride (NaCl)  2152814 Sodium chloride (NaCl)	Species Scientific Name Chironomus dilutus Chironomus dilutus Chironomus dilutus Chironomus dilutus	Endpoint IC10 IC25 IC50	Effect Measurement Biomass Biomass Biomass	Concentration (mg/L) 2,316 2,590 3,047	Minimum 80 80	Mean	Maximum 100	Units
2152811 Sodium chloride (NaCl) 2152812 Sodium chloride (NaCl) 2152813 Sodium chloride (NaCl) 2152815 Sodium chloride (NaCl) 2152814 Sodium chloride (NaCl)	Chironomus dilutus Chironomus dilutus Chironomus dilutus Chironomus dilutus	IC10 IC25 IC50 LOEC	Biomass Biomass	2,316 2,590 3,047	80		100	mg/L CaCO3
2152812 Sodium chloride (NaCl) 2152813 Sodium chloride (NaCl) 2152815 Sodium chloride (NaCl) 2152814 Sodium chloride (NaCl)	Chironomus dilutus Chironomus dilutus Chironomus dilutus	IC25 IC50 LOEC	Biomass Biomass	2,590 3,047	80		100	
2152813 Sodium chloride (NaCl) 2152815 Sodium chloride (NaCl) 2152814 Sodium chloride (NaCl)	Chironomus dilutus Chironomus dilutus	IC50	Biomass	3,047	80		77.77	mg/L CaCO3
2152815 Sodium chloride (NaCl) 2152814 Sodium chloride (NaCl)	Chironomus dilutus	LOEC	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1				100	mg/L CaCO3
2152814 Sodium chloride (NaCl)	Chironomie dilutus		Biomass	3,960	80		100	mg/L CaCO3
	כווווטווטוווט מוומנמט	NOEC	Biomass	2,133	80		100	mg/L CaCO3
201556 Sodium chloride (NaCl)	Daphnia pulex	LOEC	Length	441		96		mg/L CaCO3
201557 Sodium chloride (NaCl)		MATC	Length	372		96		mg/L CaCO3
201555 Sodium chloride (NaCl)		NOEC	Length	314		96		mg/L CaCO3
2152816 Sodium chloride (NaCl)	Hyalella azteca	IC25	Biomass	1,705	80		100	mg/L CaCO3
2152817 Sodium chloride (NaCl)	Hyalella azteca	IC50	Biomass	2,298	80		100	mg/L CaCO3
2152819 Sodium chloride (NaCl)	Hyalella azteca	LOEC	Biomass	4,237	80	Y	100	mg/L CaCO3
2152818 Sodium chloride (NaCl)	Hyalella azteca	NOEC	Biomass	2,210	80		100	mg/L CaCO3
2152752 Sodium chloride (NaCl)	Pimephales promelas	IC25	Biomass	704	80		100	mg/L CaCO3
2152753 Sodium chloride (NaCl)	Pimephales promelas	IC50	Biomass	958	80		100	mg/L CaCO3
768151 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768129 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	08		100	mg/L CaCO3
768092 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768111 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768141 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768161 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	5,000	90		100	mg/L CaCO3
768086 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	5,000	80		100	mg/L CaCO3
768094 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768149 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768078 Sodium chloride (NaCl)		LOEC	Biomass	5,000	80		100	mg/L CaCO3
768159 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768084 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768076 Sodium chloride (NaCl)		LOEC	Biomass	2,500	08		100	mg/L CaCO3
768139 Sodium chloride (NaCl)		LOEC	Biomass	2,500	08		100	mg/L CaCO3
2152755 Sodium chloride (NaCl)		LOEC	Biomass	1,058	80		100	mg/L CaCO3
768070 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	08		100	mg/L CaCO3
768068 Sodium chloride (NaCl)		LOEC	Biomass	2,500	08		100	mg/L CaCO3
768119 Sodium chloride (NaCl)		LOEC	Biomass	5,000	90		100	mg/L CaCO3
768109 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768102 Sodium chloride (NaCl)		LOEC	Biomass	2,250	80		100	mg/L CaCO3
768131 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768121 Sodium chloride (NaCl)		LOEC	Biomass	2,500	80		100	mg/L CaCO3

	Ha	ardness value	Hardness values associated with sublethal toxicity endpoints	I toxicity endpoi	nts.			
				Concentration			Hardness	
Result Number   Chemical Name	Species Scientific Name	Endpoint	Effect Measurement	(mg/L)	Minimum	Mean	Maximum	Units
Sodiu	Pimephales promelas	LOEC	Biomass	5,000	08		100	mg/L CaCO3
768060 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	2,500	80		100	mg/L CaCO3
768062 Sodium chloride (NaCl)	Pimephales promelas	LOEC	Biomass	10,000	80		100	mg/L CaCO3
	Pimephales promelas	LOEL	Biomass	2,200	90		110	mg/L CaCO3
741780 Sodium chloride (NaCl)	Pimephales promelas	LOEL	Biomass	1,140	90		110	mg/L CaCO3
768152 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768130 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768093 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768112 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768142 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768162 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	2,500	80		100	mg/L CaCO3
768087 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	2,500	80		100	mg/L CaCO3
768095 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768150 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768079 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	2,500	80		100	mg/L CaCO3
768160 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768085 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768077 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768140 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80			mg/L CaCO3
2152754 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	558	80		100	mg/L CaCO3
768071 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80			mg/L CaCO3
768069 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	250	80		100	mg/L CaCO3
768120 Sodium chloride (NaCl)	1.1	NOEC	Biomass	2,500	80		100	mg/L CaCO3
768110 Sodium chloride (NaCl)		NOEC	Biomass	1,250	80		100	mg/L CaCO3
768103 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768132 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
	Pimephales promelas	NOEC	Biomass	1,250	80		100	mg/L CaCO3
768101 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	2,500	80		100	mg/L CaCO3
768061 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	1,250	80			mg/L CaCO3
768063 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Biomass	5,000	80		100	mg/L CaCO3
14424 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Growth, general	4,000	86		94	mg/L CaCO3
42862 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Growth, general	4,000	86		94	mg/L CaCO3
14380 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Growth, general	4,000	86		94	mg/L CaCO3
42867 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Growth, general	4,000	86		94	mg/L CaCO3
96882 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Growth, general	4,000	86		94	mg/L CaCO3
108590 Sodium chloride (NaCl)	Pimephales promelas	NOEC	Growth, general	4,000	86		94	mg/L CaCO3

Passis   Sodium richoride   Nacio   Species Scientific Name   Endpoint   Effect Measurement   Concentration   Major   Major									Hardness	
Column chloride (NaCl)         Empelorides promeitos         MOEL         Biomass         (mg/L)         Minimum         Meximum           Isodium chloride (NaCl)         Impelpoles promeitos         MOEL         Biomass         210         90         110           Isodium chloride (NaCl)         Impelpoles promeitos         MOEL         Biomass         2,000         90         110           Isoficium chloride (NaCl)         Impelpoles promeitos         NOEL         Bormass         2,000         90         110           Isoficium chloride (NaCl)         Impelpoles promeitos         NOEL         Dory weight (AQUIRE only)         1,500         92         110           Isoficium chloride (NaCl)         Biomassi (1,12 Prinepholes promeitos         NOEL         Length         1,500         92         92           Isoficium chloride (NaCl)         Biomassi (1,12 Prinepholes promeitos         NOEL         Length         1,500         92         100           Sodium chloride (NaCl)         Brachibous colyciforus         ICCS         Bernoduction, general         1,503         80         100           Sodium chloride (NaCl)         Brachibous colyciforus         ICCS         Reproduction, general         1,120         80         92         100           Sodium chloride (NaCl)						Concentration				
Dimeshales promelas   NOEL   Blomass   210   90   110   Dimeshales promelas   NOEL   Blomass   210   90   110   110   Dimeshales promelas   NOEL   Blomass   210   90   110   110   200   210	Kesuit Number	cnemical Name	Species Scientific Name	Endpoint	Effect Measurement	(mg/L)	17.5	Mean	Maximum	Units
	741815	Sodium chloride (NaCl)	1-1	NOEL	Biomass	860			110	mg/L CaCO3
Ik (11.3Corbicula manilensis         IODE         Blomass         1,500         90         110           Ikt (1.3Corbicula manilensis         IODE         Dry weight (AQUIRE only)         1,500         92         110           Ikt (1.3Corbicula manilensis         IODE         Dry weight (AQUIRE only)         1,500         92         1           Ikt (1.3Corbicula manilensis         NOEC         Length         1,500         92         92           Ikt (1.3Corbicula manilensis         NOEC         Benass         1,500         92         92           Ikt (1.3Corbicula manilensis         NOEC         Bonass         1,500         92         100           Ikt (1.3Corbicula manilensis         NOEC         Bonass         1,500         92         100           Ikt (1.3Corbicula manilensis         NOEC         Bonass         1,301         80         100           Ikt (2.5Corbicula manilensis         NOEC         Reproduction, general         1,241         80         100           Ist (2.5Corbicula manilensis         ICEC         Reproduction, general         1,241         80         100           Ist (2.5Corbicula manilensis         ICEC         Reproduction, general         1,241         80         100           Int (2.5Corbicula ma	742694	Sodium chloride (NaCl)		NOEL	Biomass	210			110	mg/L CaCO3
It (1.3.Corbicula manilensis         LOEC         Condition index         1,500         92           It (1.3.Corbicula manilensis         NOEC         Dry weight (AQUIRE only)         1,500         92           It (1.3.Corbicula manilensis         NOEC         Dry weight (AQUIRE only)         1,500         92           It (1.3.Corbicula manilensis         NOEC         Dry weight (AQUIRE only)         1,500         92         92           It (1.3.Corbicula manilensis         NOEC         Dry weight (AQUIRE only)         1,500         80         92         100           Brachianus calveiflorus         NOEC         Reproduction, general         1,241         80         100           Brachianus calveiflorus         ICESO         Reproduction, general         1,505         80         100           Brachianus calveiflorus         ICESO         Reproduction, general         1,505         80         100           Brachianus calveiflorus         ICESO         Reproduction, general         1,245         80         100           Brachianus calveiflorus         ICESO         Progeny counts/numbers         2,230         80         100           Brachianus calveiflorus         ICESO         Progeny counts/numbers         2,250         94         102         100	741814	Sodium chloride (NaCl)		NOEL	Biomass	2,000			110	mg/L CaCO3
It (1:1/Corbicula manilensis         LOEC         Dry weight (AQUIRE only)         1,500         92           It (1:1/Corbicula manilensis         NOEC         Length         1,500         92           It (1:1/Corbicula manilensis         NOEC         Length         1,500         92           It (1:1/Pimephales promielus         NOEC         Blomass         1,301         80         92           J. Brachionus colyciflorus         IC10         Reproduction, general         1,241         80         100           J. Brachionus colyciflorus         IC50         Reproduction, general         1,241         80         100           J. Brachionus colyciflorus         IC50         Reproduction, general         1,241         80         100           J. Brachionus colyciflorus         IC50         Reproduction, general         1,243         80         100           J. Brachionus colyciflorus         IC50         Reproduction, general         1,240         80         100           J. Brachionus colyciflorus         IC50         Reproduction, general         1,241         80         100           J. Brachionus colyciflorus         IC50         Progeny counts/numbers         2,759         80         100           J. Brachionus colyciflorus         IC50 <td>765903</td> <td>Sulfuric acid sodium salt (1:</td> <td>Ü</td> <td>LOEC</td> <td>Condition index</td> <td>1,500</td> <td>1</td> <td>92</td> <td></td> <td>mg/L CaCO3</td>	765903	Sulfuric acid sodium salt (1:	Ü	LOEC	Condition index	1,500	1	92		mg/L CaCO3
alt (1.2) Corbicula manilensis         NOEC         Length         1,500         92           alt (1.2) Corbicula manilensis         NOEC         Dry weight (AQUIRE only)         1,500         92           alt (1.2) Emerbalies promelas         NOEC         Dry weight (AQUIRE only)         1,500         92         92           alt (1.2) Emerbalic promelas         NOEC         Beroduction, general         1,241         80         92         100           Brachionus calyciflorus         ICC50         Reproduction, general         1,505         80         100           Brachionus calyciflorus         ICC50         Reproduction, general         1,245         80         100           Brachionus calyciflorus         ICC50         Reproduction, general         1,250         80         100           Brachionus calyciflorus         ICC50         Reproduction, general         1,245         80         100           Brachionus calyciflorus         ICC50         Reproduction, general         1,250         80         100           Brachionus calyciflorus         ICC50         Progeny counts/numbers         2,750         80         100           Brachionus calyciflorus         ICC50         Progeny counts/numbers         1,250         94         102	765902	Sulfuric acid sodium salt (1:		LOEC	Dry weight (AQUIRE only)	1,500		92		mg/L CaCO3
It (1.3 Corbicula manilensis         NOEC         Dry weight (AQUIRE only)         1,500         92         100           It (1.3 Pimephales promelas         NOEC         Biomass         1,301         80         100           Brachionus callyciflorus         IC10         Beroduction, general         1,241         80         100           Brachionus callyciflorus         IC25         Reproduction, general         1,245         80         100           Brachionus callyciflorus         IC50         Reproduction, general         1,245         80         100           Brachionus callyciflorus         IC50         Reproduction, general         1,230         80         100           Brachionus callyciflorus         IC50         Reproduction, general         1,230         80         100           Brachionus callyciflorus         IC50         Reproduction, general         1,230         80         100           Brachionus callyciflorus         IC50         Progeny counts/numbers         2,750         80         100           Brachionus callyciflorus         IC50         Progeny counts/numbers         2,750         94         102         100           Ceriodaphnia dubia         IC25         Progeny counts/numbers         4,54         40         44 <td>765901</td> <td>Sulfuric acid sodium salt (1:</td> <td></td> <td>NOEC</td> <td>Length</td> <td>1,500</td> <td></td> <td>92</td> <td></td> <td>mg/L CaCO3</td>	765901	Sulfuric acid sodium salt (1:		NOEC	Length	1,500		92		mg/L CaCO3
alt (1.12) Pimephales promelas         NOEC         Biomass         1,301         80         100           b)         Brachionus calyciflorus         IC30         Reproduction, general         1,241         80         100           c)         Brachionus calyciflorus         IC30         Reproduction, general         1,241         80         100           b)         Brachionus calyciflorus         IC60         Reproduction, general         1,235         80         100           b)         Brachionus calyciflorus         IC60         Reproduction, general         2,750         80         100           ceriodaphnia dubia         EC50         Progeny counts/numbers         2,750         54         98         72           ceriodaphnia dubia         EC50         Progeny counts/numbers         1,350         54         72           ceriodaphnia dubia         IC25         Progeny counts/numbers         457         100         101         103           ceriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           ceriodaphnia dubia         IC25         Progeny counts/numbers         454         80         100           ceriodaphnia dubia         IC25         Reproduction, general	765900	Sulfuric acid sodium salt (1:		NOEC	Dry weight (AQUIRE only)	1,500		92		mg/L CaCO3
Brachionus calyciflorus   IC10   Reproduction, general   1,1241   80   100     Brachionus calyciflorus   IC25   Reproduction, general   1,505   80   100     Brachionus calyciflorus   IC6C   Reproduction, general   1,235   80   100     Brachionus calyciflorus   IC6C   Reproduction, general   1,120   80   100     Brachionus calyciflorus   IC6C   Reproduction, general   1,120   80   100     Ceriodaphnia dubia   IC650   Progeny counts/numbers   2,750   54   72     Ceriodaphnia dubia   IC650   Progeny counts/numbers   1,750   54   100     Ceriodaphnia dubia   IC25   Progeny counts/numbers   1,750   44   102   104     Ceriodaphnia dubia   IC25   Progeny counts/numbers   1,450   44   103     Ceriodaphnia dubia   IC25   Progeny counts/numbers   3,40   44   44     Ceriodaphnia dubia   IC25   Reproduction, general   454   80   40   40     Ceriodaphnia dubia   IC25   Reproduction, general   454   80   100     Ceriodaphnia dubia   IC25   Reproduction, general   5,21   45   40   40     Ceriodaphnia dubia   IC25   Reproduction, general   5,21   40   40     Ceriodaphnia dubia   IC25   Reproduction, general   5,21   40   40     Ceriodaphnia dubia   IC25   Reproduction, general   5,21   40   40     Ceriodaphnia dubia   IC30   Progeny counts/numbers   342   44   40     Ceriodaphnia dubia   IC30   Progeny counts/numbers   5,33   44   40     Ceriodaphnia dubia   IC30   Reproduction, general   5,21   40   40   48     Ceriodaphnia dubia   IC30   Reproduction, general   5,41   40   40   48     Ceriodaphnia dubia   IC30   Reproduction, general   5,41   40   40   48     Ceriodaphnia dubia   IC30   Reproduction, general   4,41   40   40   48     Ceriodaphnia dubia   IC30   Reproducing organisms   1,100   40   48	766857	Sulfuric acid sodium salt (1:		NOEC	Biomass	1,301	80		100	mg/L CaCO3
b)         Brachionus calyciflorus         IC25         Reproduction, general         1,505         80         100           b)         Brachionus calyciflorus         IC50         Reproduction, general         1,945         80         100           b)         Brachionus calyciflorus         IC6C         Reproduction, general         1,230         80         100           ceriodaphnia dubia         EC50         Reproduction, general         1,120         80         100           ceriodaphnia dubia         EC50         Progeny counts/numbers         2,750         94         102         104           ceriodaphnia dubia         EC50         Progeny counts/numbers         1,350         54         72           ceriodaphnia dubia         IC25         Progeny counts/numbers         1,47         100         101         103           ceriodaphnia dubia         IC25         Progeny counts/numbers         454         100         101         103           ceriodaphnia dubia         IC25         Reproduction, general         454         80         10           ceriodaphnia dubia         IC25         Reproduction, general         454         80         10           ceriodaphnia dubia         IC25         Reproduction, general	2152820	Sodium chloride (NaCl)	SI	IC10	Reproduction, general	1,241	80		100	mg/L CaCO3
b)         Brachionus calveiflorus         ICSO         Reproduction, general         1,945         80         100           b)         Brachionus calveiflorus         IOEC         Reproduction, general         2,230         80         100           ceriodaphnia dubia         ECSO         Progeny counts/numbers         2,750         80         100           ceriodaphnia dubia         ECSO         Progeny counts/numbers         2,750         94         102           ceriodaphnia dubia         ECSO         Progeny counts/numbers         1,750         94         102         104           ceriodaphnia dubia         IC25         Progeny counts/numbers         467         100         101         103           ceriodaphnia dubia         IC25         Progeny counts/numbers         467         100         101         103           ceriodaphnia dubia         IC25         Progeny counts/numbers         454         44         44           ceriodaphnia dubia         IC25         Reproduction, general         454         80         10           ceriodaphnia dubia         IC25         Reproduction, general         454         80         10           ceriodaphnia dubia         IC25         Reproduction, general         580         <	2152821	Sodium chloride (NaCl)		IC25	Reproduction, general	1,505	80		100	mg/L CaCO3
b)         Brachionus calyciflorus         LOEC         Reproduction, general         2,230         80         100           b)         Brachionus calyciflorus         NOEC         Reproduction, general         1,120         80         100           ceriodaphnia dubia         ECSO         Progeny counts/numbers         2,750         54         98         72           ceriodaphnia dubia         ECSO         Progeny counts/numbers         1,350         54         98         72           ceriodaphnia dubia         ECSO         Progeny counts/numbers         1,350         94         102         104           ceriodaphnia dubia         IC25         Progeny counts/numbers         467         100         101         103           ceriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           ceriodaphnia dubia         IC25         Progeny counts/numbers         345         80         93         44           ceriodaphnia dubia         IC25         Reproduction, general         454         80         40         100           ceriodaphnia dubia         IC25         Reproduction, general         521         320         40         40         40           ceriodaphnia dubia	2152822	Sodium chloride (NaCl)		IC50	Reproduction, general	1,945	80			mg/L CaCO3
b)         Brachionus calyciflorus         NOEC         Reproduction, general         1,120         80         100           ceriodaphnia dubia         EC50         Progeny counts/numbers         2,750         98         100           ceriodaphnia dubia         EC50         Progeny counts/numbers         1,330         54         72           ceriodaphnia dubia         EC50         Progeny counts/numbers         1,750         94         102         104           ceriodaphnia dubia         EC50         Progeny counts/numbers         1,750         94         102         104           ceriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           ceriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           ceriodaphnia dubia         IC25         Progeny counts/numbers         454         80         10           ceriodaphnia dubia         IC25         Reproduction, general         454         80         10           ceriodaphnia dubia         IC25         Reproduction, general         580         160         320           ceriodaphnia dubia         IC25         Reproduction, general         581         40         40 <tr< td=""><td>2152824</td><td>Sodium chloride (NaCl)</td><td></td><td>LOEC</td><td>Reproduction, general</td><td>2,230</td><td>80</td><td></td><td></td><td>mg/L CaCO3</td></tr<>	2152824	Sodium chloride (NaCl)		LOEC	Reproduction, general	2,230	80			mg/L CaCO3
Describing phinia dubia         EC50         Progeny counts/numbers         2,750         98           Describing phinia dubia         EC50         Progeny counts/numbers         1,350         54         72           Describing phinia dubia         EC50         Progeny counts/numbers         1,750         94         102         104           Describing phinia dubia         EC50         Progeny counts/numbers         447         100         101         103           Describing phinia dubia         IC25         Progeny counts/numbers         340         44         44           Describinia dubia         IC25         Progeny counts/numbers         340         44         44           Describinia dubia         IC25         Progeny counts/numbers         345         80         44           Describinia dubia         IC25         Reproduction, general         454         80         40         40           Describinia dubia         IC25         Reproduction, general         117         580         10         580         10         580         10         653         46         40         40         40         40         40         40         40         40         40         40         40         40         40 <td>2152823</td> <td>Sodium chloride (NaCl)</td> <td></td> <td>NOEC</td> <td>Reproduction, general</td> <td>1,120</td> <td>80</td> <td></td> <td></td> <td>mg/L CaCO3</td>	2152823	Sodium chloride (NaCl)		NOEC	Reproduction, general	1,120	80			mg/L CaCO3
Describidation         ECSD         Progeny counts/numbers         1,350         54         72           Describidaphnia dubia         ECSD         Progeny counts/numbers         1,750         94         102         104           Describidaphnia dubia         ECSD         Progeny counts/numbers         > 467         100         101         103           Decribidaphnia dubia         IC2S         Progeny counts/numbers         340         44         44           Decribidaphnia dubia         IC2S         Progeny counts/numbers         456         340         44         44           Decribidaphnia dubia         IC2S         Progeny counts/numbers         456         80         93         44           Decribidaphnia dubia         IC2S         Reproduction, general         454         80         100         44           Decribidaphnia dubia         IC2S         Reproduction, general         117         80         100         100           Decribidaphnia dubia         IC2S         Reproduction, general         580         160         40         40         40         40         40         40         40         40         40         40         40         40         40         40         40         40		Sodium chloride (NaCl)		EC50	Progeny counts/numbers	2,750		98		mg/L CaCO3
Describida phnia dubia         EC50         Progeny counts/numbers         1,750         94         102         104           Decriodaphnia dubia         EC50         Progeny counts/numbers         > 467         100         101         103           Decriodaphnia dubia         IC25         Progeny counts/numbers         147         44         44           Decriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           Decriodaphnia dubia         IC25         Progeny counts/numbers         456         80         93           Decriodaphnia dubia         IC25         Reproduction, general         454         80         100           Decriodaphnia dubia         IC25         Reproduction, general         454         80         100           Decriodaphnia dubia         IC25         Reproduction, general         580         160         100           Decriodaphnia dubia         IC25         Reproduction, general         581         40         40           Decriodaphnia dubia         IC25         Reproduction, general         521         320         40           Decriodaphnia dubia         IC50         Progeny counts/numbers         342         44         40           De	741661	Sodium chloride (NaCl)		EC50	Progeny counts/numbers	1,350	54			mg/L CaCO3
(Ecriodaphnia dubia         EC50         Progeny counts/numbers         > 467         100         101         103           (Ecriodaphnia dubia         IC25         Progeny counts/numbers         147         44         44           (Ecriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           (Ecriodaphnia dubia         IC25         Progeny counts/numbers         456         93         44           (Ecriodaphnia dubia         IC25         Reproduction, general         454         80         100           (Ecriodaphnia dubia         IC25         Reproduction, general         580         160         100           (Ecriodaphnia dubia         IC25         Reproduction, general         580         160         100           (Ecriodaphnia dubia         IC25         Reproduction, general         580         160         100           (Ecriodaphnia dubia         IC25         Reproduction, general         521         320         320           (Ecriodaphnia dubia         IC25         Reproduction, general         146         40         40           (Ecriodaphnia dubia         IC30         Progeny counts/numbers         342         44         44           (Eriodaphnia dubia	207720	Sodium chloride (NaCl)		EC50	Progeny counts/numbers	1,750	94	102		mg/L CaCO3
(Eeriodaphnia dubia         IC25         Progeny counts/numbers         147         44           (Eeriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           (Eeriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           (Eeriodaphnia dubia         IC25         Progeny counts/numbers         456         93         44           (Eeriodaphnia dubia         IC25         Reproduction, general         454         80         100           (Eeriodaphnia dubia         IC25         Reproduction, general         117         10         100           (Eeriodaphnia dubia         IC25         Reproduction, general         580         160         160           (Eeriodaphnia dubia         IC25         Reproduction, general         580         160         160           (Eeriodaphnia dubia         IC25         Reproduction, general         521         320         40           (Eeriodaphnia dubia         IC50         Progeny counts/numbers         342         40         20           (Eeriodaphnia dubia         IC50         Progeny counts/numbers         563         44         48           (Eeriodaphnia dubia         IC50         Reproducing o	741538	Sodium chloride (NaCl)		EC50	Progeny counts/numbers		100	101		mg/L CaCO3
Descriodaphnia dubia         IC25         Progeny counts/numbers         340         44         44           Descriodaphnia dubia         IC25         Progeny counts/numbers         456         93         93           Descriodaphnia dubia         IC25         Reproduction, general         454         80         93           Descriodaphnia dubia         IC25         Reproduction, general         454         80         100           Descriodaphnia dubia         IC25         Reproduction, general         117         454         80         100           Descriodaphnia dubia         IC25         Reproduction, general         580         160         40           Descriodaphnia dubia         IC25         Reproduction, general         581         40         40           Descriodaphnia dubia         IC25         Reproduction, general         146         40         40           Descriodaphnia dubia         IC25         Reproduction, general         264         20         44           Descriodaphnia dubia         IC50         Progeny counts/numbers         342         44         44           Descriodaphnia dubia         IC50         Reproducing organisms         750         40         48           Descriodaphnia dubia	2148464	Sodium chloride (NaCl)		C25	Progeny counts/numbers	147		44		ppm
Deriodaphnia dubia     IC25     Progeny counts/numbers     456     93       Decriodaphnia dubia     IC25     Reproduction, general     454     80     100       Decriodaphnia dubia     IC25     Reproduction, general     454     80     100       Decriodaphnia dubia     IC25     Reproduction, general     117     454     80     100       Decriodaphnia dubia     IC25     Reproduction, general     117     454     450     100       Decriodaphnia dubia     IC25     Reproduction, general     521     320     320     40       Decriodaphnia dubia     IC25     Reproduction, general     264     20     20       Decriodaphnia dubia     IC25     Reproduction, general     342     44     40       Decriodaphnia dubia     IC25     Reproduction, general     264     20     44       Decriodaphnia dubia     IC25     Reproduction, general     342     44     44       Decriodaphnia dubia     IC25     Progeny counts/numbers     563     44     44       Decriodaphnia dubia     IC20     Reproducing organisms     750     40     48       Decriodaphnia dubia     IC20     Reproducing organisms     1,100     40     48       Decriodaphnia dubia     IC	2148480 9	Sodium chloride (NaCl)		C25	Progeny counts/numbers	340		44		opm
Ceriodaphnia dubia         IC25         Reproduction, general         454         80         100           Ceriodaphnia dubia         IC25         Reproduction, general         454         80         100           Ceriodaphnia dubia         IC25         Reproduction, general         117         10         100           Ceriodaphnia dubia         IC25         Reproduction, general         580         160         160           Ceriodaphnia dubia         IC25         Reproduction, general         521         320         40           Ceriodaphnia dubia         IC25         Reproduction, general         264         20         40           Ceriodaphnia dubia         IC25         Reproduction, general         264         20         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         342         44         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         563         44         48           Ceriodaphnia dubia         IC50         Reproducing organisms         750         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,100         40         48           Ceriodaphnia dubia         IC50         Reproduc	2148474	Sodium chloride (NaCl)		C25	Progeny counts/numbers	456		93		opm
Ceriodaphnia dubiaIC25Reproduction, general45480100Ceriodaphnia dubiaIC25Reproduction, general1171010Ceriodaphnia dubiaIC25Reproduction, general580160160Ceriodaphnia dubiaIC25Reproduction, general52132040Ceriodaphnia dubiaIC25Reproduction, general2644040Ceriodaphnia dubiaIC50Progeny counts/numbers3424440Ceriodaphnia dubiaIC50Progeny counts/numbers5634444Ceriodaphnia dubiaIC50Progeny counts/numbers6534048Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms4048Ceriodaphnia dubiaIC50Reproducing organisms4048Ceriodaphnia dubiaIC50Reproducing organisms4048	2153272 9	Sodium chloride (NaCl)		C25	Reproduction, general	454		80		mg/L CaCO3
Ceriodaphnia dubia         IC25         Reproduction, general         117         10         160           Ceriodaphnia dubia         IC25         Reproduction, general         580         160         160           Ceriodaphnia dubia         IC25         Reproduction, general         521         320         320           Ceriodaphnia dubia         IC25         Reproduction, general         264         40         40           Ceriodaphnia dubia         IC50         Reproduction, general         264         20         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         342         44         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         563         44         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         653         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         750         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,100         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,130         40         48           Ceriodaphnia dubia         IC50         Reprod	2152721 9	Sodium chloride (NaCl)			Reproduction, general	454	80			ng/L CaCO3
Ceriodaphnia dubiaIC25Reproduction, general580160Ceriodaphnia dubiaIC25Reproduction, general521320Ceriodaphnia dubiaIC25Reproduction, general14640Ceriodaphnia dubiaIC25Reproduction, general26420Ceriodaphnia dubiaIC50Progeny counts/numbers34244Ceriodaphnia dubiaIC50Progeny counts/numbers56344Ceriodaphnia dubiaIC50Progeny counts/numbers6534048Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms4048Ceriodaphnia dubiaIC50Reproducing organisms4048Ceriodaphnia dubiaIC50Reproducing organisms4048Ceriodaphnia dubiaIC50Reproducing organisms4048	2152855 \$	sodium chloride (NaCl)			Reproduction, general	117		10		ng/L CaCO3
Ceriodaphnia dubia         IC25         Reproduction, general         521         320         40           Ceriodaphnia dubia         IC25         Reproduction, general         146         40         40           Ceriodaphnia dubia         IC25         Reproduction, general         264         20         20           Ceriodaphnia dubia         IC50         Progeny counts/numbers         342         44         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         563         44         44           Ceriodaphnia dubia         IC50         Progeny counts/numbers         653         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         750         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,100         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,130         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,130         40         48           Ceriodaphnia dubia         IC50         Reproducing organisms         1,130         40         48           Ceriodaphnia dubia         IC50         Reprod	2153275	sodium chloride (NaCl)			Reproduction, general	580		160	_	ng/L CaCO3
Ceriodaphnia dubiaIC25Reproduction, general14640Ceriodaphnia dubiaIC25Reproduction, general26420Ceriodaphnia dubiaIC50Progeny counts/numbers34244Ceriodaphnia dubiaIC50Progeny counts/numbers56344Ceriodaphnia dubiaIC50Progeny counts/numbers6534048Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048	2153278 \$	sodium chloride (NaCl)			Reproduction, general	521		320	-	ng/L CaCO3
Ceriodaphnia dubiaIC25Reproduction, general26420Ceriodaphnia dubiaIC50Progeny counts/numbers34244Ceriodaphnia dubiaIC50Progeny counts/numbers56344Ceriodaphnia dubiaIC50Progeny counts/numbers6534048Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048	2153269 \$	sodium chloride (NaCl)			Reproduction, general	146		40	r	ng/L CaCO3
Ceriodaphnia dubiaIC50Progeny counts/numbers34244Ceriodaphnia dubiaIC50Progeny counts/numbers56344Ceriodaphnia dubiaIC50Progeny counts/numbers65393Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048	2152881 S	odium chloride (NaCl)			Reproduction, general	264		20	r	ng/L CaCO3
Ceriodaphnia dubiaIC50Progeny counts/numbers56344Ceriodaphnia dubiaIC50Progeny counts/numbers65393Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048Ceriodaphnia dubiaIC50Reproducing organisms9604048	2148463 S	odium chloride (NaCl)			Progeny counts/numbers	342		44	7	mdc
Ceriodaphnia dubiaIC50Progeny counts/numbers65393Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048Ceriodaphnia dubiaIC50Reproducing organisms9204048	21484/9 5	odium chloride (NaCl)			Progeny counts/numbers	563		44	7	mqc
Ceriodaphnia dubiaIC50Reproducing organisms7504048Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048Ceriodaphnia dubiaIC50Reproducing organisms9204048	2148473 S	odium chloride (NaCl)			Progeny counts/numbers	653		93	77	mqc
Ceriodaphnia dubiaIC50Reproducing organisms1,1004048Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048Ceriodaphnia dubiaIC50Reproducing organisms9204048	741679 S	odium chloride (NaCl)			Reproducing organisms	750	40			ng/L CaCO3
Ceriodaphnia dubiaIC50Reproducing organisms6704048Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048Ceriodaphnia dubiaIC50Reproducing organisms9204048	741936 S	odium chloride (NaCl)			Reproducing organisms	1,100	40			ng/L CaCO3
Ceriodaphnia dubiaIC50Reproducing organisms1,1304048Ceriodaphnia dubiaIC50Reproducing organisms9604048Ceriodaphnia dubiaIC50Reproducing organisms9204048	741950 S				Reproducing organisms	670	40			ng/L CaCO3
Ceriodaphnia dubiaICSOReproducing organisms9604048Ceriodaphnia dubiaICSOReproducing organisms9204048	742564 S				Reproducing organisms	1,130	40			ng/L CaCO3
Ceriodaphnia dubia   IC50   Reproducing organisms   920   40   48	741951 S				Reproducing organisms	960	40			ng/L CaCO3
	/41929 5				Reproducing organisms	920	40			ng/L CaCO3

		7				_	Hardness	
				Concentration				
Result Number   Chemical Name	Species Scientific Name	Endpoint	Effect Measurement	(mg/L)	Minimum	Mean	Maximum	Units
742894 Sodium chloride (NaCl)	Ceriodaphnia dubia	IC50	Reproducing organisms	980	40		48	mg/L CaCO3
742895 Sodium chloride (NaCl)		IC50	Reproducing organisms	610	40			mg/L CaCO3
741958 Sodium chloride (NaCl)		IC50	Reproducing organisms	570	40		48	mg/L CaCO3
741948 Sodium chloride (NaCl)		IC50	Reproducing organisms	720	40		48	mg/L CaCO3
741943 Sodium chloride (NaCl)		IC50	Reproducing organisms	710	40		48	mg/L CaCO3
742899 Sodium chloride (NaCl)		IC50	Reproducing organisms	680	40		48	mg/L CaCO3
2153273 Sodium chloride (NaCl)		IC50	Reproduction, general	697		80		mg/L CaCO3
2152731 Sodium chloride (NaCl)		IC50	Reproduction, general	697	80		100	mg/L CaCO3
2152873 Sodium chloride (NaCl)	Ceriodaphnia dubia	IC50	Reproduction, general	161		10		mg/L CaCO3
2153276 Sodium chloride (NaCl)		IC50	Reproduction, general	895		160		mg/L CaCO3
2153279 Sodium chloride (NaCl)	Ceriodaphnia dubia	IC50	Reproduction, general	700		320		mg/L CaCO3
2153270 Sodium chloride (NaCl)	Ceriodaphnia dubia	IC50	Reproduction, general	481		40		mg/L CaCO3
2153267 Sodium chloride (NaCl)	Ceriodaphnia dubia	1050	Reproduction, general	301		20		mg/L CaCO3
767971 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
767883 Sodium chloride (NaCl)		LOEC	Progeny counts/numbers	2,500	80		100	mg/L CaCO3
767957 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
766860 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	2,500	08		100	mg/L CaCO3
767901 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
767860 Sodium chloride (NaCl)		LOEC	Progeny counts/numbers	2,500	08		100	mg/L CaCO3
767868 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
766868 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
767874 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
767893 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	2,500	08		100	mg/L CaCO3
767876 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
767858 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
742568 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	850	54		72	mg/L CaCO3
767852 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
767955 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	620	08		100	mg/L CaCO3
767850 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08			mg/L CaCO3
766862 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80			mg/L CaCO3
767866 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
766870 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
767891 Sodium chloride (NaCl)		LOEC	Progeny counts/numbers	620	08		100	mg/L CaCO3
767885 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	08		100	mg/L CaCO3
767963 Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
767899 Sodium chloride (NaCl)	Ceriodanhnia duhia	LOEC	Progeny counts/numbers	1,250	80			mg/L CaCO3

Result Number   Chemical Name   Species Scientific Name   Endpoint   Effect Messurement   Concentration   Masin   Marin   Marin   Marin   Marin   Species Scientific Name   Species Scientific Name   Concotaghnio dubic   Cerodaghnio dubic   Cerod								H	Hardness	
Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Progeny counts/numbers         1,250         actual control           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Progeny counts/numbers         2,500         80         100           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         1,000         40         43           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         1,000         40         43           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         500         40         43           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         1,000         40         43           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         1,000         40         48           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         1,000         40         48           Sodium chloride (NaCl)         Ceriodaphnia dabia         LOEC         Reproducing organisms         500         40         48           Sodium chloride (NaCl)         Ceriodaphnia dabia <th></th> <th></th> <th></th> <th></th> <th>166</th> <th>Concentration</th> <th>Minim</th> <th>No.</th> <th>Maximum</th> <th>libite</th>					166	Concentration	Minim	No.	Maximum	libite
Ceriodaphnia dubia	אפשור ואמוווספו	Coding chloride (NaCl)	Coriodanhaia dubia	LUCEC	Drogopy counts/pumbors	1 250	N&		100	ma/I (3003
Ceriodaphnia dubia         LOEC         Progeny counts/numbers         1,000         80         100           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Repr	/6/965	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,250	80		100	mg/L Cacus
Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reprod	767973	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	2,500	80		100	mg/L CaCO3
Ceriodaphnia dubia         LDEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LDEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LDEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LDEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LDEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LDEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NDEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NDEC         Progeny counts/numbers         500         40         48           Ceriodaphnia dubia         NDEC         Progeny	741947	Sodium chloride (NaCl)	Ceriodaphnia dubia	DEC	Reproducing organisms	1,000	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         520         80         40           Ceriodaphnia dubia         NOEC         Progeny cou	741938	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	1,000	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         2,160         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         2,20         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         6,20         80         100           Ceriodaphnia dubia         NOEC <td< td=""><td>741952</td><td>Sodium chloride (NaCl)</td><td>Ceriodaphnia dubia</td><td>LOEC</td><td>Reproducing organisms</td><td>250</td><td>40</td><td></td><td>48</td><td>mg/L CaCO3</td></td<>	741952	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	250	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         5,00         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reprogeny counts/numbers         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         520         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         520         80         100           Ceriodaphnia dubia         NOEC         Prog	742900	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	500	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         2,160         80         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         500         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOE	741944	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	500	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Proge	742892	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	1,000	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Pro	741957	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	1,000	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         1,000         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         520         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progen	741949	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	1,000	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         2,160         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         P	742563	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	1,000	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEL         Progeny counts/numbers         2,160         40         48           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC <td< td=""><td>742565</td><td>Sodium chloride (NaCl)</td><td>Ceriodaphnia dubia</td><td>LOEC</td><td>Reproducing organisms</td><td>500</td><td>40</td><td></td><td>48</td><td>mg/L CaCO3</td></td<>	742565	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	500	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         LOEL         Progeny counts/numbers         2,160         80         98           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC	741945	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	250	40		48	mg/L CaCO3
Ceriodaphnia dubia         LOEL         Progeny counts/numbers         2,160         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC	741959	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEC	Reproducing organisms	500	40		48	mg/L CaCO3
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Ceriodaphnia dubia         NOEC	2152727	Sodium chloride (NaCl)	Ceriodaphnia dubia	LOEL	Progeny counts/numbers	2,160		98		mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphn	767972	Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodap	767884	Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia	767958	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphn	766861	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphn	767902	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NACI)         Ceriodaphnia	767861	Sodium chloride (NaCl)	Ceriodaphnia dubia	NOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia	767869	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         620         80         100           Sodium chloride (NaCl)         Ceriodaphn	766869	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers1,25080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers440545472Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers13080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080 <td>767875</td> <td>Sodium chloride (NaCl)</td> <td></td> <td>NOEC</td> <td>Progeny counts/numbers</td> <td>620</td> <td>80</td> <td></td> <td>100</td> <td>mg/L CaCO3</td>	767875	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers4405472Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100	767894	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	1,250	80		100	mg/L CaCO3
Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers4405472Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers13080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100	767877	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers4405472Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers13080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100	767859	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80			mg/L CaCO3
Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers13080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers31080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers31080100	741688	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	440	54			mg/L CaCO3
Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     130     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     310     80     100	767853	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80			mg/L CaCO3
Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     310     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100	767956	Sodium chloride (NaCl)	ii	NOEC	Progeny counts/numbers	130	80			mg/L CaCO3
Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     310     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100	767851	sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     310     80     100       Sodium chloride (NaCl)     Ceriodaphnia dubia     NOEC     Progeny counts/numbers     620     80     100	766863	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		100	mg/L CaCO3
Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers31080100Sodium chloride (NaCl)Ceriodaphnia dubiaNOECProgeny counts/numbers62080100	767867	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80			mg/L CaCO3
Ceriodaphnia dubiaNOECProgeny counts/numbers31080100Ceriodaphnia dubiaNOECProgeny counts/numbers62080100	766871	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80		Ī	mg/L CaCO3
Ceriodaphnia dubia NOEC Progeny counts/numbers 620 80 100	767892	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	310	80			mg/L CaCO3
	767886	Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	620	80			mg/L CaCO3

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

Ceriodaphnia dubia   NOEC   Progeny counts/numbers   1,250   200		1,000	recundity			7167762 6-4:	2767762
						Sodium chloride (NaCl)	2164761
		500	Fecundity		1	2164/64 Sodium chloride (NaCl)	2164/64
Ceriodaphnia dubia   NOEC   Progeny counts/numbers   1,250   20   100   100     Ceriodaphnia dubia   NOEC   Reproducing organisms   250   40   48     Ceriodaphnia dubia   NOEC   Reproducing organisms   250   40   48     Ceriodaphnia dubia   NOEC   Reproducing organisms   250   40   48     Ceriodaphnia dubia   NOEC   Reproducing organisms   500   54   54   72     Daphnia magna   EC50   Progeny counts/numbers   467   100   101   103     Daphnia magna   IOEC   Progeny counts/numbers   4250   54   72     Daphnia magna   IOEC   Reproduction, general   1,000   150   150     Daphnia magna   IOEC   Fecundity   2,000   150   150     Daphnia magn		2,160	Progeny counts/numbers			2152715 Sodium chloride (NaCl)	2152715
Ceriodaphnia dubia   NOEC   Progeny counts/numbers   1,250   80   100	100		Reproduction, general			Sodium chloride (NaCl)	2152735
Ceriodaphnia dubia   NOEC   Reproducing organisms   S00   40   48		1,000	Fecundity	_OEC		2164728 Sodium chloride (NaCl)	2164728
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         0.20         0.00         1.00           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         1,250         80         100           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           2         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           3         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           4         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           2         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           3         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           4         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           4         Ceriodaphnia dubia         NOEC		2,000	Fecundity	_OEC		2164770 Sodium chloride (NaCl)	2164770
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           1)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           2)         Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           3)         Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           4         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           2         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           3)         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           4         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           5         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           6         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           6         Ceriodaphnia dubia         NOEL		2,000	Fecundity	OEC		2164769 Sodium chloride (NaCl)	2164769
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducin		2,000	Fecundity	LOEC	Ī	2164768 Sodium chloride (NaCl)	2164768
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         1,00           1)         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           1)         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           2         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           3         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           4         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           4         Ceriodaphnia dubia         NOEC		1,000	Fecundity	LOEC		2164767 Sodium chloride (NaCl)	2164767
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         60         1,00           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         60         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC		2,000	Fecundity	LOEC		2164766 Sodium chloride (NaCl)	2164766
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         60         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing	100		Reproduction, general	C50		2152733 Sodium chloride (NaCl)	2152733
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducin	100		Reproduction, general	IC25		2152732 Sodium chloride (NaCl)	2152732
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing		4,250	Progeny counts/numbers	EC50		Sodium chloride (NaCl)	2152713
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing or	169 172	467	Progeny counts/numbers	EC50		741530 Sodium chloride (NaCl)	741530
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing	72		Progeny counts/numbers	NOEC		741659 Sodium chloride (NaCl)	741659
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         40         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing or	72		Progeny counts/numbers	LOEC		741696 Sodium chloride (NaCl)	741696
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           100         Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           100         Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           100         Aug         NOEC         Reproducing organisms         250         40         48           101         Aug         Aug         48	72		Progeny counts/numbers	EC50		741654 Sodium chloride (NaCl)	741654
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250	101 103		Progeny counts/numbers	NOEL		741566 Sodium chloride (NaCl)	741566
Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Progeny counts/numbers         1,250         80         100           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         250         40         48           Ceriodaphnia dubia         NOEC         Reproducing organisms         500         40         48           48         48         48         48		1,296	Progeny counts/numbers	NOEL		2152726 Sodium chloride (NaCl)	2152726
Ceriodaphnia dubia       NOEC       Progeny counts/numbers       1,250       80       100         Ceriodaphnia dubia       NOEC       Progeny counts/numbers       1,250       80       100         Ceriodaphnia dubia       NOEC       Reproducing organisms       500       40       48         Ceriodaphnia dubia       NOEC       Reproducing organisms       250       40       48         Ceriodaphnia dubia       NOEC       Reproducing organisms       500       40       48         Ceriodaphnia dubia	48	250	Reproducing organisms	NOEC		742897 Sodium chloride (NaCl)	742897
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms <td< td=""><td>48</td><td></td><td>Reproducing organisms</td><td>NOEC</td><td></td><td>741942 Sodium chloride (NaCl)</td><td>741942</td></td<>	48		Reproducing organisms	NOEC		741942 Sodium chloride (NaCl)	741942
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48	48	250	Reproducing organisms	NOEC		741946 Sodium chloride (NaCl)	741946
Ceriodaphnia dubia       NOEC       Progeny counts/numbers       1,250       80       100         Ceriodaphnia dubia       NOEC       Progeny counts/numbers       1,250       80       100         Ceriodaphnia dubia       NOEC       Reproducing organisms       500       40       48         Ceriodaphnia dubia       NOEC       Reproducing organisms       250       40       48         Ceriodaphnia dubia       NOEC       Reproducing organisms       500       40       48	48		Reproducing organisms	NOEC		741649 Sodium chloride (NaCl)	741649
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48	48		Reproducing organisms	NOEC		741648 Sodium chloride (NaCl)	741648
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48	48		Reproducing organisms	NOEC		742901 Sodium chloride (NaCl)	742901
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48	48		Reproducing organisms	NOEC		742893 Sodium chloride (NaCl)	742893
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48	48		Reproducing organisms	NOEC		742891 Sodium chloride (NaCl)	742891
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48	48		Reproducing organisms	NOEC		741937 Sodium chloride (NaCl)	741937
Ceriodaphnia dubia     NOEC     Progeny counts/numbers     1,250     80     100       Ceriodaphnia dubia     NOEC     Reproducing organisms     500     40     48       Ceriodaphnia dubia     NOEC     Reproducing organisms     250     40     48	48		Reproducing organisms	NOEC		742898 Sodium chloride (NaCl)	742898
Ceriodaphnia dubia NOEC Progeny counts/numbers 1,250 80 100 100 Ceriodaphnia dubia NOEC Reproducing organisms 500 40 48	48		Reproducing organisms	NOEC		741953 Sodium chloride (NaCl)	741953
Ceriodaphnia dubia NOEC Progeny counts/numbers 1,250 80 100	48		Reproducing organisms	NOEC		742896 Sodium chloride (NaCl)	742896
ברייסמקליייים ממנים וייסרר וויספרול בסמורא/וומוויסבוז סכס סס דוס	100		Progeny counts/numbers	NOEC		767974 Sodium chloride (NaCl)	767974
Ceriodanhnia duhia NOEC Broggeny counts/numbers 630 80	100	620 8	Progeny counts/numbers	NOEC	Ceriodaphnia dubia	767966 Sodium chloride (NaCl)	767966
Ceriodaphnia dubia NOEC Progeny counts/numbers 620 80 100	100		Progeny counts/numbers	NOEC		767900 Sodium chloride (NaCl)	767900
Ceriodaphnia dubia NOEC Progeny counts/numbers	100	0	Progeny counts/numbers	NOEC	Ceriodaphnia dubia	767964 Sodium chloride (NaCl)	767964
Species Scientific Name   Endpoint   Effect Measurement   (mg/L)   Minimum   Mean   Ma	Mean	ntration g/L)	Effect Measurement	Endpoint	Species Scientific Name	Chemical Name	Result Number
Hardness	Hardness						

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

		na	i diless value	naturies values associated with subjectial toxicity	rovicità eliaboliica	ē		Hardness	
Result Number	Chemical Name	Species Scientific Name	Endpoint	Effect Measurement	Concentration	Minimum	Mean	Maximum	Units
2164765	2164765 Sodium chloride (NaCl)		NOEC	Fecundity	1,000		150		maa
2164762	2164762 Sodium chloride (NaCl)		NOEC	Fecundity	1,000		150		mdd
2164729	2164729 Sodium chloride (NaCl)		NOEC	Fecundity	500		150		ppm
2152734	2152734 Sodium chloride (NaCl)		NOEC	Reproduction, general	< 506	80		100	mg/L CaCO3
741546	Sodium chloride (NaCl)		NOEL	Progeny counts/numbers	467	166	169	172	mg/L CaCO3
2152714	2152714 Sodium chloride (NaCl)		NOEL	Progeny counts/numbers	1,296		170		mg/L CaCO3
201548	201548 Sodium chloride (NaCl)		LOEC	Fecundity	441		96		mg/L CaCO3
201551	201551 Sodium chloride (NaCl)		LOEC	Progeny counts/numbers	441		96		mg/L CaCO3
201549	201549 Sodium chloride (NaCl)		MATC	Fecundity	372		96		mg/L CaCO3
201552	201552 Sodium chloride (NaCl)		MATC	Progeny counts/numbers	372		96		mg/L CaCO3
201547	201547 Sodium chloride (NaCl)		NOEC	Fecundity	314		96		mg/L CaCO3
201550	201550 Sodium chloride (NaCl)		NOEC	Progeny counts/numbers	314		96		mg/L CaCO3
2152783	2152783 Sodium chloride (NaCl)	Lumbriculus variegatus	IC25	Reproduction, general	825	80		100	mg/L CaCO3
2152784	2152784 Sodium chloride (NaCl)		IC50	Reproduction, general	1,366	80		100	mg/L CaCO3
2152786	2152786 Sodium chloride (NaCl)		LOEC	Reproduction, general	366	80		100	mg/L CaCO3
2152785	2152785 Sodium chloride (NaCl)	Lumbriculus variegatus	NOEC	Reproduction, general	< 366	80		100	mg/L CaCO3
2152796	2152796 Sodium chloride (NaCl)	Tubifex tubifex	IC10	Reproduction, general	519	80		100	mg/L CaCO3
2152807	Sodium chloride (NaCl)	Tubifex tubifex	IC25	Reproduction, general	606	08			mg/L CaCO3
2152808	Sodium chloride (NaCl)		IC50	Reproduction, general	752	08		100	mg/L CaCO3
2152810	Sodium chloride (NaCl)	Tubifex tubifex	LOEC	Reproduction, general	964	80		100	mg/L CaCO3
2152809	2152809 Sodium chloride (NaCl)	Tubifex tubifex	NOEC	Reproduction, general	462	08		100	mg/L CaCO3
759472	759472 Sulfuric acid magnesium salt Daphnia magna	3	NOEC	Progeny counts/numbers	360		170		mg/L CaCO3
759471	Sulfuric acid magnesium salt Daphnia magna		NOEC	Progeny counts/numbers	360		170		mg/L CaCO3
2017036	Sulfuric acid sodium salt (1:2 Ceriodaphnia dubia	ibia	EC10	Progeny counts/numbers	137		40		mdd
2017047	2017047 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		EC10	Progeny counts/numbers	622		80		mg/L CaCO3
2017055	2017055 Sulfuric acid sodium salt (1:4 Ceriodaphnia dubia		EC10	Progeny counts/numbers	1,174		160		mdd
2017064	2017064 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		EC10	Progeny counts/numbers	420		320		ppm
2017035	2017035 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		EC25	Progeny counts/numbers	246		40		ppm
2017046	Sulfuric acid sodium salt (1:2		EC25	Progeny counts/numbers	855		80		ppm
2017054	2017054 Sulfuric acid sodium salt (1:2	Ceriodaphnia dubia	EC25	Progeny counts/numbers	1,212		160		ppm
2017063	2017063 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		EC25	Progeny counts/numbers	542		320		mdd
2017034	2017034 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		EC50	Progeny counts/numbers	465		40		ppm
766229	766229 Sulfuric acid sodium salt (1:2 Ceriodaphnia dubia		EC50	Progeny counts/numbers	1,458		92		mg/L CaCO3
766234	766234 Sulfuric acid sodium salt (1:2		EC50	Progeny counts/numbers	1,148		92		mg/L CaCO3
2017053	2017053 Sulfuric acid sodium salt (1:2	Ceriodaphnia dubia	EC50	Progeny counts/numbers	1,257		160		mdd
2017062	2017062 Sulfuric acid sodium salt (1:4 Ceriodaphnia dubia		EC50	Progeny counts/numbers	843		320		mdd

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

							_	Hardness	
Result Number	Chemical Name	Species Scientific Name	Endpoint	Effect Measurement	Concentration (mg/L)	Minimum	Mean	Maximum	Units
2017045	2017045 Sulfuric acid sodium salt (1:2 Ceriodaphnia dubia		EC50	Progeny counts/numbers	1,129		80		ppm
2148468	2148468 Sulfuric acid sodium salt (1:	Ceriodaphnia dubia	IC25	Progeny counts/numbers	625		44		ppm
2148476	Sulfuric acid sodium salt (1:	Ceriodaphnia dubia	IC25	Progeny counts/numbers	496		44		mdd
2148470	2148470 Sulfuric acid sodium salt (1:	Ceriodaphnia dubia	IC25	Progeny counts/numbers	1,060		93		ppm
2148475	2148475 Sulfuric acid sodium salt (1:2 <i>Ceriodaphnia dubia</i>		IC50	Progeny counts/numbers	715		44		mdd
2148469	Sulfuric acid sodium salt (1:		IC50	Progeny counts/numbers	1,252		93		mdd
2148467	Sulfuric acid sodium salt (1:	Ceriodaphnia dubia	IC50	Progeny counts/numbers	766		44		mdd
2017032	2017032 Sulfuric acid sodium salt (1:4 <i>Ceriodaphnia dubia</i>	1	LOEC	Progeny counts/numbers	150		40		mdd
2017043	2017043 Sulfuric acid sodium salt (1:2 <i>Ceriodaphnia dubia</i>		LOEC	Progeny counts/numbers	1,250		80		ppm
766734	766734 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		LOEC	Progeny counts/numbers	934	80		100	mg/L CaCO3
766737 5	766737 Sulfuric acid sodium salt (1:2 <i>Ceriodaphnia dubia</i>		LOEC	Progeny counts/numbers	1,195	80		100	mg/L CaCO3
766354	766354 Sulfuric acid sodium salt (1:	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,000		92		mg/L CaCO3
766344	766344 Sulfuric acid sodium salt (1::	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,000		92		mg/L CaCO3
766355	766355 Sulfuric acid sodium salt (1:7 <i>Ceriodaphnia dubia</i>		LOEC	Progeny counts/numbers	1,000		92		mg/L CaCO3
766230 5	Sulfuric acid sodium salt (1:2 Ceriodaphnia dubia		LOEC	Progeny counts/numbers	1,236		92		mg/L CaCO3
766351 S	Sulfuric acid sodium salt (1:		LOEC	Progeny counts/numbers	1,000		92		mg/L CaCO3
766356	Sulfuric acid sodium salt (1:2	Ceriodaphnia dubia	LOEC	Progeny counts/numbers	1,000		92		mg/L CaCO3
766232 S	Sulfuric acid sodium salt (1:4 Ceriodaphnia dubia	1	LOEC	Progeny counts/numbers	899		92		mg/L CaCO3
2017051 5	2017051 Sulfuric acid sodium salt (1:3 <i>Ceriodaphnia dubia</i>	-	LOEC	Progeny counts/numbers	1,300		160		ppm
2017060 5	2017060 Sulfuric acid sodium salt (1:4 <i>Ceriodaphnia dubia</i>		LOEC	Progeny counts/numbers	480		320		ppm
2017033 5	2017033 Sulfuric acid sodium salt (1:4 Ceriodaphnia dubia		NOEC	Progeny counts/numbers	< 150		40		ppm
2017044 S	2017044 Sulfuric acid sodium salt (1:‡ <i>Ceriodaphnia dubia</i>		NOEC	Progeny counts/numbers	645		80		ppm
766735 S	766735 Sulfuric acid sodium salt (1:4 Ceriodaphnia dubia		NOEC	Progeny counts/numbers	780	80		100	mg/L CaCO3
766738 S	766738 Sulfuric acid sodium salt (1:4 Ceriodaphnia dubia	P= 1	NOEC	Progeny counts/numbers	906	80		100	mg/L CaCO3
766855 S	Sulfuric acid sodium salt (1:2		NOEC	Progeny counts/numbers	1,301	80		100	mg/L CaCO3
766231 S	766231 Sulfuric acid sodium salt (1:7 Ceriodaphnia dubia	-2	NOEC	Progeny counts/numbers	618		92		mg/L CaCO3
766233 S	766233 Sulfuric acid sodium salt (1:2 Ceriodaphnia dubia		NOEC	Progeny counts/numbers	450		92		mg/L CaCO3
2017052 S	2017052 Sulfuric acid sodium salt (1:2 <i>Ceriodaphnia dubia</i>			Progeny counts/numbers	775		160		mdd
2017061 S	2017061 Sulfuric acid sodium salt (1:3 Ceriodaphnia dubia		NOEC	Progeny counts/numbers	420		320		mdd
2017147 S	2017147 Sulfuric acid sodium salt (1:4Hyalella azteca		EC10	Progeny counts/numbers	380		80		ppm
2017148 S	2017148 Sulfuric acid sodium salt (1:2 Hyalella azteca		EC25	Progeny counts/numbers	1,056		80		ppm
2017149 S	2017149 Sulfuric acid sodium salt (1:2 Hyalella azteca		EC50	Progeny counts/numbers	> 2,412		80		ppm
2017151 S	2017151 Sulfuric acid sodium salt (1:3 Hyalella azteca		LOEC	Progeny counts/numbers	2,412		80		ppm
2017150 S	2017150 Sulfuric acid sodium salt (1:7 Hyalella azteca		NOEC	Progeny counts/numbers	1,637		80		ppm
759464 S	759464 Sulfuric acid, Calcium salt (1 Daphnia magna		NOEC	Progeny counts/numbers	360		170		mg/L CaCO3
759022 S	759022 Sulfuric acid, Calcium salt (1 Daphnia magna		NOEC	Progeny counts/numbers	1,600	256		1,468	mg/L CaCO3

Supplemental Information in Support of a Use Attainability Analysis for Dissolved Minerals in the Red River in Arkansas Prepared by FTN Associates, Ltd. for Southwestern Electric Power Company, September 27, 2016

		На	rdness value	Hardness values associated with sublethal toxicity	toxicity endpoints	ts.			
							#	Hardness	
					Concentration				
<b>Result Number</b>	Chemical Name	Species Scientific Name Endpoint	Endpoint	Effect Measurement	(mg/L)	Minimum Mean Maximum	Mean	Maximum	Units
759463	759463 Sulfuric acid, Calcium salt (1 <i>Daphnia magna</i>	Daphnia magna	NOEC	Progeny counts/numbers	360		170		mg/L CaCO3
759484	759484 Sulfuric acid, Calcium salt (1 Daphnia magna	Daphnia magna	NOEC	Progeny counts/numbers	1,600	256		1,468	mg/L CaCO3

			Reference Information		
	Reference				Publication
Result Number	Number	Author	Title	Source	Year
2152811	158449	Elphick,J.R.F., K.D. Bergh, a	Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol	Environ. Toxicol. Chem.30(1): 239-246	2011
2152812	158449	Elphick,J.R.F., K.D. Bergh, a	158449 Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol		2011
2152813	158449	Elphick,J.R.F., K.D. Bergh, a	158449 Elphick,J.R.F., K.D. Bergh, ad Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol	Environ. Toxicol. Chem.30(1): 239-246	2011
2152815	158449	Elphick,J.R.F., K.D. Bergh, a	Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S		2011
2152814		Elphick,J.R.F., K.D. Bergh, a	Elphick, J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S	Environ. Toxicol.	2011
201556	45826	Birge, W.J., J.A. Black, A.G. V	Birge, W.J., J.A. Black, A.G. VRecommendations on Numerical Values for University of Kentucky, Lexington, KY:73	University of Kentucky, Lexington, KY:73 p.	1985
201557	45826	Birge, W.J., J.A. Black, A.G. \	45826 Birge, W.J., J.A. Black, A.G. √Recommendations on Numerical Values for University of Kentucky, Lexington, KY:73 p	University of Kentucky, Lexington, KY:73 p.	1985
201555	45826	Birge, W.J., J.A. Black, A.G. V	Birge, W.J., J.A. Black, A.G. VRecommendations on Numerical Values for University of Kentucky, Lexington, KY:73 p	University of Kentucky, Lexington, KY:73 p.	1985
2152816	158449	Elphick,J.R.F., K.D. Bergh, a	Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol.	Environ. Toxicol. Chem.30(1): 239-246	2011
2152817	158449	Elphick,J.R.F., K.D. Bergh, a	Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S	Environ. Toxicol. Chem.30(1): 239-246	2011
2152819	158449	Elphick,J.R.F., K.D. Bergh, a	158449 Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol		2011
2152818	158449	Elphick,J.R.F., K.D. Bergh, a	158449 Elphick,J.R.F., K.D. Bergh, ar Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol		2011
2152752	158449	Elphick,J.R.F., K.D. Bergh, ar	Elphick,J.R.F., K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol	Environ. Toxicol. Chem.30(1): 239-246	2011
2152753	158449	Elphick,J.R.F., K.D. Bergh, ar	K.D. Bergh, an Chronic Toxicity of Chloride to Freshwater S Environ. Toxicol	Environ. Toxicol. Chem.30(1): 239-246	2011
768151	116830		Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
768129	116830	Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
768092	116830	116830 Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
768111	116830	116830 Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
768141	116830	Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
768161	116830	Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco	Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	1993
768086	116830	116830 Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
768094	116830	116830 Sanders, D.F.	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	IPS Environmental and Analytical Services,	1993
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2017061	153684	G.	G An Aquatic Toxicological Evaluation of Sulfat	Environ. Toxicol.	2011
2017147	153684	Elphick,J.R., M. Davies, G.	GAn Aquatic Toxicological Evaluation of Sulfal Environ.	Environ. Toxicol.	2011
2017148	153684	Elphick,J.R., M. Davies, G. G	153684 Elphick, J.R., M. Davies, G. GAn Aquatic Toxicological Evaluation of Sulfatenviron. Toxicol.		2011
2017149	153684	Elphick,J.R., M. Davies, G. G	153684 Elphick, J.R., M. Davies, G. GAn Aquatic Toxicological Evaluation of Sulfatenviron. Toxicol.	Environ. Toxicol. Chem.30(1): 247-253	2011
2017151	153684	Elphick, J.R., M. Davies, G. G	Elphick, J.R., M. Davies, G. G An Aquatic Toxicological Evaluation of Sulfa Environ. Toxicol.	Environ. Toxicol. Chem.30(1): 247-253	2011
2017150	153684	Elphick,J.R., M. Davies, G. G	An Aquatic Toxicological Evaluation of Sulfat Environ. Toxicol.		2011
759464	107318	Leblanc, G.A., and D.C. Surp	The Influence of Mineral Salts on Fecundity	Hydrobiologia108:25-31	1984
759022	115903	115903 E.G. and G. Bionomics	The Chronic Toxicity of Sulfate to the Water Res. Rep. No. BN-7	Res.Rep.No.BN-79-10-546, Submitted to St	1979

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1984	Hydrobiologia108:25-31	107318 Leblanc, G.A., and D.C. Surp The Influence of Mineral Salts on Fecundity Hydrobiologia 108:25-31	Leblanc, G.A., and D.C. Surp	107318	759484
1984	Hydrobiologia108:25-31	107318 Leblanc, G.A., and D.C. Surp The Influence of Mineral Salts on Fecundity Hydrobiologia108:25-31	Leblanc,G.A., and D.C. Surp		759463
Year	Source	Title	Author	Number	Result Number
Publication				Reference	
		Reference Information			

0		The second second	7	Title Charles Tart Carrier		l'annonie management	100000 / Calcium chicacho /Cacion	JUNISCHULL	136063
	Fresh water Lab	Unmeasured	Static	rish; Standard Test Species	Bluegill	repomis macrochirus	Calciniti ciliotide (cacis)	7 47CC+00T	750077
5			Static				10043524 Calcium chloride (CaCl2)	10043524 C	126057
Ď	Fresh water Lab		Static		I Catfish	Ictalurus punctatus	10043524 Calcium chloride (CaCl2)	10043524 C	740777
Ď	Fresh water Lab	Unmeasured	Static	Fish; Standard Test Species	Channel Catfish F	Ictalurus punctatus	Calcium chloride (CaCl2)	10043524 C	740765
Ď	Fresh water Lab	Unmeasured	Static	Western Mosqu Fish; U.S. Exotic/Nuisance Species	Western Mosqu F	Gambusia affinis	10043524 Calcium chloride (CaCl2)	10043524 C	2063735
b	Fresh water Lab	Unmeasured	Static	Western Mosqu Fish; U.S. Exotic/Nuisance Species	Western Mosqu F	Gambusia affinis	10043524 Calcium chloride (CaCl2)	10043524 C	2063726
ιb	Fresh water Lab	Unmeasured	Static	Western Mosqu Fish; U.S. Exotic/Nuisance Species	Western Mosqu F	Gambusia affinis	Calcium chloride (CaCl2)	10043524 C	2063733
b		Unmeasured	Static	Molluscs; U.S. Exotic/Nuisance Speci-Static		Dreissena polymorpha	10043524 Calcium chloride (CaCl2)	10043524 C	740761
Ь	Fresh water Lab	Unmeasured	Static	Molluscs; U.S. Exotic/Nuisance Speci-	Zebra Mussel N	Dreissena polymorpha	Calcium chloride (CaCl2)	10043524 C	2153697
ď			Static	Molluscs; U.S. Exotic/Nuisance Speci Static		Dreissena polymorpha	Calcium chloride (CaCl2)	10043524 C	2153700
б	Fresh water Lab		Static	Molluscs; U.S. Exotic/Nuisance Speci	$\Xi$	Dreissena polymorpha	10043524 Calcium chloride (CaCl2)	10043524 C	740732
d	Fresh water Lab	Unmeasured	Static	Molluscs; U.S. Exotic/Nuisance Speci Static	Zebra Mussel N	Dreissena polymorpha	Calcium chloride (CaCl2)	10043524 C	740760
Б		Unmeasured	Static	Molluscs; U.S. Exotic/Nuisance Speci Static		Dreissena polymorpha	Calcium chloride (CaCl2)	10043524 C	740759
Ď		Unmeasured	Static	Molluscs; U.S. Exotic/Nuisance Speci		Dreissena polymorpha	10043524 Calcium chloride (CaCl2)	10043524 C	2153676
ğ		Unmeasured	Static		Zebra Mussel N	Dreissena polymorpha	Calcium chloride (CaCl2)	10043524 C	2153763
ð	Fresh water Lab	Measured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)	10043524 C	768398
ğ	Fresh water Lab	Measured	Static	Crustaceans; Standard Test Species	Water Flea C	Daphnia magna	Calcium chloride (CaCl2)	10043524 C	768460
1b	Fresh water Lab	Measured	Static		Water Flea C	Daphnia magna	10043524 Calcium chloride (CaCl2)	10043524 C	768402
3b	Fresh water Lab	Unmeasured	Renewal	-	Water Flea C	Daphnia magna	10043524 Calcium chloride (CaCl2)	10043524 C	2152736
Lab	Fresh water La	Measured	Static		Water Flea C	Daphnia magna	Calcium chloride (CaCl2)	10043524 C	768462
dr	Fresh water Lab	Measured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)	10043524 C	768457
Lab		Measured	Static		Water Flea C	Daphnia magna	Calcium chloride (CaCl2)	10043524 C	768396
Lab	Fresh water La	Measured	Static		Water Flea C	Daphnia magna	Calcium chloride (CaCl2)	10043524 0	768399
ge	Fresh water Lab	Unmeasured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)		119478
Lab		Unmeasured	Static	Crustaceans; Standard Test Species	Water Flea C	Daphnia magna	Calcium chloride (CaCl2)	10043524	32466
Lab	Fresh water La	Unmeasured	Static	Crustaceans; Standard Test Species	Water Flea	Daphnia magna	Calcium chloride (CaCl2)	10043524	160621
Lab	Fresh water La	Unmeasured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)		742079
Lab	Fresh water La	Unmeasured	Static	- 1	Water Flea (	Daphnia magna	10043524 Calcium chloride (CaCl2)	ď	18687
Lab		Unmeasured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)		18683
Lab		Unmeasured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)		18684
Lab	Fresh water La	Unmeasured	Static	-		Daphnia magna	10043524 Calcium chloride (CaCl2)	Ũ	18688
qe		Unmeasured	Static			Daphnia magna	10043524 Calcium chloride (CaCl2)		18685
Lab	Fresh water La	Unmeasured	Static	-		Daphnia magna	Calcium chloride (CaCl2)	A	18686
qe	Fresh water Lab	Unmeasured	Renewal	Crustaceans; Standard Test Species		Daphnia magna	10043524 Calcium chloride (CaCl2)		2152737
Lab	Fresh water La	Unmeasured	Renewal			Daphnia magna	10043524 Calcium chloride (CaCl2)		2152738
Lab	Fresh water Li	Unmeasured	Static	Standard Test Species		Daphnia magna	Calcium chloride (CaCl2)	10043524	160622
ab	Fresh water Lab	Unmeasured	Static		Water Flea (	Daphnia hyalina	Calcium chloride (CaCl2)	T	44241
Lab		Unmeasured	Static	- 1	Sheepshead Mir F	Cyprinodon variegatus	10043524 Calcium chloride (CaCl2)		742988
Lab	Fresh water L	Measured	Static	50.3	Water Flea	Ceriodaphnia dubia	Calcium chloride (CaCl2)		768286
de		Measured	Static	- 4	Ĭ	Ceriodaphnia dubia	10043524 Calcium chloride (CaCl2)		768181
Lab		Measured	Static			Ceriodaphnia dubia	Calcium chloride (CaCl2)	10043524	768183
Lab	Fresh water L	Measured	Static	- 1		Ceriodaphnia dubia	Calcium chloride (CaCl2)		768283
ab		Measured	Static	- 1	Ĭ	Ceriodaphnia dubia	10043524 Calcium chloride (CaCl2)	A	767342
Lab	Fresh water L	Measured	Static		Water Flea (	Ceriodaphnia dubia	Calcium chloride (CaCl2)	10043524	768182
Lab	Fresh water L	Measured	Static	100		Ceriodaphnia dubia	Calcium chloride (CaCl2)		768284
Lab	Fresh water L	Measured	Static	Crustaceans; Standard Test Species	Water Flea (	Ceriodaphnia dubia	10043524 Calcium chloride (CaCl2)		768180
Lab	Fresh water L	Unmeasured	Renewal	Crustaceans; Standard Test Species	Water Flea (	Ceriodaphnia dubia	Calcium chloride (CaCl2)	10043524	2152765
Lab	Fresh water L	Unmeasured	Static			Ceriodaphnia dubia	10043524 Calcium chloride (CaCl2)	10043524	160620
Lab	Fresh water L	Unmeasured	Static	Crustaceans; Standard Test Species	Water Flea (	Ceriodaphnia dubia	Calcium chloride (CaCl2)	10043524	160619
Lab	Fresh water L	Unmeasured	Renewal	Crustaceans; Standard Test Species	Water Flea (	Ceriodaphnia dubia	Calcium chloride (CaCl2)	10043524	2152764
Lab	Fresh water	Unmeasured	Renewal	Molluscs	_	Biomphalaria pfeifferi	10043524 Calcium chloride (CaCl2)	7	2147187
Test Location	Media Type	Chemical Analysis	Exposure Type	Species Group	Common Name	Species Scientific Name	Chemical Name	Result Number CAS Number	esult Numbe

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r CAS Number	Chemical Name	Species Scientific Name	Species Common Name	Species Group	Exposure Type	Chemical Analysis	Media Type	Test Locati
	sodium chloride (NaCl)	Brachionus calyciflorus	Rotifer	Invertebrates; Standard Test Species	Static	Measured	Fresh water	Lab
741752 7647145 S	odium chloride (NaCl)	Brachycentrus numerosus	Caddisfly	piders	_	Unmeasured	Fresh water	Lab
764481 7647145 S	odium chloride (NaCl)	Bulinus truncatus	Snail	Molluscs	Static	Unmeasured	Fresh water	Lab
7647145	odium chloride (NaCl)	Caenorhabditis elegans	Nematode		Static	Unmeasured		Lab
	odium chloride (NaCl)	Caenorhabditis elegans			Static	Unmeasured	Fresh water	Lab
	sodium chloride (NaCl)	Caenorhabditis elegans			Static	Unmeasured	Fresh water	Lab
187901 7647145 S	odium chloride (NaCl)	Caenorhabditis elegans	100		Static	Unmeasured	Fresh water	Lab
187906 7647145 S	odium chloride (NaCl)	Caenorhabditis elegans	Nematode	Worms	Static	Unmeasured	Fresh water	Lab
7647145	odium chloride (NaCl)	Caenorhabditis elegans			Static	Unmeasured		Lab
7647145	odium chloride (NaCl)	Caenorhabditis elegans			Static	Unmeasured		Lab
	odium chloride (NaCi)	Caeroniabalis elegans			Static	Inmeasured	Fresh water	Lab
742918 7647145 S	odium chloride (NaCl)	Caenorhabditis elegans			Static	Jnmeasured	Fresh water	Lab
7647145	odium chloride (NaCl)	Carassius auratus		S. Exotic/Nuisance Species	Renewal	Unmeasured		Lab
	odium chloride (NaCl)	Carassius auratus				Unmeasured		Lab
196831 7647145 S	odium chloride (NaCl)	Carassius auratus	Goldfish			Unmeasured		Lab
196814 7647145 S	odium chloride (NaCl)	Carassius auratus	Goldfish	Fish; U.S. Exotic/Nuisance Species		Unmeasured		Lab
196791 7647145 S	odium chloride (NaCl)	Carassius auratus	Goldfish	Fish; U.S. Exotic/Nuisance Species		Unmeasured	Fresh water	Lab
196776 7647145 S	odium chloride (NaCl)	Carassius auratus	Goldfish	Fish; U.S. Exotic/Nuisance Species		Unmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus	Goldfish			Unmeasured	Fresh water Lab	Lab
	odium chloride (NaCi)	Carassius auratus	Goldfish			Jameasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus	Goldfish			Unmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus	Goldfish			Not Reported	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus	Goldfish			Measured	Fresh water	Lab
196826 7647145 S	odium chloride (NaCl)	Carassius auratus	Goldfish	Fish; U.S. Exotic/Nuisance Species	Ē	Jnmeasured	Fresh water	Lab
5	odium chloride (NaCl)	Carassius auratus	Goldfish			Jnmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured		Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured		Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water	Lab
7647145	odium chloride (NaCl)	Carassius auratus				Inmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water 1	Lab
196780 7647145 St	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water I	Lab
7647145	odium chloride (NaCl)	Carassius auratus	Goldfish	-		Jnmeasured		Lab
7647145	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water I	Lab
7647145	odium chloride (NaCl)	Carassius auratus		Exotic/Nuisance Species		Inmeasured		Lab
	odium chloride (NaCI)	Carassius auratus		Exotic/Nuisance Species		Inmeasured	Fresh water	Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured	Fresh water L	Lab
7647145	odium chloride (NaCl)	Carassius auratus		T <sub>m</sub>		Jnmeasured		Lab
	odium chloride (NaCl)	Carassius auratus				Jnmeasured		Lab
196810 7647145 50	odium chloride (NaCl)	Carassius auratus	Goldfish	114		Jnmeasured	Fresh water L	Lab
196822 7647145 Sc	odium chloride (NaCl)	Carassius auratus	Goldfish			Jnmeasured	Fresh water L	Lab
7647145	odium chloride (NaCl)	Carassius auratus	Goldfish			Jnmeasured	Fresh water L	Lab
196798 7647145 50	odium chloride (NaCl)	Carassius auratus	Goldfish	(1)		Jnmeasured	Fresh water L	Lab
1	odium chloride (NaCl)	Carassius auratus				Inmeasured	Fresh water Lab	ab
7647145	odium chloride (NaCl)	Carassius auratus				Inmeasured	Fresh water L	Lab
	odium chloride (NaCI)	Carassius auratus				Inmeasured	Fresh water	LdD
	Towns the same of	Contraction of the Contraction		L			i cair trace.	-000
Number 2152847 741752 7641752 7641752 7641752 7641752 7641752 7641752 7641752 7642918 187902 187903 187904 196811 196793 196795 196828 196829	CAS	rCAS Number 7647145 Sodium Chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCI)         Brachinas cohyliforus         Rollifer         Invertebrates; Sandard Test Species Group           Sodium chloride (NaCI)         Brachinas cohyliforus         Snall         Mollisscs           Sodium chloride (NaCI)         Brachinas cohyliforus         Snall         Mollisscs           Sodium chloride (NaCI)         Caenorhabditis elegons         Nematode         Worms           Sodium chloride (NaCI)         Caenosius auratus         Goldfish         Fish; U.S. Exactic/Nuisance Species           Sodium chloride (NaCI)	Species Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Grandi Marchinde (NaCl) Grandi Marchinde (NaCl) Grandiad Test Species Group Sodium chloride (NaCl) Grandiad Test Species Sodium chloride (NaCl) Grandiad Grandiad Test Species Sodium chloride (NaCl) Grandiad Grandiad Species Species Sodium chloride (NaCl) Grandiad Grandiad Species Sp	Species   Spec	Chemical Name         Species Sciuntific Name         Species Sciuntific Name         Expecies Group         Expoure Yepe Chemical Analysis Media Type           Socium chioride (Naci)         Borchivorsic coloristis         Borchivorsic coloristis         Common Chioride (Naci)         Expoure Yepe Chemical Analysis Media Type           Socium chioride (Naci)         Gordon Chioride (Naci)

Depart Number   Colonian Num		Frech water I		Renewal		Snake-Head Cattle	Channa nunctata	Sodium chloride (NaCl)	7647145	741635
Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Wat	30	LIESH MATEL TO			stacealls	Andrei Lica	cerioaabinina reticatata	Sociality contract (seact)	1011210	1 14100
Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Water Flea Crustaceans; Sandard Test Species Sodium chloride (NaCl) Ceriodophina dubio Wat	5 6	Fresh water La			brancala icacapacica		Ceriodaphnia reticulata	Sodium chloride (NaCl)	7647145	741435
Sodium chloride (NaCl)	ab de	Fresh water La		Static			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	768175
Sodium chloride (NaCl)	de	Fresh water La		Static			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	768266
Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species Sodum chloride (NaCl) ( <i>Ceriodophnia dubia</i> Water Flea Crustaceans; Sandard Test Species	de			Renewal	Standard Test Species		Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	742530
Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl)	qe	Fresh water La	ed	Renewal			Ceriodaphnia dubia			2152722
Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodum chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustacea	de	Fresh water La		Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	207721
Sodium chloride (Nacl)  Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nacl)  Ceriodophnio dubio  Water Flea  Crustaceans, Standard Test Species Sodium chloride (Nac	qe	Fresh water La	Measured	Renewal			Ceriodaphnia dubia			741660
Sodium chloride (NaCl)  Sodium chloride (NaCl)  Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Spec	qe	Fresh water La		Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	207719
Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Stand	de						Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	207718
Sodium chloride (NaCl)	ab	Fresh water La					Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	768173
Sodium chloride (NaCl)	de	Fresh water La					Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	768268
Sodium chloride (NaCl)	ab		Measured	Static			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	768340
Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio  Water Flea  Crustaceans; Standard Test Species Sodium chloride (NaCl)  Ceriodophnio dubio	ab	Fresh water La		Static			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	768272
Sodium chloride (NaCl)	ab	Fresh water La		Renewal	_		Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	741671
Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl)	ab		Measured	Renewal			Ceriodaphnia dubia			741699
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnio dubio         Water Flea         Crustaceans; Stand	ab	Fresh water La	Measured	Static	_		Ceriodaphnia dubia			741695
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodophnio dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Stand	ab	Fresh water La	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	251006
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Stand	ab		Unmeasured	Renewal			Ceriodaphnia dubia			251008
Chemical Name Ceriodophnia dubia Sodium chloride (NaCl) Sodium chlor	ab	Fresh water La		Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	251002
Chemical Name Ceriodophnia dubia Custaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Ceriodophnia dubia Custaceans; Standard Test Species Codium chloride (NaCl) Ceriodophnia dubia Ceriodophnia dubia Custaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia	ab		Unmeasured	Renewal			Ceriodaphnia dubia		7647145	251004
Species Scientific Name Common Name   Species Group	ab		Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	120855
Species Scientific Name Common Name Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodophnia dub	ab	Fresh water La	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	251000
Chemical Name  Ceriodophnio dubia  Sodium chloride (NaCl)  Sodium chloride (Na	ab		Unmeasured	Renewal			Ceriodaphnia dubia		ij	250997
Chemical Name  Species Scientific Name  Common Name  Species Group  Sodium chloride (NaCl)  Sodium chl	ab			Renewal	-		Ceriodaphnia dubia			767849
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Stand	ab			Renewal	-		Ceriodaphnia dubia	Sodium chloride (NaCl)		767863
Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species	ah	Fresh water L	Unmeasured	Renewal			Ceriodaphnia duhia			767865
Chemical Name  Species Scientific Name  Common Name  Species Group  Sodium chloride (NaCl)  Sodium chl	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia		7647145	767976
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodi	ah l		Unmeasured	Renewal			Ceriodaphnia dubia			767882
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Stand	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia			767888
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodi	ab	Fresh water L	Unmeasured	Renewal	112		Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	767954
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodi	ab		Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	767847
Chemical Name Species Scientific Name Common Name Sodium chloride (NaCl) Sodium chloride (N	аь	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	767970
Chemical Name         Species Scientific Name         Common Name           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species	ah	Fresh water	Unmeasured	Renewal			Ceriodaphnia duhia	Sodium chloride (NaCl)		767857
Chemical Name  Chemical Name  Common Name  Species Group  Sodium chloride (NaCl)  Ceriodophnia dubia  Common Name  Coustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Common Name  Coustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Common Name  Coustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Coustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Coustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Ceriodophnia dubia  Ceriodophnia dubia  Ceriodophnia dubia  Coustaceans; Standard Test Species  Sodium chloride (NaCl)  Ceriodophnia dubia  Coustaceans; Standard Test Species  Colium chloride (NaCl)  Ceriodophnia dubia  Cerio	du		Unmeasured	Renewal	-4:		Ceriodaphnia dubia	Sodium chloride (NaCl)		767890
Chemical Name         Species Scientific Name         Common Name           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species	db	Fresh Water L	Unmeasured	Kenewai	_		cerioaapnnia aubia	sodium chioride (NaCi)		/800/3
Chemical Name         Species Scientific Name         Common Name           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species	ab		Unmeasured	Renewal			ceriodapnnia dubia	Sodium chioride (NaCI)		306/9/
Chemical Name         Species Scientific Name         Common Name           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)		767871
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodi	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	7647145	767960
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodium chloride (NaCl) Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Sodium chloride (NaCl) Sodiu	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)		767873
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Vater Flea Crustaceans; Standard Test Species Vater Flea Crustaceans; Standard Test Species Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Codium chloride (NaCl) Ceriodaphnia dubia Vater Flea Crustaceans; Standard Test Species Codium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Codium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Crustaceans; Standard Test Species Codium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Crustaceans; Standard Test Species Crustaceans; Standard Test Species Crustaceans; Standard Test Species	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)		767896
Chemical Name Species Scientific Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Sodium chloride (NaCl) Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Sodium chloride (NaCl) Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Sodium chloride (NaCl) Sodium chloride (NaCl) Sodium chloride (NaCl) Ceriodaphnia dubia Sodium chloride (NaCl) Sodium chloride	.ab		Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	21	767880
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species	ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)		767962
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species	.ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)	1	767898
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species	.ab		Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)		766865
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodaphnia dubia         Water Flea         Crustaceans; Standard Test Species	.ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	Sodium chloride (NaCl)		767904
Chemical Name         Species Scientific Name         Common Name         Species Group           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species           Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species	.ab		Unmeasured	Renewal		9	Ceriodaphnia dubia	5 Sodium chloride (NaCl)	100	767855
Chemical Name Species Scientific Name Common Name Species Group  Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species  Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species	.ab		Measured	Renewal	Crustaceans; Standard Test Species		Ceriodaphnia dubia	Sodium chloride (NaCl)		741687
Chemical Name Species Scientific Name Common Name Species Group Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species	.ab	Fresh water L	Unmeasured	Renewal			Ceriodaphnia dubia	S Sodium chloride (NaCl)	11 1 2	251005
Chemical Name Species Scientific Name Common Name Species Group	.ab	Fresh water L	Unmeasured	Renewal		Ť.	Ceriodaphnia dubia			250998
	Test Locatio	Media Type 1	Chemical Analysis	Exposure Type	Species Group	Species Common Name	Species Scientific Name		CAS Number	ult Number

7667135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Renew 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Renew 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Renew 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Renew 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodium chloride (NaCl) Dophnia magina Water Flea Custaceans. Standard Test Species Static 767135 Sodiu	Result Number	CAS Number	Chemical Name	Species Scientific Name	Species Common Name	Species Group	Exposure Type	ure Type   Chemical Analysis   Media Type   Test Location	Media Type T	est Loc
7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants, Standard Test Species   Static Unmeasured   7-867125 Soalum chloride (NaCl) Dophnic magne   Water Fiela Coutaceants,	18464		m chloride (NaCl)	Daphnia magna				Unmeasured	Fresh water Lab	ab
1 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Rerewal Unmeasured Frotal Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Rerewal Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Unmeasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Water Fila Crustaeans; Standard Test Species Static Weasured 7 7621745 Soulum chloride (MaCl) Duphnin magino Wat	2152826		m chloride (NaCl)	Daphnia magna	F	100		Measured	Fresh water Lab	ab
1 P621745 Sodalm chloride (NaCl) Daphnio magno Water File Custaceans; Standard Test Species   Sank: Unmeasured   February   File   February   F	759179	E	m chloride (NaCl)	Daphnia magna	Ē		<u>a</u>	Unmeasured		ab
Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Ummeasured   Pol 1745 Sodium chloride (NaCl)   Daphnia magna   Water Fea   Custaceans; Standard Test Species Static   Weasured   Pol 1745 Sodium chloride (NaCl)   Daphni	2152709		m chloride (NaCl)	Daphnia magna			2 12	Unmeasured	Fresh water L	Lab
Februs   Sedium chloride (NaCl)   Dalphini magina   Water Files   Constituents, Standard Text Species Static Unmeasured   Februs   Sedium chloride (NaCl)   Dalphini magina   Water Files   Constituents, Standard Text Species Static Unmeasured   Februs	144721		m chloride (NaCl)	Danhnia magna			/di	Inmeasured	Fresh water Lab	Lab
7-647745 Sodium chloride (NaCl) Daphina magna Mare Files Coustreams; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Mare Files Coustreams; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustreams; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustreams; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustreams; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Measured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Umneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Unneasured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Measured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Species Static Measured 7-7647745 Sodium chloride (NaCl) Daphina magna Water Files Coustream; Sandard fest Speci	144220	7647145	m chloride (NaCl)	Daphnia magna		- 1		Unmeasured		Lab
7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Umneasured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Measured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Measured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Measured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Measured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standard Test Species Static Measured (7647145 Sedium chloride (NaCl) Daphina magna Water Flee Crustrecens; Standar	144218		m chloride (NaCl)	Daphnia magna				Unmeasured	Fresh water L	Lab
Ted 7145 Sedium chloride (NaCl) Dophnia mogna Water Flee Crustreems; Standard Test Species (Static Umneasured Test 7547145) Sedium chloride (NaCl) Dophnia mogna Water Flee Crustreems; Standard Test Species (Static Umneasured Test 7547145) Sedium chloride (NaCl) Dophnia mogna Water Flee Crustreems; Standard Test Species (Static Umneasured Test 7547145) Sedium chloride (NaCl) Dophnia mogna Water Flee Crustreems; Standard Test Species (Static Umneasured Dophnia mogna Water Flee) Crustreems; Standard Test Species (Static Umneasured Dophnia mogna Water Flee) Crustreems; Standard Test Species (Static Umneasured Crustreems) Standard Test Species (Static Umneasured Dophnia mogna Water Flee) Crustreems; Standard Test Species (Static Umneasured Crustreems) Standard Test Species (Static Measured Crustreems) Standard Test Species (Static Measured Crustreems) Standard Test Species	144219		m chloride (NaCl)	Daphnia magna				Unmeasured	Fresh water L	Lab
VS47135 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Ummeasured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured  VS47145 Sodium chloride (NaCl)  Dispinio magno  Water Files  Custaceans; Standard Test Species Static  Measured	160322		m chloride (NaCl)	Daphnia magna		- 1		Jnmeasured	Fresh water Lab	Lab
7-6273-5 Sodium chloride (NaCl) Dophnio mogno Water Flea Crustaceans; Standard Test Species Static Measured Fresh (Crustaceans; Standard Test Species Static Measured Dophnio mogno Water Flea Crustaceans; Standard Test Species Static Measured Fresh (Crustaceans; Standard Test Species Static Measured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Fresh (Crustaceans; Standard Test Species Static Unmeasured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Polytria Static Unmeasured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Polytria Static Unmeasured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Polytria Static Unmeasured Polytria mogno Water Flea Crustaceans; Standard Test Species Static Unmeasured Polytria Static Unmeasured	742529		m chloride (NaCl)	Daphnia magna			a	Jnmeasured	Fresh water L	Lab
7647145   Sodium chloride (NaCl)   Daphnia magna	160321	-	m chloride (NaCl)	Daphnia magna	à	1.22		Unmeasured	Fresh water Lab	ab
7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Standard Test Spe	767302	7647145	n chloride (NaCl)	Daphnia magna				Measured	Fresh water L	Lab
Tel-1145 Sodium chloride (NaCl)   Daphina magna	772103		m chloride (NaCl)	Daphnia magna				Measured	Fresh water L	Lab
7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Dophnia mogna Water Flea Crustaceans; Standard Test Species Static Measured 76471	742019	7647145	m chloride (NaCl)	Daphnia magna				Jnmeasured	Fresh water L	Lab
7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Umnessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Umnessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Umnessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured   7647145   Sodium chloride (NaCl)   Dophnia magna   Water Flea   Crustaceans; Sandard Test Species   Static   Wessured	18460		n chloride (NaCl)	Daphnia magna	2			Jnmeasured	Fresh water Lab	ab
Tel-1145   Sodium chloride (NaCl)   Daphnia magna	119469	7647145	m chloride (NaCl)	Daphnia magna		100		Jnmeasured	Fresh water La	Lab
Tright   Sodium chloride (Nacl)   Daphnia magna	2164731	7647145	m chloride (NaCl)	Daphnia magna			_	Inmeasured	Fresh water Lab	ah
Te47145 Sodium chloride (NaCl)   Dophnia magna	768426	7647145	n chloride (NaCl)	Daphnia magna		-		Measured	Fresh water La	Lab
7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static	768527		n chloride (NaCl)	Daphnia magna				Measured	Fresh water Lab	ab
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7647145   Sodium chloride (NaCl)   Daphria magna	768445		n chloride (NaCl)	Daphnia magna				Measured	Fresh water Lab	ab
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7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   Unmeasured   Unmeasured   Unmeasured   Unmeasured   Unmeasured   Unmeasured   Unmeasured   Unmeasur	768486		n chloride (NaCl)	Daphnia magna				Measured	Fresh water La	Lab
7647145 Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   Total Static   Measured   Total Static   Tota	2164732		n chloride (NaCl)	Daphnia magna				Inmeasured	Fresh water Lab	ab
Total 145   Sodium chloride (NaCl)   Daphnia magna	2152707	7647145	n chloride (NaCl)	Daphnia magna			/al	Inmeasured	Fresh water Lab	ab
Total 145   Sodium chloride (NaCl)   Dophnia magna	768428	764/145	n chloride (NaCl)	Daphnia magna				Measured	Fresh water Lab	ab
7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145     Sodium chloride (NaCl)     Daphnia magna     Water Flea     Cr	768500	7647145	n chloride (NaCl)	Daphnia magna				Measured		Lab
7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Weasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Te	768431		n chloride (NaCl)	Daphnia magna				Neasured		de de
Total Titals   Sodium chloride (NaCl)   Daphnia magna   Water Flea   Crustaceans; Standard Test Species   Static   Measured	768498	7647145 Sodiur	n chloride (NaCl)	Daphnia magna				Neasured		Lab
7647145 Sodium chloride (NaCl) Daphnia magna Water Flea Crustaceans; Standard Test Species Static Measured 7647145 Sodium chloride (NaCl) Daphnia magna Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Daphnia magna Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Daphnia magna Water Flea Crustaceans; Standard Test Species Static Measured Crustaceans; Standard Test Species Static Measured Water Flea Crustaceans; Standard Test Species Static Measured Water Flea Crustaceans; Standard Test Species Static Measured Opphnia magna Water Flea Crustaceans; Standard Test Species Static Unmeasured Test Species Test Spec	768450	7647145 Sodium	n chloride (NaCl)	Daphnia magna			/al	Jnmeasured Measured	Fresh water Lab	ah ab
7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Measured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium chloride (NaCl)     Dophnia magna     Water Flea     Crustaceans; Standard Test Species     Static     Unmeasured       7647145 Sodium	768523		n chloride (NaCl)	Daphnia magna				Neasured	Fresh water Lab	qE
7647145 Sodium chloride (NaCl)  Dophnio magna  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Static  Measured  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Valer Flea  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Static  Unmeasured  Valer Flea  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Static  Unmeasured  Valer Flea  Crustaceans; Standard Test Species  Crustaceans; St	768530	7647145	n chloride (NaCl)	Daphnia magna				Neasured	Fresh water Lab	qe
Total   Tota	2210504	7647145	n chloride (NaCl)	Daphnia magna				Inmeasured		de
7647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  7647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Static  Unmeasured  Crustaceans; Standard Test Species  Static  Unmeasured  7647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Crustacea	768488	7647145	n chloride (NaCl)	Daphnia magna				Measured	Fresh water Lab	5 6
7647145 Sodium chloride (NaCl)  Dophnia magna  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Crustaceans; Standard Test Species	768526		n chloride (NaCl)	Daphnia magna		- 1		Aeasured	Fresh water Lab	qE
7647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Crustaceans; Standard Test Species  Static  Unmeasured  Crustaceans; Standard Test Species  Static  Unmeasured  T647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Crustaceans; Standard Test Speci	2210505	7647145 Sodiun	n chloride (NaCl)	Daphnia magna				Inmeasured	Fresh water Lab	qe
7647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Vater Flea  Crustaceans; Standard Test Species  Static  Unmeasured  T647145 Sodium chloride (NaCl)  Daphnia magna  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Crustaceans; Standard Test Species  Static  Unmeasured  Unmeasured	742069	7647145 Sodiun	n chloride (NaCl)	Daphnia magna					Fresh water Lab	g
7647145 Sodium chloride (NaCl) Dabhila magan Water Flea Crustaceans, Standard Test Species Static Unmasured	742926	7647145 Sodium	n chloride (NaCl)	Daphnia magna					Fresh water Lab	5 8
The state of the s	742068	7647145 Sodiun	n chloride (NaCl)	Daphnia magna					Fresh water Lab	Ď

Result Number	r CAS Number	Chemical Name	Species Scientific Name	Species	Species Group		Chamical Application	Modia Tuno	Tost long
196762	7647145	Sodium c	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Renewal	Renewal Unmeasured	Fresh water	Lab
196719	7647145	7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnoy Fish; Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
2055542	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
160325		7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Static	Unmeasured	Fresh water	Lab
160324		Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Static	Unmeasured	Fresh water Lab	Lab
167861	7647145	7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Renewal	Not Reported	Fresh water	Lab
160323	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Static	Unmeasured	Fresh water Lab	Lab
201537		Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Flow-through	Measured	Fresh water Lab	Lab
196769	7647145	7647145 Sodium chloride (NaCl)	Pimephales prometas	Fathead Minnoy	Fathead Minnoy Fish; Standard Test Species	Renewal	Unmeasured	Fresh water	Lab
196765	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish: Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
196734	37	7647145 Sodium chloride (NaCI)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Renewal	Unmeasured	Fresh water	Lab
196761	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Renewal	Unmeasured	Fresh water	Lab
196738		Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnov Fish; Standard Test Species	Renewal	Unmeasured	Fresh water	Lab
196/18		7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnoy Fish; Standard Test Species	Renewal	Unmeasured	Fresh water	Lab
196757	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov	Fathead Minnoy Fish: Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
196749	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov P	Fathead Minnov Fish; Standard Test Species	Renewal	Unmeasured	Fresh water	Lab
196722	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnoy Fish; Standard Test Species	Renewal	Unmeasured	Fresh water I	Lab
196742	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species		Unmeasured	Fresh water I	Lab
196773	T	7647145 Sodium chloride (NaCl)	Pimenhales prometas	Eathead Minnou	Fathead Minnoy Fish; Standard Test Species			Fresh water	Lab
196730		7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish: Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
167862	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnoy Fish; Standard Test Species				ab
42866	7647145	7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fish; Standard Test Species			Fresh water Lab	ab
10000	2647145	Sodium chloride (Naci)	Pimephales promeias	Fathead Minnovi	Fathead Winnov Fish; Standard Test Species		Unmeasured	Fresh water Lab	ab
96906		7647145 Sodium chloride (NaCI)	Pimephales promelas	Fathead Minnov F	Fathead Minnoy Fish; Standard Test Species  Fathead Minnoy Fish: Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
14389		Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water Lab	ah
68598	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water Lab	ab
768104	l v	7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water Lab	ab
768072	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water Lab	.ab
768066	7647145	7647145 Sodium chloride (NaCl)	Pimephales prometas	Fathead Minnov	Fathead Minnoy Fish: Standard Test Species			Fresh water Lab	ab
768058	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnoy Fish: Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
768096	7647145 9	7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnoy F	Fathead Minnov Fish; Standard Test Species			Fresh water L	Lab
768127		7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water L	Lab
768143	7647145 \$	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species		A	Fresh water Lab	ab
768088	7647145	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species				Lab
768123	7647145 S	7647145 Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnoy Fish: Standard Test Species	Renewal	Unmeasured	Fresh water Lab	Lab
768163	7647145 S	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water L	Lab
768153	7647145 S	Sodium chloride (NaCl)	Pimephales promelas	Fathead Minnov F	Fathead Minnov Fish; Standard Test Species			Fresh water L	Lab
768098	7647145 S	7647145 Sodium chloride (NaCl)		Fathead Minnov Fi	Fathead Minnov Fish; Standard Test Species			Fresh water Lab	ab
/680/4		Sodium chloride (NaCl)		Fathead Minnov Fi	Fathead Minnov Fish; Standard Test Species	Renewal	Unmeasured F	Fresh water L	Lab
760113	7647145 S	Sodium chloride (NaCl)		Fathead Minnov Fi	Fathead Minnov Fish; Standard Test Species		7	Fresh water L	Lab
768107	7647145 5	7647145 Sodium chloride (NaCl)	Pimephales promeias	Fathead Minnov F	Fathead Minnoy Fish; Standard Test Species			Fresh water L	Lab
768082	7647145 S	7647145 Sodium chloride (NaCl)		Fathead Minnov Fi	Fathead Minnov Fish: Standard Test Species	Renewal	Unmeasured F	Fresh water L	Lab
768137	7647145 S	Sodium chloride (NaCl)		Fathead Minnov Fi	Fathead Minnov Fish; Standard Test Species			Fresh water La	Lab
768147	7647145 S	Sodium chloride (NaCl)		Fathead Minnov Fi	Fathead Minnov Fish; Standard Test Species			Fresh water La	Lab
100004	C C+1 / +0/	764/145 Sodium chloride (Naci)	Pimephales promelas	Fathead Minnov Fi	Fathead Minnoy Fish; Standard Test Species		Unmeasured F	Fresh water Lab	ab

2	Trock mater lob		Ctotio	NACTION OF THE PROPERTY OF THE			77E 7076 Culfurie acid codium calt (1.7)	775707616	767505
Lab	Fresh water La		Static		Snail	Biomphalaria alexandrina		1/5/8/6/1	104340
Lab				ans	Order			7757826 5	181/9
ab						Amphipoda		7757826 S	18177
Lab	Fresh water La	Unmeasured	Static	Crustaceans	Scud Order C	Amphipoda	7757826 Sulfuric acid sodium salt (1:2)	7757826 5	18176
Lab	Fresh water La	Unmeasured		Crustaceans	Scud Order C	Amphipoda		7757826 S	18178
Lab	Fresh water La	Measured			Fathead Minnoy F	Pimephales promelas		7487889 S	767519
Lab	Fresh water La	Unmeasured	Static	Fish; Standard Test Species	Fathead Minnoy F	Pimephales promelas		7487889 S	160293
ab					Fathead Minnov F	Pimephales promelas		7487889 S	160292
Lab		Unmeasured	val		Fathead Minnov F	Pimephales promelas	_	7487889 S	759181
Lab	Fresh water La	Unmeasured			Fathead Minnov F	Pimephales promelas	_	7487889 S	160291
Lab		Unmeasured			Japanese Medal F	Oryzias latipes		7487889 \$	89660
Lab	Fresh water La	Unmeasured			Japanese Medal F	Oryzias latipes			89662
Lab	Fresh water L	Unmeasured			Japanese Medal H	Oryzias latipes		7487889 S	61289
Lab	Fresh water L	Unmeasured	Static	Japanese Medal Fish; Standard Test Species	Japanese Medal F	Oryzias latipes		7487889 S	61288
Lab	Fresh water L	Unmeasured	Static	Japanese Medal Fish; Standard Test Species	Japanese Medal I	Oryzias latipes	Sulfuric acid magnesium salt (1:1)	7487889 S	61287
Lab	Fresh water L	Unmeasured	Static	Japanese Medal Fish; Standard Test Species	Japanese Medal I	Oryzias latipes		7487889 S	89658
Lab	Fresh water L	Unmeasured	Static	Molluscs	Pond Snail	Lymnaea sp.	-	7487889 S	24915
Lab	Fresh water L	Unmeasured	Static	Molluscs		Lymnaea sp.	7487889 Sulfuric acid magnesium salt (1:1)	7487889 S	24913
Lab	Fresh water L	Unmeasured	Static	Molluscs	Pond Snail	Lymnaea sp.		7487889 S	24912
Lab	Fresh water L	Unmeasured	Static	Molluscs	Pond Snail	Lymnaea sp.	Sulfuric acid magnesium salt (1:1)	7487889 S	24914
Lab	Fresh water L	Unmeasured	Static	Fish; Standard Test Species	Bluegill F	Lepomis macrochirus		7487889 S	24911
Lab	Fresh water L	Unmeasured	Static	Worms		Heterorhabditis bacterioph (Nematode		7487889 S	220044
Lab	Fresh water L	Unmeasured	Static	Fish; U.S. Exotic/Nuisance Species	Western Mosqu F	Gambusia affinis	_	7487889 S	2064015
Lab	Fresh water L	Unmeasured	Static		Western Mosqu	Gambusia affinis		7487889 S	2064016
Lab	Fresh water L	Unmeasured	Static	Fish; U.S. Exotic/Nuisance Species	Western Mosqu	Gambusia affinis		7487889 S	2064012
Lab	Fresh water L	Unmeasured				Daphnia magna		7487889 9	759470
Lab		Unmeasured	/al			Daphnia magna		7487889 S	759473
Lab	Fresh water L	Measured				Daphnia magna		7487889 \$	767490
Lab		Measured				Daphnia magna	_	7487889 \$	767487
Lab		Measured		- 1		Daphnia magna			767471
Lab	Fresh water L	Measured	Static			Daphnia magna		7487889 9	767485
Lab	Fresh water L	Measured				Daphnia magna		7487889 \$	767488
Lab		Unmeasured		_		Daphnia maana	_	7487889	24909
Lab	Fresh water L	Unmeasured	Static			Daphnia magna		7487889 9	24906
Lab		Unmeasured	Static			Daphnia magna		7487889 9	24908
Lab		Unmeasured	Static			Daphnia magna		7487889 9	24907
Lab		Unmeasured	Static	-		Daphnia maana		7487889 9	24910
Lab		Unmeasured	Static			Daphnia magna		7487889	160290
Lab		Unmeasured	Static			Daphnia magna	Sulfuric acid magnesium salt (1:1)	7487889	160289
Lah		Unmeasured	Renewal	Crustaceans: Standard Test Species		Danhnia maana	7487889 Sulfuric acid magnesium salt (1:1)	7487889	759182
lah		Measured	Static			Ceriodaphnia dubia	Sulfuric acid magnesium salt (1:1)	7487889	767465
Lab	Fresh water L	Measured	Static			Ceriodaphnia dubia	Sulfuric acid magnesium salt (1:1)	7487889	767461
Lab		Measured	Static			Ceriodaphnia dubia	7487889 Sulfuric acid magnesium salt (1:1)	7487889	767405
Lab	Fresh water L	Measured	Static	-		Ceriodaphnia dubia	Sulfuric acid magnesium salt (1:1)	7487889	767347
Lab		Measured	Static	-		Ceriodaphnia dubia		7487889	767410
Lab		Measured	Static		2	Ceriodaphnia dubia	_	7487889	767366
Lab	Fresh water L	Measured	Static	Crustaceans; Standard Test Species		Ceriodaphnia dubia	Sulfuric acid magnesium salt (1:1)	7487889	767364
Lab		Measured	Static	100		Ceriodaphnia dubia	7487889 Sulfuric acid magnesium salt (1:1)	7487889	767407
Lab	Fresh water L	Measured	Static	Crustaceans; Standard Test Species	Water Flea	Ceriodaphnia dubia	Sulfuric acid magnesium salt (1:1)	7487889	767365
Lab	Fresh water L	Measured	Static	Crustaceans; Standard Test Species	Water Flea	Ceriodaphnia dubia	7487889 Sulfuric acid magnesium salt (1:1)	7487889	767402
Lab	Fresh water L	Measured	Static	Crustaceans; Standard Test Species	Water Flea	Ceriodaphnia dubia	Sulfuric acid magnesium salt (1:1)	7487889	767348
Test Location	Media Type	Chemical Analysis Media Type	Exposure Type	Species Group	Species Common Name	Species Scientific Name	Chemical Name	Result Number CAS Number	sult Number

Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobbs   Water Files   Crustacents; Standard Test Species   Renewal   Unmeasured   Sedium ribindie Naci)   Cerodophino dobb	ult Number	Result Number   CAS Number	Chemical Name	Species Scientific Name	Species Common Name	Species Group	Exposure Type	re Type   Chemical Analysis   Media Type		Test Location
7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina dubles Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal Unmeasured 7-867135 (Sodium chloride (MisCl) (Circidalphina duble) Water Flas Crustacents; Standard Test Species Rerewal	2153267		Sodium c	Ceriodaphnia dubia	Water Flea	Crustaceans; Standard Test Species	Renewal	Measured	Fresh water	Lab
1967/14/5 Sondim chloride (NaCl) (Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test Species Bereval Unmeasured Circidaphina dubia Mater Files Custaceanis, Standard Test	767971	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia	Water Flea	Crustaceans; Standard Test Species	Renewal			Lab
Part 145 Sodium chloride (NaCl) (Ceriodophino dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional dubio Water Fiela Custaceants, Standard Test Speciels Renewal Unmeasured (Professional	767883	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia	Water Flea		Renewal			Lab
Territo Sodium chloride (NaCl) (Ecrolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured (Errolospinia dubio Water Fela Custateanis, Standard Est Species Renewal Unmeasured	767957		Sodium chloride (NaCl)	Ceriodaphnia dubia	Water Flea		Kenewal		Fresh water	Lab
7-627135 Scalum ribioride (NaCl) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Sodium chloride (NaCl) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Sodium chloride (NaCl) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Sodium chloride (NaCl) (Ceriodiphino dubio Water Files (Custaceans, Standard Test Species) Renewal Unmeasured (PRCL) (Sodium chloride (NaCl) (Ceriodiphino dubio Water Files (Cust	767901		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
T-9617145 Sedium chloride (NaCl) Cerrodophnia dubbie Water Flas Constaenns; Standard Test Species Renewal Unmeasured restriction of the Control of the Species Renewal Unmeasured control of the Control of the Species Renewal Unmeasured control of the Control of the Species Renewal Unmeasured control of the Control of the Species Renewal Unmeasured control of the Control of the Species Renewal Unmeasured control of the Control of the Species Renewal Unmeasured Control of the Species Renewa	767860	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia		-	Renewal		Fresh water Lab	Lab
7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7-647745 Sodium chloride (NaCl) Ceriodophnia dublia Water	767868		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal	i	Fresh water	Lab
1. 7647745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Renewal Unmeasured (174745 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sandard Test Species Static Unm	766868		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7617145 Sodium chioride (NaC)	767893		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Coustaceans; Sandard Test Species Salatic Unmea	767876		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnio dubio   Water Flea   Ciustaceans; Sandard rest Species   Static   Unmeasured	767858		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal	i	Fresh water	Lab
7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Cerodophnio dubio   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured	742568		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species Renewal Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species Renewal   Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species Renewal   Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species Renewal   Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species   Static   Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina dubio   Water Flea   Crustaceans, Standard Test Species   Static   Unmeasured   Total 145 Sodium chloride (NaCl)   Ceriodophina du	767955		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water Lab	Lab
7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species Renewal   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (Naci)   Ceriodophnia dubio   Water Flea   Ciustaceans; Standard Test Species   Static   Unmeasured   7647145	767850		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal			Lab
Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmessured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmessured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmessured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmessured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmessured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured Ted-1145 Sodium chloride (NaCl) Ceriodophnio dubio Water Flea Crustaceans; Sta	766862		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species	767866		Sodium chloride (NaCl)	Ceriodaphnia dubia	Ĭ		Renewal		Fresh water	Lab
7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Specie	767891		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water Lab	Lab
7647145 Sodium chloride (NaCl)	767885		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubio Water Flea Crustaceans; Standard Test Species S	767963		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water Lab	Lab
7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Re	767899		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145 Sodium chloride (NaCl)	767973	7647145	Sodium chloride (NaCI)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   77647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species	251022	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal			Lab
7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewal Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodophnia dubia Water Flea Crustaceans; Standard Test Species Renewa	251024		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal	Ī	Fresh water	Lab
7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmea	251014	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal	Ī	Fresh water	Lab
7647145 Sodium chloride (NaCl)	251020		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
17647145 Sodium chloride (NaCl)	251016		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           8         7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured           7647145         Sodium chloride (NaCl)         Ceriodophnia dubia         Water Flea         Crustaceans; Standard Test Species         Static         Unmeasured	251018		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal			Lab
7647145 Sodium chloride (NaCl)	251012		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal		Fresh water	Lab
T647145 Sodium chloride (NaCl) T647146 Sodium chloride (NaCl) T647147 Sodium chloride (NaCl) T647148 Sodium chloride (NaCl) T647149 Sodium chloride (NaCl) T647145 Sodium chloride (NaCl) T647145 Sodium chloride (NaCl) T647146 Sodium chloride (NaCl) T647147 Sodium chloride (NaCl) T647148 Sodium chloride (NaCl) T647148 Sodium chloride (NaCl) T647149 Sodium chloride (NaCl)	741947	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia			Static		Fresh water	Lab
7647145 Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145 Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145 Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145 Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   Test Species   Static   Unmeasured   Unmeasured   Test Species   Static   Unmeasured   Unmeasured   Test Species   Static   Unmeasured   Test Species   Static   Unmeasured   Unmeasured   Unmeasured   Unmeasured   Test Species   Unmeasured   Unmeasured   Unmeasured   Test Species   Unmeasured   Unmeasured   Test Species   Unmeasured   Unmeasured   Unmeasured   Test Species   Unmeasured   U	741938	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia		1	Static		Fresh water	Lab
7647145 Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured 7647145 Sodium chloride (NaCl) Ceriodaphnia dubia Water Flea Crustaceans; Standard Test Species Static Unmeasured Crustaceans; Standard Test Species Static Unmeasured Water Flea Crustaceans; Standard Test Species Static Unmeasured Crustaceans; Standard Test Species Static Unmeasured Water Flea Crustaceans; Standard Test Species Static Unmeasured Crustaceans; Standard Test Species Static Unmeasured Water Flea Crustaceans; Standard Test Species Static Unmeasured Crustaceans; Standard Test Species Renewal Unmeasured Crustaceans;	741952	Ī	Sodium chloride (NaCl)	Ceriodaphnia dubia			Static		Fresh water Lab	Lab
7647145 Sodium chloride (NaCl)  7647146 Sodium chloride (NaCl)  7647147 Sodium chloride (NaCl)  7647148 Sodium chloride (NaCl)  7647149 Sodium chloride (NaCl)  7647149 Sodium chloride (NaCl)  7647145 Sodium chloride (NaCl)  7647145 Sodium chloride (NaCl)  7647146 Sodium chloride (NaCl)  7647147 Sodium chloride (NaCl)  7647148 Sodium chloride (NaCl)  7647149 Sodium chloride (NaCl)	741944	7	Sodium chloride (NaCl)	Ceriodaphnia dubia			Static	1	Fresh water	Lab
7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   Variation   Variatio	742892	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia			Static		Fresh water Lab	Lab
T647145 Sodium chloride (NaCl)  Ceriodaphnia dubia  Water Flea  Crustaceans; Standard Test Species  Static  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Static  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Renewal  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Unmeasured  Unmeasured  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured	741957	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia			Static	Ī	Fresh water Lab	Lab
7647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   Unmeasured   T647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   Unmeasu	741949	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia		1	Static		Fresh water	Lab
T647145   Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   T647145   Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   T647145   Sodium chloride (NaCl)   Ceriodaphnia dubia   Water Flea   Crustaceans; Standard Test Species   Static   Unmeasured   Unmeasure	742563	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia	E		Static		Fresh water Lab	Lab
7647145 Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Johnneasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Unmeasured  Total Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Unmeasured  Unmeasured  Unmeasured	741945	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia		- 1	Static		Fresh water	Lab
7647145 Sodium chloride (NaCl)  Ceriodophnia dubia  Ceriodophnia dubia  Ceriodophnia dubia  Todes (NaCl)  Ceriodophnia dubia  Coustaceans; Standard Test Species  Renewal  Unmeasured  Unmeasured  Coustaceans; Standard Test Species  Coustaceans; Standard Test	741959	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia			Static		Fresh water Lab	Lab
7647145 Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Water Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Valer Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Valer Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Valer Flea  Crustaceans; Standard Test Species  Renewal  Unmeasured  Crustaceans; Standard Test Species  Renewal  Unmeasured  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Crustaceans; Standard Test Species  Unmeasured  Unmeasured	2152727	7647145	Sodium chloride (NaCl)	Ceriodaphnia dubia		_	Renewal		Fresh water I	Lab
7647145 Sodium chloride (NaCl)  Ceriodophnia dubia  Water Flea  Crustaceans; Standard Test Species Renewal  Unmeasured  Valet Flea  Crustaceans; Standard Test Species Renewal  Unmeasured  Crustaceans; Standard Test Species Renewal  Unmeasured  Valet Flea  Crustaceans; Standard Test Species Renewal  Unmeasured	767972		Sodium chloride (NaCl)	Ceriodaphnia dubia		1	Renewal			Lab
7647145 Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chloride (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Water Flea   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Crustaceans; Standard Test Species   Renewal   Unmeasured   7647145   Sodium chlorida (NaCl)   Ceriodophnia dubia   Crustaceans; Standard Test Species   Renewal   Unmeasured   Crustaceans; Standard Test Species   Crustaceans; Standard Test Species   Crustaceans; Standard Test Species   Cru	767884		Sodium chloride (NaCl)	Ceriodaphnia dubia			Renewal			Lab
7627145 (Addium chloride (NaCi)) Ceriodonhio dishiri Water Elaa Crustanaana Taet Spacias Ranawal Illomassured	766861		Sodium chloride (NaCl)	Ceriodaphnia dubia		-	Renewal		Fresh water	Lab
	767902		Sodium chloride (NaCl)	Ceriodaphnia dubia		-	Renewal		Fresh water Lab	- Fac

	Mortality	Mortality	50	121/15				2000		
MSMT/TLM///	Mortality	Mortality	LC50	Day(s)	NR	NR	4	Not reported		126052
MISMIT/ILMI///		Mortality	LC20	Day(s)		NR	. 4	Not reported		126054
		Mortality	NR-LETH				2	Not reported		740777
	Mortality	Mortality	NR-LETH			NR	2	Not reported		740765
ENDPT/TLM/	Mortality EN	Mortality	LC50	Day(s)		NR	4	Not reported		2063735
ENDPT/TLM/	Mortality EN	Mortality	LC50	Day(s)	NR	NR	1	Not reported		2063726
ENDPT/TLM/	ĺ	Mortality	LC50			NR	2	Not reported		2063733
	Mortality	Mortality	NR-LETH			NR	1.5	Not reported		740761
	Mortality	Mortality	NR-LETH	Day(s)		NR	1.5	Not reported		2153697
	Mortality	Mortality	NR-LETH	Day(s)		NR	2	Not reported		2153700
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	2	Not reported		740732
	Mortality	Mortality	NR-LETH	Day(s)		NR	2	Not reported		740760
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1.125	Not reported		740759
	Mortality	Mortality	NR-LETH	Day(s)	NR	1.25 NR	1.25	Not reported		2153676
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	2	Not reported		2153763
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768398
	Mortality	Mortality	NR-ZERO	Day(s)		NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768460
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768402
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	10	Not reported		2152736
	Mortality	Mortality	NR-ZERO	Day(s)		NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768462
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768457
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1	Not reported	768396 FED, MODERATELY HARD RECONSTITUTED WATER	768396
	Mortality	Mortality	NR-LETH	Day(s)		NR	1	Not reported		768399
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported		119478
	Mortality	Mortality	TC50		NR	NR	1	Not reported		32466
	Mortality	Mortality	TC50		NR	NR	1	Not reported		160621
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported		742079
MSMT/TLM///	Mortality MS	Mortality	LCS0	Day(s)	NR	NR	3	Not reported		18687
MSMT/TLM///	Mortality Ms	Mortality	LC50	Day(s)	NR	NR	1.0417	Not reported		18683
MSMT/TLM///	Mortality Ms	Mortality	LC50	Day(s)	NR		2.0833 NR	Not reported		18684
MSMT/TLM///	Mortality Ms	Mortality	LC50	Day(s)			4.2	Not reported		18688
MSMT/TLM///	Mortality Ms	Mortality	LC50	Day(s)	NR	NR	1	Not reported	5	18685
MSMT/TLM///	Mortality MS	Mortality	LC50	Day(s)	NR		2	Not reported		18686
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported		2152737
	Mortality	Mortality	LC50	Day(s)	NR	NR	10	Not reported		2152738
	Mortality	Mortality	LC50	Day(s)			N	Not reported		160622
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	CA, MG, NA+K, SO4, CL2	44241
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported		742988
	Mortality	Mortality	NR-ZERO	Day(s)		NR		Not reported	UNFED. MODERATELY HARD RECONSTITUTED WATE	768286
	Mortality	Mortality	NR-ZERO	Day(s)		NR	2	Not reported		768181
	Mortality	Mortality	NR-ZERO	Dav(s)		NR		Not reported	UNFED. MODERATELY HARD RECONSTITUTED WATE	768183
	Mortality	Mortality	NR-ZERO	Day(s)				Not reported	FED. MODERATELY HARD RECONSTITUTED WATER	768283
	Mortality	Mortality	NR-ZERO	Day(s)			N)	Not reported		767342
	Mortality	Mortality	NR-LETH	Day(s)				Not reported	UNFED, MODERATELY HARD RECONSTITUTED WATE	768182
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1	Not reported	UNFED, MODERATELY HARD RECONSTITUTED WATE	768284
	Mortality	Mortality	NR-LETH	Day(s)				Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768180
	Mortality	Mortality	LC50	Day(s)		9 NR	2	Not reported	5	2152765
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported		160620
	Mortality	Mortality	LC50	Day(s)	NR	NR		Not reported		160619
	Mortality	Mortality	LC50	Day(s)	NR	NR		Not reported	4	2152764
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR NR		Not reported		2147187
EE Comment	Effect Measurement	Effect*	Endpoint	Dr	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	er Experimental Design	Result Number
				Observed	Observed	Observed	Observed			

Result Number Experimental Design	Response Site	Observed Observed Duration Duration Mean (Days) Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
	Not reported	1	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
196145	Not reported	4 NR	NR	Day(s)	LC50	Mortality	Survival	
19522 WATER CHEM PARAMETERS RPTD	Not reported	5 NR	NR	Day(s)	LC50	Mortality	Mortality	
	Not reported	4 NR	NR	Day(s)	LC50	Mortality	Mortality	
740778	Not reported	2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
741717	Not reported	1.5 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
2153801	Not reported	1.5 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
18693	Not reported	4 NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
18691	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
18690	Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
18692	Not reported	3 NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
2153798	Not reported	1.5 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
740767	Not reported	2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
184730 RECIRCULATING	Not reported	42 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
99472	Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	
99476	Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	
99478	Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	
61681	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
99477	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
61680	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
160623	Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	
160624	Not reported	2 NR	N. N.	Day(s)	LC50	Mortality	Mortality	
160625	Not reported	, 4 2 2 2 3	2 2	Day(s)	1000	Mortality	Mortality	
741718	Not reported	2 NR	NR N	Day(s)	NR-I FTH	Mortality	Mortality	
2153836	Not reported		NR :	Day(s)	NR-LETH	Mortality	Mortality	
2153835	Not reported	1.125 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
768541 FED, MODERATELY HARD RECONSTITUTED WATER	-	1 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
768539 FED, MODERATELY HARD RECONSTITUTED WATER		1 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
742032	Not reported	4 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
742060 REP 3	Not reported	4 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
742031 REP 7	Not reported	4 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
740738	Not reported	2 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
742933 REP 10	Not reported	4	N.R.	Day(s)	NR-LEIH	Mortality	Mortality	
740779	Not reported	1.125 NR	N. N.	Day(s)	NR-LEIH	Mortality	Mortality	
2152902	Not reported	1 5 NR	N. P.	Day(s)	NR-ZERO	Mortality	Mortality	
740734	Not reported	1 NR	NR.	Dav(s)	NR-ZERO	Mortality	Mortality	
22954	Not reported	30 NR	NR .	Day(s)	LC50	Mortality	Mortality	
22953	Not reported	30 NR	NR	Day(s)	LC50	Mortality	Mortality	
22952	Not reported	4 NR	NR	Day(s)	LC50	Mortality	Mortality	
160513	Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	
160514	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
768178 UNFED, MODERATELY HARD RECONSTITUTED WAT	WAT Not reported	1 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
768176 FED, MODERATELY HARD RECONSTITUTED WATER	ATER Not reported	1 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
768177 FED, MODERATELY HARD RECONSTITUTED WATER	ATER Not reported	2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
768277 FED, MODERATELY HARD RECONSTITUTED WATER		2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
22960 CA, MG, NA+K, SULPHATE, CL2	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
160516	Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
25475	Not reported		Z Z	Day(s)	LCSU	Mortality	Mortality	MSMI/ILM///
254/4	Not reported	1.U41/ NR	Z Z	Day(s)	1050	Mortality	Mortality	MSMT/TIM///
2470	inor ichoicea			- Lallan		(trimer service)	Tanana da maria	100000000000000000000000000000000000000

Result Number	Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
160515	T and the second	Not reported	_	NR		Day(s)	LC50	Mortality	Mortality	ON THE ABOUT THE
	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1	NR		Day(s)	NR-LETH	Mortality	Mortality	
768393 F		Not reported	Ţ.	NR		Day(s)	NR-LETH	Mortality	Mortality	
768501 FI		Not reported	1	NR		Day(s)	NR-LETH	Mortality	Mortality	
768451 F		Not reported	1	NR		Day(s)	NR-LETH	Mortality	Mortality	
768390 FI		Not reported	1	NR		Day(s)	NR-LETH	Mortality	Mortality	
768392 FI	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	2	NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
768395 F.		Not reported	2	NR		Day(s)	NR-ZERO	Mortality	Mortality	
		Not reported	2	NR		Day(s)	NR-ZERO	Mortality	Mortality	
2064007	FED; MODERATELY HAND RECONSTITUTED WATER	Not reported	2	NR NR	NR NR	Day(s)	ICSO INK-ZEKO	Mortality	Mortality	ENDRT/TIM/
2064009		Not reported	4			Day(s)	LC50	Mortality	Mortality	ENDPT/TLM/
2064004		Not reported	1			Day(s)	LC50	Mortality	Mortality	ENDPT/TLM/
22955		Not reported	4	NR	NR	Day(s)	LC50	Mortality	Mortality	
22957		Not reported	30 NR		NR	Day(s)	LC50	Mortality	Mortality	
22956		Not reported	30 NR			Day(s)	LC50	Mortality	Mortality	
160518		Not reported	2	2 NR	ZR Z	Day(s)	1050	Mortality	Mortality	
160519		Not reported	4			Day(s)	LC50	Mortality	Mortality	
2055980 3	2055980 3 RÉPLICATES	Not reported	6			Day(s)	NR-ZERO	Mortality	Mortality	
741753 A	ACUTE STUDY	Not reported	4	4 NR	NR	Day(s)	LC50	Mortality	Mortality	
		Not reported	. 1			Day(s)	NR-ZERO	Mortality	Mortality	
741707 11		Not reported	<u> </u>			Day(s)	NR-ZERO	Mortality	Mortality	
741741 TF	TRIALB	Not reported	1 4	1 AND	NR Z	Day(s)	NR-ZERO	Mortality	Mortality	
741740 TF		Not reported	1			Day(s)	NR-ZERO	Mortality	Mortality	
2455		Not reported	4	NR		Day(s)	LC50	Mortality	Mortality	
12210		Not reported	4			Day(s)	LC50	Mortality		
231580		Not reported	·ω			Day(s)	LC50	Mortality		MSMT/TLM/
231578		Not reported	. 1			Day(s)	LC50	Mortality	Mortality	MSMI/ILM/
231577		Not reported	4	N. N.		Day(s)	TC20	Mortality	Mortality	MSMT/TIM/
231575		Not reported	21		NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231576		Not reported	з			Day(s)	LC50	Mortality		MSMT/TLM/
231579		Not reported	2			Day(s)	LC50	Mortality		MSMT/TLM/
231581		Not reported	3			Day(s)	LC50	Mortality		MSMT/TLM/
231569		Not reported	7		NR	Day(s)	LC50	Mortality		MSMT/TLM/
231571		Not reported	2 NR			Day(s)	TC20	Mortality		MSMT/TIM/
231567		Not reported	5 NR			Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231573		Not reported	4 1		NR [	Day(s)	LC50	Mortality	Ì	MSMT/TLM/
231572		Not reported	3.1			Day(s)	LC50	Mortality		MSMT/TLM/
231563		Not reported	11	NR	NR [	Day(s)	LC50	Mortality		MSMT/TLM/
231566		Not reported	41			Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231570		Not reported	1 NR			Day(s)	LC50	Mortality		MSMT/TLM/
231567		Not reported	u u			Day(s)	LC50	Mortality		MSM1/1LM/
764333		Not reported	1 2	NR NR	NR I	Day(s)	LC20	Mortality	Mortality	KISMI/ILIVI/
764325		Not reported	11				NR-LETH	Mortality	Mortality	
764330		Not reported	17				NR-LETH	Mortality	Mortality	
764326		Not reported	11	NR	NR E	Day(s)	NR-LETH	Mortality	Mortality	
764329		Not reported	1	NR	NR I	Dav(s)	NRLIETH	Mortality	Mortality	

				NAV S			4	VOT TABOUTAGE		196
	Mortality	Mortality	LCSO	Day(s)	No.		4 4	Not reported	106800	100
	Mortality	Mortality	LCSO	Day(s)			2 +	Not reported		106
	Mortality	Mortality	LC50	Day(s)			a 1-	Not reported		106
	Mortality	Mortality	LCSO	Day(s)			4 L	Not reported		106
	Mortality	Mortality	LC50	Day(s)			غر د	Not reported		196
	Mortality	Mortality	LC20	Day(s)			٠ <del>١</del>	Not reported		100
	Mortality	Mortality	LC50	Day(s)			, ,	Not reported		106
	Mortality	Mortality	LC50	Day(s)			, <sub>1</sub>	Not reported		196
	Mortality	Mortality	LC50	Day(s)			2	Not reported		196
	Mortality	Mortality	LC50	Day(s)			1	Not reported		196
	Mortality	Mortality	LC50	Day(s)		1 NR	1	Not reported		196
	Mortality	Mortality	LC50	Day(s)	NR [	NR	4	Not reported		196
	Mortality	Mortality	LC50	Day(s)		2 NR	2	Not reported	196811	196
	Mortality	Mortality	LC50	Day(s)		4 NR	4	Not reported		196
	Mortality	Mortality	LC50	Day(s)	NR	NR	1	Not reported	196790	196
	Mortality	Mortality	LC50	Day(s)		4 NR	4	Not reported		196
	Mortality	Mortality	LC50	Day(s)	NR	2 NR	2	Not reported		196
	Mortality	Mortality	LC50	Day(s)	NR I	4 NR	4	Not reported	196836	196
	Mortality	Mortality	LC50	Day(s)		2 NR	2	Not reported		196
	Mortality	Mortality	LC50	Day(s)	NR	NR	4	Not reported	196808	196
	Mortality	Mortality	LC50	Day(s)	NR	NR	1	Not reported	196834	196
	Mortality	Mortality	LC50	Day(s)		2 NR	2	Not reported	196775	196
	Mortality	Mortality	LC50	Day(s)		NR	4	Not reported	196784	196
	Mortality	Mortality	LC50	Day(s)			4	Not reported	196788	196
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	196803	196
	Mortality	Mortality	LC50	Day(s)		NR	1	Not reported	196826	196
	Mortality	Mortality	LC50	Day(s)			10	Not reported		75
	Mortality	Mortality	LC50	Day(s)			4	Not reported		167
	Mortality	Mortality	LC50	Day(s)			4	Not reported		196
	Mortality	Mortality	LC50	Dav(s)			4	Not reported		196
	Mortality	Mortality	LC50	Dav(s)			2	Not reported	196795	196
	Mortality	Mortality	1050	Dav(s)			2	Not reported	196783	196
	Mortality	Mortality	1050	Day(s)			4	Not reported	196776	196
	Mortality	Mortality	1050	Dav(s)			2	Not reported	196791	196
	Mortality	Mortality	LC50	Dav(s)		NR	1	Not reported	196814	196
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	196831	196
	Mortality	Mortality	1020	Day(s)			2	Not reported	196799	196
	Mortality	Mortality	NOEL	Day(s)	NR X	NR NR	1	Not reported	196818	196
	Mortality	Mortality	NOEL	Day(s)		Z Z		Not reported	MEDIUM HARD RECONSTITUTED WATER (MHRW)	74,
	Mortality	Mortality	LOEL	Day(s)			4	Not reported		74.
	Mortality	Mortality	LOEL	Day(s)		NR	1	Not reported	MEDIUM HARD RECONSTITUTED WATER (MHRW)	747
	Mortality	Mortality	LC50	Day(s)		NR	14	Not reported	MINERAL WATER	187
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	187906 MINERAL WATER, FED	187
	Mortality	Mortality	LC50	Day(s)	NR	1 NR	1	Not reported	187901 RECON WATER	187
	Mortality	Mortality	LC50	Day(s)	NR	NR	1	Not reported	187902 RECON WATER, FED	187
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	187903 RECON WATER	187
	Mortality	Mortality	LC50	Day(s)		NR	1	Not reported	187905 MINERAL WATER, FED	187
	Mortality	Mortality	NR-LETH	Day(s)			1	Not reported	764481	764
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	1	Not reported	741752 TRIAL A	74:
	Mortality	Mor	LC50	Day(s)			1	Not reported	4 REPLICATES	215
EE Comment	Effect Measurement	Effect*	Endpoint	Duration Units (Days)	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	Number Experimental Design	Result Number
				Observed	Observed	Observed	Observed			

Result Number	Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
196812	-	Not reported	45	NR		Day(s)	LC50	Mortality	Mortality	
196779		Not reported	2 1	NR	NR	Day(s)	LC50	Mortality	Mortality	
196815 196792		Not reported	2 4	NR NR	Z. Z.	Day(s)	1020	Mortality	Mortality	
196835		Not reported	21	NR NR	NR	Day(s)	LC20	Mortality	Mortality	
196820		Not reported	4	NR	NR	Day(s)	LC50	Mortality	Mortality	
196774		Not reported	1 NR	VR	NR	Day(s)	LC50	Mortality	Mortality	
196806		Not reported	1 7	NR	NR	Day(s)	LC50	Mortality	Mortality	
196832		Not reported	4 NR	VR	NR	Day(s)	LC50	Mortality	Mortality	
196840		Not reported	3 NR	VR.	NR	Day(s)	LC50	Mortality	Mortality	
196819		Not reported	2 NR	NR.	NR	Day(s)	LC50	Mortality	Mortality	
196787		Not reported	2 NR	X	N.	Day(s)	LC20	Mortality	Mortality	
196801		Not reported	6 NR	5 5	Z	Day(s)	LETC	Mortality	Mortality	
196817		Not reported	NA VINA	NA NA	20 2	Day(s)	LEIC	Mortality	Mortality	
167863		Not reported	6 NR	NR S		Dav(s)	1FTC	Mortality	Mortality	
196793		Not reported	5 NR	R		Day(s)	LETC	Mortality	Mortality	
196805		Not reported	6 NR	VR.		Day(s)	LETC	Mortality	Mortality	
196777		Not reported	6 1	NR	NR	Day(s)	LETC	Mortality	Mortality	
196785		Not reported	5 7	NR		Day(s)	LETC	Mortality	Mortality	
196797		Not reported	5 7	NR		Day(s)	LETC	Mortality	Mortality	
196825		Not reported	n > 7	No No	NR NR	Day(s)	LETC	Mortality	Mortality	
196789		Not reported	4	NR S		Dav(s)	LETC	Mortality	Mortality	
196821		Not reported	6 NR	R		Day(s)	LETC	Mortality	Mortality	
196833		Not reported	5 7	NR		Day(s)	LETC	Mortality	Mortality	
196829		Not reported	6 7	NR		Day(s)	LETC	Mortality	Mortality	
196781		Not reported	6 NR	5 5		Day(s)	LETC	Mortality	Mortality	
C113C		Not reported	<u> </u>			Day(s)	LEIC	Mortality	Mortality	MSNAT/TINA///
241686 VCIITE	ITE	Not reported	2 NR		NR	Day(s)	1050	Mortality	Mortality	INISINITY I CINUTY
2152825 4 R	4 REPLICATES	Not reported	2 7			Dav(s)	LC50	Mortality	Mortality	
2075467		Not reported	2 7		NR	Day(s)	LC50	Mortality	Mortality	
2153271 HAI	HARDNESS OF 40 MG/L CACO3	Not reported	7 7			Day(s)	LC50	Mortality	Mortality	
2153268 HAI	HARDNESS OF 20 MG/L CACO3	Not reported	7 7			Day(s)	LC50	Mortality	Mortality	
2075442		Not reported	27			Day(s)	LC50	Mortality	Mortality	
2075434		Not reported	27	NR.		Day(s)	LC50	Mortality	Mortality	
2075410		Not reported	2 NR	NR	ZR	Day(s)	1050	Mortality	Mortality	
2075471		Not reported	2 1			Day(s)	LC50	Mortality	Mortality	
2153274 HAF	HARDNESS OF 80 MG/L CACO3	Not reported	7 NR			Day(s)	LC50	Mortality	Mortality	
2152724		Not reported	9 NR	JR:	NR	Day(s)	LC50	Mortality	Mortality	
2152723		Not reported	2 7	NR		Day(s)	LC50	Mortality	Mortality	
2152874 HAR	HARDNESS OF 10 MG/L CACO3	Not reported	7 7	NR		Day(s)	LC50	Mortality	Mortality	
2153280 HAR	HARDNESS OF 320 MG/L CACOS	Not reported	7 NR	NR Z	NR P	Day(s)	LC20	Mortality	Mortality	
2075466		Not reported	2 NR	D S		Day(s)	LC50	Mortality	Mortality	
2075469		Not reported	2 NR			Day(s)	LC50	Mortality	Mortality	
2075470		Not reported	2 NR			Day(s)	LC50	Mortality	Mortality	
2074934		Not reported	2 NR			Day(s)	LC50	Mortality	Mortality	
2075426		Not reported	2 NR		N. N.	Day(s)	LC20	Mortality	Mortality	
The second second second		The second secon				100		A TO A COLOR OF A	THE PERSON NAMED IN COLUMN TWO IS NOT THE OWNER.	

		4000	· · · · · · · · · · · · · · · · · · ·	-0-0					The state of the s	0.000	
Chapter   Chap		Survival		LOFC	Dav(s)			7	Not reported	251009 TEST MATERIAL 3	2
Departmental Design		Sunival	Mortality	LOEC	Day(s)			7	Not reported	250999 TEST MATERIAL 1	2
Page-parameter   Page-parameter   Page-page-page-page-page-page-page-page-p		Survival	Mortality	LOEC	Day(s)			7	Not reported	251001 TEST MATERIAL 3	2
Page-internal Besign		Survival	Ì	LOEC	Day(s)			7	Not reported	251003 TEST MATERIAL 3	2
Experimental Design		Survival		LOEC	Day(s)			7	Not reported	251007 TEST MATERIAL 3	1
December		Survival	Mortality	LOEC	Day(s)	VR		7	Not reported	767854	7
District		Survival	Mortality	LOEC	Day(s)			7	Not reported	767848	7
District		Survival	Mortality	LOEC	Day(s)	VR		7	Not reported	767862	7
		Survival	Mortality	LOEC	Day(s)			7	Not reported	767864	7
Experimental Design         Experimental Design         Observed Duration Duration Duration May Duration Ma		Survival	Mortality	LOEC	Day(s)			7	Not reported	767975	7
District		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767881	2
Description		Survival		LOEC	Day(s)	NR		7	Not reported	767887	e.i
Description		Survival		LOEC	Day(s)			7	Not reported	767953	
Charment   Cheaned   Che		Survival		LOEC	Day(s)	NR	Ĭ	7	Not reported	767846	
Experimental Design		Survival		LOEC	Day(s)	NR		7	Not reported	767969	
Distance    Dist		Survival		LOEC	Day(s)	NR		7	Not reported	767856	
Disease   Dise		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767889	0.0
Disease   Disease   Disease   Disease   Disease   Disease   Effect   Experimental Design   Part   Disease   Diseas		Survival		LOEC	Day(s)	NR		7	Not reported	768866	
Distance		Survival		LOEC	Day(s)	NR		7	Not reported	/668/2	
Description		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	/6/96/	
Description		Survival		LOEC	Day(s)	NR		7	Not reported	/6/8/0	
District		Survival		LOEC	Day(s)	NR		7	Not reported	767959	
Experimental Design		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767872	
Experimental Design		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767895	
Diseased		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767879	24
Diserved		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767961	
Diserved   Day(5)   LC50   Mortality   Mortality   Mortality   Mortality   Diserved		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	767897	200
Dissave		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	766864	
Diserved   Direction Max   Direction M		Survival	Mortality	LOEC	Day(s)	NR		7	Not reported		
Doserved		Survival		LOEC	Day(s)	NR		7	Not reported		
Diserved		Survival		LC50	Day(s)	NR		00	Not reported	207717	
Diserved   Diserve		Survival		LC50	Day(s)	NR		2		207716	
Diserved   Diration Max   Diration Max   Diration Max   Direct   Effect   Effect   Effect   Direct		Mortality		LC50	Day(s)	NR		2		741288 REFERENCE TOXICANT TEST, VALUE DETERMIN	
Duration		Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	160320	
Experimental Design		Mortality	Mortality	LC50	Day(s)	NR		1	Not reported		
Diserved		Mortality		LC50	Day(s)	NR		7	Not reported	LAB D,	
Experimental Design		Mortality		LC50	Day(s)	NR		7	Not reported	120825 LAB D. TEST 1. TEST MATERIAL 1	
Cobserved   Cobs		Mortality		LC50	Day(s)	NR		7	Not reported	120828 LAB H, TEST 1, TEST MATERIAL 1	
Experimental Design		Mortality		LC50	Day(s)	NR		7	Not reported	120839 LAB D, TEST 2, TEST MATERIAL 3	
Cobserved   Cobs		Mortality		LC50	Day(s)	NR		7	Not reported	120824 LAB A, TEST MATERIAL 1	
Cobserved   Cobs		Mortality		LC50	Day(s)	NR		7	Not reported	120841 LAB G, TEST 1, TEST MATERIAL 3	
Column		Mortality		LC50	Day(s)	NR		7	Not reported	LAB G,	
Experimental Design         Response Site         Mean (Days)         Min (Days)         Unration (Days)         Duration Max (Days)         Duration Max (Days)         Duration Units         Effect*         Measurement           LAB H, TEST 1, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality           LAB H, TEST 2, TEST MATERIAL 1         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB H, TEST 2, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB H, TEST 2, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB H, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB H, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB G, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB J, TES		Mortality		LC50	Day(s)	NR		7	Not reported		
Experimental Design         Response Site         Observed Duration Duration Duration Max         Observed Duration Max         Observed Duration Max         Duration Units         Effect*         Measurement           LAB H, TEST 1, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality           LAB F, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB F, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB F, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB G, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB G, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB G, TEST MATERIAL 3         Not reported         7 NR         NR         Day(s)         LC50         Mortality         Mortality           LAB G, TEST MATERIAL 1         Not reported		Mortality		LC50	Dav(s)	NR		7	Not reported		
Contain   Cont		Mortality		LC50	Day(s)	NR	NR	7	Not reported		
Cobserved   Cobs		Mortality		LC50	Day(s)	NR	NR	9	Not reported	741547 THREE BROOD TEST	
Coloration   Col		Mortality		LC50	Day(s)	NR		7	Not reported	LAB G,	
Observed		Mortality		LC50	Day(s)	NR S		7	Not reported	120840 LAB F, TEST MATERIAL 3	
Observed Obs		Mortality		LCSO	Day(s)	NA		7	Not reported	120844 I AB H TEST 3 TEST MATERIAL 1	
Observed Obs		Mostality		LCSO	Day(s)	20 27		7 /	Not reported	AB II,	
Observed Observed Observed Observed Observed Observed Steeperimental Design Response Site Mean (Days) Min (Days) (Days) (Days) Endpoint Effect* Measurement		Modelity		LCSU	Day(s)	25		7	Not reported		7
Observed Observed Observed Observed  Duration Duration Max Duration Units  Effect	EE Comment	Measurement	DOINT	End	(Days)		in (Days)	١,٠	Response Site		Result
		Effect		n	Observed  Duration Unit	Observed Duration Max		Observed Duration			

Result Number	· Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
250998	TEST MATERIA	Not reported	7	NR		Day(s)	LOEC	Mortality	Survival	
251005		Not reported	7	NR	NR	Day(s)	LOEC	Mortality	Survival	
741687	CHRONIC -3 BROOD STDY	Not reported	7	NR NR		Day(s)	NOEC	Mortality	Survival	
767904		Not reported	7	NR NR	N. R.	Day(s)	NOEC	Mortality	Survival	
766865		Not reported	7	NR :		Day(s)	NOEC	Mortality	Survival	
767898		Not reported	7	NR		Day(s)	NOEC	Mortality	Survival	
767962		Not reported	7	NR	NR	Day(s)	NOEC	Mortality	Survival	
767880		Not reported	7	7 NR		Day(s)	NOEC	Mortality	Survival	
767896		Not reported	7	7 NR		Day(s)	NOEC	Mortality	Survival	
767960		Not reported	7	7 NR	Z.R	Day(s)	NOEC	Mortality	Survival	
767871		Not reported	7	NR		Day(s)	NOEC	Mortality	Survival	
767968		Not reported	7	7 NR		Day(s)	NOEC	Mortality	Survival	
766873		Not reported	7	7 NR	NR	Day(s)	NOEC	Mortality	Survival	
766867		Not reported	7	7 NR	NR	Day(s)	NOEC	Mortality	Survival	
767890		Not reported	7	NR			NOEC	Mortality	Survival	
767970		Not reported	7	7 NR	N Z		NOEC	Mortality	Survival	
767847		Not reported	7	NR :		Day(s)	NOEC	Mortality	Survival	
767954		Not reported	7	NR			NOEC	Mortality	Survival	
767888		Not reported	7	NR			NOEC	Mortality	Survival	
767976		Not reported	7	NR NR	Z	Day(s)	NOEC	Mortality	Survival	
767865		Not reported	7	NR			NOEC	Mortality	Survival	
767863		Not reported	7	NR	NR		NOEC	Mortality	Survival	
767849		Not reported	7			Day(s)	NOEC	Mortality	Survival	
250997		Not reported	7			Day(s)	NOEC	Mortality	Survival	MSMT/FOUND BETWEEN 10 AND 20% OF
251000	TEST MATERIAL 3	Not reported	7	NR NR	NR	Day(s)	NOEC	Mortality	Survival	MSMT/FOUND IN ~10% OF LABS///
251004		Not reported	7			Dav(s)	NOEC	Mortality	Survival	MSMT/FOUND IN ~10% OF LABS///
		Not reported	7			Day(s)	NOEC	Mortality	Survival	MSMT/FOUND IN ~10% OF LABS///
251008	251008 TEST MATERIAL 3	Not reported	7	NR	NR	Day(s)	NOEC	Mortality	Survival	MSMT/FOUND BETWEEN 50 AND 60% OF
	TEST MATERIAL 3	Not reported	7			Day(s)	NOEC	Mortality	Survival	MSMT/FOUND IN ~10% OF LABS///
741699	741699 CHRONIC -3 BROOD STDY	Not reported	Z NR		NR NR	Day(s)	NR-LETH	Mortality	Mortality	
741671		Not reported	14 NR			Day(s)	NR-LETH	Mortality	Mortality	
768272	768272 UNFED, MODERATELY HARD RECONSTITUTED WAT Not reported	ATI Not reported	1 NR				NR-LETH	Mortality	Mortality	
768340	FED, MODERATELY HARD RECONSTITUTED WATER	R Not reported	1 NR				NR-LETH	Mortality	Mortality	
768173	768268 UNFED, MODERATELY HARD RECONSTITUTED WATENOT reported	All Not reported	1 NR			Day(s)	NR-LETH	Mortality	Mortality	
207718		Not reported	2 NR		NR N	Day(s)	NR-ZERO	Mortality	Mortality	
207719		Not reported	8 NR				NR-ZERO	Mortality	Mortality	
741660	CHRONIC -3 BROOD STDY	Not reported	7 NR		NR		NR-ZERO	Mortality	Mortality	
207721		Not reported	8			Day(s)	NR-ZERO	Mortality	Mortality	MSMT/NOEL///
2152/22	THREE BROOD TEST	Not reported	9 NR		N. R.	Day(s)	NR-ZERO	Mortality	Mortality	
768266	UNFED, MODERATELY HARD RECONSTITUTED WAT Not reported	AT(Not reported				Day(s)	NR-ZERO	Mortality	Mortality	
768264	FED, MODERATELY HARD RECONSTITUTED WATER	R Not reported	2 1	NR	NR		NR-ZERO	Mortality	Mortality	
768175	768175 UNFED, MODERATELY HARD RECONSTITUTED WAT		2 7				NR-ZERO	Mortality	Mortality	
741635	741635	Not reported	27				LC50	Mortality	Mortality	
CCOT+1/		Morrieborren	4	NN	NN I	Day(s)	LCSU	MOLITAILLY	INIOITAIILY	

Result Number	Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
741678		Not reported	4	NR		Day(s)	NR-ZERO	Mortality	Mortality	
2060209 4	2060209 4 REPLICATES	Not reported	2	NR		Day(s)	EC01	Mortality	Survival	
2060210 4	2060210 4 REPLICATES	Not reported	2 1	NR	NR	Day(s)	EC05	Mortality	Survival	
2060211 4	4 REPLICATES	Not reported	21	NR		Day(s)	EC10	Mortality	Survival	
2060212 4	2060212 4 REPLICATES	Not reported	21	NR	NR	Day(s)	EC15	Mortality	Survival	
2060213 4	4 REPLICATES	Not reported	2 1	NR	NR	Day(s)	EC20	Mortality	Survival	
2060214 4	4 REPLICATES	Not reported	2 1	NR	NR	Day(s)	EC25	Mortality	Survival	
2060216 4	2060216 4 REPLICATES	Not reported	21	NR		Day(s)	EC40	Mortality	Survival	
2060217 4	20602174 REPLICATES	Not reported	2	NR R	NR NR	Day(s)	FC75	Mortality	Survival	
2060220 4	2060220 4 REPLICATES	Not reported	21			Day(s)	EC80	Mortality	Survival	
2060221 4	4 REPLICATES	Not reported	21			Day(s)	EC85	Mortality	Survival	
2060222 4	4 REPLICATES	Not reported	21			Day(s)	EC90	Mortality	Survival	
2060223 4	4 REPLICATES	Not reported	2 1			Day(s)	EC95	Mortality	Survival	
2060225 4	2060225 4 REPLICATES 2152838 A REBLICATES	Not reported	A NR		N.R.	Day(s)	FC99	Mortality	Mortality	
2060201 4	4 REPLICATES	Not reported	2 7			Dav(s)	LC50	Mortality	Mortality	
2060236 4	2060236 4 REPLICATES	Not reported	2 NR			Day(s)	LOEC	Mortality	Survival	
2060237 4	4 REPLICATES	Not reported	2 NR		NR	Day(s)	MATC	Mortality	Survival	
2060228 4	4 REPLICATES	Not reported	2 NR			Day(s)	NOEC	Mortality	Survival	
20602474	A REPLICATES	Not reported	2 NR		NR NR	Day(s)	NR-LETH	Mortality	Mortality	MSMT/REPLICATE D/
	4 REPLICATES	Not reported				Day(s)	NR-LETH	Mortality	Mortality	MSMT/REPLICATE B/
	4 REPLICATES	Not reported	1 7			Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE B/
2060270 4	4 REPLICATES	Not reported	1 7			Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE C/
2211037 S	SAND AND SILICA SEDIMENT, 2 REPLICATES	Not reported				Day(s)	NR-ZERO	Mortality	Mortality	
741489 FI	FIBERGLASS TANKS FILLED WITH 20L OF TEST SOLU	Not reported	2 NR			Day(s)	LC50	Mortality	Mortality	
2162290 3	3 REPLICATES	Not reported	1 NR		NR S	Day(s)	LC50	Mortality	Mortality	
2211025		Not reported	4 NR				NR-LETH	Mortality	Mortality	
2211024		Not reported	4 NR				NR-ZERO	Mortality	Mortality	
16495		Not reported	2 7			Day(s)	LC50	Mortality	Mortality	
26114		Not reported	2 7			Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
2152587		Not reported	11 NR		ZR ZR	Day(s)	TD50	Mortality	Mortality	INDIVITY I FINITY
2152588		Not reported	11 NR				LD99	Mortality	Mortality	
2152592		Not reported	11 NR				NR-LETH	Mortality	Mortality	
2060148 2	2 REPLICATES	Not reported	4 7		NR	Day(s)	LC50	Mortality	Mortality	MSMT/TRIM LEVEL 0.0%/
2060157 2	2060157 2 REPLICATES	Not reported	4 NR				LC50	Mortality	Mortality	MSMT/TRIM LEVEL 5.0%/
2060161 2	2060161 2 REPLICATES	Not reported	4 NR		Z <sub>R</sub>	Day(s)	1020	Mortality	Mortality	MSMT/TRIM LEVEL 20.0%/
2060198 2	2 REPLICATES	Not reported	1 NR				NR-LETH	Mortality	Mortality	MSMT/REPLICATE 1/
2060191 2	2 REPLICATES	Not reported	1 NR				NR-LETH	Mortality	Mortality	MSMT/REPLICATE 2/
2060190 2	2060190 2 REPLICATES	Not reported	4 NR			Day(s)		Mortality	Mortality	MSMT/REPLICATE 2/
	2 REPLICATES	Not reported	4 NR				NR-ZERO	Mortality	Mortality	MSMT/REPLICATE 1/
742566 At	ACUTE	Not reported	2 NR				LC50	Mortality	Mortality	
7/1600 (1	741698 CHRONIC 3 BROOD STDY	Not reported	10 NR				LOEC	Mortality	Survival	
	10 REPLICATES	Not reported	21 NR		NR S	Day(s)		Mortality	Survival	
		Not reported	4.2 NR				LC50	Mortality	Mortality	MSMT/TLM///
18463		Not reported	1 NR		NR I	Day(s)		Mortality	Mortality	MSMT/TLM///
18461		Not reported	2.0833 NR		NR I	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///

	Mortality	2001	NB 7500	Day/6)		NR		Not reported		742068
	INIOI Lairty	INIOLEGIILA	IAIV-CEIVO				7	Not reported		
	Mortality	Mortality	NIB ZEBO			ND	2 1	Not reported		747976
	Mortality	Mortality	NR-ZERO			NR	2	Not reported		742931
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	Not reported		742069
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	Not reported	RANGEFINDING ACUTE	2210505
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	_	FED, MODERATELY HARD RECONSTITUTED WATER	768526
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768488
	Mortality	Mortality	NR-ZERO		NR	NR	2	R Not reported	FED,	
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	Not reported	2210504 DEFINITIVE ACUTE, 4 REPLICATES	2210504
	Mortality	Mortality	NR-ZERO	Day(s)	NR C	NR	2	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768530
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2			
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768450
	Mortality	Mortality	NR-ZERO	Day(s)	NR	10 NR	10	Not reported	THREE BROOD TEST	741531
	Mortality	Mortality	NR-ZERO	Day(s)		2 NR	2	R Not reported		768498
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	2	R Not reported		768431
	Mortality	Mortality	NR-ZERO	Day(s)		2 NR	2			
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	2	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768533
	Mortality	Mortality	NR-ZERO	Day(s)	NR E	2 NR	2	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768428
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	10 NR	10	Not reported		2152707
	Mortality	Mortality	NR-ZERO	Day(s)	NR D	NR	21	Not reported	2 10 REPLICATES	2164732
	Mortality	Mortality	NR-LETH	Day(s)	NR L	NR	1	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768486
	Mortality	Mortality	NR-LETH	Day(s)	NR E	1 NR	1			768521
	Mortality	Mortality	NR-LETH	Day(s)	NR E	NR	1	R Not reported	7 FED, MODERATELY HARD RECONSTITUTED WATER	768467
	Mortality	Mortality	NR-LETH	Day(s)	NR E	NR	1	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768499
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1			
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1	R Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768496
	Mortality	Mortality	NR-LETH	Day(s)	NR [	NR	1		1 FED, MODERATELY HARD RECONSTITUTED WATER	768531
	Mortality	Mortality	NR-LETH	Day(s)		NR	1		768475 FED, MODERATELY HARD RECONSTITUTED WATER	768475
	Mortality	Mortality	NR-LETH	Day(s)	NR [	NR	1	_	9 FED, MODERATELY HARD RECONSTITUTED WATER	768489
	Mortality	Mortality	NR-LETH	Day(s)		NR	1	100		768527
	Mortality	Mortality	NR-LETH	Day(s)		NR	I	-		768426
	Mortality	Mortality	NR-LETH	Day(s)		NR	21	Not reported	1 10 REPLICATES	
	Mortality	Mortality	LC50	Day(s)			14	Not reported	۵	31699
		Mortality	LC50	Dav(s)		NR	2	Not reported	ñ	119469
MSMT/TLM///		Mortality	LC50	Dav(s)		NR	1.0417	Not reported	3	18460
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	$\rightarrow$	742019
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		
	Mortality	Mortality	LC50	Day(s)		N <sub>R</sub>	2	Not reported	2 CA:MG MOLAR RATIO 1.8	
	Mortality	Mortality	LC50	Day(s)		1 NR	1	Not reported	.1	160321
	Mortality	Mortality	1050	Day(s)		10 NR	10	Not reported	9 THREE BROOD TEST	742529
	Mortality	Mortality	1050	Day(s)			2 1	Not reported	2	160322
	Mortality	Mortality	1050	Dav(s)		NR	2	Not reported	7	144222
	Mortality	Mortality	LC50	Dav(s)			2	Not reported	9	144219
	Mortality	Mortality	LC50	Day(s)			. 2	Not reported	00	144218
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	0	144220
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	1	144221
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	8	2152708
	Mortality	Mortality	LC50	Day(s)	NR	NR	10	Not reported	9	2152709
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	9	759179
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	6 4 REPLICATES	2152826
MSMT/TLM///		Mortality	LC50	Day(s)	NR	NR	2	Not reported		18464
EE Comment	Effect Measurement	t Effect*	Endpoint	Duration Units (Days)	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	er Experimental Design	Result Number
				Observed	Observed	Observed	Observed			

Result Number Experimental Design	Response Site	Observed Observed Duration Duration Mean (Days) Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
	Not reported	12		Day(s)	NR-ZERO	Mortality	Mortality	יייייייייייייייייייייייייייייייייייייי
2224820 3 REPLICATES	Not reported	2 NR	NR NR	Day(s)	EC50	Mortality	Survival	MSMT/BASED ON FOOT MOVEMENT/
231519	Not reported	2 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231583	Not reported	7 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231522	Not reported	5 NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231584	Not reported	10.875 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231582	Not reported	6 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TIM/
231521	Not reported	1 NR	NR X	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231520	Not reported	3 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
203677	Not reported	4 NR		Day(s)	LC50	Mortality	Survival	9
203678	Not reported	4 NR		Day(s)	NR-LETH	Mortality	Mortality	
2152829 4 REPLICATES	Not reported	4 NR		Day(s)	LC50	Mortality	Mortality	ASSAT/TBINA LEVEL E OO/
2060288 4 REPLICATES	Not reported	4 NR		Day(s)	LC50	Mortality	Mortality	MISMIT/TRIMITENEL 10 08/
2060289 4 REPLICATES	Not reported	4 NR	N X	Day(s)	LC20	Mortality	Mortality	MSMT/TRIM LEVEL 20.0%/
2060290 4 REPLICATES	Not reported	4 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TRIM LEVEL 0.0%/
2060292 4 REPLICATES	Not reported	4 NR		Day(s)	LOEC	Mortality	Survival	
2060293 4 REPLICATES	Not reported	4 NR	NR	Day(s)	MATC	Mortality	Survival	
2060291 4 REPLICATES	Not reported	4 NR		Day(s)	NOEC	Mortality	Survival	MSMT/REDITCATE C/
2060317 4 REPLICATES	Not reported	1 NR	NR R	Day(s)	NR-LETH	Mortality	Mortality	MSMT/REPLICATE D/
206030714 REPLICATES	Not reported	1 NR		Day(s)	NR-LETH	Mortality	Mortality	MSMT/REPLICATE B/
2060306 4 REPLICATES	Not reported	1 NR	NR	Day(s)	NR-LETH	Mortality	Mortality	MSMT/REPLICATE A/
2060315 4 REPLICATES	Not reported	4 NR		Day(s)	NR-ZERO	Mortality	Mortality	MSM1/REPLICATE C/
2060319 4 REPLICATES	Not reported	u I NR		Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE A/
2060302 4 REPLICATES	Not reported	N NR	NR N	Dav(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE B/
18185	Not reported	1 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
18186	Not reported	2 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
764505	Not reported	1 NR		Day(s)	NR-LETH	Mortality	Mortality	
741739 TRIALB	Not reported	1 NR		Day(s)	NR-ZERO	Mortality	Mortality	
2164606 FALL	Not reported	14 NR		Day(s)	LOEC	Mortality	Survivorship	
	Not reported	14 NR		Day(s)	LOEC	Mortality	Survivorship	
2164592 WINTER	Not reported	14 NR		Day(s)	LOEC	Mortality	Survivorship	1
	Not reported	14 NR	N Z	Day(s)	LOEC	Mortality	Survivorship	
2164603 SPRING	Not reported	14 NR		Day(s)	LOEC	Mortality	Survivorship	
2164595 WINTER	Not reported	14 NR		Day(s)	LOEC	Mortality	Survivorship	1
2164605 FALL	Not reported	14 NR		Day(s)	NOEC	Mortality	Survivorship	
	Not reported	14 NR		Day(s)		Mortality	Survivorship	
2164591 WINTER	Not reported	14 NR		Day(s)		Mortality	Survivorship	
	Not reported	14 NR		Day(s)		Mortality	Survivorship	
	Not reported	14 NR		Day(s)		Mortality	Survivorship	
2164602 SPRING	Not reported	14 NR		Day(s)	NOEC	Mortality	Survivorship	
2164594 WINTER 802/193 2 REDITCATES	Not reported	4 NR	NR N	Day(s)	ICO	Mortality	Mortality	
	Not reported	4 NR		Day(s)	LC0	Mortality	Mortality	
802492 2 REPLICATES	Not reported	3 NR	NR	Day(s)		Mortality	Mortality	
802504 2 REPLICATES	Not reported	3 NR		Day(s)	LC0	Mortality	Mortality	
	Not reported	1 NR	NR	Dav(s)	100	Mortality	Mortality	

802506 2 REPLICATES 802491 2 REPLICATES 802494 2 REPLICATES 802495 2 REPLICATES 802495 2 REPLICATES 802500 2 REPLICATES 802501 2 REPLICATES 802501 2 REPLICATES 802502 2 REPLICATES 802502 2 REPLICATES 802502 2 REPLICATES 802503 2 REPLICATES 802503 3 REPLICATES 6170 6174 6169 6173 6167 6173 6168 2075478 116494 2060327 3 REPLICATES 2060327 3 REPLICATES	Not reported	2 NR 3 NR 1 NR 2 NR 2 NR 3 NR 1 NR 2 NR 2 NR 3 NR 4 NR 4 NR 3 NR 1 NR 4 NR 4 NR 4 NR 4 NR 4 NR 4 NR 6 NR 6 NR		Day(s)	1020   1020	Mortality	Mortality	MSMT/TRIM LEVEL 10.0%/ MSMT/TRIM LEVEL 0.0%/ MSMT/TRIM LEVEL 5.0%/ MSMT/TRIM LEVEL 5.0%/
2060327 3 REPLICATES 2060325 3 REPLICATES 2060326 3 REPLICATES	Not reported Not reported	4 NF		Day(s) Day(s)	LC50	Mortality Mortality	Mortality Mortality	MSMT/TRIM LEVEL 10.0%/ MSMT/TRIM LEVEL 0.0%/
2060329 3 REPLICATES 2060329 3 REPLICATES 2060335 3 REPLICATES	Not reported  Not reported	4 NR 4 NR 4 NR		Day(s) Day(s) Day(s)	LOEC LOSC	Mortality  Mortality	Mortality Survival	MSMT/TRIM LEVEL 20.0%/
2060333   3 REPLICATES 2060352   3 REPLICATES	Not reported  Not reported	4 NR		Day(s) Day(s)	NOEC NR-LETH	Mortality	Survival	MSMT/REPLICATE B/
2060350 3 REPLICATES 2060361 3 REPLICATES	Not reported  Not reported	4 NR 3 NR		Day(s)	NR-LETH	Mortality Mortality	Mortality Mortality	MSMT/REPLICATE A/ MSMT/REPLICATE C/
2060342 3 REPLICATES 2060354 3 REPLICATES	Not reported  Not reported	3 NR		Day(s) Day(s)	NR-ZERO NR-ZERO	Mortality	Mortality Mortality	MSMT/REPLICATE A/ MSMT/REPLICATE B/
740693 741709	Not reported Not reported	2 NR 2 NR		Day(s) Day(s)	NR-ZERO NR-ZERO	Mortality Mortality	Mortality Mortality	
201542 201543	Not reported Not reported	4 NR 1 NR		Day(s) Day(s)	NR-LETH	Mortality	Survival Mortality	
231545	Not reported	4 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231549	Not reported	3 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
232524	Not reported	3 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231547	Not reported	1 NR	NR C	Day(s)	TC20	Mortality	Mortality	MSMT/TLM/
231542	Not reported	1 NR		Day(s)	TC20	Mortality	Mortality	MSMT/TLM/
231548	Not reported	2 NR	NR R	Day(s)	TC20	Mortality	Mortality	MSMT/TLM/
231540	Not reported	3 NR			LC50	Mortality	Mortality	MSMT/TLM/
231539	Not reported	2 NR	NR NR	Day(s)	LC20	Mortality	Mortality	MSM1/1LM/

(Days)         Duration Units           (Days)         (Day(s)           8         Day(s)           8         Day(s)           8         Day(s)           9         Day(s)           10         Day(s)		1			71. 11				120 T. C.		
Exportmental Design		Mortality	Mortality	LC50	Day(s)			2	Not reported		196747
Exportmental Design		Mortality	Mortality	LC50	Day(s)			4	Not reported		196717
Experimental Design		Mortality	Mortality	LC50				i i	Not reported		196754
Experimental Design		Mortality	Mortality	LC50				1	Not reported		196739
Experimental Design		Mortality	Mortality	LC50				4	Not reported		196721
Experimental Design		Mortality	Mortality	LC50				2	Not reported		196771
Experimental Design		Mortality	Mortality	LC50	Day(s)			1	Not reported		196727
Experimental Design		Mortality	Mortality	1020	Dav(s)			1 1	Not reported		196715
Experimental Design		Mortality	Mortality	LC50	Day(s)			4 4	Not reported		196725
Experimental Design		Mortality	Mortality	LC50				. 1	Not reported		196758
Experimental Design   Response Site   Monaton   Duration   Durat		Mortality	Mortality	LC50				4	Not reported		196733
Experimental Design		Mortality	Mortality	LC50				4	Not reported		196764
Experimental Design		Mortality	Mortality	LC50	Day(s)			2	Not reported		196712
Experimental Design         Responses Site         Mend Days (Min Days)         Duration (Days)         Duration (Days)         Effect         Meximum (Min Days)		Mortality	Mortality	LC50	Day(s)			4	Not reported		196737
Experimental Design   Responses Site   Mean Days   Maria   Days		Mortality	Mortality	LC50	Day(s)			1	Not reported		196711
Experimental Design		Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		196751
Experimental Design		Mortality	Mortality	LC50	Day(s)			2	Not reported		196716
Experimental Design         Response Site Mean (Days)         Moration (Days)         (Days)         (Days)         (Casy)         (Moration Units         Effect         Measurement Moratiny Moratiny         Moration Moratiny Moratiny         Moration Moratiny Moratiny         Moratiny Moratiny Moratiny Moratiny         Moratiny Moratiny Moratiny Moratiny Moratiny Moratiny         Moratiny M		Mortality	Mortality	LC50	Day(s)		NR.	2 4	Not reported	LICATES	4
Experimental Design         Response Site Mean (Darys)         Duration Max Duration Units         Duration Units         Effect         Measurement Measu		Mortality	Mortality	1050	Day(s)		NR.	2	Not reported	1	
Experimental Design         Response Site         Mean (Days)         Duration Max         Duration Units         Effect         Measurement           Not reported         1 NR         NR         Day(s)         CSO         Mortality         Mortality         Most reported         Mortality         <		Mortality	Mortality	LC50	Day(s)			2	Not reported		196/32
Experimental Design         Response Site         Mean (Days)         Duration Duration Mark (Days)         (Days)         (Edge)         Effect*         Mortality         Most reported         Effect*         Mean (Days)         (Coays)         (Coays)         Effect*         Most reported         Mortality         Most reported         Mortality         Mortality         Most reported         Mortality		Mortality	Mortality	LC50	Day(s)		NR	1	Not reported		196723
Experimental Design         Response Site         Mean (Days)         Unation of Management         Duration of Management         Duration of Management         Effect*         Measurement           Not reported         1 NR         0 JNR         0 Jay(s)         C.50         Mortality         Mortality         MSM/TIAM/           Not reported         4 NR         NR         0 Jay(s)         C.50         Mortality         Mortality         MSM/TIAM/           Not reported         4 NR         NR         0 Jay(s)         C.50         Mortality         Mortality         MSM/TIAM/           Not reported         4 NR         NR         0 Jay(s)         C.50         Mortality         Mortal		Mortality	Mortality	LC50	Day(s)			1	Not reported		196766
Experimental Design         Response Site         Mean (Days)         Unation of Man (Days)         Unation of Man (Days)         Unation of Man (Days)         Effect*         Mean (Days)         (Days)         (C30)         Mortality         Most reported           Not reported         1 NR         NR         Day(s)         (C50)         Mortality         Mortality         MSM/TIAM/           Not reported         4 NR         NR         Day(s)         (C50)         Mortality         Mortality         MSM/TIAM/           Not reported         4 NR         NR         Day(s)         (C50)         Mortality         Mortality         Mortality         MSM/TIAM/           Not reported         2 NR         NR         Day(s)         (C50)         Mortality		Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		196720
Experimental Design         Aesponse Site         Mean (Days)         Min (Days)         Duration Max         Duration Initis         Effect         Measurement           Not reported         Not reported         2 INR         NR         Day(s)         LC50         Mortality         Mortality         MSM1/TLM/           Not reported         2 INR         NR         Day(s)         LC50         Mortality         Mortality         MSM1/TLM/           Not reported         4 INR         NR         Day(s)         LC50         Mortality         MSM1/TLM/           Not reported         4 INR         NR         Day(s)         LC50         Mortality         MSM1/TLM/           Not reported         4 INR         NR         Day(s)         LC50         Mortality         MSM1/TLM/           Not reported         4 INR         NR         Day(s)         LC50         Mortality         MSM1/TLM/           Not reported         4 INR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         4 INR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         4 INR         NR         Day(s)         LC50         Mortality         <		Mortality	Mortality	LC50	Day(s)			1	Not reported		196770
Experimental Design         Response Site         Man (Days)         Min (Days)         (Days)         (Days)         Effect         Measurement           Not reported         Not reported         2 NR         NR         Day(s)         LCSO         Mortality         Mortality         Mostality         MSM1/TLM/           Not reported         2 NR         NR         Day(s)         LCSO         Mortality         Mortality         Msm1/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality         Msm1/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Morta		Mortality	Mortality	LC50	Day(s)			3	Not reported		196839
Experimental Design         Response Site         Mean (Days)         Lowation May         Lowation May         Logy         Logy         Effect         Measurement           Not reported         1 NR         0 Ayrs         (Days)         LCSO         Mortality         Mostality         MSM1/TLM/           Not reported         2 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSM1/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSM1/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mo		Mortality	Mortality	LC50	Day(s)			1	Not reported		196731
Experimental Design         Response Site Not reported         Munation / Image: Not reported / Image: Not Report		Mortality	Mortality	LC50	Day(s)			2	Not reported		196759
Experimental Design         Response Site (paration Long)         Duration (paration Long)         Duration (paration Long)         Duration (paration Long)         Duration (paration Long)         Effect (paratic)         Medical (paratic)         Effect (paratic)         Medical (paratic)         Mort (paratic)         Logy()         LC50         Mortality         MSMI/TLM/         MSMI/TLM/         MSMI/TLM/         Mortality         Mortality         MSMI/TLM/         Mortality         Mortality         MSMI/TLM/         MSMI/TLM/         Mortality         Mortality         Mortality         Mortality         Mortality         MSMI/TLM/         MSMI/TLM/         MSMI/TLM/         Mortality		Mortality	Mortality	LC50	Day(s)			4	Not reported		196760
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Duration (Days)         Unation (Days)         Unation (Days)         Unation (Days)         Effect*         Measurement           Response Site         Mean (Days)         Mini (Days)         (Days)         (LSO)         Mortality         Measurement           Not reported         1 NR         NR         Day(s)         (LSO)         Mortality         Mortality </td <td></td> <td>Mortality</td> <td>Mortality</td> <td>LC50</td> <td>Day(s)</td> <td></td> <td></td> <td>4</td> <td>Not reported</td> <td></td> <td>196729</td>		Mortality	Mortality	LC50	Day(s)			4	Not reported		196729
Experimental Design         Response Site Mort Days (Not reported)         Duration Days (Days)         Duration (Days)         Duration (Days)         Duration (Days)         Endpoint Effect*         Effect*         Measurement Most reported         Mn (Days)         (Days)         Duration (Days)         Day(s)         LCSO         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LCSO         Mortality		Mortality	Mortality	LC50	Day(s)			4	Not reported		196756
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Duration (Days)         Unity         Effect         Modality         Modality         Modality         Mortality         Mortality         Mortality         Mortality         Mortality         Mortality         Most/7/LM/           Not reported         2 NR         4 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortali		Mortality	Mortality	LC50	Day(s)		NR	4	Not reported		196741
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Duration (Days)         Duration (Days)         Duration (Days)         Endpoint         Effect*         Mean (Pays)         Mean (Days)         Endpoint         Effect*         Mestirement           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSM1/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSM1/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         MSM1/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mort		Mortality	Mortality	LC50	Day(s)			2	Not reported		196728
Experimental Design         Response Site         Muration (Days)         Duration (Days)         Duration (Days)         Duration (Days)         Effect*         Measurement           Response Site         Mont (Days)         Min (Days)         (Days)         (Days)         Endpoint         Effect*         Measurement           Response Site         Mont (Days)         Min (Days)         (Days)         LC50         Mortality         Mort		Mortality	Mortality	LC50	Day(s)			4	Not reported		196748
Experimental Design         Response Site         Mean (Days)         Munit (Days)         Duration (Days)         Duration (Days)         Duration (Days)         Endpoint         Effect*         Mean (Bays)         Effect*         Mean (Days)         Effect*         Mean (Days)         Mortality         Mortality         MSMT/TLM/           Response Site         Not reported         1 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Response Site         Not reported         2 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Response Site         Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Response Site         Not reported         1 NR         NR         Day(s)         LC50         Mortality		Mortality	Mortality	LC50	Day(s)			4	Not reported		196714
Experimental Design         Response Site         Mean (Days)         Duration Value         Duration Value         Duration Value         Effect*         Measurement           Response Site         Mean (Days)         VIII (Days)         (Days)         Unation Value         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality		Mortality	Mortality	LC50	Day(s)			4	Not reported		196745
Experimental Design         Response Site         Mean (Days)         Min (Days)         Quration Max (Days)         Duration Units         Endpoint         Effect*         Measurement           Response Site         Response Site         Men (Days)         Min (Days)         (Days)         (Days)         Endpoint         Effect*         Measurement           Response Site         Mean (Days)         Min (Days)         (Days)         (Days)         LC50         Mortality         Measurement           Response Site         Mean (Days)         NR         Day(s)         LC50         Mortality         Mortality <td< td=""><td></td><td>Mortality</td><td>Mortality</td><td>LC50</td><td>Day(s)</td><td></td><td></td><td>4</td><td>Not reported</td><td></td><td>759180</td></td<>		Mortality	Mortality	LC50	Day(s)			4	Not reported		759180
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Quration (Days)         Quration (Days)         Unration (Days)         Propried (Days)         Effect*         Measurement           Response Site         Mean (Days)         Min (Days)         (Days)         (Days)         Endpoint         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortalit		Mortality	Mortality	LC50	Day(s)			2	Not reported		196763
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Quaration Max (Days)         Duration Units         Effect*         Measurement           Most reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality           Not		Mortality	Mortality	LC50	Day(s)			2	Not reported		196724
Experimental Design         Response Site         Mean (Days)         Duration Max         Duration Max         Duration Units         Endpoint         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortalit		Mortality	Mortality	LC50	Day(s)			2	Not reported		196744
Experimental Design         Response Site         Mean (Days)         Duration Max         Duration Max         Duration Units         Effect*         Measurement           Mot reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortali		Mortality	Mortality	LC50	Day(s)			4	Not reported		196713
Experimental Design         Response Site         Mean (Days)         Duration Max         Duration Max         Duration Units         Effect*         Measurement           Mot reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality		Mortality	Mortality	LC50	Day(s)			2	Not reported		196736
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Duration Max (Days)         Duration Montality         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LCSO         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LCSO         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LCSO         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LCSO         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LCSO         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality           Not reported         1 NR         NR         Day(s)         LCSO         Mortality         Mortality     <		Mortality	Mortality	LC50	Day(s)			2	Not reported		196767
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Duration Max (Days)         Duration Units         Endpoint         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality		Mortality	Mortality	LC50	Day(s)			4	Not reported		196768
Experimental Design   Response Site   Mean (Days)   Min (Days)		Mortality	Mortality	LC50	Day(s)			1	Not reported		196735
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Duration Max (Days)         Duration Units         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality         Mortality           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality		Mortality	Mortality	LC50	Day(s)			2	Not reported		196755
Experimental Design         Response Site         Mean (Days)         Duration (Days)         Quration Max (Days)         Duration Units         Endpoint         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         Mortality         Mortality         Mortality		Mortality	Mortality	LC50	Day(s)			4	Not reported		196772
Experimental Design         Response Site         Mean (Days)         Min (Days)         (Days)         (Days)         Unration Max         Duration Units         Effect*         Measurement           Not reported         1 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         2 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/           Not reported         4 NR         NR         Day(s)         LC50         Mortality         Mortality         MSMT/TLM/		Mortality	Mortality	LC50	Dav(s)				Not reported		196750
Experimental Design  Response Site Not reported  Not repor	MSMT/TLM/		Mortality	LC50	Day(s)		Î	4	Not reported		231541
Experimental Design Response Site Mean (Days) Min (Days) (Days) (Days) Endpoint Effect*  Not reported 1 NR NR Days(s) LC50 Mortality Montality MSMT/TLM/	MSMT/TLM/	3	Mortality	LC50	Day(s)			2	Not reported		231552
Experimental Design Response Site Mean (Days) Min (Days) (Days) (Days) Endpoint Effect* Measurement	- 1	//	Mor	LC50	Day(s)			1	Not reported		231538
	EE Comr	Effect Measurement		Endpoin	Duration Units (Days)	Duration Max (Days)		Duration Mean (Days)	Response Site	Experimental Design	Result Number

	no to lite	Mortality	NID ZEDO	021/2	7	700	_		/42932	
	Mortality	Mortality	NR-ZERO	Day(s)		4 NR	4	Not reported		742077
	Mortality	Mortality	NR-ZERO	Day(s)	NR C	NR	4	Not reported		/420/0
	Mortality	Mortality	NR-ZERO	Day(s)	NR	4 NR	4	Not reported		742057
	Mortality	Mortality	NR-ZERO	Day(s)			4	Not reported		742029
	Mortality	Mortality	NR-ZERO	Day(s)	NR	4 NR	4	Not reported	59	742059
	Mortality	Mortality	NR-ZERO	Day(s)		2 NR	2	Not reported		740696
	Mortality	Mortality	NR-ZERO	Day(s)		NR	2	Not reported		740694
	Mortality	Mortality	NR-LETH	Day(s)		NR	1	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	768558
	Mortality	Mortality	NR-LETH	Day(s)	NR [	NR	ω	Not reported		201538
	Mortality	Mortality	NR-LETH	Day(s)	NR [	.25 NR N	1.25	Not reported	97	740697
	Survival	Mortality	NOEL	Day(s)	NR [	4 NR	4	Not reported	66 24 HOUR PULSE DURATION	742666
MSMT/CONC/ AVERAGE 7 DAY CONCENTRA	Survival	Mortality	NOEL	Day(s)	NR [	NR P	7	Not reported		74140
	Survival	Mortality	NOEC	Day(s)		NR	7	Not reported	18	768118
	Survival	Mortality	NOEC	Day(s)	NR		7	Not reported	34	768134
	Survival	Mortality	NOEC	Day(s)	NR I	NR 7	7	Not reported	65	768065
	Survival	Mortality	NOEC	Day(s)			7	Not reported	48	768148
	Survival	Mortality	NOEC	Day(s)			7	Not reported	38	768138
	Survival	Mortality	NOEC	Day(s)	NR	NR	7	Not reported	83	768083
	Survival	Mortality	NOEC	Day(s)			7	Not reported	08	768108
	Survival	Mortality	NOEC	Day(s)		NR 7	7	Not reported	14	768114
	Survival	Mortality	NOEC	Day(s)			7	Not reported	81	768081
	Survival	Mortality	NOEC	Day(s)			7	Not reported	75	768075
	Survival	Mortality	NOEC	Day(s)	NR	NR	7	Not reported	99	768099
	Survival	Mortality	NOEC	Day(s)		NR	7	Not reported	.54	768154
	Survival	Mortality	NOEC	Day(s)		NR I	7	Not reported	.64	768164
	Survival	Mortality	NOEC	Day(s)		NR	7	Not reported	.24	768124
	Survival	Mortality	NOEC	Day(s)			7	Not reported	189	768089
	Survival	Mortality	NOEC	Day(s)			7	Not reported	91	768091
	Survival	Mortality	NOEC	Day(s)		NR		Not reported	.44	768144
	Survival	Mortality	NOEC	Day(s)			7	Not reported	.28	768128
	Survival	Mortality	NOEC	Day(s)			7	Not reported	97	768097
	Survival	Mortality	NOEC	Day(s)			7	Not reported	950	768059
	Survival	Mortality	NOEC	Day(s)			7	Not reported	)67	768067
	Survival	Mortality	NOEC	Day(s)		1	7	Not reported	.58	768158
	Survival	Mortality	NOEC	Day(s)		NR	7	Not reported	)73	768073
	Survival	Mortality	NOEC	Day(s)		NR	33	Not reported	560	201560
	Survival	Mortality	NOEC	Day(s)			7	Not reported	105	768105
	Mortality	Mortality	NOEC	Day(s)			7	Not reported	907	96907
	Mortality	Mortality	NOEC	Day(s)	NR	NR		Not reported	357	69057
	Mortality	Mortality	NOEC	Day(s)				Not reported	385	96885
	Mortality	Mortality	NOEC	Day(s)		NR		Not reported	383	14383
	Mortality	Mortality	NOEC	Day(s)	NR	NR		Not reported	365	42865
	Mortality	Mortality	NOEC	Day(s)	NR	NR		Not reported	599	68599
	Survival	Mortality	MATC	Day(s)	NR	NR	33	Not reported	562	201562
	Survival	Mortality	LOEL	Day(s)	NR	NR		Not reported	110 96 HOUR PULSE DURATION	741410
	Survival	Mortality	LOEL	Day(s)	NR.	NR		Not reported	742691 3 HOUR PULSE DURATION	74269
	Survival	Mortality	LOEL	Day(s)	NR	1 NR	2	Not reported	744 24 HOUR PULSE DURATION	741744
	Survival	Mortality	LOEC	Day(s)	NR	NR	4	Not reported	509 DEFINITIVE CHRONIC, 4 REPLICATES	2210509
	Survival	Mortality	LOEC	Day(s)	NR	NR	33	Not reported	561	201561
	Survival	Mortality	LOEC	Day(s)	NR	7 NR		Not reported	117	768117
	Survival	Mortality	LOEC	Day(s)	NR			Not reported	133	768133
EE Comment	Measurement	Effect*	Endpoint	(Days)	(Days)	Min (Days)	Mean (Days)	Response Site	ber Experimental Design	Result Number
	176			_	Observed		Observed			

Result Number	Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
	777	Not reported	+2	D.		Day(s)	NR-ZERO	Mortality	Mortality	
	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	4 NR	R		Day(s)	NR-ZERO	Mortality	Mortality	
742027		Not reported	4 NR	R		Day(s)	NR-ZERO	Mortality	Mortality	
742030		Not reported	4 NR	, 20		Day(s)	NR-ZERO	Mortality	Mortality	
020247		Not reported	4 NR	7		Day(s)	NK-ZEKO	Mortality	Mortality	
231532		Not reported	1 NR	Z Z	20 2	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231531		Not reported	4 NR	D Z		Day(s)	LC50	Mortality	Mortality	MSMT/TLMZ/
231530		Not reported	3 NR	D		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231529		Not reported	2 NR	R		Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
26115		Not reported	1.0417 NR	R	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
26116		Not reported	2 NR	R	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
	2 REPLICATES	Not reported	4 NR	R	NR	Day(s)	LC50	Mortality	Mortality	
	2 REPLICATES	Not reported	4 NR	R	NR	Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE B/
2060323	2 REPLICATES	Not reported	4 NR	R		Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE A/
741795 /	ACUTE STUDY	Not reported	4 NR	R		Day(s)	LC50	Mortality	Mortality	
742665 1	TRIALA	Not reported	1 NR	R	NR	Day(s)	NR-ZERO	Mortality	Mortality	
742664 1	TRIAL B	Not reported	1 NR	D		Day(s)		Mortality	Mortality	
741756 /	ACUTE STUDY	Not reported	4 NR	R	NR	Day(s)	LC50	Mortality	Mortality	
741783 1	TRIALA	Not reported	1 NR			Day(s)	NR-ZERO	Mortality	Mortality	
741751 T	TRIALB	Not reported	1 NR			Day(s)	NR-ZERO	Mortality	Mortality	
2170552		Not reported	30 NR			Day(s)	NR-ZERO	Mortality	Mortality	
740686		Not reported	2 NR			Day(s)	NR-ZERO	Mortality	Mortality	
740685		Not reported	2 NR			Day(s)	NR-ZERO	Mortality	Mortality	
2075473		Not reported	4 NR	3 - 6	NR	Day(s)	LC50	Mortality	Mortality	
2075472		Not reported	4 NR			Day(s)	LC50	Mortality	Mortality	
231556		Not reported				Day(s)	LC50	Mortality		MSMT/TLM/
231560		Not reported	2 NR		NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231561		Not reported	3 NR			Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231559		Not reported	1 NR		NR [	Day(s)	LC50	Mortality		MSMT/TLM/
231557		Not reported	3 NR		NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231558		Not reported	4 NR			Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
231562		Not reported	4 NR			Day(s)	LC50	Mortality		MSMT/TLM/
231555		Not reported	1 NR		NR [	Day(s)	LC50	Mortality	Mortality	MSMT/TLM/
741781 T	TRIALB	Not reported	1 NR			Day(s)	NR-ZERO	Mortality		
741788 A	ACUTE STUDY	Not reported	4 NR		NR L	Day(s)	LC50	Mortality	Mortality	
741743 T	TRIAL B	Not reported	1 NR			Day(s)	NR-ZERO	Mortality	Mortality	
741742 T	TRIALA	Not reported	1 NR			Day(s)	NR-ZERO	Mortality	Mortality	
2075476		Not reported	4 NR				LC50	Mortality	Mortality	
2075477		Not reported	4 NR				LC50	Mortality	Mortality	
2152833 4	4 REPLICATES	Not reported	4 NR			Day(s)	LC50	Mortality	Mortality	
803853		Not reported	1 NR			Day(s)	EC50	Mortality	Survival	
741562		Not reported	2 NR				EC50	Mortality	Survival	
741561		Not reported	1 NR				EC50	Mortality	Survival	
741583		Not reported	4 NR				FC50	Mortality	Survival	
803850		Not reported					EC20	Mortality	Survival	
759169		Not reported	78 NR				בכטט	Mortality	DUIVIVAI	
764615		Not reported	1 NR				NR-LETH	Mortality	Mortality	
160288		Not reported	ON C				INV-FE ILI	Mortality	Mortality	
160287		Not reported	1 NR		NR Z		1050	Mortality	Motolity	
767346 FE	ED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1 NR				NRLIETH	Mortality	Mortality	
767362 FI	767362 FED. MODERATELY HARD RECONSTITUTED WATER Not reported	Not reported	1 NR				ND LETE	Mortality	Mortality	
101001	בם, אוספבות וברו וואוס וובכסווטווו סובט אאובוג	Aprieboiled	TINA		NX.	Day(s)	2 スートーエ	MOLTAILLA	INIORTALITY	

Result Number	Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
767348	767348 UNFED, MODERATELY HARD RECONSTITUTED WAT Not reported	Not reported	=	NR	NR TELE	Day(s)	NR-LETH	Mortality	Mortality	
767402	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1 7	NR		Day(s)	NR-LETH	Mortality	Mortality	
	UNFED, MODERATELY HARD RECONSTITUTED WATENOT reported	Not reported	17	NR		Day(s)	NR-LETH	Mortality	Mortality	
767407	UNFED, MODERATELY HARD RECONSTITUTED WAT	Not reported	1 7	NR	NR	Day(s)	NR-LETH	Mortality	Mortality	
767364	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	21	NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
767366	UNFED, MODERATELY HARD RECONSTITUTED WAT	Not reported	2 1	NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
767410	UNFED, MODERATELY HARD RECONSTITUTED WAT	Not reported	2 1	NR	NR	Day(s)		Mortality	Mortality	
767347	767347 FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	21			Day(s)	NR-ZERO	Mortality	Mortality	
767461	FED. MODERATELY HARD RECONSTITUTED WATER	Not reported	21	NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
767465	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	2 1			Day(s)	NR-ZERO	Mortality	Mortality	
759182		Not reported	2 1	NR	NR	Day(s)	LC50	Mortality	Mortality	
160289		Not reported	1 NR		NR	Day(s)	LC50	Mortality	Mortality	
160290		Not reported	21	NR	NR	Day(s)	LC50	Mortality	Mortality	
24910		Not reported	4 NR			Day(s)	LC50	Mortality		MSMT/TLM///
24907		Not reported	2 2	NR R	NR	Day(s)	1050	Mortality	Mortality	MSMT/TIM///
24906		Not reported	1 NR			Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
24909		Not reported	4 7	NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
767488	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1 1	NR		Day(s)	NR-LETH	Mortality	Mortality	
	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1 NR			Day(s)	NR-LETH	Mortality	Mortality	
767/97	EED MODERATELY HARD RECONSTITUTED WATER	Not reported	2 2	NR NR	NR NR	Day(s)	NR-ZERO	Mortality	Mortality	
	FED. MODERATELY HARD RECONSTITUTED WATER	Not reported	2 NR			Day(s)	NR-ZERO	Mortality	Mortality	
759473		Not reported	21 N			Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE B///
759470		Not reported	21 NR		NR	Day(s)	NR-ZERO	Mortality	Mortality	MSMT/REPLICATE A///
2064012		Not reported	P			Day(s)	LC50	Mortality	Mortality	ENDPT/TLM/
2064016		Not reported			N.R.	Day(s)	LC50	Mortality	Mortality	ENDRI/TIM/
220044		Not reported	4 1			Day(s)	NOEC	Mortality	Mortality	MSMT/CORRECTED FOR CONTROL///
24911		Not reported				Day(s)	LC50	Mortality		MSMT/TLM///
24914		Not reported	37	NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
24912		Not reported	1 NR			Day(s)	LC50	Mortality		MSMT/TLM///
24913		Not reported	2 7	NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
24915		Not reported	4 NR			Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
89658		Not reported	2 NR			Day(s)	LC50	Mortality	Mortality	
6128/		Not reported	1 NR			Day(s)	LC50	Mortality	Mortality	
61786		Not reported	1 NR			Day(s)	LCSO	Mortality	Motality	
89662		Not reported	2 NR		NR NR	Day(s)	1020	Mortality	Mortality	
89660		Not reported	1 NR			Day(s)	LC50	Mortality	Mortality	
160291		Not reported	1 NR			Day(s)	LC50	Mortality	Mortality	
759181		Not reported	4 NR		NR	Day(s)	LC50	Mortality	Mortality	
160292		Not reported	2 NR			Day(s)	LC50	Mortality	Mortality	A
		Not reported	4 NR			Day(s)	LC50	Mortality	Mortality	
/6/519	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	4 NR			Day(s)	NR-ZERO	Mortality		ACDAT/TINA///
18176		Not reported	1 NR		NR NR	Day(s)	1050	Mortality	Mortality	MSMT/TIM///
18177		Not reported	2 NR			Day(s)	LC50	Mortality		MSMT/TLM///
18179		Not reported	4 NR		NR	Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
764340		Not reported	1 NR			Day(s)	NR-LETH	Mortality	Mortality	
COLLOS		Morreborren	T INI		INIV	Day(s)	TAIN-FF III	iviolitality	INIOITAIITY	

MSMT/MEAN OF 3 TESTS/	Mortality MS	Mortality	LCOO	Cay(3)						-
			CEO	Dav(s)	NR I	NR	2	Not reported	CA:MG RATIO 2.33	766718 CA
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	CA:MG RATIO 2.33	
	Mortality	Mortality	LC50	Day(s)	NR I	NR	2	Not reported	CA:MG RATIO 2.33	766555 CA
	Mortality	Mortality	LC50	Day(s)	NR I	NR	2	Not reported	CA:MG RATIO 1.46	766610 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 1.46	766586 CA
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	CA:MG RATIO 1.46	766595 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	766578 CA
		Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	766573 CA
MSMT/MEAN OF 3 TESTS/	1	Mortality	LC50	Day(s)	NR		2	Not reported	CA:MG RATIO 2.33	766722 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	CA:MG RATIO 2.33	766575 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 1.46	766603 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	66560 CA
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	CA:MG RATIO 2.33	766577 CA
MSMT/MEAN OF 3 TESTS/		Mortality	LC50	Day(s)		NR	2	Not reported	CA:MG RATIO 2.33	766717 CA
		Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	766574 CA
MSMT/MEAN OF 3 TESTS/		Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	766720 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	766556 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 1.46	766607 CA
	Mortality	Mortality	LC50	Day(s)	NR	2 NR	2	Not reported	CA:MG RATIO 2.33	766565 CA
	Mortality	Mortality	LC50	Day(s)			2	Not reported	CA:MG RATIO 2.33	766580 CF
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	CA:MG RATIO 2.33	
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	CA:MG RATIO 2.33	766554 CA
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	CA:MG RATIO 1.46	766609 C/
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	CA:MG RATIO 2.33	766576 CA
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		766434
		Mortality	LC50	Day(s)		NR	2	Not reported		160331
MSMT/MEAN OF 3 TESTS/		Mortality	LC50	Day(s)		NR	2	Not reported		766215
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	REFORMULATED MODERATELY HARD WATER	
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		766430
	Mortality	Mortality	LC50	Day(s)		1 NR	1	Not reported		160330
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		766432
	Mortality	Mortality	LC50	Brood or litter		NR	ω	Not reported		759078
	Mortality	Mortality	LC50	Day(s)		NR	7	Not reported	REFORMULATED MODERATELY HARD WATER	766226 RI
	Mortality	Mortality	LC50	Day(s)		NR	7	Not reported	MODERATELY HARD WATER	766221 M
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	MODERATELY HARD WATER	766227 M
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	REFORMULATED MODERATELY HARD WATER	766228 RI
	Mortality	Mortality	LC50	Dav(s)		NR	2	Not reported		
	Mortality	Mortality	1020	Dav(s)		2 NR	2	Not reported	CA:MG RATIO 1.46	
	Mortality	Mortality	1010	Day(s)		N.	2	Not reported		766216
	Mortality	Mortality	1010	Dav(s)		2 NR	2	Not reported		766429
	Mortality	Mortality	LC10	Dav(s)			2	Not reported		766431
	Mortality	Mortality	LC10	Dav(s)			2	Not reported	REFORMULATED MODERATELY HARD WATER	
	Mortality	Mortality	LC10	Day(s)		NR	N	Not reported		766433
	Mortality	Mortality	LC10	Day(s)		NR		Not reported		766427
	Mortality	Mortality	LC10	Day(s)	NR	NR	2	Not reported		766435
	Survival	Mortality	EC50	Day(s)		NR	7	Not reported		2017029
	Survival	Mortality	EC50	Day(s)		NR	7	Not reported		2017040
	Survival	Mortality	EC50	Day(s)	NR	NR	-	Not reported		2017057
	Survival	Mor	EC50	Day(s)		NR	7	Not reported		2017048
EE Comment	Measurement	Effect*	Endpoint	(Days)	(Days)	Min (Days)	Mean (Days)	Response Site	Experimental Design	Result Number
	Effort			Observed	Observed	Observed	Observed			

	Tarion solita	Mortality	מוסק מוא	201(2)			,	- contract	CED MODED ATELY HADD DECOMISTITUTED MATER
	Mortality	Mortality	NB ZERO	Day(s)	ND NO	ND	4 +	Not reported	767351 EED MODERATELY HARD RECONSTITUTED WATER
	Mortality	Mortality	NB IETH	Day(s)			4 4	Not reported	
	Mortality	Mortality	NR-LETH	Day(s)	NR		1	Not reported	767411 FED. MODERATELY HARD RECONSTITUTED WATER
	Mortality	Mortality	NR-LETH	Day(s)	NR		1	Not reported	
	Mortality	Mortality	NR-LETH	Day(s)	NR		1	Not reported	767380 UNFED, MODERATELY HARD RECONSTITUTED WATE
	Mortality	Mortality	NR-LETH	Day(s)	NR			Not reported	
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1	Not reported	767457 FED, MODERATELY HARD RECONSTITUTED WATER
	Survival	Mortality	NOEC	Day(s)	NR	NR	7	Not reported	766854
MSMT/MEAN OF 3 TESTS/	Survival	Mortality	NOEC	Day(s)	NR	NR	7	Not reported	766731 MODERATELY HARD RECONSTITUTED WATER
MSMT/MEAN OF 3 TESTS/	Survival	Mortality	NOEC	Day(s)	NR	NR	7	Not reported	766740 REFORMULATED MODERATELY HARD RECONSTITUT
	Survival	Mortality	NOEC	Day(s)	NR	NR	7	Not reported	761596
	Survival	Mortality	NOEC	Day(s)	NR	NR.	7	Not reported	2017030
	Survival	Mortality	NOEC	Day(s)	NR		7	Not reported	2017041
	Survival	Mortality	NOEC	Day(s)	NR		7	Not reported	2017058
	Survival	Mortality	NOEC	Day(s)	NR		7	Not reported	2017049
	Mortality	Mortality	NOEC	Brood or litter	NR		i w	Not reported	759073
	Mortality	Mortality	NOEC	Day(s)	N.R.		,	Not reported	766223 MODERATELY HARD WATER
	Mortality	Mortality	NOEC	Day(s)	NR.		7	Not reported	766225 REFORMULA IEU MODERA IELY HARD WATER
MSM1/MEAN OF 3 TESTS/	Survival	Mortality	LOEC	Day(s)	N. N.		1	Not reported	766730 MODERATELY HARD RECONSTITUTED WATER
MSMI/MEAN OF 3 TESTS/	Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	766739 REFORMULATED MODERATELY HARD RECONSTITU
	Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	
	Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	2017042
	Survival	Mortality	LOEC	Day(s)	NR		7	Not reported	2017059
	Survival	Mortality	LOEC	Day(s)	N.X		-	Not reported	201/050
	Nortality	Mortality	LOEC	Brood or litter	ZZ		1 4	Not reported	/590/2
	Mortality	Mortality	LOEC	Day(s)	N. N.		7	Not reported	/66222 MODERATELY HARD WATER
	Modelity	Mortality	LOCK	Day(s)	NA		,	Not reported	700224 REFORMODATED MODERATED TAKE WATER
MISINIT/ MEAN OF 3 TESTS/	SULVIVAL	Mortality	LCSU	Day(s)	NA.		2	Not reported	
MENAT/MEAN OF 3 TESTS/	Survival	Modelity	LCJO	Day(s)	NIN IN		,	Morrieborren	700/43 NECONIVIOLATED INTOCENATELY HAND NECONSTITU
MSMT/MEAN OF 3 TESTS/	Suprival	Mortality	1050	Day(s)	ND NA		7	Not reported	766374 BEECOBALLI ATED MODEBATELY HARD BECONSTITUT
	Mortality	MOLIGITY	LCSU	Day(s)	27		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR.		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR	100	2	Not reported	766612 CA:MG RATIO 1.46
	Mortality	Mortality	LC50	Day(s)	NR	ľ	2	Not reported	
MSMT/MEAN OF 3 TESTS/	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	766726 CA:MG RATIO 2.33
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	766563 CA:MG RATIO 2.33
	Mortality	Mortality	LC50	Day(s)	NR.		2	Not reported	766551 CA:MG RATIO 2.33
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NX		2	Not reported	
	Mortality	Mortality	LC50	Day(s)	NX	NR	2	Not reported	/66606 CA:MG RATIO 1.46
	Mortality	Mortality	LCSU	Day(s)	NR.	N.X	2	Not reported	
	Mortality	Mortality	LC50	Day(s)	ZX		2	Not reported	766572 CA:MG RATIO 2.33
MSM1/MEAN OF 3 TESTS/	Mortality	Mortality	LCSU	Day(s)	NR.			Not reported	
	Mortality	Mortality	1050	Day(s)	NR		2	Not reported	766591 CA:MG RATIO 5.4
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported	
MSMT/MEAN OF 3 TESTS/	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	766723 CA:MG RATIO 2.33
EE Comment	Effect Measurement	Effect*	Endpoint	Duration Units (Days)	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	Result Number Experimental Design
				Observed	Observed	Chacinen	Observed		

Result Number 767456 FE	Experimental Design ED, MODERATELY HARD RECONSTITUTED WATER	Response Site	Observed O Duration C Mean (Days) M 2 NR	bserved Duration in (Days)	Observed Duration Max (Days) NR	Observed Duration Units (Days) Day(s)	Endpoint NR-ZERO	Effect*	Effect Measurement Mortality
	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	2 N			Day(s)	NR-ZERO	Mortality	Mortality
767353 UI	UNFED. MODERATELY HARD RECONSTITUTED WATEN NOT REPORTED  UNFED. MODERATELY HARD RECONSTITUTED WATENOT REPORTED	Not reported	2 NR		NR	Day(s)	NR-ZERO	Mortality	Mortality
	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported				Day(s)	NR-ZERO	Mortality	Mortality
766218	7	Not reported	2 NR			Day(s)	LC10	Mortality	Mortality
766217		Not reported	2 NR		NR	Day(s)	LC50	Mortality	Mortality
18144		Not reported	1 NR			Day(s)	LC50	Mortality	Mortality
18145		Not reported					LC50	Mortality	Mortality
767794		Not reported	2 NR		NR Z	Day(s)	LC50	Mortality	Mortality
	CA:MG MOLAR RATIO 7.0	Not reported	2 NR				LC50	Mortality	Mortality
767306 C/	CA:MG MOLAR RATIO 7.0	Not reported	2 NR				LC50	Mortality	Mortality
767296		Not reported					LC50	Mortality	Mortality
	CA:MG MOLAR RATIO 1.8	Not reported	2 NR		NR	Day(s)	LC50	Mortality	Mortality
		Not reported	2 NR		NR		LC50	Mortality	Mortality
767298 C	CA:MG MOLAR RATIO 0.7	Not reported	2 NR		NR		LC50	Mortality	Mortality
18452		Not reported	3 NR			Day(s)	LC50	Mortality	Mortality
18147		Not reported	2 NR				LC50	Mortality	Mortality
18453		Not reported	4 NR				LC50	Mortality	Mortality
18451		Not reported	2 NR		NR S	Dav(s)	LC50	Mortality	Mortality
18450		Not reported	1 NR				LC50	Mortality	Mortality
18146		Not reported	1 NR				LC50	Mortality	Mortality
759092		Not reported	2 NR		NR		LC50	Mortality	Mortality
767297		Not reported	2 NR				LC50	Mortality	Mortality
767305 CA	CA:MG MOLAR RATIO 1.8	Not reported	2 NR		NR		LC50	Mortality	Mortality
_	CA:MG MOLAR RATIO 0.7	Not reported	2 NR				LC50	Mortality	Mortality
160332		Not reported	1 NR				LC50	Mortality	Mortality
110333		Not reported	NR NR				LC20	Mortality	Mortality
31687		Not reported	1 NR		Z.R	Day(s)	1050	Mortality	Mortality
759090		Not reported	2 NR				LOEC	Mortality	Mortality
759091		Not reported	2 NR				NOEC	Mortality	Mortality
767473 FE	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1 NR		NR I	Day(s)	NR-LETH	Mortality	Mortality
767496 FE		Not reported	1 NR			Day(s)	NR-LETH	Mortality	Mortality
		Not reported	1 NR				NR-LETH	Mortality	Mortality
767491 FE	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	1 NR		N.R.	Day(s)	NR-LETH	Mortality	Mortality
		Not reported	2 NR				NR-ZERO	Mortality	Mortality
		Not reported	2 NR				NR-ZERO	Mortality	Mortality
		Not reported	6 NR				LC50	Mortality	Mortality
2064204		Not reported	1 NR				LC50	Mortality	Mortality
2064206		Not reported	4 NR				LC50	Mortality	Mortality
2064205		Not reported	2 NR		NR [	Day(s)	LC50	Mortality	Mortality
2017113 4 6	4 REPLICATES	Not reported	4 NR		NR	Day(s)	EC10	Mortality	Survival
2017143 4 F	4 REPLICATES	Not reported	4 NR		NR [		EC25	Mortality	Survival
	4 REPLICATES	Not reported	4 NR			Day(s)	EC50	Mortality	Survival
766367		Not reported	4 NR			Day(s)	LC10	Mortality	Mortality
766638 CA		Not reported	4 NR			Day(s)	LC50	Mortality	Mortality
766630 CA		Not reported	4 NR				LC50	Mortality	Mortality
766713 CA	CA:MG RATIO 2.33	Not reported	4 NR			Day(s)	LC50	Mortality	Mortality
766712 CA	766712 CA:MG RATIO 2.33	Not reported	4 NR		ND		CEO	200+0 +0	

	Mortality	Mortality	LC50	Day(s)	NR I	NR		Not reported	6215	
				2.5 %						
	Mortality	Mortality	1050	Dav(s)			2	Not reported	6216	
	Mortality	Mortality	LC50	Day(s)	NR	NR	4	Not reported	6214	
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported	41640	4
	Mortality	Mortality	LC50	Day(s)	NR	NR	4	Not reported	41642	4
	Mortality	Mortality	LC50	Day(s)		NR	w	Not reported	6217	
	Mortality	Mortality	LC50	Day(s)	NR	NR	w	Not reported	41641	4
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	41636	4
	Mortality	Mortality	LC50	Day(s)			1	Not reported	6211	
	Mortality	Mortality	LC100	Day(s)			w	Not reported	802673 2 REPLICATES	80
	Mortality	Mortality	LC100	Day(s)		NR	4	Not reported		80
	Mortality	Mortality	LC100	Day(s)		NR	2	Not reported	802672 2 REPLICATES	80
	Mortality	Mortality	LC100	Day(s)		NR	2	Not reported	802669 2 REPLICATES	80
	Mortality	Mortality	LC100	Day(s)		NR	4	Not reported	802667 2 REPLICATES	80
	Mortality	Mortality	LC100	Day(s)			1	Not reported		80
	Mortality	Mortality	LC100	Day(s)			1	Not reported	802670 2 REPLICATES	80
	Mortality	Mortality	LC100	Day(s)			ω ω	Not reported	12	80
	Mortality	Mortality	LC0	Day(s)			4	Not reported	802666 2 REPLICATES	80
	Mortality	Mortality	LC0	Day(s)			3	Not reported	802676 2 REPLICATES	80
	Mortality	Mortality	TC0	Day(s)		NR	1	Not reported	802663 2 REPLICATES	80
	Mortality	Mortality	LC0	Day(s)			1	Not reported	802678 2 REPLICATES	80
	Mortality	Mortality	LC0	Day(s)			3	Not reported	802665 2 REPLICATES	80
	Mortality	Mortality	LC0	Day(s)		NR	2	Not reported	802664 2 REPLICATES	80
	Mortality	Mortality	LC0	Day(s)	NR	NR	2	Not reported	802677 2 REPLICATES	80
	Mortality	Mortality	LC0	Day(s)	NR	4 NR	4	Not reported	802675 2 REPLICATES	80
	Mortality	Mortality	NR-LETH	Day(s)	NR	NR	1	Not reported	764588	76
MSMT/TLM///	Mortality N	Mortality	LC50	Day(s)		NR	1	Not reported	18149	
MSMT/TLM///		Mortality	LC50	Day(s)			4	Not reported	18152	
MSMT/TLM///		Mortality	LC50	Day(s)		NR	ω	Not reported	18151	
MSMT/TLM///		Mortality	LC50	Day(s)			2	Not reported	18150	
	Mortality	Mortality	LC50	Day(s)			4	Not reported	13415 8 PARAMETERS, SEE PAPER	
	Mortality	Mortality	LC50	Day(s)			4	Not reported	40508 COMPOSITION OF WATER REPORTED	7
	Mortality	Mortality	LC50	Day(s)			4	Not reported	40509 COMPOSITION OF WATER REPORTED	7
	Survival	Mortality	LC50	Day(s)			4	Not reported	200196	20
MSMT/TLM///		Mortality	LC50	Day(s)			4	Not reported	126070	17
MSMT/TLM///		Mortality	LC50	Day(s)			4	Not reported	126072	12
MSMT/TLM///		Mortality	LC50	Day(s)			4	Not reported	126071	17
MSMT/TLM///		Mortality	LC50	Day(s)			1	Not reported		
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		76
	Mortality	Mortality	LC50	Day(s)	NR		2	Not reported		76
	Mortality	Mortality	LC50	Day(s)		NR	2	Not reported		76
	Mortality	Mortality	LC50	Day(s)			4	Not reported		76
	Mortality	Mortality	LC50	Day(s)			4	Not reported		76
	Mortality	Mortality	LC50	Day(s)			4	Not reported		76
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	766658 CA:MG RATIO 2.33	76
	Mortality	Mortality	LC50	Day(s)		NR	.4	Not reported		76
	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported		76
	Mortality	Mortality	LC50	Day(s)		NR	4	Not reported	766659 CA:MG RATIO 2.33	76
	Mortality	Mortality	NR-LETH	Day(s)		NR	4	Not reported	766440	76
	Survival	Mortality	NOEC	Day(s)	NR	NR	3	Not reported	766445	76
	Survival	Mortality	NOEC	Day(s)	NR	NR	2	Not reported	766444	76
	Survival	Mor	NOEC	Day(s)		NR	4	Not reported	4 REPLICATES	20:
EE Comment	Measurement	Effect*	Endpoint	(Days)	(Days)	Min (Days)	Mean (Days)	Response Site	Result Number Experimental Design	ult Nu
				Observed	Observed	Observed	Observed			

Result Number	Experimental Design	Response Site	Observed Observed Duration Duration Mean (Days) Min (Days)	Observed Duration Max (Davs)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
41635	0	Not reported	_	NR	Day(s)	LC50	Mortality	Mortality	
6212		Not reported	2 NR	NR	Day(s)	LC50	Mortality	Mortality	
6213		Not reported	3 NR	NR	Day(s)	LC50	Mortality	Mortality	
41637		Not reported	3 NR	NR	Day(s)	LC50	Mortality	Mortality	
41638		Not reported	4 NR	NR	Day(s)	LC50	Mortality	Mortality	
41639		Not reported	1 NR	NR	Day(s)	LC50	Mortality	Mortality	
		Not reported	4 NR	NR	Day(s)	LC50	Mortality	Mortality	
2017209	3 REPLICATES	Not reported	7 NR	N. N.	Day(s)	EC10	Mortality	Survival	
20171/8	2017178 3 REPLICATES	Not reported	7 NR	NR R	Day(s)	FC10	Mortality	Survival	
2017168	2017168 3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC10	Mortality	Survival	
2017210	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC25	Mortality	Survival	
2017179	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC25	Mortality	Survival	
2017189	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC25	Mortality	Survival	
2017169	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC25	Mortality	Survival	
2017211	2017211 3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC50	Mortality	Survival	
		Not reported	7 NR	NR	Day(s)	EC50	Mortality	Survival	
2017190	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC50	Mortality	Survival	
2017170	2017170 3 REPLICATES	Not reported	7 NR		Day(s)	EC50	Mortality	Survival	
160335		Not reported	2 NR	NR R	Day(s)	LC50	Mortality	Mortality	
160334		Not reported	1 NR		Day(s)	LC50	Mortality	Mortality	
160336		Not reported	4 NR	N <sub>R</sub>	Day(s)	LC50	Mortality	Mortality	
	3 REPLICATES	Not reported	7 NR		Day(s)	LOEC	Mortality	Survival	
2017182	3 REPLICATES	Not reported	7 NR	NR	Day(s)	LOEC	Mortality	Survival	
2017172	3 REPLICATES	Not reported	7 NR		Day(s)	LOEC	Mortality	Survival	
2017192	2017192 3 REPLICATES	Not reported	7 NR		Day(s)	LOEC	Mortality	Survival	
761598	111111111111111111111111111111111111111	Not reported	7 NR		Day(s)	NOEC	Mortality	Mortality	
701/191	201/191 3 REPLICATES	Not reported	/ NR		Day(s)	NOEC	Mortality	Survival	
2017712	מ הרהונה דרה	Not reported	1 NX		Day(s)	NOEC	Mortality	SULVIVAI	
2017101	3 REPLICATES	Not reported	7 NR		Day(s)	NOEC	Mortality	Survival	
2017171	2017171 3 REPLICATES	Not reported	7 NR	NR S	Day(s)	NOFC	Mortality	Survival	
18153		Not reported	2 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
18148		Not reported	1 NR		Day(s)	LC50	Mortality	Mortality	MSMT/TLM///
		Not reported	4 NR		Day(s)	LC10	Mortality	Mortality	MSMT/MEAN OF THREE TESTS/
	CA:MG RATIO 2.33	Not reported	4 NR		Day(s)	LC50	Mortality	Mortality	MSMT/MEAN OF 2 TESTS/
766363	CHING RATIO 1.40	Not reported	4 NR	NB	Day(s)	1050	Mortality	Mortality	MSMT/MEAN OF THREE TESTS/
766707	766707 CA:MG RATIO 5.4	Not reported	4 NR		Dav(s)	LC50	Mortality	Mortality	MSMT/MEAN OF 2 TESTS/
766703	CA:MG RATIO 1.46	Not reported	4 NR		Day(s)	LC50	Mortality	Mortality	MSMT/MEAN OF 3 TESTS/
766704	CA:MG RATIO 1.46	Not reported	4 NR		Day(s)		Mortality	Mortality	MSMT/MEAN OF 2 TESTS/
766708 (	CA:MG RATIO 1.46	Not reported	4 NR	NR	Day(s)	LC50	Mortality	Mortality	MSMT/MEAN OF 3 TESTS/
160500		Not reported	2 NR		Day(s)		Mortality	Mortality	
160498		Not reported	2 NR		Day(s)		Mortality	Mortality	
767797	777	Not reported	1 NR		Day(s)		Mortality	Mortality	
767360 F	767360 EED MODERATELY HARD RECONSTITUTED WATER	Not reported	2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
767399 L	UNFED, MODERATELY HARD RECONSTITUTED WATE	Not reported	NR 1	2	Day(s)		Mortality	Mortality	
	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	
767462 F	FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	2 NR			ZERO	Mortality	Mortality	
/6/31/1	/6/31/ FED, MODERATELY HARD RECONSTITUTED WATER	Not reported	2 NR	NR	Day(s)	NR-ZERO	Mortality	Mortality	

	Biomass	Growth	LOEC	Day(s)	NA IC	NA IN	/	ואטרובטטונבט		100001
	Biomass	Growth					1 /	Not reported		250000
	Biomass	Growth					7	Not reported		UC1097
	Biomass	Growth		Day(s)			33	Whole organism	5 REPLICATES	2152753
	Biomass	Growth		Day(s)			33	Whole organism	5 REPLICATES	
	Biomass	Growth	IC25	Day(s)			7	Not reported	1 7 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	
	Biomass	Growth	IC25	Day(s)	NR E	NR	7	Not reported		761554
	Biomass	Growth		Day(s)		NR	7	Not reported		761562
	Biomass	Growth		Day(s)		NR 1	7	Not reported		761551
	Biomass	Growth	IC25	Day(s)	NR I	NR	7	Not reported		761557
	Biomass	Growth	IC25	Day(s)		NR	7	Not reported		761561
	Biomass	Growth	IC25	Day(s)		NR I	7	Not reported		761569
	Biomass	Growth	IC25	Day(s)	NR [	NR I	7	Not reported		761566
	Biomass	Growth	IC25	Day(s)		NR I	7	Not reported		761553
	Biomass	Growth	IC25	Day(s)		NR I	7	Not reported		761565
	Biomass	Growth	IC25	Day(s)	NR	NR	7	Not reported		761558
	Biomass	Growth	IC25	Day(s)			7	Not reported		761548
	Biomass	Growth	NOEC	Day(s)		NR I	28	Not reported	4 REPLICATES	2152818
	Biomass	Growth	LOEC	Day(s)			28 NR	Not reported	4 REPLICATES	2152819
	Biomass	Growth		Day(s)			28 NR	Not reported		2152817
	Biomass	Growth	IC25	Day(s)	NR		28 NR	Not reported	4 REPLICATES	2152816
MSMT/DRY WEIGHT/		Growth		Day(s)			28	Whole organism	4 TO 6 REPLICATES, CONDUCTED IN STANDARD ART	2164559
	Length	Growth	NOEC	Day(s)			21 NR	Not reported		201555
MSMT/CHRONIC VALUE///	Length	Growth	MATC	Day(s)			21	Not reported		201557
	Length	Growth		Day(s)			21	Not reported		201556
	Biomass	Growth	NOEC	Day(s)			20	Not reported	4 REPLICATES	2152814
	Biomass	Growth	LOEC	Day(s)			20	Not reported		2152815
	Biomass	Growth	IC50	Day(s)			20	Not reported	4 REPLICATES	2152813
	Biomass	Growth	IC25	Day(s)	NR	20 NR	20	Not reported	4 REPLICATES	2152812
	Biomass	Growth	IC10	Day(s)		20 NR	20	Not reported	4 REPLICATES	2152811
	Weight	Growth	NOEC	Day(s)	NR	7 NR	7	Not reported		761769
	Weight	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		761768
MSMT/TLM///	Mortality	Mortality	LC50	Day(s)			1	Not reported		67862
MSMT/TLM///	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported		67863
		Mortality	NR-ZERO	Day(s)		NR	4	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	767515
	Mortality	Mortality	NR-ZERO	Day(s)			4	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	767512
	Mortality	Mortality	LC50	Day(s)	NR	NR	1	Not reported		160501
	Mortality	Mortality	LC50	Day(s)	NR	2 NR	2	Not reported		160502
	Mortality	Mortality	LC50	Day(s)		4 NR	4	Not reported		160503
	Mortality	Mortality	LC50	Day(s)	NR	NR	4	Not reported	8 PARAMETERS, SEE PAPER	26240
		Mortality	LC50	Day(s)		4 NR	4	Not reported		196198
ENDPT/TLM/		Mortality	LC50	Day(s)	NR	NR	4	Not reported		2063750
ENDPT/TLM/	Mortality	Mortality	LC50	Day(s)	NR	NR	2	Not reported	9	2063749
ENDPT/TLM/		Mortality	LC50	Day(s)			1	Not reported		2063748
MSMT/REPLICATE B///	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	21	Not reported		759465
MSMT/REPLICATE A///		Mortality	NR-ZERO	Day(s)	NR	21 NR	21	Not reported	2	759462
	Mortality	Mortality	NR-ZERO	Day(s)	NR	2 NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	767484
	Mortality	Mortality	NR-ZERO	Day(s)	NR	NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	767469
	Mortality	Mortality	NR-ZERO	Day(s)	NR	2 NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	767467
	,	Mortality	NR-ZERO	Day(s)	NR	NR	2	Not reported	FED, MODERATELY HARD RECONSTITUTED WATER	767483
MSMT/F/F2 SURVIVAL/	Survival	Mortality	NOEC	Day(s)	NR	NR	42	Not reported	1 CACL2 FOR HARDNESS CONTROL	759021
	Mortality	Mortality	LC50	Day(s)			1	Not reported		160499
EE Comment	Effect Measurement	Effect*	Endpoint	Duration Units (Days)	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	er Experimental Design	Result Number
				Observed	Observed	Chacived	Observed			

	Biomass	Growth	NOEC	Day(s)	NR	NR	/	Not reported		768071
		) (	1000				1	C		
	Biomass	Growth	NOEC				33 NR	Whole organism	2152754 5 REPLICATES	2152754
	Biomass	Growth	NOEC	Day(s)	NR D	NR	7	Not reported		768140
	Biomass	Growth	NOEC	Day(s)	NR D	NR 7	7	Not reported		768077
	Biomass	Growth	NOEC	Day(s)	NR	NR	7	Not reported		768085
	Biomass	Growth	NOEC	Day(s)	NR D	NR	7	Not reported		768160
	Biomass	Growth	NOEC	Day(s)	NR D	NR 7	7	Not reported		768079
	Biomass	Growth	NOEC	Day(s)	NR D	NR 1	7	Not reported		768150
	Biomass	Growth	NOEC	Day(s)	NR D	NR 7	7	Not reported		768095
	Biomass	Growth	NOEC	Day(s)	NR C	NR I	7	Not reported		768087
	Biomass	Growth	NOEC	Day(s)	NR D	NR I	7	Not reported		768162
	Biomass	Growth	NOEC	Day(s)	NR	NR I	7	Not reported		768142
	Biomass	Growth	NOEC	Day(s)			7	Not reported		768112
	Biomass	Growth	NOEC	Day(s)			7	Not reported		768093
	Biomass	Growth	NOEC	Day(s)			7	Not reported		768130
	Biomass	Growth	NOEC	Day(s)			7	Not reported		/68152
	Biomass	Growth	NOEC				7	Not reported		/615/1
	Biomass	Growth	NOEC		NR		/	Not reported		/61563
	Biomass	Growth	NOEC				7	Not reported		462556
	Biomass	Growth	NOEC				1	Not reported		201550
	Biomass	Growth	NOEC				7	Not reported		761560
	Biomass	Growth	NOEC				7	Not reported		761554
	Biomass	Growth	NOEC				7	Not reported		/6155U
	Biomass	Growth	NOEC	Day(s)	NR		7	Not reported		761555
	Biomass	Growth	NOEC	Day(s)		NR	7	Not reported		761560
	Biomass	Growth	NOEC	Day(s)			7	Not reported		761567
	Biomass	Growth	NOEC	Day(s)	NR		7	Not reported		761549
	Biomass	Growth	LOEL	Day(s)		NR	7	Not reported	3 HOUR PULSE DURATION	741780
	Biomass	Growth	LOEL	Day(s)			7	Not reported	96 HOUR PULSE DURATION	741831
	Weight	Growth	LOEC	Day(s)	NR I	NR	7	Not reported		761764
	Biomass	Growth	LOEC	Day(s)	NR	NR	7	Not reported		768062
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768060
	Biomass	Growth	LOEC	Day(s)	NR I	NR	7	Not reported		768100
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768121
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768131
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768102
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768109
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768119
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768068
	Biomass	Growth	LOEC	Day(s)		NR	7	Not reported		
	Biomass	Growth	LOEC	Day(s)		NR	33	Whole organism	5 REPLICATES	2152755
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768139
	Biomass	Growth	LOEC	Day(s)			7	Not reported		768076
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768084
	Biomass	Growth	LOEC	Day(s)		NR	7	Not reported		768159
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768078
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768149
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768094
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768086
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768161
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768141
	Biomass	Growth	LOEC	Day(s)	NR	7 NR	7	Not reported		768111
EE Comment	Measurement	Effect*	Endpoint	(Days)	(Days)	Min (Days)	Mean (Days)	Response Site	Experimental Design	Result Number
				Observed	Observed		Observed			

represent of principal (Not the Out of the Modernor									T T T T T T T T T T T T T T T T T T T	747007
MT/ALITHOR REPORTS REPRODUCTION	Reproducing orgaMSI	Reproduction	NOEC	Day(s)	NR [	NR	7	Not reported	TEST 9	741942
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orgaMSI	Reproduction	NOEC	Day(s)	NR I	NR	7	Not reported	EST 10	
Reproducing orga MSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orgaMSI		NOEC	Day(s)			7	Not reported	TEST 3	741649
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION (	Reproducing orga MSI	Reproduction	NOEC	Day(s)			7	Not reported	EST 1	741648 TEST 1
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orga MSI	Reproduction	NOEC	Day(s)	NR	NR	7	Not reported	TEST 12	742901 1
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orgaMSI	Reproduction	NOEC	Day(s)	NR I	NR	7	Not reported	TEST 6	742893 1
Reproducing orga MSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orga MSI	Reproduction	NOEC	Day(s)	NR [	NR	7	Not reported	EST 2	742891 TEST 2
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orga MSI		NOEC	Day(s)	NR [	NR	7	Not reported	TEST 5	741937
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orgaMSI	Reproduction	NOEC	Day(s)		NR	7	Not reported	TEST 8	742898 1
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orgaMSI	Reproduction	NOEC	Day(s)	NR	NR	7	Not reported	EST 11	741953 TEST 11
Reproducing orgaMSMT/AUTHOR REPORTS REPRODUCTION	Reproducing orga MSI	X.	NOEC	Day(s)		NR	7	Not reported	TEST 4	742896 1
pers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR		7	Not reported		761581
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR	7	Not reported		761593
pers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported		761573
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported		761589
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported		761577
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported		761585
Progeny counts/n/MSMT/FOUND IN ~10% OF LABS///	Progeny counts/nMS	Reproduction	NOEC	Day(s)			7	Not reported	TEST MATERIAL 3	251021
Progeny counts/nMSMT/FOUND BETWEEN 10 AND 20% OF L	Progeny counts/rMS	Reproduction	NOEC	Day(s)			7	Not reported	TEST MATERIAL 1	
Progeny counts/n MSMT/FOUND BETWEEN 30 AND 40% OF L	Progeny counts/rMS	Reproduction	NOEC	Day(s)			7	Not reported	TEST MATERIAL 1	251015
Progeny counts/nMSMT/FOUND BETWEEN 40 AND 50% OF L	Progeny counts/nMS	Reproduction	NOEC	Day(s)			7	Not reported	TEST MATERIAL 1	251013
Progeny counts/nMSMT/FOUND BETWEEN 10 AND 20% OF L	Progeny counts/r MS	Reproduction	NOEC	Day(s)		NR	7	Not reported	TEST MATERIAL 1	120856
Progeny counts/nMSMT/FOUND IN ~20% OF LABS///	Progeny counts/nMS	Reproduction	NOEC	Day(s)		7 NR	7	Not reported	251017 TEST MATERIAL 3	251017
Progeny counts/nMSMT/FOUND BETWEEN 60 AND 70% OF L	Progeny counts/nMS	Reproduction	NOEC	Day(s)	NR	7 NR	7	Not reported	TEST MATERIAL 3	251023
Progeny counts/dMSMT/FOUND IN ~10% OF LABS///	Progeny counts/nMS	Reproduction	NOEC	Day(s)			7	Not reported	TEST MATERIAL 3	251019
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR	7	Not reported		767974
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported		767966
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		7 NR	7	Not reported		767900
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR	7	Not reported		767964
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR	7	Not reported		767886
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		7 NR	7	Not reported		767892
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR	7	Not reported		766871
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR	7	Not reported		767867
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		7 NR	7	Not reported		766863
hers	Progeny counts/numbers	Reproduction	NOFC	Day(s)		NR		Not reported		767851
hers	Progeny counts/numbers	Reproduction	NOFC	Day(s)		7 NR		Not reported		767956
bers	Progeny counts/numbers	Reproduction	NOEC	Dav(s)		7 NR		Not reported		767853
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR		Not reported		761590
borr	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NB	NB		Not reported		761574
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR		Not reported		761506 07CT07
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR		Not reported		761579
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR		Not reported	CHRONIC -3 BROOD STDY	
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR NR		Not reported		
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR		Not reported		767877
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR NR		Not reported		767894
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR NR		Not reported		767875
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR NR		Not reported		766869
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR NR		Not reported		767869
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		7 NR		Not reported		767861
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		7 NR		Not reported		761582
EE Comment	Effect Measurement	Effect*	Endpoint	Duration Units (Days)	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	Experimental Design	Result Number
				02000		1	7.70 97 0 4-7			

INNEE BIOCOD STOY	Result Number	r Experimental Design	Response Site	Observed Duration Mean (Days)	Observed Duration Min (Days)	Observed Duration Max (Days)	Observed Duration Units (Days)	Endpoint	Effect*	Effect Measurement	EE Comment
	2152726		Not reported	9	NR		Day(s)	NOEL	Reproduction	Progeny counts/numb	
Mol Report	741566		Not reported	9	NR	j	Day(s)	NOEL	Reproduction	Progeny counts/numb	ers
CHRENNIC_3 BRODD SIDY   Mot reported   10 NR   NR   Day(s)   Egg. Reproduction	76175	2	Not reported	7	NR		Day(s)	LOEC	Reproduction	Fecundity	
CHRONIC 3 BRODD STDY    Mot reported   10 NR   NR   Day(s)   Day(s)   ECS0   Reproduction   CHRONIC 3 BRODD STDY   Mot reported   10 NR   NR   Day(s)   Day(s)   ECS0   Reproduction   CHRONIC 3 BRODD STDY   Mot reported   10 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   10 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   Mot reported   21 NR   NR   Day(s)   ECS0   Reproduction   CREENICATES   NR   Reported   END   CREENICATES   CREENICATES   NR   REPORTED   END   CREENICATES   CREENICATES   CREENICATES   END   CREENICATES   CREENICATES   END   CREENICATES   CREENICATES   END   CREENICATES   END	76175		Not reported		NR		Day(s)	NOEC	Reproduction	Fecundity	
CHRONIC 3 BRODD STOY   Not reported   10 NR   NR   Day(s)   CUEC   Reproduction   CHRONIC 3 BRODD STOY   Not reported   10 NR   NR   NR   Day(s)   CUEC   Reproduction   CHRONIC 3 BRODD STOY   Not reported   10 NR   NR   Day(s)   CECD   Reproduction   Not reported   10 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   Not reported   21 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   Not reported   21 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   NR reported   21 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   NR reported   21 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   NR reported   21 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   NR reported   21 NR   NR   Day(s)   CECD   Reproduction   CHRONIC 3 REPRODUCTION   NR REPORTED   CHRONIC 3	741654		Not reported	10	NR		Day(s)	EC50	Reproduction	Progeny counts/numb	ers
	741696	6 CHRONIC -3 BROOD STDY	Not reported	10	NR		Day(s)	LOEC	Reproduction	Progeny counts/numb	ers
	741659	9 CHRONIC -3 BROOD STDY	Not reported	10	NR		Day(s)	NOEC	Reproduction	Progeny counts/numb	ers
DREPICATES   Not reported   21 NR	741530		Not reported	10	NR		Day(s)	ECS0	Reproduction	Progeny counts/numb	ers
DREPUCATES   Motreported   Z1 NR NR   Day(s)   C55   Regroduction   DREPUCATES   Motreported   Z1 NR NR   Day(s)   C55   Regroduction   DREPUCATES   Motreported   S1 NR NR   Regroduction   Regroduction   Motreported   S1 NR NR   Regroduction   Regroduc	2152713		Not reported	10	NR		Day(s)	EC50	Reproduction	Progeny counts/numb	ers
	2152732	_	Not reported	21	NR		Day(s)	IC25	Reproduction	Reproduction, general	
	2152733		Not reported	21	NR			IC50	Reproduction	Reproduction, general	
DREPUCATES   Not reported   5 NR   NR   Brood or litter   LOEC   Reproduction   DREPUCATES   Not reported   3 NR   NR   Brood or litter   LOEC   Reproduction   DREPUCATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   DREPUCATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   DREPUCATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   DREPUCATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   DREPUCATES   NR   NR   Brood or litter   NR   LOEC   Reproduction   NR   REPRODUCTION   NR   NR   Brood or litter   NR   NR   REPRODUCTION   NR   REPRODUCTION   NR   NR   REPRODUCTION   NR   REPRODUCTI	2164766		Not reported	6	NR			LOEC	Reproduction	Fecundity	
DREPLICATES   Not reported   4 NR   NR   Brood or litter   LOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   LOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   5 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   5 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   5 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   Not reported   2 NR   NR   Brood or litter   NOEC   Reproduction   10 REPLICATES   NR   NR   Day(s)   NOEC   Reproduction   NR   REPLICATES   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   NR   NR   Day(s)   NOEC   Reproduction   NR   REPLICATES   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   NR   NR   Day(s)   NOEC   Reproduction   NR   REPLICATES   NR   Reproduction   NR   Reproduction   NR   Reproduction   NR   Reproduction   NR   Reproduction	2164767		Not reported	5	NR		Ť	LOEC	Reproduction	Fecundity	
DREPUCATES   Not reported   2 NR   NR   Brood orliter   LOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   LOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   LOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   LOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   LOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Brood orliter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Brood orliter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Brood orliter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Brood orliter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Brood orliter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Brood orliter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPORTED   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPORTED   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPORTED   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPORTED   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   10 REPORTED   NOEC   Reproduction   Not reported   Not reported   NR   NR   Day(s)   NOEC   Reproductio	2164768		Not reported	4	NR			LOEC	Reproduction	Fecundity	
DR REPUCATES   Not reported   21 NR	2164769		Not reported	3	NR			LOEC	Reproduction	Fecundity	
DREPUCATES   Not reported   21 NR	2164770	0 10 REPLICATES	Not reported	2	NR			LOEC	Reproduction	Fecundity	
10 REPUCATES   Not reported   10 NR   NR   Day(s)   DEC   Reproduction   10 REPUCATES   Not reported   10 NR   NR   Broad or litter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Broad or litter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Broad or litter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Broad or litter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Broad or litter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Broad or litter   NOEC   Reproduction   10 REPUCATES   Not reported   2 NR   NR   Day(s)   NOEC   Reproduction   Not reported   NOE reported   NOE Reproduction   Not reported   NOE Rep	2164728	8 10 REPLICATES	Not reported	21	NR			LOEC	Reproduction	Fecundity	
DREPUCATES   Not reported   S NR   NR   Day(s)   DEL   Reproduction   ID REPUCATES   Not reported   S NR   NR   Brood or litter   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Brood or litter   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Brood or litter   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Brood or litter   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Brood or litter   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Brood or litter   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Day(s)   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Day(s)   NOEC   Reproduction   ID REPUCATES   Not reported   A NR   NR   Day(s)   NOEC   Reproduction   Not reported   A NR   NR   Day(s)   N	2152735		Not reported	21	NR			LOEC	Reproduction	Reproduction, general	
DREPUCATES   Not reported   S   NR   NR   Broad or litter   NOEC   Reproduction   DREPUCATES   Not reported   4   NR   NR   Broad or litter   NOEC   Reproduction   DREPUCATES   Not reported   4   NR   NR   Broad or litter   NOEC   Reproduction   Not reported   3   NR   NR   Broad or litter   NOEC   Reproduction   Not reported   3   NR   NR   Broad or litter   NOEC   Reproduction   Not reported   3   NR   NR   Broad or litter   NOEC   Reproduction   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   Not reported   22   NR   NR   Day(s)   NOEC   Reproduction   Not reported   23   NR   NR   Day(s)   NOEC   Reproduction   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   N	2152715	5	Not reported	10				LOEL	Reproduction	Progeny counts/numb	ers
INBEPLICATES   Not reported   2   NR   NR   Broad or litter   NOEC   Reproduction   INDEPLICATES   Not reported   6   NR   NR   Broad or litter   NOEC   Reproduction   INDEPLICATES   Not reported   5   NR   NR   Broad or litter   NOEC   Reproduction   INDEPLICATES   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   INDEPLICATES   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   INDEPLICATES   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   Not reported   22   NR   NR   Day(s)   NOEC   Reproduction   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   Not reported   27   NR   NR   Day(s)   NOEC   Reproduction   Not reported   7   NR   NR   Day(s)   ECIO   Reproduction   Not reported   7   NR   NR   Day(s)   ECIO	2164764		Not reported	5	NR			NOEC	Reproduction	Fecundity	
Not reported   A NR   NR   Broad or litter   NOEC   Reproduction   Not reported   A NR   NR   Broad or litter   NOEC   Reproduction   Not reported   A NR   NR   Broad or litter   NOEC   Reproduction   Not reported   A NR   NR   Broad or litter   NOEC   Reproduction   Not reported   A NR   NR   Broad or litter   NOEC   Reproduction   Not reported   A NR   NR   Day(s)   NR   NR   Day(s)   NOEC   Reproduction   Not reported   A NR   NR   Day(s)   NOEC   Reproduction   NR   NR   Day(s)   NOEC   Reproduction   NR   NR   Day(s)   NOEC   Reproduction   NR   NR   Day(s)   NR	2164761		Not reported	2				NOEC	Reproduction	Fecundity	
	2164763		Not reported	4				NOEC	Reproduction	Fecundity	
IOREDICATES   Not reported   21 NR	2164765		Not reported	6				NOEC	Reproduction	Fecundity	
	2164762		Not reported	3				NOEC	Reproduction	Fecundity	
INREPLICATES   Not reported   21 NR	2164725		Not reported	21				NOEC	Reproduction	Fecundity	
ITHREE BROOD TEST   Not reported   10 NR   NR   Day(s)   NOEL   Reproduction	2152734		Not reported	21				NOEC	Reproduction	Reproduction, general	
Not reported   21 NR   NR   Day(s)   NOEL   Reproduction	741546		Not reported	10				NOEL	Reproduction	Progeny counts/numb	ers
Not reported   21 NR	2152714	4	Not reported	10				NOEL	Reproduction	Progeny counts/numb	ers
Not reported   21 NR	201548		Not reported	21				LOEC	Reproduction	Fecundity	
Not reported   21   NR   NR   Day(s)   MATC   Reproduction   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   Not reported   21   NR   NR   Day(s)   NOEC   Reproduction   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   REPLICATES   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   Not reported   28   NR   NR   Day(s)   NOEC   Reproduction   Not reported   7   NR   NR   Day(s)   EC10   Reproduction   Not reported   7   NR   NR   Day(s)   EC25   Reproduction   NR   NR   Day(s)	201551		Not reported	21				LOEC	Reproduction	Progeny counts/numb	ers
Not reported   21 NR	201549	9	Not reported	21				MATC	Reproduction		NT/CHRONIC VALUE///
REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           Not reported         21 NR	201552	2	Not reported	21				MATC	Reproduction	Progeny counts/nMSN	1T/CHRONIC VALUE///
Not reported   21 NR	201547	7	Not reported	21				NOEC	Reproduction	Fecundity	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR	201550		Not reported	21				NOEC	Reproduction	Progeny counts/nMSN	IT/CHRONIC VALUE///
4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC10         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR	2152783	4 REPLICATES	Not reported	28				IC25	Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC10         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Da	2152784		Not reported	28				IC50	Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC10         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LC5C         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LC5C         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LC5C         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LC5C         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         7 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day	2152786	4 REPLICATES	Not reported	28					Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC10         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day	2152785	4 REPLICATES	Not reported	28					Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NNR         Day(s)         IC25         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         7 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day	2152796	4 REPLICATES	Not reported	28 1				IC10	Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         IC50         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         7 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2152807	4 REPLICATES	Not reported	28 1				IC25	Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         LOEC         Reproduction           4 REPLICATES         Not reported         28 NR         NR         Day(s)         NOEC         Reproduction           4 REPLICATES         Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)	2152808	4 REPLICATES	Not reported	28 1				IC50	Reproduction	Reproduction, general	
4 REPLICATES         Not reported         28 NR         NR         Day(s)         NOEC         Reproduction           Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2152810	4 REPLICATES	Not reported	28 1					Reproduction	Reproduction, general	
Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2152809	4 REPLICATES	Not reported	28 1					Reproduction	Reproduction, general	
Not reported         21 NR         NR         Day(s)         NOEC         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	759472		Not reported	21		7		NOEC	Reproduction	Progeny counts/nMSN	IT/REPLICATE B///
Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	759471		Not reported	21 1					Reproduction	Progeny counts/nMSN	IT/REPLICATE A///
Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2017036		Not reported	7 7					Reproduction	Progeny counts/numbe	SLS
Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2017047		Not reported	7 7					Reproduction	Progeny counts/numbe	SIS
Not reported         7 NR         NR         Day(s)         EC10         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2017055		Not reported	7 7					Reproduction	Progeny counts/numbe	ers
Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2017064		Not reported	7.1					Reproduction	Progeny counts/numbe	ers
Not reported         7 NR         NR         Day(s)         EC25         Reproduction           Not reported         7 NR         NR         Day(s)         EC25         Reproduction	2017035		Not reported	717				EC25	Reproduction	Progeny counts/numbe	ers
Not reported / NR NR Day(s) EC25 Reproduction	201/046		Not reported	717					Reproduction	Progeny counts/number	ers
	707705		INOTEPHOTEC				12///21	/5	E DOTO CITICO	Drogany collete/allmag	100

DESCRIPTION TO SIGNIFICANT INCREA				17/17/				000000000000000000000000000000000000000	751707
Progeny counts/nMSMT/STATISTICALLY SIGNIFICANT INCREA	Progeny counts/nMS	Reproduction	NOEC	Day(s)	NR	NR	21	Not reported	759463
Progeny counts/r MSMT/F1 AND F2 CUMULATIVE NUMBER/	Progeny counts/nMS	Reproduction		Day(s)	NR	NR	42 NR	Not reported	759022 CACL2 FOR HARDNESS CONTROL
Progeny counts/nMSMT/STATISTICALLY SIGNIFICANT INCREA	Progeny counts/n MS	Reproduction			NR	NR	21	Not reported	759464
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR	NR	14	Not reported	2017150 4 REPLICATES, SEDIMENT
bers	Progeny counts/numbers	Reproduction	LOEC	Day(s)	NR	NR	14	Not reported	2017151 4 REPLICATES, SEDIMENT
bers	Progeny counts/numbers	Reproduction		Day(s)			14	Not reported	2017149 4 REPLICATES, SEDIMENT
bers	Progeny counts/numbers	Reproduction		Day(s)			14	Not reported	
bers	Progeny counts/numbers	Reproduction				14 NR	14	Not reported	2017147 4 REPLICATES, SEDIMENT
	Fecundity	Reproduction					7	Not reported	761751
ā	Fecundity	Reproduction		Dav(s)				Not reported	761750
s (general)	Reproductive success (general)	Reproduction	NOEC	Brood or litter			3	Not reported	759075
bers	Progeny counts/numbers	Reproduction		Day(s)			7	Not reported	2017061
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported	2017052
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7	Not reported	766233 MODERATELY HARD WATER
bers	Progeny counts/numbers	Reproduction		Day(s)			7	Not reported	766231 REFORMULATED MODERATELY HARD WATER
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)			7		766855
Progeny counts/n/MSMT/MEAN OF 3 TESTS/	Progeny counts/nMs	Reproduction	NOEC	Dav(s)		NR.	7		766738 REFORMULATED MODERATELY HARD RECONSTITUT
Progeny counts/dMSMT/MEAN OF 3 TESTS/	Progeny counts/nMs	Reproduction	NOEC	Day(s)		NR	7	Not reported	766735 MODERATELY HARD RECONSTITUTED WATER
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)		NR	7	Not reported	2017044
bers	Progeny counts/numbers	Reproduction	NOEC	Day(s)	NR.	NR	7	Not reported	2017033
s (general)	Reproductive success (general)	Reproduction	LOEC	Brood or litter		NR	ω	Not reported	759074
bers	Progeny counts/numbers	Reproduction	LOEC	Day(s)		NR	7	Not reported	2017060
bers	Progeny counts/numbers	Reproduction	LOEC	Dav(s)		NR	7	Not reported	
bers	Progeny counts/numbers	Reproduction	LOEC	Dav(s)	NR.	NR	7	Not reported	
Progeny counts/nMSMT/MEASURED IN F5 GENERATION/	Progeny counts/nM	Reproduction	LOEC	Generation	NR R	NR	5	Not reported	
Progeny counts/dMSMT/MEASURED IN F2 GENERATION/	Progeny counts/nM	Reproduction	LOEC	Generation		NR :	2	Not reported	
bers	Progeny counts/numbers	Reproduction	TOEC	Dav(s)		NR N	7	Not reported	766230 REFORMILI ATED MODERATELY HARD WATER
Progeny counts/d MSMT/MFASURED IN F4 GENERATION/	Progeny counts/nMSMT/	Reproduction	LOEC	Generation		NR 25	4	Not reported	766355 MODERATELY HARD WATER
Progeny counts/plimbers	Progeny counts/num	Reproduction	LOEC	Day/s)	N. N.	NR NR	× 4	Not reported	766344 MODERATELY HARD WATER
Progeny counts/n MSM1/MEAN OF 3 TESTS/	Progeny counts/nM:	Reproduction	LOEC	Day(s)		NR.	2 /	-	/66/3/ REFORMULATED MODERATELY HARD RECONSTITU
Progeny counts/n MSWI/MEAN OF 3 TESTS/	Progeny counts/nM	Reproduction	LOEC	Day(s)		N.R.	7		766/34 MODERATELY HARD RECONSTITUTED WATER
bers	Progeny counts/numbers	Reproduction	LOEC	Day(s)		NR.	7	Not reported	
bers	Progeny counts/numbers	Reproduction	LOEC	Day(s)		NR	7	Not reported	2017032
s (general)	Reproductive success (general)	Reproduction	ICS0	Brood or litter	NR	NR	<sub>3</sub>	Not reported	759077
bers	Progeny counts/numbers	Reproduction	IC50	Day(s)	7	6	NR	Not reported	
bers	Progeny counts/numbers	Reproduction	IC50	Day(s)	7	6	NR	Not reported	
bers	Progeny counts/numbers	Reproduction	IC50	Day(s)	7	6	NR	Not reported	2148475 10 REPLICATES
s (general)	Reproductive success (general)	Reproduction	IC25	Brood or litter	NR		3	Not reported	759076
ibers	Progeny counts/numbers	Reproduction	IC25	Day(s)	NR	NR	7	Not reported	761597
bers	Progeny counts/numbers	Reproduction	IC25	Day(s)	7	6	NR	Not reported	
bers	Progeny counts/numbers	Reproduction	IC25	Day(s)	7	6	NR	Not reported	
1bers	Progeny counts/numbers	Reproduction	IC25	Day(s)	7	6	NR	Not reported	2148468 10 REPLICATES
bers	Progeny counts/numbers	Reproduction	EC50	Day(s)		NR	7	Not reported	2017045
ibers	Progeny counts/numbers	Reproduction	EC50	Day(s)		NR	7	Not reported	2017062
bers	Progeny counts/numbers	Reproduction	EC50	Day(s)	NR	NR	7	Not reported	2017053
bers	Progeny counts/numbers	Reproduction	ECSO	Day(s)	NR	NR	7	Not reported	766234 MODERATELY HARD WATER
bers	Progeny counts/numbers	Reproduction	ECSO	Day(s)	NR	NR	7	Not reported	766229 REFORMULATED MODERATELY HARD WATER
bers	Progeny counts/numbers	Reproduction	EC50	Day(s)		NR	7	Not reported	2017034
nbers	Progeny counts/numbers	Reproduction	EC25	Day(s)	NR	NR	7	Not reported	
EE Comment	Effect Measurement	Effect*	Endpoint	Duration Units (Days)	Duration Max (Days)	Duration Min (Days)	Duration Mean (Days)	Response Site	Result Number Experimental Design
				Coociaca	0000000	0000000	Carriera		

Conc 1  Result Number Mean Op 2147187 2152764 160619 160620	(m)	san Statistical (L)(1) Significance (L)(2) Not applicable (L)(2) Not applicable (L)(3) Not applicable (L)(3) Not applicable (L)(3) Not applicable (L)(3) Not applicable	Significance Level Mean Op	Significance Level Mean NA NA	Hardness Mean	Hardness Min 80	Max 100	lits	General Comments  TMETH/APHA, 1989// CONC1/FROM GRAPH/ HARD/MODERATELYHARD// HARD/MODERATELYHARD//
160619	2,260	Not applicable		NA		80	100		HARD/MODERATELYHARD//
160620	1,830	1,830 Not applicable		NA		80	100	100 mg/L CaCO3	HARD/MODERATELYHARD//
2152765	1,250	Not applicable		NA		2	***		TMETH/APHA, 1989// CONC1/FROM GRAPH//
768180	1,599	1,599 Not applicable		NA NA		80	100	100 mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2 CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768182	1,599	,599 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
767342	800	800 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768283	800	Not applicable		NA		80	100	mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768183	800	800 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768181	800	800 Not applicable		NA		80	100	mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768286	401	401 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
742988	4,970	4,970 Not applicable		NA					DNUM/6//DOSES/4190/4310/4810/5250/5430 mg/L//TE
44241	3,000	Not applicable		NA					
160622	2,770	2,770 Not applicable		NA		80	100	100 mg/L CaCO3	HARD/MODERATELYHARD//
2152738	1,750	Not applicable		NA					TMETH/APHA, 1989// CONC1/FROM GRAPH//
2152737	2,250	2,250 Not applicable		NA					TMETH/APHA, 1989// CONC1/FROM GRAPH//
18686	759	759 Not applicable		NA					
18685	1,838	,838 Not applicable		NA					
18684	3.005	3.005 Not applicable		NA S					
18683	3,526	Not applicable		NA					
18687	759	759 Not applicable		NA					
742079	590	590 Not applicable		NA					DNUM/6//DOSES/410/470/580/660/760 mg/L//TESTID/2
160621	3,250	3,250 Not applicable		NA		80	100	mg/L CaCO3	HARD/MODERATELYHARD//
32466	1,838	1,838 Not applicable		NA		ĺ			
119478	759	Not applicable		NA		3	4		100000000000000000000000000000000000000
768399	3,196	3,196 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768396	3,196	3,196 Not applicable		NA NA		80	100	mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//PO1/2 CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//PO1/2
768462	1.196	1.196 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
2152736	172	172 Not applicable		NA					TMETH/APHA, 1989//
768402	800	800 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
768460	1,196	,196 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
752752	10.000	800 Not applicable		NA	100	80	DOT		CA/14.0" mg/L//NG/12.0" mg/L//NA/26.3" mg/L//PO1/2
2153/63	10,000	10.000 Not applicable		NA NA	180			mg/L CaCO3	CONC1/ONLY CONC TESTED//
740759	10,000	10,000 Not applicable		NA	180				CONC1/ONLY CONC TESTED//
740760	10,000	10,000 Not applicable		NA	180				CONC1/ONLY CONC TESTED//
740732	10,000	10,000 Not applicable		NA	180			ű	CONC1/ONLY CONC TESTED//
2153700	10,000	10,000 Not applicable		NA	180				CONC1/ONLY CONC TESTED//
2153697	10,000	10,000 Not applicable		NA	180			mg/L CaCO3	CONC1/ONLY CONC TESTED//
740761	10,000	10,000 Not applicable		NA	180			Ę	CONC1/ONLY CONC TESTED//
2063733	13,400	13,400 Not applicable	-	NA					
2063726	13,400	13,400 Not applicable	-	NA					
2063735	13,400	13,400 Not applicable		NA				L	
740765	10,000	10,000 Not applicable		NA	140				
13605/	10,000	10,000 Not applicable		NA	140			mg/L CaCO3	CONCI/ONLY CONC TESTED//
126057	9 500	9 500 Not reported		NR S					
126052	9,500	9,500 Not reported		200					
176057	0 500	Not roported	-	i b					

TESTID/10//MSMT/NR/DOSES/NR//SOURCE/NR//WINT TESTID/10//MSMT/NR/DOSES/NR//SOURCE/NR//WINT TESTID/10//MSMT/NR//DOSES/NR//SOURCE/NR//WINT DOSES/NA ppm//TESTID/1//OEF/MIXTURE,TEMP EFCTS// DOSES/NA ppm//TESTID/1//OEF/MIXTURE,TM/ DOSES/NA ppm//TESTID/1//OEF/MIXTURE,TM/ DOSES/NA ppm//TESTID/1//OEF/MIXTURE,TM/ DOSES/NA pm//TESTID/1//OEF/MIXTURE,TM/ DOSES/NA pm//TESTID/1//DEF/MIXTURE,TM/ DOSES/NA PM//TESTID/1//DEF/MIXTURE,TM// DOSES/NA PM//TESTID/1//DEF/MIXTURE,TM// DOSES/NA PM//TESTID/1//DEF/MIXTURE,TM// DOSES/NA PM//TESTID/I// D				Abri		INOL applicable	1		075407
TESTID/10//MSMT/NR//DOSES/NR//SOUR TESTID/10//MSMT/NR//DOSES/NR//SOUR TESTID/10//MSMT/NR//DOSES/NR//SOUR DOSES/NA ppm//TESTID/1//OEF/MIXTURE DOSES/NA ppm//TESTID/1//OEF/MIXTURE				Z NA		3,900 Not applicable	3,900		764330
TESTID/10//MSMT/NR//DOSES/NR//SOUFI TESTID/10//MSMT/NR//DOSES/NR//SOUFI TESTID/10//MSMT/NR//DOSES/NR//SOUFI DOSES/NA ppm//TESTID/5//OEF/MIXTURI				NA		4,600 Not applicable	4,600		764325
TESTID/10//MSMT/NR//DOSES/NR//SOUR TESTID/10//MSMT/NR//DOSES/NR//SOUR				NA		3,700 Not applicable	3,700		764333
TESTID/10//MSMT/NR//DOSES/NR//SOUI	mg/L		100	NA		Not applicable	10,000	V	231564
1	mg/L		100	NA		8,250 Not applicable	8,250		231565
TESTID/11//MSMT/NR//DOSES/NR//SOLIRCE/NR//W/TINT	mg/L		20	NA		5,600 Not applicable	5,600	>	231570
TESTID/10//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		Not applicable	8,250		231566
TESTID/10//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		10,000 Not applicable	10,000	V	231563
TESTID/11//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		20	NA		5,100 Not applicable	5,100		231572
TESTID/11//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		20	NA		5,100 Not applicable	5,100		231573
TESTID/10//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		7,200 Not applicable	7,200		231567
TESTID/11//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		20	NA		5,600 Not applicable	5,600	V	231571
TESTID/10//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		6,800 Not applicable	6,800		231568
TESTID/10//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		6,150 Not applicable	6,150		231569
TESTID/13//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L	100	100	NA		23,000 Not applicable	23,000		231581
TESTID/13//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		20	NA		32,000 Not applicable	32,000		231579
TESTID/12//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		Not applicable	24,000		231576
TESTID/12//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		26,000 Not applicable	26,000		231575
TESTID/12//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		24,000 Not applicable	24,000		231577
TESTID/12//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		32,000 Not applicable	32,000	>	231574
	mg/L		20	NA		32,000 Not applicable	32,000	٧	231578
TESTID/13//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L		100	NA		26,000 Not applicable	26,000		231580
3	48 mg/L CaCO3	40		NA		21,450 Not applicable	21,450		12210
	48 mg/L CaCO3	40		NA		17,880 Not applicable	17,880		2455
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/				NR		10,000 Not reported	10,000		741740
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/				NR		10,000 Not reported	10,000		741741
DNUM/4//DOSES/1000/5000/10000 mg/I//TESTID/16//				NA		10,000 Not applicable	10,000	٧	741787
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/				NR		10,000 Not reported	10,000		741782
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/				NR		10,000 Not reported	10,000		741747
DNUM/4//DOSES/1000/5000/10000 mg/I//TESTID/15//				NA		10,000 Not applicable	10,000	V	741753
	mg/L CaCO3		22	NA		616 Not applicable	616		2055980
	100 mg/L CaCO3	80		NA		2,120 Not applicable	2,120		160519
	100 mg/L CaCO3	80		NA		2,840 Not applicable	2,840		160518
)3 HARD/MODERATELYHARD//	100 mg/L CaCO3	80		NA		Not applicable	3,520		160517
				NA		440 Not applicable	440		22956
				NA		570 Not applicable	570		22957
				NA :		760 Not applicable	760		22955
				NA		18 750 Not applicable	18 750		2064004
				N 5		16 500 Not applicable	16 500		2064009
27 CD/ TT: 0 1118/11/110/ TE: 0 1118/11/110/ TE: 0 1118/11/11/11/11/11/11/11/11/11/11/11/11	100 mg/r cacon	00		N Z	1	Not applicable	17 750		200007
		00		NA NA		Not applicable	27.5		768503
	100 mg/l CaCO3	00		200		F12 Not applicable	104		760456
13 CA/14 0* mg/L//WG/13 0* mg/L//WA/26.3 mg/L// OT/2	100 mg/l CaCO3	00		NA NA		467 Not applicable	707		760007
		80		NA NA		1,864 Not applicable	1,864		768390
	100 mg/L CaCO3	00		NA		2,044 Not applicable	1,004		769300
	100 mg/l Cacc	00		NA 3		1,401 Not applicable	1,404		760751
	100 mg/l CaCO3	08		NA		1 401 Not applicable	1 /01		769501
		80		NA		Not applicable	1 864		768393
	100 mg/L CaCO3	80 00		NA NA		1,560 Not applicable	7,56U		7507E7
Onits General Comments	Max Hardness Units	Vin	Wean	Level Mean	Op	Significance	(mg/L)	Mean Op	Result Number
	SS	SS	Hardness		Level Mean	Statistical	Mean	Conc 1	

Conc 1 Result Number Mean Op 2152847 741752 764481	Mean (mg/L) 1,645 10,000 2,600	Itean Statistical Ng/L) Significance 1,645 Not applicable 10,000 Not reported 2,600 Not applicable	Level Mean Op	Significance Level Mean NA NR	Hardness Mean	Hardness Min 80	Hardness Max 100	Hardness Units 100 mg/L CaCO3	General Comments  DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/ DOSES/NA ppm//TESTID/20//OEF/MIXTURE,TEMP EFCTS,
764481	2,600	Not applicable		NA NA					DOSES/NA ppm//TESTID/20//OEF/MIXTURE
187905	23,817	23,817 Not applicable		NA		80	90		
187903	24,829	24,829 Not applicable		NA		80	90	90 mg/L CaCO3	
187902	25,064	25,064 Not applicable		NA		80	90	90 mg/L CaCO3	
187901	25,190	25,190 Not applicable		NA		80	90	90 mg/L CaCO3	
187906	22,457	22,457 Not applicable		NA		80	90	90 mg/L CaCO3	
187904	25,786	25,786 Not applicable		NA		80	90	90 mg/L CaCO3	
742006	20,550	20,550 Significant		0.05		80	100	100 mg/L CaCO3	
742038	21,000	21,000 Significant		0.05		80	100	mg/L CaCO3	
742918	20,500	20,500 No significance		0.05		80	100	100 mg/L CaCO3	
742919	20,950	20,950 No significance		0.05		80	100		TESTID/4//OEF/PH, HARDNESS////
196818	10,400	10,400 Not applicable		NA					
196799	8,200	8,200 Not applicable		NA					
196831	7,150	7,150 Not applicable		NA					
196814	10,450	10,450 Not applicable		NA					
196791	7,950	7,950 Not applicable		NA					
196776	7,000	7,000 Not applicable		NA					
196783	7,200	7,200 Not applicable		NA					
196/95	7,300	7,400 Not applicable		NA NA					
196816	7,950	7,950 Not applicable		NA					
167864	7,341	Not applicable		NA	210			mg/L CaCO3	
75451	4,324	4,324 Not applicable		NA	148.8				12 H PHOTO//NOT FED//AERATED//DECHLORINATED, AG
196826	9,600	9,600 Not applicable		AN					
196803	8,400	8,400 Not applicable		NA					
196788	6,800	6,800 Not applicable		NA					
196784	7,050	7,050 Not applicable		NA					
196775	7,550	7,550 Not applicable		NA					
196834	9,950	9,950 Not applicable		NA					
196808	7,200	7,200 Not applicable		NA					
196827	7,850	7,850 Not applicable		NA					
196000	0,000	5,500 Not applicable		NA					
196623	6,900	7,900 Not applicable		NA					
196790	0,950	o,950 Not applicable		NA NA					
196804	8.050	8.050 Not applicable		NA S					
196811	7,400	7,400 Not applicable		NA					
196824	7,650	7,650 Not applicable		NA					
196830	8,800	8,800 Not applicable		NA					
196778	9,750	9,750 Not applicable		NA					
196807	7,900 1	7,900 Not applicable		NA					
196802	11,050	11,050 Not applicable		NA					
196810	10,200	10,200 Not applicable		NA					
196822	9,980 r	9,980 Not applicable		NA					
196794	10,000 r	Not applicable		NA					
196798	11,050	11,050 Not applicable		NA					
196786	8,350	8,350 Not applicable		NA					
196782	9,800 1	9,800 Not applicable		NA					
196/96	7,200	7,200 Not applicable		NA					
196800	7 000	7 goo Not applicable		NA					

2075426	2075402	20/4934	2074074	2075470	2075460	2075766	2153280	2153277	2152874	2152723	2152724	2153274	2075471	2075410	2075468	2075434	2075442	2153268	2153271	2075467	2152825	741686	26112	196809	196781	196829	196833	106971	196789	196813	106975	196707	196777	1966777	196905	167863	196837	196817	196801	196787	196819	196840	196832	196806	196774	196820	196835	196815	196792	196779		Result Number
																																																				Conc 1 Mean Op
1,589	861	9//	1,154	1 15/	1 357	1 256	1.303	1 240	132	2,750	2,750	1,134	1,192	1,249	1,317	1,779	1,836	316	540	1,489	1,068	1,590	13.750	7.200	6950	7 300	7,000	7,000	6 800	7,050	7,500	7 700	7,000	2,000	7,600	7,322	6,800	7,750	7,900	6,800	8,500	7,388	7,000	10,100	9,900	7,950	6,950	8,800	7,600	7,350	7,050	Mean (mg/L)
1,589 Not applicable	Not applicable	9// Not applicable	1,154 Not applicable	1 154 Not applicable	1,356 Not applicable	Not applicable	1.303 Not applicable	1 240 Not applicable	Not applicable	2,750 Not applicable	2,750 Not applicable	Not applicable	1,192 Not applicable	1,249 Not applicable	1,317 Not applicable	1,779 Not applicable	1,836 Not applicable	316 Not applicable	540 Not applicable	1,489 Not applicable	1,068 Not applicable	1,590 Not applicable	13.750 Not applicable	7,200 Not applicable	Not applicable	7 300 Not applicable	7 000 Not applicable	7 050 Not applicable	7,050 Not applicable	7,050 Not applicable	7,200 Not applicable	7 200 Not applicable	7,000 Not applicable	7 000 Not applicable	7,600 Not applicable	7,322 Not applicable	6,800 Not applicable	7,750 Not applicable	7,900 Not applicable	6,800 Not applicable	8,500 Not applicable	7,388 Not applicable	7,000 Not applicable	10,100 Not applicable	9,900 Not applicable	7,950 Not applicable	6,950 Not applicable	8,800 Not applicable	7,600 Not applicable	7,350 Not applicable	7,050 Not applicable	Statistical Significance
																																																				Significance Level Mean
NA	NA	NA	NA	NA NA	NA		NA :	NA	NA	NA	NA	NA	NA	AN	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA 3	200	N	NA	NA	NA	N		NA	NA NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Significance Level Mean
388	47	28	285	787	280	020	320	160	10	98	98	80	279	96	282	565	796	20	40	278																210																Hardness
375	44	25	780	087	1000	270							278	95	280	560	792			276	80	54																														Hardness Min
400	49	30	290	187	280								280	96	283	570	800			280	100	72																														Hardness Max
400 mg/L CaCO3	49 mg/L CaCO3	30 mg/L CaCO3	290 mg/L CaCO3	281 mg/L CaCO3	mg/L CaCO3	1115/1-0000	mg/L CaCO3	mg/L CaCO3	mg/L Cacos	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	280 mg/L CaCO3	96 mg/L CaCO3	283 mg/L CaCO3	570 mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	280 mg/L CaCO3	100 mg/L CaCO3	72 mg/L CaCO3														mg/L CaCO3																Hardness Units
										TMETH/APHA 1989// CONC1/FROM GRAPH//	TMETH/APHA, 1989// CONC1/FROM GRAPH//											DNUM/6//DOSES/0.479/0.879/1.27/1.79/2.26 g/L//TEST																														s General Comments

Conc 1 Result Number Mean Op 2075418	(mg	Statistical Significance Not applicable	Significance Level Mean Op	Significance Level Mean NA	Hardness Mean 187	Hardness Min 180	Hardness Max 194	Hardness Units	General Comments
120843	1,170	1,170 Not applicable		NA	707	TOO	104	iiig/ r cacos	TESTID/33//
120826	1,940	1,940 Not applicable		NA					TESTID/22//
120844	910	910 Not applicable		NA					TESTID/34//
120840	280	280 Not applicable		NA					TESTID/30//
120842	1,940	,940 Not applicable		NA					TESTID/32//
741547 >	467	467 Not applicable		NA	101	100	103	mg/L CaCO3	
120829	1,830	,830 Not applicable		NA					TESTID/25//
120845	1,430	Not applicable		NA					TESTID/35//
120837 <	330	330 Not applicable		NA					TESTID/27//
120827	1,830	,830 Not applicable		NA					TESTID/23//
120841	1,940	1,940 Not applicable		NA					TESTID/31//
120824	1,740	1,740 Not applicable		NA					TESTID/20//
120839	1,940	1,940 Not applicable		NA					TESTID/29//
120828	1,710	1,710 Not applicable		NA					TESTID/24//
120825	1,640	1,640 Not applicable		NA					TESTID/21//
120838	1,830	1,830 Not applicable		NA					TESTID/28//
160319	3,380	3,380 Not applicable		NA		80	100	mg/L CaCO3	HARD/MODERATELYHARD//
741788	1,000	1 042 Not applicable		NA S		C	100	100 mg/ r cacon	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
207716	2,250	2,250 Not applicable		NA	102	94	104	mg/L CaCO3	CONC/FROM GRAPH//
207717 ~	2,000	2,000 Not applicable		NA	102	94	104	mg/L CaCO3	CONC/FROM GRAPH//
742567	2,200	2,200 Significant	٨	0.05		54	72	mg/L CaCO3	DNUM/7//DOSES/0.21/0.44/0.85/1.3/1.7/2.2 g/L//TESTID
767903	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/20//
766864	5,000	5,000 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/4//
767897	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/19//
767961	2,500	2,500 Not reported		NR NR		80	100	NR	DOSES/NA g/L//TESTID/23//
767805	2,500	Not reported		No No		000	100		DOSES/NA g/L//TESTID/18//
767873	2,500	2,500 Not reported		NB NA		00	100 NB		DOSES/NA 8/1//TESTID/13//
767959	2,500	2,500 Not reported		NR S		80	100 NR		DOSES/NA g/L//TESTID/22//
767870	2,500	2,500 Not reported		NR		80	100 NR		DOSES/NA g/L//TESTID/12//
767967	2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/24//
766872	2,500	2,500 Not reported		NR		80			DOSES/NA g/L//TESTID/6//
766866	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/5//
767889	2,500	2,500 Not reported		NR		80	100 NR		DOSES/NA g/L//TESTID/17//
767856	2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/9//
767846	2,500	2,500 Not reported		NR P		80	100 NR		DOSES/NA g/1//TESTID/3//
767953	2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/21//
767887	2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/16//
767881	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/15//
767975	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/26//
767864	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/11//
767862	2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/10//
767848	2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/7//
767854	2,500	2,500 Not reported		NR		80	100	NR	DOSES/NA g/L//TESTID/8//
251007	T,500	,500 Significant		25					TESTID/37//DOSES/<0.3/0.3/0.3/0.3/1.3/2.3 G/L//WSWIT
251001	300	300 Significant		NR NR					TESTID/37//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT/
250999	2,500	2,500 Significant		NR :					TESTID/36//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT/
351009	2 EDD Significant			5					

	18463 3 417	18462 3,114	4,			742566 2,000	2060189 4,940	2060190 3,100	2060191 7,810		2060161 6,137		2060157 6,137	2060148 6,070		2152588 11,000	2152587 4,700	26113 10,500	26114 10,200	16495 6,221	2211024 4,850	2211025 10,800	2162290 700	2162289 610	741489 14,040	2211037 10,000	2060270 4,940				1				2060201 6.033					2060220 6,898			2060216 5,79:		2060213 5,274	2060212 5,11:	2060211 4,917	2060210 4,64	2060209 4,16	741678 1,20	Result Number   Mean Op (mg/L)
5 874 Not applicable	3 412 Not applicable	3,114 Not applicable		ance		2,000 Not applicable	4,940 Not applicable	3,100 Not applicable	7,810 Not applicable	7,810 Not applicable	6,137 Not applicable	6,137 Not applicable	6,137 Not applicable	6,070 Not applicable	13,000 Not applicable	11,000 Not applicable	4,700 Not applicable	10,500 Not applicable	10,200 Not applicable	6,221 Not applicable	4,850 Not applicable	10,800 Not applicable	700 Not applicable	610 Not applicable	14,040 Not applicable	10,000 Not applicable	4,940 Not applicable	7,810 Not applicable	7,810 Not applicable	7,810 Not applicable	11,600 Not applicable	4,940 No significance	6.211 Not applicable	7 010 Cignificant	6 032 Not applicable	8,740 Not applicable	7,840 Not applicable	7,399 Not applicable	7,116 Not applicable	Not applicable	6,717 Not applicable	6,280 Not applicable	5,793 Not applicable	5,417 Not applicable	Not applicable	5,113 Not applicable	Not applicable	4,640 Not applicable	4,163 Not applicable	1,200 Not applicable	Statistical Significance
7 -		7		٨	^	7	7	7	7	7	-	-																				î	1																		Level Mean Op
No.	No	NA	NA	0.05	0.05	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.01	NA C.O.	001	NA	NA	NA	NA	NA	NA	NA	NA	AN	NA	NA	NA	NA	NA	NA	NA	Significance Level Mean
			149.6				296	296	296	296	296	296	296	296						124							296	296	296	296	296	296	296	305	206	2967	296	296	296	296	296	296	296	296	296	296	296	296	296		Hardness Mean
				54	54	54															140	140				140									80	0															Hardness Min
			,	72	72	72															160	160				160									DOT																Hardness Max
		ç					Н							11						mg/L CaCO3	mg/L	160 mg/L					mg/L CaCO3	mg/L CaCO3	mg/I CaCO3			mg/L CaCO3	mg/L CaCO3		1	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3		Hardness Units				
			1.5	DNUM/7//DOSES/0.21/0.44/0.85/1.3/1.7/2.2 g/1//TESTID	DNUM/7//DOSES/0.21/0.44/0.85/1.3/1.7/2.2 g/L//TESTID	DNUM/6//DOSES/0.479/0.879/1.27/1.79/2.26 g/L//TESTI	CONTR/B,C//											TIME/ADDITIONAL DURATIONS REPORTED//DNUM/6//DC		CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B.C//	CONTR/B.C//	CONTR/B/C//	CONTRIB (I)	CONTRIB CIT	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	LIFESTG/INITIAL BW, AVG LENGTH 148MM//TIME/OTHER	General Comments							

CONICT /ONLY CONIC TESTED //				140	NA		20,000 Not applicable	20,000 N		740725
	mg/L CaCO3			124	NA		6,621 Not applicable	6,621 N		16496
					NA		1,240 Not applicable	1,240 N		2164557
					NA		1,510 Not applicable	1,510 N		2164555
					NA		1,200 Not applicable	1,200 N		2164558
					NA		1,280 Not applicable	1,280 N		2164556
	mg/L CaCO3	100 r	80	1	NA		1,382 Not applicable	1,382 N		2152842
					NA		954 Not applicable	954 N		2164554
					NA		1,050 Not applicable	1,050 N		2164552
					NA		998 Not applicable	998 N		2164553
					NA		,140 Not applicable	1,140 N		2164551
					NA		733 Not applicable	733 N		2164550
					NA		781 Not applicable	781 N		2164549
					NA		831 Not applicable	831 N		2164547
					NA		842 Not applicable	842 N		2164548
	mg/L CaCO3			212	NA		3,009 Not applicable	3,009 N		20/54/5
	mg/L CaCO3			56	NA		3,078 Not applicable	3,078 N		2075474
TESTID/4//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L TE			100	NA		10,000 Not applicable	10,000 N	V	231534
TESTID/4//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		Not applicable	3,200 N		231536
TESTID/4//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L TE			100	NA		10,000 Not applicable	10,000	V	
TESTID/4//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		3,700 Not applicable	3,700		231535
TESTID/4//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA NA		3,200 Not applicable	3,200 1		231537
					NA		12,600 Not applicable	12,600 N		205381/
					NA		12,000 Not applicable	12,000 1		2053819
					NA		12,110 Not applicable	12,110		2053814
					NA		8,530 Not applicable	8,530		2053818
					NA		10,040 Not applicable	10,040		2053813
					NA		9,260 Not applicable	9,260 1		2053816
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/	D				NR		10,000 Not reported	10,000 1		742663
DNUM/4//DOSES/1000/5000/10000 mg/I//TESTID/18//	D				NA		7,700 Not applicable	7,700 1		741789
					NA		11,540 Not applicable	11,540		118481
					NA		18,100 Not applicable	18,100		2064154
					NA		17,550 Not applicable	17,550 [		2064156
					NA		18,100 Not applicable	18,100		2064155
	mg/L CaCO3	256.5	171	193.9	NA		16,000 Not applicable	16,000		2062416
	CaCO3	256.5	171	193.9	NA		24,000 Not applicable	24,000		2062417
TESTID/2//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		7,500 Not applicable	7,500		231526
TESTID/2//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		7,500 Not applicable	7,500		231525
TESTID/2//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		7,500 Not applicable	7,500		231527
				100	NA		10,000 Not applicable	10,000	٧	231523
TESTID/2//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L T			100	NA		7,500 Not applicable	7,500		231524
Q J					NA		244 Not applicable	244		2052775
DNUM/6-7//DOSES/0.25 to 8.0 g/L//TESTID/19//CONTR/0		192	170		NA		2,670 Not applicable	2,670		803856
TIME/OTHER DURATIONS ALSO REPORTED//DNUM/6-7//		192	170		NA		2,230 Not applicable	2,230		741563
	mg/L CaCO3			180	NA		20,000 Not applicable	20,000		740662
	mg/L CaCO3			180	NA		10,000 Not applicable	10,000		740690
	mg/L CaCO3			96.3	NA		441 Not applicable	441		201558
					NA		3,050 Not applicable	3,050		201566
	mg/L CaCO3			92.8	NA		1,470 Not applicable	1,470		201545
	CaCO3	82			NA		1,760 Not applicable	1,760		2055812
TESTID/1//	100 mg/L T	100	80	84	NA		1,250 Not applicable	1,250		741434
DNUM/2//DOSES/1770 mg/L//TESTID/9//					NA		Not applicable	1,770		742927
General Comments	Hardness Units	Hardness Max	Hardness Min	Hardness Mean	Significance Level Mean	Level Mean Op	Statistical Significance	(mg/L)	Mean Op	Result Number

33 TIME/OTHER DURATIONS ALSO REPORTED//DNUM/6-7// 33 DNUM/6-7//DOSES/0.25 to 8.0 g/L/TESTID/23//CONTR/C 33 DNUM/6-7//DOSES/0.5 to 8.0 g/L/TESTID/22//CONTR/C 33 DNUM/6-7//DOSES/0.5 to 8.0 g/L/TESTID/22//CONTR/C 33 AERATION//CONTR/NO MORTI//		360					2,220 INOT applicable	3.220		C+00177
TIME/O DNUM/ DNUM/ DNUM/	וווק/ ב נמנטט		140	674	200		5,103 Not applicable	CO1,0		270702
TIME/O DNUM/ DNUM/ DNUM/	Ba/1 0000			400	NA NA		12,946 Not applicable	12,946		730765
TIME/O DNUM/ DNUM/	2 mg/L CaCO3	52	39.2		NA		1,295 Not applicable	1,295		40512
TIME/O DNUM/ DNUM/	mg/L CaCO3	,		101.7	NA		5,840 Not applicable	5,840		201540
TIME/O DNUM/	mg/L CaCO3			10	NA		12,946 Not applicable	12,946		196203
TIME/O DNUM/					NA		Not applicable	14,125		18184
TIME/O DNUM/	180 mg/L CaCO3	18	160		NA		3,570 Not applicable	3,570		2224806
TIME/O	192 mg/L CaCO3	19	170		NA		Not applicable	4,560		741594
	192 mg/L CaCO3		170		NA		560 Not applicable	560		803860
03	192 mg/L CaCO3		170		NA		550 Not applicable	550		741568
	mg/L CaCO3			99	NA		Not applicable	1,430		2052771
33	mg/L CaCO3			47	NA		763 Not applicable	763		2052776
					NA		168 Not applicable	168		2052770
33	mg/L CaCO3			322	NA		1,870 Not applicable	1,870		2052778
)3	mg/L CaCO3			172	NA		1,962 Not applicable	1,962		2052777
03		180	160		NA		2,630 Not applicable	2,630		2224815
33				278	NA		1,313 Not applicable	1,313		2052779
DNUM/6-7//DOSES/0.25 to 8.0 g/L//TESTID/20//CONTR/0		192	170		NA		1,840 Not applicable	1,840		803857
DNOM	192 mg/L cacus		T/0		NA		3,980 Not applicable	3,980		/41584
	mg/L CaCO3			306	NA		1,265 Not applicable	1,265		2052/85
				300	NA		Not applicable	285		2052774
	I				NA		113 Not applicable	113		2052//3
J3 IIME/OTHER DURATIONS ALSO REPORTED//DNOW/6-///	2 mg/L Cacus	192	1/0		NA		1,740 Not applicable	1,/40		741564
	mg/L Cacos		170	322	NA		1,391 Not applicable	1,391		2052/83
3 3	mg/L Cacos			292	NA		1,559 Not applicable	1,559		18/7507
3 2	mg/L Cacos			322	NA		Not applicable	261		2052784
	mg/L Cacus			306	NA		153 Not applicable	153		2052786
	mg/L CaCU3			292	NA		403 Not applicable	403		2052782
	mg/L CaCO3			278	NA		432 Not applicable	432		2052780
					NA		817 Not applicable	817		2052772
					0.05		2,000 No significance	2,000		2164623
					0.05		2,000 No significance	2,000		2164/10
					0.05		2,000 No significance	2,000		2164710
					0.05		2,000 No significance	2,000		2164677
					0.05		500 No significance	3 000		2164699
					0.05		2,000 No significance	2,000		2164692
					0.05		2,000 No significance	2,000		2164695
					0.05	concentration	8,000 Not significant at all concentration	8,000		2164618
					0.05		1,000 No significance	T,000		2164645
					0.05	concentration	8,000 Not significant at all concentration	8,000		2164613
					0.05		2,000 Significant	2,000		2164625
					0.05		4,000 Significant	4,000		2164641
					0.05		1,000 Significant	1,000		2164700
					0.05		2,000 Significant	2,000		2164628
					0.05		4,000 Significant	4,000		2164693
					0.05		4,000 Significant	4,000		2164696
					0.05		2,000 Significant	2,000		2164646
					0.05		4,000 Significant	4,000		2164711
ONC1/ONLY CONC TESTED//	mg/L CaCO3			140	NA		10,000 Not applicable	10,000		740689
CONCI	mg/L CaCO3			140	NA		10,000 Not applicable	10,000		741710
		Max	Min	Mean	Level Mean	Op	Significance	(mg/L)	-	Result Number
		Hardness	Hardness	Hardness	Significance	Significance Level Mean	Statistical	Mean	Conc 1	

The second of th								1000		
ESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L			100	NA		5,600 Not applicable	5,600	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	231543
TESTID/5//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		Not applicable	3,500		231540
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		6,950 Not applicable	6,950		231548
TESTID/7//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT				20	NA		4,800 Not applicable	4,800		231551
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,				100	NA		Not applicable	5,600	ľ	231542
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,				100	NA		7,500 Not applicable	7,500		231547
TESTID/7//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	mg/L T			20	NA		4,250 Not applicable	4,250		231553
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		5,600 Not applicable	5,600	V	231544
TESTID/7//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L T			20	NA		4,100 Not applicable	4,100		231554
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L T			100	NA		6,200 Not applicable	6,200		231549
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		6,200 Not applicable	6,200		231550
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	mg/L T			100	NA		5,100 Not applicable	5,100		231545
TESTID/6//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA	7	5,100 Not applicable	5,100		231546
	mg/L CaCO3			100.1	NA		5,060 Not applicable	5,060		201543
	mg/L CaCO3			100.1	NA		2,540 Not applicable	2,540		201542
CONC1/ONLY CONC TESTED//				140	NA		10,000 Not applicable	10,000		741709
	mg/L CaCO3 (			140	NA		10,000 Not applicable	10,000		740693
CONTR/B,C//		292	288	290	NA		5,340 Not applicable	5,340		2060354
CONTR/B,C//		292		290	NA		5,340 Not applicable	5,340		2060342
CONTR/B,C//	mg/L CaCO3	292	288	290	NA		5,340 Not applicable	5,340		2060360
CONTR/B,C//		292	288	290	NA		7,720 Not applicable	7,720		2060361
CONTR/B,C//	mg/L CaCO3	292	288	290	NA		Not applicable	5,340		2060350
CONTR/B,C//		292	288	290	NA		5,340 Not applicable	5,340		2060352
CONTR/B,C//		292	288	290	0.01	â	3,320 No significance	3,320		2060333
CONTR/B,C//	Ť	292	288	290	NA		4,211 Not applicable	4,211		2060336
CONTR/B,C//	mg/L CaCO3	292	288	290	0.01	î	5,340 Significant	5,340		2060335
CONTR/B,C//	mg/L CaCO3	292	288	290	NA		4,270 Not applicable	4,270		2060329
CONTR/B,C//		292	288	290	NA		4,270 Not applicable	4,270		2060326
CONTR/B,C//	mg/L CaCO3	292	288	290	NA		4,310 Not applicable	4,310		2060325
CONTR/B,C//	mg/L CaCO3	292	288	290	NA		4,270 Not applicable	4,270		2060327
CONTR/11 % MORT//				124	NA		2,569 Not applicable	2,569		16494
	mg/L CaCO3			48	NA		Not applicable	1,930		2075478
					NA		1,500 Not applicable	1,500	u	6168
					NA	1	7,000 Not applicable	7,000	V	6171
~					NA		Not applicable	3,000		6167
					NA		5,000 Not applicable	5,000		6173
					NA		1,000 Not applicable	1.000		6169
S 1					NA :		5,000 Not applicable	5,000		6174
			1		NA		1 000 Not applicable	1,000		6170
CONTR/< 10 % MORT//					NA :		Not applicable	5,000		6172
					NA		6,000 Not applicable	6,000		802502
DNUM/11//DOSES/NA ppm//TESTID/27//OEF/NO FOAM					NA		2,000 Not applicable	2,000		802496
					NA		4,000 Not applicable	4,000		802497
					AN		6,000 Not applicable	6,000		802501
					NA		Not applicable	6,000		802500
DNUM/11//DOSES/NA ppm//TESTID/28//OEF/NO FOAM					NA		7,000 Not applicable	7,000	٧	802499
					AN		2,000 Not applicable	2,000		802495
DNUM/11//DOSES/NA ppm//TESTID/27//OEF/NO FOAM					NA A		2,000 Not applicable	2,000		802494
DNUM/11//DOSES/NA ppm//TESTID/27//OEF/NO FOAM					NA		1,000 Not applicable	1,000	٨	802491
					NA		7,000 Not applicable	7,000		802506
DNUM/11//DOSES/NA ppm//TESTID/28//OEF/NO FOAM					NA		4,000 Not applicable	4,000		802505
General Comments	Hardness Units	Hardness Max	Hardness Min	Hardness Mean	Significance Level Mean	Level Mean Op	Statistical Significance	Mean (mg/L)	Conc 1 Mean Op	Result Number
						C				

							8 150 Not applicable	0,000		196747
					100		7,100 Not applicable	7,550		10777
					NA		7 100 Not applicable	3,100		196717
					20		o,300 Not applicable	0,300		106754
					200		200 Not applicable	000,400		196720
					NA		7,300 Not applicable	7,000		196771
					NA		7,730 Not applicable	7,700		106771
					NA		7,200 Not applicable	7,200		CT / 06T
					20		Not applicable	7,700		106715
							7 AEO Not applicable	7,000		106763
					200		Not applicable	2,650		106705
					2 3		7,200 Not applicable	0,000		106750
					NA.		Not applicable	7 700		106733
					NA		8.150 Not applicable	8.150		196764
					NA		7,050 Not applicable	7,050		196712
					NA		8,300 Not applicable	8,300		196737
					NA		7,100 Not applicable	7,100		196711
					NA		7,550 Not applicable	7,550		196751
					NA		7,100 Not applicable	7,100		196716
	mg/L CaCO3	100	80		NA		4,079 Not applicable	4,079		2152828
					NA		8,200 Not applicable	8,200		196740
					NA		Not applicable	7,300		196/32
					NA		/,/UU Not applicable	/,/00		196723
					NA		7,500 Not applicable	7,500		19676b
					2 2		7,400 Not applicable	7,400		196760
					No.		7 400 Not applicable	7 400		007700
					NA		Not applicable	7,600		196770
					NA A		7.650 Not applicable	7.650		196839
					NA		7.500 Not applicable	7.500		196731
					NA		Not applicable	8.700		196759
				TV	NA		8,400 Not applicable	8,400		196760
					NA		7,400 Not applicable	7,400		196729
					NA		7,950 Not applicable	7,950		196756
				8 8 9	NA		8,200 Not applicable	8,200		196741
					NA		Not applicable	7,400		196728
					NA		7,650 Not applicable	7,650		196748
					NA		7,050 Not applicable	7,050		196714
					NA		7,800 Not applicable	7,800		196745
DOSES/NA mg/L//TESTID/6//OEF/LEACHATE STUDY, MIXT	mg/L CaCO3	694	404	563	NA		11,400 Not applicable	11,400		759180
					NA		8,300 Not applicable	8,300		196763
					NA		7,650 Not applicable	7,650		196724
					AN		7,800 Not applicable	7,800		196744
					NA		7,050 Not applicable	7,050		196713
					NA		8,300 Not applicable	8,300		196736
					NA		7,200 Not applicable	7,200		196767
					NA		7,200 Not applicable	7,200		196768
					NA		9,000 Not applicable	9,000		196735
					NA		7,950 Not applicable	7,950		196755
					NA		7,500 Not applicable	7,500		196772
					NA		7,750 Not applicable	7,750		196750
TESTID/S//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L			100	NA		3,500 Not applicable	3,500		231541
TESTID/7//MSMT/NR//DOSES/NR//SOURCE/NR//WIINI/	mg/L			20	NA		4,250 Not applicable	4,250		231552
TESTID/5//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L			100	NA		4,200 Not applicable	4,200		231538
	Hardness Units	Hardness Max	Hardness Min	Hardness Mean	Significance Level Mean	Level Mean Op	Statistical Significance	Mean (mg/L)	Conc 1 Mean Op	Result Number
						Significance				

Conc 1 Mean Op	Mean Statistical (mg/L) Significance	Significance Level Mean Significance Op Level Mean	an Mean	Hardness Min	Hardness Max	Hardness Units	General Comments
	00 Not		П				
196719	7,400 Not applicable	NA		J			
160375	8,360 Not applicable	NA		52	96		
160325	6,390 Not applicable	NA NA		80	100		
167861	7 650 Not applicable	NA	210	ŠÚ.	UUT	mg/L CaCO3	HARD/MODERATELYHARD//
160323	8,280 Not applicable	NA NA	210	80	100		HARD/MODERATE YHARD//
201537	6,570 Not applicable	NA	96.3	1		- 1	
196726	7,650 Not applicable	NA				Q	
196769	7,200 Not applicable	NA		I			
196765	8,150 Not applicable	NA					
196734	7,200 Not applicable	NA					
100700	8,400 Not applicable	NA					
196/38	8,300 Not applicable	NA NA					
196753	7 ASO Not applicable						
196757	7,950 Not applicable	NA :					
196749	7,650 Not applicable	NA					
196722	7,400 Not applicable	NA					
196742	8,200 Not applicable	NA					
196746	7,800 Not applicable	NA					
196730	7 400 Not applicable	N N					
167862	7 650 Not applicable	NA S	210			2000	
42866	8,000 Significant	0.05		86	94	94 mg/L CaCO3	
96881	8,000 Significant	0.05	05	86	94	94 mg/L CaCO3	
42423	8,000 Significant	0.05	05	86	94	mg/L CaCO3	
96906	8,000 Significant	0.05	05	86	94 1	94 mg/L CaCO3	
14389	8,000 Significant	0.05	05	86	94	94 mg/L CaCO3	
68598	8,000 Significant	0.05	05	86	94 1	/L CaCO3	
768071	2,250 Not reported	NR		80	100 NR		DOSES/NA g/L//TESTID/40//
768157	3 500 Not reported	No.		80	100		DOSES/NA g/L//TESTID/32//
768066	5,000 Not reported	NR .		80	100 NR		DOSES/NA g/1//TESTID/31//
768058	5,000 Not reported	NR		80	1001		DOSES/NA g/L//TESTID/29//
768096	5,000 Not reported	NR		80	100 NR		DOSES/NA g/L//TESTID/38//
768173	2,500 Not reported	NR		80	100 r		DOSES/NA g/L//TESTID/45//
768090	2 500 Not reported	NID		80	100		DOSES/NA g/L//IESIID/48//
768088	5,000 Not reported	NR NR		80	100 NR		DOSES/NA @/L//TESTID/3///
768123	2,500 Not reported	NR		80	100		DOSES/NA g/L//TESTID/44//
768163	5,000 Not reported	NR		80	100 NR		DOSES/NA g/L//TESTID/52//
768153	2,500 Not reported	NR		80	100 N		DOSES/NA g/L//TESTID/50//
768098	5,000 Not reported	NR		80			DOSES/NA g/L//TESTID/39//
768080	2,500 Not reported	NR		80	100 N		DOSES/NA g/L//TESTID/33//
768113	2.500 Not reported	NR		80 00	100 NR		DOSES/NA g/L//TESTID/34//
768107	2,500 Not reported	NR		80	100 NR		DOSES/NA g/1//TESTID/42//
768082	2,500 Not reported	NR		80	100 NR		DOSES/NA g/L//TESTID/35//
768137	2,500 Not reported	NR		80	100 N	NR D	DOSES/NA g/L//TESTID/47//
10014/1	2,500 Not reported	NR		80	100 NR		DOSES/NA g/L//TESTID/49//
760064	1000			2			

7/1937	770077	742070	742057	742029	742059	740696	740694	768558	201538	740697	742666	741400 TION///	768118	768134	768065	768148	768138	768083	768108	768114	768081	768075	768099	768154	768164	768124	768089	768091	768144	768128	768097	768059	768067	768158	768073	201560	768105	96907	69057	18890	1/200	68599	201562	741410	742691	741744	2210509	201561	768117		Conc 1 Result Number Mean Op
4,450	7 700	4.490	4,490	4,490	4,490	10,000	10,000	6,069	7,310	20,000	860	2,000	1,250	2,500	2,500	1,250	1,250	1,250	1,250	1,250	2,500	1,250	2,500	1,250	2,500	1,250	2,500	1,250	1,250	1,250	2,500	2,500	2,500	1,250	2,500	252	1 250	4.000	4 000	4,000	4,000	4,000	298	2,200	210	1,140	201	352	2,500	5,000	Mean (mg/L)
4,430 Not applicable	Not applicable	4,490 Not applicable	4,490 Not applicable	4,490 Not applicable	4,490 Not applicable	10,000 Not applicable	10,000 Not applicable	6,069 Not applicable	7,310 Not applicable	20,000 Not applicable	860 No significance	2,000 Not significant at all concentration	1,250 Not reported	2,500 Not reported	2,500 Not reported	1,250 Not reported	2,500 Not reported	1,250 Not reported	1,250 Not reported	Not reported	2,500 Not reported	2,500 Not reported	2,500 Not reported	1,250 Not reported	2,500 Not reported	252 No significance	1 250 Not reported	4,000 No significance	No significance	4 000 No significance	No significance	4,000 No significance	Not applicable	2,200 Significant at all concentrations	210 Significant	1,140 Significant	201 Significant at all concentrations	Significant	2,500 Not reported	5,000 Not reported	Statistical Significance										
												concentration																								^								centrations			centrations	۸		1000	Significance Level Mean Op
100	NA	NA A	NA	NA	NA	NA	NA	NA	NA	NA	0.05	0.05	NR	NR	NR	NR	0.05	NR	0.05	0.05	0.05	0.05	0.05	NA	0.05	0.05	0.05	0.05	0.05	NR	NR	Significance Level Mean																			
						140	140		96.3	140																										96.9							96.9					96.9			Hardness Mean
								80			90	90	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80	80		80	86	86	86	28	00		90	90	90			80	80	Hardness Min
								100			110	110	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100														100	100	Hardness Max
						mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3		mg/L	NR	NR	NR	NR		NR	NR	NR	NR	100 NR	NR	NR	NR	NR	NR	NR	100 NR	NR	NR		NR	100 NR		mg/L CaCO3	100 NR	mg/L CaCO3	94 mg/l CaCO3	mg/L Cacos	94 mg/L CaCO3	94 mg/L CaCO3	mg/L CaCO3	TIU mg/L	110 mg/L	mg/L		mg/L CaCO3	NR	NR	Hardness Units
DNI IM/3//DOSES/A480 mg/1//TESTID/18//	DNI IM/2//DOSES/MAGO mg/I //TESTID/21//	DNUM/2//DOSES/4490 mg/L//TESTID/18//	2	DNUM/2//DOSES/4490 mg/L//TESTID/12//	12	CONC1/ONLY CONC TESTED//	CONC1/ONLY CONC TESTED//	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2		CONC1/ONLY CONC TESTED//	TIME/MEASURED DURING RECOVERY.//DNUM/3//DOSES	DNUM/2//DOSES/2 g/L//TESTID/1//	DOSES/NA g/L//TESTID/43//	DOSES/NA g/L//TESTID/46//	DOSES/NA g/L//TESTID/30//	DOSES/NA g/L//TESTID/49//	DOSES/NA g/L//TESTID/47//	DOSES/NA g/L//TESTID/35//	DOSES/NA g/L//TESTID/41//	DOSES/NA g/L//TESTID/42//	DOSES/NA g/L//TESTID/34//	DOSES/NA g/L//TESTID/33//	DOSES/NA g/L//TESTID/39//	DOSES/NA g/L//TESTID/50//	DOSES/NA g/L//TESTID/52//	DOSES/NA g/L//TESTID/44//	DOSES/NA g/L//TESTID/36//	DOSES/NA g/L//TESTID/37//	DOSES/NA g/L//TESTID/48//	DOSES/NA g/L//TESTID/45//	DOSES/NA g/L//TESTID/38//	DOSES/NA g/L//TESTID/29//		DOSES/NA g/L//TESTID/51//	DOSES/NA g/L//TESTID/32//	0.77	DOSES/NA g/L//TESTID/40//							DNUM/2//DUSES/2,2 g/L//TESTID/2//CUNC/ AVERAGE /	TIME/MEASURED DURING RECOVERY//DNUM/3//DOSES/	TIME/MEASURED DURING RECOVERY.//DNUM/3//DOSES	CONC1/ONLY CONC TESTED//		DOSES/NA g/L//TESTID/43//	DOSES/NA 8/L//TESTID/46//	

1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1		100 // 6-603	08		NA	2	2,076 Not applicable	2.076 N		767362
CA/14 0* mg/l //MG/17 0* mg/l //N/ /76 2* mg/l //DOT/7		100	0.8		NA	7	2,076 Not applicable	2,076 N	1.0	767346
HARD/MODERATELYHARD//		100	80		NA	7	1,770 Not applicable	1,770 N		160287
HARD/MODERATELYHARD//	100 mg/L CaCO3 H	100	80		NA	7	1,770 Not applicable	1,770 N		160288
DOSES/NA ppm//TESTID/30//OEF/MIXTURE, TEMP EFCTS,	D				NA	7	4,000 Not applicable	4,000 N		/64615
DNUM/3//DOSES/1000/2000 ppm//TESTID/1//OEF/RECO					NA	7	1,000 Not applicable	1,000 N		/59169
DNUM/6-7//DOSES/0.25 to 8.0 g/L//TESTID/16//CONTR/	192 mg/L CaCO3 D	192	170		NA	7	3,630 Not applicable	3,630 N		803850
DNUM/6-7//DOSES/0.5 to 8.0 g/L//TESTID/17//CONTR/C	mg/L CaCO3 D	192	170		NA	7	5,230 Not applicable	5,230 N		741583
TIME/OTHER DURATIONS ALSO REPORTED//DNUM/6-7//		192	170		NA	7	Not applicable	3,310 N		/41561
TIME/OTHER DURATIONS ALSO REPORTED//DNUM/6-7//	mg/L CaCO3 T	192	170		NA	2	2,590 Not applicable	2,590 N		741562
NUM/6-7//DOSES/0.25 to 8.0 g/L//TESTID/18//CONTR		192	170		NA	7	Not applicable	2,760 N		803853
	100 mg/L CaCO3	100	80		NA	7	5,648 Not applicable	5,648 N		2152833
	mg/L CaCO3			220	NA	7	6,008 Not applicable	6,008 N		2075477
	mg/L CaCO3			52	NA	7	4,278 Not applicable	4,278 N		2075476
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/	0				NR	7	10,000 Not reported	10,000 N		741742
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/	0				NR	7	10,000 Not reported	10,000 N		741743
DNUM/4//DOSES/1000/5000/10000 mg/I//TESTID/17//	0				NA	-	10,000 Not applicable	10,000 N	V	741788
DNUM/5//DOSES/1000/2500/5000/10000 mg/I//TESTID/	0				NR		10,000 Not reported	10,000 N		/41/81
TESTID/8//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	mg/L 1			100	NA		2,250 Not applicable	2,250 N		231555
TESTID/9//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT	mg/L T			20	NA		1,150 Not applicable	1,150 N		231562
TESTID/8//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT				100	NA		1,100 Not applicable	1,100 1		231558
TESTID/8//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	mg/L T			100	NA		1,250 Not applicable	1,250 N		231557
TESTID/9//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	mg/L 1			20	NA		2,400 Not applicable	2,400 1		231559
TESTID/9//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,				20	NA		1,250 Not applicable	1,250 N		231561
TESTID/9//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	mg/L 1			20	NA		1,550 Not applicable	1,550 1		231560
TESTID/8//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT,	- 1			100	NA		1,950 Not applicable	1,950 1		231556
	mg/L CaCO3			51	NA		740 Not applicable	740 N		2075472
	mg/L CaCO3			192	NA		1,100 Not applicable	1,100 1		2075473
				140	NA		10,000 Not applicable	10,000 1		740685
	CaCO3			140	NA		10,000 Not applicable	10,000 1		740686
	mg/L (	205	132		NA		4,000 Not applicable	4,000 r		2170552
					NR R		10,000 Not reported	10,000		741751
DNUM/5//DOSES/1000/2500/5000/10000 mg/l//TESTID					NR		10,000 Not reported	10,000		741783
DNUM/4//DOSES/1000/5000/10000 mg/l//TESTID/20/					NA		3,526 Not applicable	3,526		741756
DNUM/5//DOSES/1000/2500/5000/1000 mg////TESTID/					NR		10,000 Not reported	10,000		742664
DNUM/5//DOSES/1000/2500/5000/10000 mg/l//TESTID					NR		10,000 Not reported	10,000		742665
DNUM/4//DOSES/1000/5000/10000 mg/l//TFSTID/19/					NA		3,526 Not applicable	3,526		741795
CONTR/B.C//		292	288	290	NA		7,720 Not applicable	7,720		2060323
CONTR/B.C//	mg/L CaCO3	292	288	290	NA		11,700 Not applicable	11,700		2060324
CONTR/B.C//	mg/L CaCO3	292	288	290	NA		11,700 Not applicable	11,700	٧	2060321
					NA		Not applicable	16,595		26116
					NA		Not applicable	18,735		26115
TESTID/3//MSMIT/NR//DOSES/NR//SOURCE/NR//WTINT				100	NA		Not applicable	7,500		231529
				100	NA		6,150 Not applicable	6,150		231530
TESTID/3//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	AN		6,150 Not applicable	6,150		231531
TESTID/3//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/				100	NA		Not applicable	10,000	V	231528
TESTID/3//MSMT/NR//DOSES/NR//SOURCE/NR//WTINT/	mg/L			100	NA		6,150 Not applicable	6,150		231532
DNUM/2//DOSES/4490 mg/L//TESTID/13//					NA		4,490 Not applicable	4,490		742026
DNUM/2//DOSES/4490 mg/L//TESTID/14//					NA		4,490 Not applicable	4,490		742030
DNUM/2//DOSES/4490 mg/L//TESTID/20//					NA	Y	4,490 Not applicable	4,490		742027
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	100	80		NA		Not applicable	3,035		768559
DNUM/2//DOSES/4490 mg/L//TESTID/16//	100				NA		4,490 Not applicable	4,490		742058
General Comments	Hardness Units	Max Max	Min Min	Mean Mean	Significance Level Mean	Op Op	Significance	(mg/L)	Mean Op	Result Number
						0.0			,	

CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2		100	80		NA		2.178 Not applicable	2 178 N		767459
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/		100	80		NA		1,772 Not applicable	1,772 N		767351
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2		100	80		NA	1	2,989 Not applicable	2,989 N		767454
		100	80		NA		3,463 Not applicable	3,463 N		767411
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	mg/L CaCO3	100	80		NA		3,463 Not applicable	3,463 N		767350
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2		100	80		NA		Not applicable	3,463 N		767380
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/	100 mg/L CaCO3	100	80		NA		3,463 Not applicable	3,463 N		767375
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	mg/L CaCO3		80		NA		2,989 Not applicable	2,989 N		767457
DNUN/8//DOSES/594.9/710.4/851.7/873.4/1011.6/1272			80		VR		1,301 Not reported	1,301 N		766854
DNUM/7//DOSES/780/1011/1318/1727/2273/3000 mg/L		100	80		NR		1,727 Not reported	1,727 N		766731
DNUM/7//D0SES/7742/989/1299/1713/2264/3000 mg/L	mg/L CaCO3	100	80		NR		2,264 Not reported	2,264 1		766740
DNUM/6//DOSES/854/1181/1259/1355/1381 mg/L//T					NA		1,381 Not applicable	1,381		761596
				40	NR		610 No significance	610 1		2017030
	mg/L			80	NR		1,250 No significance	1,250 1		2017041
	mg/L			320	NR		1,450 No significance	1,450 1		2017058
	mg/L			160	NR		1,300 No significance	1,300 1		2017049
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TEST					NR		1,060 Not reported	1,060 1		759073
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/1//DOSES	mg/L CaCO3			92	0.05	۸	1,750 No significance	1,750 1		766223
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/2//DOSES				92	0.05	^	2,250 No significance	2,250 1		766225
DNUM/7//DOSES/780/1011/1318/1727/2273/3000 mg/L	1	100	80		NR		2,273 Not reported	2,273 1		766730
DNUM/7//DOSES/7742/989/1299/1713/2264/3000 mg/L	CaCO3	100	80		NR		3,000 Not reported	3,000 1		766739
	mg/L			40	NR		1,300 Significant	1,300 5		2017031
	mg/L			80	NR		2,000 Significant	2,000 9		2017042
	mg/L			320	NR		2,700 Significant	2,700 9		2017059
	mg/L			160	NR		2,600 Significant	2,600 9		2017050
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TEST					NR		Not reported	3,650		759072
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/1//DOSES	mg/L CaCO3			92	0.05	۸	2,216 Significant	2,216		766222
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/2//DOSES	mg/L CaCO3			92	0.05	۸	3,000 Significant	3,000 9		766224
DNUM/7//DOSES/780/1011/1318/1727/2273/3000 mg/L	mg/L CaCO3	100	80		NR		2,250 Not reported	2,250		766748
DNUM/7//DOSES/7742/989/1299/1713/2264/3000 mg/L	mg/L CaCO3	100	80		NR		3,250 Not reported	3,250		766745
CL/497.7 mg/L//DNUM/6-7//DOSES/NA mg/I //TESTID/23	mg/L CaCO3			94	NA		Not applicable	1,116		766584
CL/24.6 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/13/	mg/L CaCO3			96	NA		2,472 Not applicable	2,472		766571
CL/1.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/32/	mg/L CaCO3			194	NA		3,265 Not applicable	3,265		766594
CL/1.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/31//	mg/L CaCO3			194	NA		2,706 Not applicable	2,706		766593
C1/34 mg/1 //DNUM/6-7//DOSES/NA mg/1 //TESTID/28//C	mg/L CaCO3			107	NA :		2,607 Not applicable	2,607		766590
Ct/13 mg/t/ DNUM/6-7//DOSES/NA mg/t//TESTID/42//	mg/L CaCO3			94	NA S		2.470 Not applicable	2.470		766561
CI/1 9 mg/l //DNI IM/6-7//DOSES/NA mg/l //TESTID//	mg/I CaCO3			484	NA		3 338 Not applicable	3 338		766612
CI/1 9 mg/l //DNI IM/6-7//DOSES/NA mg/l //TESTID/26//	mg/L CaCO3			94	NA :		2.269 Not applicable	2.269		766587
Ct/1:9 mg/l //DNU IM/6-7//DOSES/NA mg/l //TESTID/36/	mg/L CaCO3			100	N 2		1.496 Not applicable	1,496		766726
Ct/20 thg/t//DNUM/6-///DOSES/NA mg/t//TESTID/8//Ot	mg/L Cacos			104	200		Not applicable	2,050		766597
CL/10 mg/L//UNUM/6-///UOSES/NA mg/L// IESIIU/2//O	mg/L CaCO3			OUT	NA		2,357 Not applicable	2,337		766563
CL/1.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/39/	mg/L CaCO3			390	NA		Not applicable	3,091		766608
CL/34 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/27//C	mg/L CaCO3			107	NA		2,538 Not applicable	2,538		766588
CL/1.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/24//	mg/L CaCO3			94	NA		2,030 Not applicable	2,030		766585
CL/1.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/37/	mg/L CaCO3			390	NA		Not applicable	3,369		766606
	mg/L CaCO3			100	NA		2,526 Not applicable	2,526		766548
CL/24.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/14/	mg/L CaCO3			96	NA		2,481 Not applicable	2,481		766572
CL/300 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/25/	mg/L CaCO3			100	NA		Not applicable	1,914		766724
CL/34 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/29//C	mg/L CaCO3			107	NA		2,436 Not applicable	2,436		766591
CL/1.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/36/	mg/L CaCO3			288	NA		3,297 Not applicable	3,297		766605
CL/100 mg/L//DNL	mg/L CaCO3			100	NA		,417 Not applicable	2,417		766723
General Comments	Hardness Units	Hardness Max	Hardness Min	Hardness Mean	Significance Level Mean	Level Mean Op	Statistical Significance	Mean (mg/L)	Conc 1 Mean Op	Result Number
						Significance				

C1 / 25 - 11 / (DNI INV ) C 3 / (DOCED) NO 118/ C/ ) 12 / (DECT) (05/			100	NA		1,854 Not applicable	1 854 1		766712
1/25.4 mg/1 //DNI IM/6-7//DOSES/NA mg/1 //TESTID/65			392	NA	V	2,724 Not applicable	2,724 N		766639
CL/24.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/68/	G.		486	NA		4,145 Not applicable	4,145 N		766642
CL/26.6 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/64/			292	NA		2,840 Not applicable	2,840 N		766638
CA/17.6 mg/L//MG/12.1 mg/L//NA/26.3 mg/L//POT/2.1 n	CaCO3		94	NA		262 Not applicable	262 N		766367
	mg/L		80	NA		Not applicable	2,461 N		2017144
	mg/L		80	NA		2,246 Not applicable	2,246 N		2017143
	mg/L		80	NA		2,069 Not applicable	2,069 1		2017113
				NA		17,500 Not applicable	17,500 1		2064205
				NA		16,500 Not applicable	16,500 1		2064206
				NA		24,000 Not applicable	24,000 1		2064204
				NA		10,000 Not applicable	10,000 1		2064207
	100 mg/L CaCO3	80		NA		1,434 Not applicable	1,434		767507
	100 mg/L CaCO3	80		NA		Not applicable	927 [		767494
	100 mg/L CaCO3	80		NA		5,492 Not applicable	5,492 1		767505
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	80		NA		6,844 Not applicable	6,844		767491
		80		NA		6,844 Not applicable	6,844		767475
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	80		NA		6,844 Not applicable	6,844		767496
	100 mg/L CaCO3	80		NA		6,844 Not applicable	6,844		767473
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI				NR		3,650 Not reported	3,650		759091
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI				NR		7,460 Not reported	7,460		759090
				NA		8,384 Not applicable	8,384		31684
				NA		2,564 Not applicable	2,564		119467
HARD/MODERATELYHARD//	100 mg/L CaCO3	80		NA		4,580 Not applicable	4,580		160333
HARD/MODERATELYHARD//	100 mg/L CaCO3	80		NA		6,290 Not applicable	6,290		160332
CA/17.4 mg/L//MG/15.1 mg/L//NA/32.8 mg/L//POT/2.6 n	mg/L CaCO3		106	NA		3,203 Not applicable	3,203		767299
DOSES/NA mg/L//TESTID/17//	mg/L CaCO3		25	NA		1,563 Not applicable	1,563		767305
DOSES/NA mg/L//TESTID/9//	mg/L CaCO3		100	NA		3,203 Not applicable	3,203		767297
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI				NR		Not reported	5,218		759092
				NA		6,800 Not applicable	6,800		18146
				NA		8,384 Not applicable	8,384		18450
				NA		2,564 Not applicable	2,564		18451
				NA		4,547 Not applicable	4,547		18454
				NA		630 Not applicable	630		18453
				NA		6,100 Not applicable	6,100		18147
	Ç			NA		725 Not applicable	725		18452
DOSES/NA mg/L//TESTID/10//	mg/L CaCO3		25	NA		,194 Not applicable	1,194		767298
DOSES/NA mg/1 //TESTID/7//	mp/I CaCO3		50	NA		1,551 Not applicable	1,551		767295
CA/27.2 mg/L//MG/9.2 mg/l //NA/32.8 mg/l //POT/2.6 m	mg/L CaCO3		106	NA		3,808 Not applicable	3,808		767303
DOSES/NA mg/l //TESTID/8//	mg/L CaCO3		75	NA :		3,342 Not applicable	3,342		767296
DOSES (NA ma/I //TESTID/19//	mg/L Cacos		35	NA S		1.985 Not applicable	1.985		767306
DOSES/NA mg/L//TESTID/6//	mg/L CaCO3		106	NA		4 395 Not applicable	4 395		767307
DUSES/NA mg/L//IESIID/1//OEF/LEACHAIE SIUDY, MIXI	b94 mg/L CaCU3	404	503	Z NA		1 100 Not applicable	0,000		767704
			100	NA		13,350 Not applicable	13,350		750175
				NA		11,430 Not applicable	11,430		18144
CA/17.6 mg/L//MG/12.1 mg/L//NA/26.3 mg/L//POT/2.1 r	mg/L CaCO3		94	NA		Not applicable	14,134		766217
CA/17.6 mg/L//MG/12.1 mg/L//NA/26.3 mg/L//POT/2.1 n	mg/L CaCO3		94	NA		11,682 Not applicable	11,682		766218
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	80		NA		1,772 Not applicable	1,772		767376
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	80		NA		1,772 Not applicable	1,772		767353
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	80		NA		1,772 Not applicable	1,772		767412
CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2	100 mg/L CaCO3	80		NR		178 Not reported	2,178		767456
General Comments	Max Hardness Units	Min Hardness	Mean	Level Mean	Op Op	Significance	(mg/L)	Mean Op	Result Number
		_		_				1	

CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//POT/2 CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//POT/2 CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//POT/2 CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//POT/2 CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//POT/2	1 6-603			200			1,400 DIRTHICATE AL ALL COLL	1,400		744007
CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//PO CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//PO CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//PO CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//PO CA/17.6* mg/L//MG/12.1* mg/L//NA/26.3* mg/L//PO	IIIB/L CACOS			200	0.05		1,460 Significant at all con	1,400		200443
CA/17.6* mg/L//MG/12.1* mg/L//MG/26.3* mg/L//PO	mg/L Cacos			00T	0.05		1,400 Significant at all con	1,400		766443
CA / 27 C* / (10 C / 2 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	mg/L Cacos			100 TOO	0.05		1,460 Significant at all con	1,460		766730
(A/1/6* mg/ //M(5/1) 1* mg/ //NA//6 3* mg/ //PO	mg/L CaCO3			200	0.05	^	1,460 Significant at all con	1,460		766438
CA/1/.6° mg/L//MG/12.1° mg/L//NA/26.3° mg/L//PO1/2	mg/L CaCO3			300	0.05	^	Significant at all con	1,460		/66446
	mg/L			080	NX.		2,412 Significant	2,412		201/146
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI				3	NR		Not reported	3,650		759094
CA/32.7 mg/L//MG/6.1 mg/L//NA/26.3 mg/L//POT/2.1 m		100	80	107	NA		Not applicable	2,855		766415
				94	NA		512 Not applicable	512		766365
DOSES/NA mg/L//TESTID/4//	mg/L CaCO3			123	NA		Not applicable	3,144		767292
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TEST					NR		1,226 Not reported	1,226		759093
DOSES/NA mg/L//TESTID/1//	mg/L CaCO3			25	NA		Not applicable	569		767289
CA/16.8 mg/L//MG/14.6 mg/L//NA/27.7 mg/L//POT/2.2 n	mg/L CaCO3			102	NA		Not applicable	2,101		767312
CA/26.3 mg/L//MG/8.9 mg/L//NA/27.7 mg/L//POT/2.2 m	mg/L CaCO3			102	NA		2,240 Not applicable	2,240		767311
DOSES/NA mg/L//TESTID/3//	mg/L CaCO3			75	NA		Not applicable	1,580		767291
DOSES/NA mg/L//TESTID/5//	mg/L CaCO3			250	NA		5,259 Not applicable	5,259		767293
CA/35.9 mg/L//MG/3.1 mg/L//NA/27.7 mg/L//POT/2.2 m	mg/L CaCO3			102	NA		Not applicable	2,725		767310
DOSES/NA mg/L//TESTID/2//	mg/L CaCO3			50	NA		1,448 Not applicable	1,448		767290
CL/105.5 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/47	mg/L CaCO3			100	NA		1,830 Not applicable	1,830		766621
CL/26.3 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/60/	mg/L CaCO3			192	NA		Not applicable	2,203		766634
CL/26.1 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/58/	mg/L CaCO3			102	NA		2,002 Not applicable	2,002		766632
CL/24.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/	mg/L CaCO3			296	NA		2,121 Not applicable	2,121		766636
CL/485 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/52/	mg/L CaCO3			98	NA		1,433 Not applicable	1,433		766626
CL/445 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/54/	mg/L CaCO3			100	NA		1,438 Not applicable	1,438		766628
CL/10 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/12/	mg/L CaCO3			100	NA		1,387 Not applicable	1,387		766709
CL/105.7 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/48	mg/L CaCO3			100	NA		2,002 Not applicable	2,002		766622
CL/25.3 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/63.	mg/L CaCO3			292	NA		3,462 Not applicable	3,462		766637
CL/20 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/14//	mg/L CaCO3			100	NA		1,562 Not applicable	1,562		766711
CL/25.6 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/61/	mg/L CaCO3			196	NA		2,955 Not applicable	2,955		766635
CL/25.6 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/56,	mg/L CaCO3			100	NA		1,901 Not applicable	1,901		766630
CL/25 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/69//C	mg/L CaCO3			482	NA		4,345 Not applicable	4,345		766643
CL/308 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/49/	mg/L CaCO3			98	NA		1,679 Not applicable	1,679		766623
CL/15 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/13/	mg/L CaCO3			100	NA		Not applicable	1.563		766710
CL/25.5 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/55/	mg/L CaCO3			106	NA		2.000 Not applicable	2.000		766629
CL/300 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/:	mg/L CaCO3			100	NA		.691 Not applicable	1.691		766715
CL/100 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/17/	mg/L CaCO3			100	NA		1,938 Not applicable	1,938		766714
CL/108 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/46/	mg/L CaCO3			98	NA		1.879 Not applicable	1.879		766619
CL/310.2 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/50	mg/L CaCO3			96	NA		1.616 Not applicable	1.616		766624
C1/33.7 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/44.	mg/1 CaCO3			100	NA		1,820 Not applicable	1,820		766617
CI /525.5 mg/l //DNIJM/6-7//DOSES/NA mg/l //TESTID/53	mg/L CaCO3			100	NA		Not applicable	1 480		766627
CI /25 3 mg/l //DNI IM/6-7//DOSES/NA mg/l //TESTID/70/	mg/I CaCO3			487	NA		3 796 Not applicable	3 796		766644
	mg/L CaCO3			396	NA		4.046 Not applicable	4.046		766640
CL/25.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/57/	mg/L CaCO3			98	NA		Not applicable	1.621		766631
CL/25.1 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/59,	mg/L CaCO3			194	NA		2,740 Not applicable	2,740		766633
CL/303.7 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/51	mg/L CaCO3			100	NA		1,684 Not applicable	1,684		766625
CL/33.6 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/43/	mg/L CaCO3			96	NA		1,767 Not applicable	1,767		766616
CL/25.5 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/67	mg/L CaCO3			396	NA		4.336 Not applicable	4.336		766641
CL/33 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/16//	mg/L CaCO3			100	NA		1,799 Not applicable	1,799		766713
CL/33.9 mg/L//DNUM/6-7//DOSES/NA mg/L//TESTID/45	mg/L CaCO3			96	NA		1,779 Not applicable	1,779		766618
CL/500 mg/L//DNU	mg/L CaCO3			100	NA		1,470 Not applicable	1,470		766716
its General Comments	Hardness Units	Max	Min	Mean	Level Mean	Level Mean Op	Statistical Significance	(mg/L)	Conc 1 Mean Op	Result Number
						Significance				

57	Conc 1 Mean Op	Mean (mg/L) 1,970	an Statistical (/L) Significance (1,970 Not applicable		Significance Level Mean	Hardness Mean	Hardness Min 80	Hardness Max 100	lits	HARD/I
160499 > 759021		1,9/0	1,970 Not applicable   1,600 Not significant at all concentration		0.05		256	1468	11	CHAR/56% TO 71% SULFATE//S04/160* mg/L//DNUM/6/
767483		1,470	1,470 Not applicable		NA		80	100		CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
767467		1,470	1,470 Not applicable		NA		80	100	mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
767469		860	Not applicable		NA		80	100	mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
767484		1,470	470 Not applicable		NA		80	100	mg/L CaCO3	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3* mg/L//POT/2
759462		360	360 Not applicable		NA	170				DNUM/2//DOSES/360 mg/L//TESTID/1//CONC/ONLY CON
		360	360 Not applicable		NA	170			mg/L CaCO3	DNUM/2//DOSES/360 mg/L//TESTID/2//CONC/ONLY CON
2063749 >		56,000	56,000 Not applicable		NA S					
		56,000	56,000 Not applicable		NA					
196198		2,980	2,980 Not applicable		NA	10			mg/L CaCO3	
26240		2,980	2,980 Not applicable		NA					
160503 >		1,970	1,970 Not applicable		NA		80	100	100 mg/L CaCO3	HARD/MODERATELYHARD//
160502 >		1,970	1,970 Not applicable		NA		80	100	100 mg/L CaCO3	HARD/MODERATELYHARD//
160501 >		1,970	1,970 Not applicable		NA		80	100	100 mg/L CaCO3	HARD/MODERATELYHARD//
/6/512		1,4/0	1,4/0 Not applicable		NA		080	100	mg/L CaCO3	CA/14.0° mg/L//MG/12.0° mg/L//NA/26.3° mg/L//PO1/2
67863		300 074/T	1,470 Not applicable		NA		00	700	TOO IIIB/ L CACOS	CA/14.0 INB/C//MIG/12:0 INB/C//MA/20:3 INB/C//FO//
67862		300	300 Not applicable		NA :					
761768 >		1,410	1,410 Not applicable		NA					DOSES/NA mg/L//TESTID/10//OEF/MICROTOX STUDY///
761769		1,410	1,410 Not applicable		NA					DOSES/NA mg/L//TESTID/10//OEF/MICROTOX STUDY///
2152811		2,316	2,316 Not applicable		NA		80	100	100 mg/L CaCO3	
2152812		2,590	2,590 Not applicable		NA		80	100	100 mg/L CaCO3	
2152815		3,960	3.960 Significant	٨	0.01		80	100	100 mg/L CaCO3	
2152814		2,133	No significance	٨	0.01		80	100	mg/L CaCO3	
201556				۸	0.05	96.3			mg/L CaCO3	
201557		372	372 Not applicable		NA	96.3			mg/L CaCO3	
201555		314	314 No significance	۸	0.05	96.3			mg/L CaCO3	
2164559		421	421 Not applicable		NA					
2152816		1,705	1,705 Not applicable		NA		80	100	mg/L CaCO3	
2152817		2,298 I	2,298 Not applicable	`	NA OO1		80 80	100	100 mg/L CaCO3	
2152818		2,210	ance	۸	0.01		80	100	100 mg/L CaCO3	
761548		1,360	1,360 Not applicable		NA					DOSES/NA g/L//TESTID/1//
761558		1,330	1,330 Not applicable		NA					
761565		1,730	1,730 Not applicable		NA					DOSES/NA 8/L//TESTID/9//
761566		1,090	940 Not applicable		NA NA					DOSES/NA B/L//TESTID/3//
761569		1,470	1,470 Not applicable		NA					
761561		1,020 1	1,020 Not applicable		NA					DOSES/NA g/L//TESTID/7//
761557		1,430 1	1,430 Not applicable		NA					
761551		1,610	Not applicable		NA					DOSES/NA g/L//TESTID/2//
761562		1,090	1,090 Not applicable		NA					DOSES/NA g/L//TESTID/8//
761570		2 070	2,070 Not applicable		N 5					DOSES/NA s/1//TESTID/13//
2152752		704 1	704 Not applicable		NA		80	100	100 mg/L CaCO3	Of all sections
2152753		958	958 Not applicable		NA		80	100	mg/L CaCO3	
768151		2,500 1	2,500 Not reported		NR		80		E	DOSES/NA g/L//TESTID/50//
768129		2,500 N	2,500 Not reported		NR		80	100		
768092		2 500 1	2 500 Not reported		NR .		08	100	100 mg/l CaCO3	DOSES (NIA #/ I / / TESTID / 37 / /

	Conc 1 Mean Op	Mean (mg/L)	Statistical Significance	Significance Level Mean Op	Significance Level Mean	Hardness Mean	Hardness Min	Hardness Max	Hardness Units	
768111		2,500	,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/42//
768161		5,000	5,000 Not reported		NR		80	100	100 mg/L CaCO3	DOSES/NA g/L//TESTID/52//
768086		5,000	5,000 Not reported		NR		80			DOSES/NA g/L//TESTID/36//
768094		2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/38//
768149		2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/49//
768078		5,000	5,000 Not reported		NR		80	100		DOSES/NA g/L//TESTID/34//
768159		2,500	2,500 Not reported		NR NR		80	100	mg/L CaCO3	DOSES/NA g/L//TESTID/31//
768076		2,500	2,500 Not reported		NR		80	100		
768139		2,500	2,500 Not reported		NR		80	100		
2152755		1,058	1,058 Significant	۸	0.01		80	100	25	
768070		2,500	2,500 Not reported		NR NR		80	100	100 mg/L CaCO3	DOSES/NA g/L//TESTID/32//
768119		5,000	5,000 Not reported		NR		80	100	100 mg/L CaCO3	DOSES/NA g/L//TESTID/43//
768109		2,500	2,500 Not reported		NR		80	100	8	
768102		2,250	2,250 Not reported		NR		80	100		DOSES/NA g/L//TESTID/40//
768131		2,500	2,500 Not reported		NR		80	100		DOSES/NA g/L//TESTID/46//
768100		2,500	2,500 Not reported		NB NR		00	100	mg/L CaCO3	DOSES/NA 8/L//TESTID/44//
768060		2,500	2,500 Not reported		NR S		80	100	mg/L CaCO3	DOSES/NA g/L//TESTID/29//
768062		10,000	10,000 Not reported		NR		80	100		DOSES/NA g/L//TESTID/30//
761764		1,000	1,000 Not applicable		NA					DOSES/NA mg/L//TESTID/8//OEF/MICROTOX STUDY////
741831		2,200	2,200 Significant at all concentrations	centrations	0.05		90	110		TIME/MEASURED DURING RECOVERY//DNUM/2//DOSES
761549		1,000	1,140 Significable		NA O.U.S		90	OTT	TIO III B/ L CACOS	DOSES/NA g/L//TESTID/1//
761567		500	500 Not applicable		NA					DOSES/NA g/L//TESTID/10//
761560		1,000	1,000 Not applicable		NA					DOSES/NA g/L//TESTID/7//
761555		1,000	1,000 Not applicable		NA					DOSES/NA g/L//TESTID/4//
761567		1,000	1,000 Not applicable		N N					DOSES/NA g/L//TESTID/8//
761552		1,000	1.000 Not applicable		NA S					DOSES/NA g/L//TESTID/3//
761568		1,000 1	1,000 Not applicable		NA					DOSES/NA g/L//TESTID/11//
761556		1,000	Not applicable		NA					DOSES/NA g/L//TESTID/5//
761563		1,000	1,000 Not applicable		NA					DOSES/NA g/L//TESTID/8//
761571		1,000	1,000 Not applicable		NA		3			DOSES/NA g/L//TESTID/12//
768130		1,250	1,250 Not reported		NR NR		80	100	mg/L CaCO3	DOSES/NA g/L//TESTID/45//
768093		1,250	1,250 Not reported		NR		80	100	, Yr	DOSES/NA g/L//TESTID/37//
768112		1,250	1,250 Not reported		NR		80		ň	DOSES/NA g/L//TESTID/42//
768142		1,250	1,250 Not reported		NR		80			DOSES/NA g/L//TESTID/48//
768162		2,500	Not reported		NR		80	100		DOSES/NA g/L//TESTID/52//
768095		1 250	1 250 Not reported		ZB S		80	100	mg/I CaCO3	DOSES/NA g/L//TESTID/38//
768150		1,250	1,250 Not reported		NR		80	100		DOSES/NA g/L//TESTID/49//
768079		2,500 1	2,500 Not reported	-	NR		80	100		DOSES/NA g/L//TESTID/34//
768160		1,250 1	Not reported	7	NR		80	100		DOSES/NA g/L//TESTID/51//
768085		1,250 1	1,250 Not reported	7	NR		80	100		DOSES/NA g/L//TESTID/35//
768077		1,250 1	Not reported		NR NR		80	100		DOSES/NA g/L//TESTID/33//
2152754		558 1	558 No significance	٨	0.01		80	100	100 mg/L CaCO3	000-01 NO 8/4/1-00-01-01
768071		1 250 1			NR		US	400		b comp (i.e. ), ( function) (co. )

7148474	2148480	2148464	761576	761587	761575	761580	741538 >	207720	741661	2152725	2152823	2152824	2152822	2152821	2152820	2152740	2152741	2152739	761757 <	761756	2152767	2152768	2152766	761763 <	766857		761599 >	765000	765901	765003	765000 CUT/	742694	741815	761765	108590	96882	42867	14380	42862	14424	768063	768061	768101	768122	768132	768103	761559	768110	768120		Conc 1 Result Number   Mean Op
			1	1	1	1		1,	1	2,	1	2	1	1	1		1	1,				1			1		1								4	4	4	4	4	4	CH.									$\exists$	Conc 1 Mean Mean Op (mg/L)
	340 Not applicable	147 Not applicable	1,200 Not applicable	1,170 Not applicable	1,160 Not applicable	1,060 Not applicable	467 Not applicable	1,750 Not applicable	1,350 Not applicable	2,750 Not applicable	1,120 No significance	2,230 Significant	1,945 Not applicable	1,505 Not applicable	1,241 Not applicable	668 No significance	1,198 Significant	1,750 Not applicable	305 Not applicable	305 Not applicable	638 No significance	1,106 Significant	750 Not applicable	220 Not applicable	1,301 Not reported	220 Not applicable	1.381 Not applicable	1 500 Not significant at al	1 500 Not significant at all concentration	1,500 Significant at all concentrations	2,000 Not significant at all concentrations	210 Not significant at all concentration	860 No significance	500 Not applicable	4,000 No significance	5,000 Not reported	1,250 Not reported	2,500 Not reported	1,250 Not reported	1,250 Not reported	1,250 Not reported	1,000 Not applicable	1,250 Not reported	2,500 Not reported	No.	an Statistical /L) Significance					
											۸	۸															Control of the control	concentration	I concentration	Centrations	Centrations	concentration																			Level Mean Op
200	NA	NA	NA	NA	NA	NA	NA	NA	NA	AN	0.01	0.01	NA	NA	NA	NR	NR	NA	NA	AN	NR	NR	NA	NA	NR	NA A	NA A	I	0.8333	70.010	0.03			NA	0.05	0.05	0.05	0.05	0.05	0.05	NR	NR	NR	NR	NR	NR	NA	NR	NR	NR	Significance Level Mean
2	44	44					101	102		98																		92	92	92	97																				Hardness Mean
							100	94	54		80	80	80	80	80										80						20	00	90		86	86	86	86	86	86	80	80	80	80	80	80		80	80	08	Hardness Min
							103	104	72		100	100	100	100	100										100						011	110	110		94	94	94	94	94	94	100	100	100	100	100	100		100	100	100	Hardness Max
ma/1	mg/L	mg/L					mg/L CaCO3	104 mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	100 mg/L CaCO3	mg/L CaCO3	100 mg/L CaCO3	100 mg/L CaCO3	mg/L CaCO3										mg/L CaCO3		0/	mg/l CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	110 mg/L CaCO3	110 mg/L CaCO3		94 mg/L CaCO3	94 mg/L CaCO3	mg/L CaCO3	94 mg/L CaCO3	94 mg/L CaCO3	94 mg/L CaCO3	100 mg/L CaCO3	mg/L CaCO3	100 mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	100 mg/L CaCO3		100 mg/L CaCO3		mg/L CaCO3	Hardness Units
			DOSES/NA g/L//TESTID/15//	DOSES/NA g/L//TESTID/20//	DOSES/NA g/L//TESTID/14//	DOSES/NA g/L//TESTID/17//		CONC/FROM GRAPH//	DNUM/7//DOSES/0.21/0.44/0.85/1.3/1.7/2.2 g/L//TESTID	TMETH/APHA, 1989// CONC1/FROM GRAPH//						TMETH/APHA, 1989//	TMETH/APHA, 1989//	TMETH/APHA, 1989// CONC1/FROM GRAPH//	DOSES/NA mg/L//TESTID/4//OEF/MICROTOX STUDY////	DOSES/NA mg/L//TESTID/4//OEF/MICROTOX STUDY////	TMETH/APHA, 1989//	TMETH/APHA, 1989//	TMETH/APHA, 1989// CONC1/FROM GRAPH//	DOSES/NA mg/L//TESTID/7//OEF/MICROTOX STUDY////	DNUM/8//DOSES/594.9/710.4/851.7/873.4/1011.6/1272	DOSES/NA mg/L//TESTID/7//OEF/MICROTOX STUDY////	DNUM/6//DOSES/854/1181/1259/1355/1381 mg/L//TES	DNIIM/2//DOSES/1500 mg/l //TESTID/1//CONC/ONLY CO	DNUM/2//DOSES/1500 mg/L//TESTID/1//CONC/ONLY CO	DNI IM/2// DOSES/1500 mg/I //TESTID/1//CONC/ONLY	DNITN/2//DOSES/1500 mg/1 //TESTID/1//CONC/ONLY CO	DNIIINA/3//DOSES/3 7/1 //TESTID/10//	TIME/MEASURED DURING RECOVERY//DNUM/3//DOSES	DOSES/NA mg/L//TESTID/8//OEF/MICROTOX STUDY////							DOSES/NA g/L//TESTID/30//	DOSES/NA g/L//TESTID/29//	DOSES/NA g/L//TESTID/39//	DOSES/NA g/L//TESTID/44//	DOSES/NA g/L//TESTID/46//	DOSES/NA g/L//TESTID/40//	DOSES/NA g/L//TESTID/6//	DOSES/NA g/L//TESTID/41//	DOSES/NA g/L//TESTID/43//	DOSES/NA g/L//TESTID/31//	s General Comments

Conc 1 Result Number Mean Op	Mean (mg/L)	Statistical Significance	Significance Level Mean Op	Significance Level Mean	Hardness Mean	Hardness Min	Hardness Max	Hardness Units	
761579 761588	1,140	1,140 Not applicable		NA					DOSES/NA g/L//TESTID/16//
761572	600	600 Not applicable		NA					DOSES/NA g/L//TESTID/13//
761584	670	670 Not applicable		NA					DOSES/NA g/L//TESTID/19//
761591	690	690 Not applicable		NA					DOSES/NA B/L//TESTID/22//
761592	710	710 Not applicable		NA					DOSES/NA g/L//TESTID/23//
2153272	454	454 Not applicable		NA	80			mg/L CaCO3	
2152721	454	454 Not applicable		NA		80	100	100 mg/L CaCO3	
2152855	117	Not applicable		NA	10			mg/L CaCO3	
2153275	580	580 Not applicable		NA	160			mg/L CaCO3	
2153278	521	521 Not applicable		NA	320			mg/L CaCO3	
2153269	146	146 Not applicable		NA	40			mg/L CaCO3	
2152881	264	264 Not applicable		NA	20			mg/L CaCO3	
2148463	342	Not applicable		NA	44			mg/L	
2148479	563	563 Not applicable		NA	44			mg/L	
2148473	653	653 Not applicable		NA	93			mg/L	
120834	1,500	1,500 Not applicable		NA					TESTID/32//
120833	1,760	Not applicable		NA					TESTID/22//
120830	1,600	1,600 Not applicable		NA					TESTID/20//
120832	1,290	1,290 Not applicable		NA					TESTID/21//
120852	740	740 Not applicable		NA					TESTID/33//
120853	650	650 Not applicable		NA					TESTID/34//
120850	1,340	1,340 Not applicable		NA					TESTID/31//
120849	300	300 Not applicable		NA					TESTID/30//
120846 <	330	330 Not applicable		NA					TESTID/27//
120835	720	720 Not applicable		NA					TESTID/24//
120848	1,390	1,390 Not applicable		NA					TESTID/29//
120847	1,690	1,690 Not applicable		NA					TESTID/28//
120854	730	730 Not applicable		NA					TESTID/35//
120831 <	330	330 Not applicable		NA					TESTID/26//
741679 DF ORGANIS		750 Not applicable		NA S		40	48	48 mg/L CaCO3	DNUM/5//DOSES/0.25/0.5/1.0/2.0 e/L//TESTID/2//
741936 DF ORGANIS		1,100 Not applicable		NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/4/
741950 DF ORGANIS		670 Not applicable		NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/12//
742564 DF ORGANIS	1	,130 Not applicable		NA		40	48	mg/L CaCO3	DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/1//
741951DF ORGANIS		960 Not applicable	Y	NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/8//
741929 DF ORGANIS		920 Not applicable		NA		40	48	1	DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/3//
742894 DF ORGANIS		980 Not applicable		NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/5//
741958 DE ORGANIS		570 Not applicable		NA NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/7//
741948 DF ORGANIS		720 Not applicable		NA		40	48	48 mg/l CaCO3	DNIJM/5//DOSES/0.25/0.5/1.0/2.0 g/1//TESTID/11//
741943 DF ORGANIS		710 Not applicable	7	NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/9//
		680 Not applicable	7	NA		40	48		DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/10//
2153273	697	697 Not applicable	7	NA	80			mg/L CaCO3	
2152731	697	Not applicable	7	NA		80	100	mg/L CaCO3	
2152873	161	161 Not applicable	7	NA	10			mg/L CaCO3	
2153279	700	700 Not applicable	2 -	NA	320			mg/L CaCO3	
C12C12		The second second second			0.00			10/ - 00000	

Result Number 761582 767861 767869 766869 767875 767877 767877 767879 741688	Conc 1  Wean Op  1  1  1  1  1  1  1  1  1  1  1  1  1	Mean (mg/L) 500 620 620 620 620 620 620 620 620 620 6	an Statistical /L) Significance 500 Not applicable 1,250 Not reported 620 Not applicable	Significance Level Mean Op	Significance Level Mean NA NR NR NR NR NR NR NR	Hardness Mean	Hardness Min 80 80 80 80 80 80 80 80 80 80 80 80 80	Hardness Max 100 100 100 100 100 100 100 100 100 72	Hardness Units Hardness Units 100 mg/L CaCO3	General (  DOSES/NA g/L/TESTID/18//  DOSES/NA g/L/TESTID/12//  DOSES/NA g/L/TESTID/12//  DOSES/NA g/L/TESTID/13//  DOSES/NA g/L/TESTID/13//  DOSES/NA g/L/TESTID/13//  DOSES/NA g/L/TESTID/14//  DOSES/NA g/L/TESTID/14//
761578 761586 761574		1,500	1,500 Not applicable 1,000 Not applicable 1,000 Not applicable		NA NA					DOSES/ DOSES/
761590 767853	3 0 4	2005	500 Not applicable		NA NA		Ug	100		
767853 767956	01 00	130	130 Not reported		NR NR		80	100		mg/L CaCO3 DOSES/NA g/L//TESTID/8//
767851	1	620	620 Not reported		NR		80		100	mg/L CaCO3
766863 767867	3	620	620 Not reported		NR NR		80		100	mg/L CaCO3
766871		620	Not reported		NR		80		100	
767892	2	310	310 Not reported		NR		80		100	mg/L CaCO3
767964	44 0	620	620 Not reported		NR NR		80		100	100 mg/L CaCO3 DOSES/NA g/L//TESTID/23//
767900		620	620 Not reported		NR		80		100	mg/L CaCO3
767966		620	Not reported		NR NR		80		100	100 mg/L CaCO3 DOSES/NA g/L//TESTID/24//
251019		300	300 No significance		NR NR		00		5	TESTID/37//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT/
251023	3ABS///	900	No significance		NR					TESTID/
	۸	300	300 No significance		NR					TESTID/37//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT/
120856 < 251013 ABS///	ARS///	300	300 No significance		NR R					TESTID/36//COMPEP/3// TESTID/36//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT/
251015	ABS///	1,500	,500 No significance		NR					TESTID/36//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT
251021	1	500	500 No significance		NR					TESTID/37//DOSES/<0.3/0.3/0.5/0.9/1.5/2.5 G/L//MSMT
761585 761577	4 0.	1 500	500 Not applicable		NA NA					DOSES/NA g/L//TESTID/19//
761589		500	500 Not applicable		NA					DOSES/NA g/L//TESTID/21//
761593		500	500 Not applicable		NA S					DOSES/NA g/L//TESTID/23//
761581		1,500	500 Not applicable		NA					
742896	742896DF ORGANIS	250	500 No significance	^ ^	0.05		40		48	48 mg/L CaCO3
742898	DF ORGANIS	250	250 No significance	^ -	0.05		40		48	
741937	741937 DF ORGANIS	500	500 No significance		0.5		40		48	48 mg/L CaCO3 DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/4//
742893		500	500 No significance				40		48	mg/L CaCO3
742901 741648	742901 DF ORGANIS 741648 DF ORGANIS	500	500 No significance	^	0.05 NR		40		48	48 mg/L CaCO3 DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/12// 48 mg/L CaCO3 DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/1//
741649	741649 DF ORGANIS	250	250 No significance	^ ^	0.05		40		48	48 mg/L CaCO3 DNUM/5//DOSES/0.25/0.5/1.0/2.0 g/L//TESTID/2//
741942	741942 DF ORGANIS	250	250 No significance	^	0.05		40		48	mg/L CaCO3
742897	٨	250	No significance	۸	0.05		40		48	48 mg/L CaCO3 DNUM//

	mg/L CaCO3 DN			+10		Olicelle and				
				170	0.05	nocentration	360 Not significant at all concentration	360 N	759463 SE OVER CO	7594635
		1468 r	256		0.05	oncentration	,600 Not significant at all concentration	1,600 N		759022
DNUM/2//DOSES/360 mg/L//TESTID/2//CONC/ONLY CON	CaCO3			170	0.05	oncentration	360 Not significant at all concentration	360 N	759464 SE OVER CO	759464
	mg/L	7		80	NR		1,637 No significance	1,637 N		2017150
	mg/L			80	NR		Significant	2,412 Si		2017151
	mg/L			80	NA		Not applicable	2,412 N	V	2017149
	mg/L			80	AN		1,056 Not applicable	1,056 N		2017148
	mg/L	_		80	AN		Not applicable	380 N		2017147
DOSES/NA mg/L//TESTID/1//OEF/MICROTOX STUDY////	DC				NA		2,220 Not applicable	2,220 N	٨	761751
DOSES/NA mg/L//TESTID/1//OEF/MICROTOX STUDY////	DC				NA		2,220 Not applicable	2,220 N		761750
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TEST					NR		Not reported	1,060 N		759075
	mg/L			320	NR	7 5 77	420 No significance	420 N		2017061
	mg/L			160	NR		775 No significance	775 N		2017052
				92	0.05	^	No significance	450 N		766233
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/2//DOSES				92	0.05	٨		818 N		766231
DNUM/8//DOSES/594.9/710.4/851.7/873.4/1011.6/127	mg/L CaCO3 DI	100	80		NR		1,301 Not reported	1,301 N		/66855
DNUM/7//DOSES/7742/989/1299/1713/2264/3000 mg/L		100	80		NR		Not reported	906 N		/66/38
DNUM/7//DOSES/780/1011/1318/1727/2273/3000 mg/L		100	80		NR		780 Not reported	780 N		766735
	mg/L			80	NR		645 No significance	645 N		2017044
	mg/L			40	NR		150 No significance	150 N	۸	
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI					NR		,650 Not reported	3,650 N		759074
	mg/L			320	NR		480 Significant	480 S		2017060
				160	NR		,300 Significant	1,300 S		2017051
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/1//DOSES		100		92	0.05	٨		899 S		766232
	mg/L CaCO3 D			92	0.05	^	at all con	1,000 S		766356
DNUM/2//DOSES/1000 mg/L//TESTID/3//CONC/ONLY CO	mg/L CaCO3 D			92	0.05	۸	at all con	1,000 S		766351
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/2//DOSES				92	0.05	۸	1,236 Significant	1,236 S		766230
DNUM/2//DOSES/1000 mg/L//TESTID/3//CONC/ONLY CO				92	0.05	^	at all con	1,000 S		766355
DNUM/2//DOSES/1000 mg/L//TESTID/3//CONC/ONLY CO				92	0.05	٨	1,000 Significant at all con	1,000 S		766344
NUM/2//DOSES/1000 mg/L//TESTID/3//CONC/ONLY C				92	0.05	۸	all con	1,000 S		766354
DNUM/7//DOSES/7742/989/1299/1713/2264/3000 mg/L		100	80		NR		1,195 Not reported	1,195		766737
DNUM/7//DOSES/780/1011/1318/1727/2273/3000 mg/L	CaCO3	100	80		NR		934 Not reported	934		766734
	mg/L			80	NR		,250 Significant	1,250 9		2017043
Oi -11	mg/L			40	NR		150 Significant	150 9		2017032
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI					NA		2,061 Not applicable	2,061		759077
	mg/L			44	NA		766 Not applicable	766 1		2148467
	mg/L			93	NA		1,252 Not applicable	1,252		2148469
(i) -	mg/L			44	NA		Not applicable	715		2148475
DNUM/6//DOSES/200/500/1000/3000/6000 mg/L//TESTI	0				NA		1,267 Not applicable	1,267		759076
DNUM/6//DOSES/854/1181/1259/1355/1381 mg/L//TES					NA		1,381 Not applicable	1,381	>	761597
	mg/L			93	AN		,060 Not applicable	1,060		2148470
	mg/L			44	NA		496 Not applicable	496		2148476
	mg/L			44	NA		625 Not applicable	625		2148468
	mg/L			80	NA		,129 Not applicable	1,129		2017045
	mg/L	7		320	NA		843 Not applicable	843		2017062
				160	NA		1,257 Not applicable	1,257		2017053
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/1//DOSES				92	NA		Not applicable	1,148		766234
DNUM/7//DOSES/>500 to <3500 mg/L//TESTID/2//DOSES	CaCO3			92	NA		1,458 Not applicable	1,458		766229
	mg/L		1	40	AN		465 Not applicable	465		2017034
	mg/L			320	NA		542 Not applicable	542		2017063
General Comments	Hardness Units	Hardness Max	Hardness Min	Hardness Mean	Significance Level Mean	Level Mean Op	Statistical Significance	Mean (mg/L)	Conc 1 Mean Op	Result Number
						Significance				

1959 1959 1959	The Relationship of Bluegill Sunfish Body Sid In: Proc.13th Ind.Waste Conf., Series No.95, The Relationship of Bluegill Sunfish Body Sid In: Proc.13th Ind.Waste Conf., Series No.95, The Relationship of Bluegill Sunfish Body Sid In: Proc.13th Ind.Waste Conf., Series No.95,	The Relationship of Bluegill Sunfish Bod The Relationship of Bluegill Sunfish Bod The Relationship of Bluegill Sunfish Bod	930 Cairns,J.,Jr., and A. Scheier	930 930	126052
1959 1959	<pre>jy SizIn: Proc.13th Ind.Waste Conf.,Series No.95, ly SizIn: Proc.13th Ind.Waste Conf.,Series No.95,</pre>	The Relationship of Bluegill Sunfish Bod	Cairns, J., Jr., and A. Scheier	930	126052
1959	dy Sialn: Proc.13th Ind.Waste Conf.,Series No.95,	The Relationship of BluegIII Sunfish Boo	(21110)010) 2112 11 (2110)	930	10001
	-0		Cairns.JJr., and A. Scheier		126054
1996	and Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Prog. Fish-Cult. 58(2): 77-84	76054 Waller D.L. S.W. Fisher, and H. Dabrov	76054	740777
1996	and Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Prog. Fish-Cult.58(2): 77-84	Waller,D.L., S.W. Fisher, and H. Dabro	76054	740765
1957	Wallen, I.E., W.C. Greer, and R. Lasater Toxicity to Gambusia affinis of Certain Pure Sewage Ind. Wastes29(6): 695-711	Toxicity to Gambusia affinis of Certain I	Wallen,I.E., W.C. Greer, and R. Lasater	508	2063735
1957	508 Wallen, I.E., W.C. Greer, and R. Lasater Toxicity to Gambusia affinis of Certain Pure Sewage Ind. Wastes29(6): 695-711	Toxicity to Gambusia affinis of Certain I	Wallen,I.E., W.C. Greer, and R. Lasate	508	2063726
1957	Wallen,I.E., W.C. Greer, and R. Lasated Toxicity to Gambusia affinis of Certain Pure Sewage Ind. Wastes29(6): 695-711	Toxicity to Gambusia affinis of Certain I	Wallen, I.E., W.C. Greer, and R. Lasate	508	2063733
1996	and Prog. Fish-Cult.58(2): 77-84	76054 Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult. 58(2): 77-84	Waller,D.L., S.W. Fisher, and H. Dabro	76054	740761
1996	and Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult. 58(2): 77-84	Waller,D.L., S.W. Fisher, and H. Dabro	76054	2153697
1996	and Prog. Fish-Cult.58(2): 77-84	Prevention of Zebra Mussel Infestation and Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro	76054	2153700
1996	and Prog. Fish-Cult.58(2): 77-84	76054 Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro	76054	740732
1996	and Prog. Fish-Cult.58(2): 77-84	76054 Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult.58(2): 77-84	Waller,D.L., S.W. Fisher, and H. Dabro	76054	740760
1996	and Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult. 58(2): 77-84	Waller,D.L., S.W. Fisher, and H. Dabro	76054	740759
1996	and Prog. Fish-Cult.58(2): 77-84	76054 Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult. 58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro	76054	2153676
1996	and Prog. Fish-Cult.58(2): 77-84	76054 Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro	76054	2153763
1992	Development of a Salinity/Toxicity Relation: Final Rep. GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount,D.R., and D.D. Gulley	14761	768398
1992	Development of a Salinity/Toxicity Relation: Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	14761 Mount, D.R., and D.D. Gulley	14761	768460
1992	Development of a Salinity/Toxicity Relation: Final Rep. GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount,D.R., and D.D. Gulley	14761	768402
1991	HardArch. Hydrobiol.122(1): 33-56	Demographic Effects of Salinity, Water Hard Arch. Hydrobiol.122(1): 33-56	Cowgill, U.M., and D.P. Milazzo	119414	2152736
1992	Development of a Salinity/Toxicity Relation Final Rep. GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela		14761	768462
1992	Development of a Salinity/Toxicity Relation Final Rep. GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount,D.R., and D.D. Gulley	14761	768457
1992	Development of a Salinity/Toxicity Relation; Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	14761 Mount, D.R., and D.D. Gulley	14761	768396
1992	ation Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount,D.R., and D.D. Gulley	14761	768399
1961	ic Sall Proc. La. Acad. Sci.23:77-85	Cumulative Toxicities of Some Inorganic Sall Proc. La. Acad. Sci. 23:77-85	Dowden,B.F.	2465	119478
1961	ic Sall Proc. La. Acad. Sci.23:77-85	Cumulative Toxicities of Some Inorganic Sall Proc. La. Acad. Sci.23:77-85	Dowden, B.F.	2465	32466
1997	Statistical Models to Predict the Toxicity of Environ, Toxicol, Chem.16(10): 2009-2019	Statistical Models to Predict the Toxicit	Mount.D.R., D.D. Gulley, J.R. Hockett.	18272	160621
1980	Resonance of Freshwater and Saltwater Toxid Ph D Thesis   Injuercity of North Texas   Deni	Response of Freshwater and Saltwater	Drice E E	195/19	742079
1065	Toxicity of Selected Chemicals to Certain Apl. Water Pollut. Control Fed. 37(9): 1308-131	Toxicity of selected chemicals to certa	915 Downden B.E. and H.J. Beilliett	915	10607
1965	in AnJ. Water Pollut. Control Fed. 37(9): 1308-131	Toxicity of Selected Chemicals to Certa	Dowden, B.F., and H.J. Bennett	915	18684
1965	in AnJ. Water Pollut. Control Fed.3/(9): 1308-131	loxicity of selected Chemicals to Certa	Dowden, B.F., and H.J. Bennett	915	18688
1965	Toxicity of Selected Chemicals to Certain AnJ. Water Pollut. Control Fed. 37(9): 1308-131	Toxicity of Selected Chemicals to Certa	915 Dowden, B.F., and H.J. Bennett	915	18685
1965	iin AnJ. Water Pollut. Control Fed.37(9): 1308-131	Toxicity of Selected Chemicals to Certa	Dowden,B.F., and H.J. Bennett	915	18686
1993	· HardArch. Hydrobiol.122(1): 33-56	Demographic Effects of Salinity, Water Hard Arch. Hydrobiol.122(1): 33-56	119414 Cowgill, U.M., and D.P. Milazzo	119414	2152737
1991	· HardArch. Hydrobiol.122(1): 33-56	Demographic Effects of Salinity, Water Hard Arch. Hydrobiol.122(1): 33-56	Cowgill, U.M., and D.P. Milazzo	119414	2152738
1997	Statistical Models to Predict the Toxicity of Environ. Toxicol. Chem.16(10): 2009-2019	_	Mount,D.R., D.D. Gulley, J.R. Hockett,	18272	160622
1974	Acute Toxicity of Various Metals to Freshwa Bull. Environ. Contam. Toxicol.12(6): 745-79	Acute Toxicity of Various Metals to Fre	5339 Baudouin, M.F., and P. Scoppa	5339	44241
1989	Toxi Ph.D.Thesis, University of North Texas, Den	Response of Freshwater and Saltwater	Price,E.E.	19549	742988
1992	ation Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela		14761	768286
1992	ation Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount,D.R., and D.D. Gulley	14761	768181
1992	Development of a Salinity/Toxicity Relations Final Rep. GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount,D.R., and D.D. Gulley	14761	768183
1992	ation Final Rep. GRI-92/0301. FNSR Contract No. 5	Development of a Salinity/Toxicity Bela	14761 Mount D.B. and D.D. Gullev	14761	768283
1993	ation Final Ren GRI-92/0301 FNSR Contract No. 5	Development of a Salinity/Toxicity Rela	Mount D.B. and D.D. Gulley	14761	767342
1992	ation: Final Rep. GRI-92/0301. ENSR Contract No.5	Development of a Salinity/Toxicity Rela	Mount.D.R and D.D. Gullev	14761	768182
1992	Development of a Salinity/Toxicity Relation Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela	14761 Mount D.R., and D.D. Gullev	14761	768284
1992	ation Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Rela		14761	768180
199	HardArch, Hydrobiol.122(1): 33-56	Demographic Effects of Salinity. Water Hard Arch. Hydrobiol.122(1): 33-56	19414 Coweill U.M., and D.P. Milazzo	119414	2152765
1997	Charitatical Models to Predict the Tovisity of Environ Tovisol Chem 16/10): 2003 2003	Statistical Models to Predict the Toxici	Mount DR DD Gulley IR Hockett	10272	160620
199	ty of [Fnyiron Toxical Chem 16/10]: 2009-2019	Statistical Models to Predict the Toxici	Mount D.B. D.D. Gulley I.B. Hockett	18272	160619
1991	Hard Arch Hydrohiol 122(1): 33-56	Demographic Efforts of Calinity, Water Hard Arch Hydrohiol 122/11: 33-56	110000 Clasellinge, G.I., and D.P. Milazzo	1100114	2147207
7007		Effect of Calcium Chloride on Growth	Olacabinda G L and LB O Overinda	110666	21/7107
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1992	DeGraeve,G.M., J.D. Cooney, B.H. Mat Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem.11(6): 851-866	Variability in the	eGraeve,G.M., J.D. Cooney, B.H. Mar	11152 D	251001
1992	11152 DeGraeve,G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem.11(6): 851-866	Variability in the	eGraeve,G.M., J.D. Cooney, B.H. Mar	11152 D	251003
1992	e Performance of the 7-D Ce Environ. Toxicol. Chem.11(6): 851-866	Variability in the	DeGraeve, G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce	11152 D	251007
1993	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioass	Sanders, D.F.	116830 S	767854
1993	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioass		116830 S	767848
1993	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioass		116830 S	767862
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioass		116830 S	767864
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767975
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767881
1993	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas:		116830 S	767887
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767953
1993	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas	Sanders, D.F.	116830 S	767846
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas.		116830 S	767969
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas	Sanders, D.F.	116830 S	767856
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas	Sanders,D.F.	116830 S	767889
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	766866
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas	Sanders,D.F.	116830 S	766872
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767967
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767870
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas	Sanders, D.F.	116830 S	767959
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767872
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767895
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas	Sanders,D.F.	116830 S	767879
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767961
	ssays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioas		116830 S	767897
1993	Site Water Bioassays for Huff and Huff, IncolPS Environmental and Analytical Services, A	Site Water Bioas			766864
	scave for Huff and Huff Incolps Environmental and Analytical Services A	Site Water Binas	Sanders D.F.	116830	767903
	A Comparison of the Daphnids Ceriodaphnid Arch. Environ. Contam. Toxicol. 45(1): 79-85	A Comparison of	_1	71674	742567
199	of the Three Brood Ceriodapi Arch. Environ. Contam. Toxicol.21(1): 35-40	The Response of	45168 Cowgill, U.M., and D.P. Milazzo	45168 (	207717
199	of the Three Brood Ceriodap Arch. Environ. Contam. Toxicol.21(1): 35-40	The Response of	Cowgill,U.M., and D.P. Milazzo	45168 (	207716
199	f a Salinity/Toxicity Relations Final Rep. GRI-92/0301. ENSR Contract No. 5		Mount,D.R., and D.D. Gullev	14761	741288
199	Statistical Models to Predict the Toxicity of Environ. Toxicol. Chem.16(10): 2009-2019		Mount,D.R., D.D. Gulley, J.R. Hockett,	18272	160320
199	Mount D.R., D.D. Gulley J.R. Hockett and Control of the Control of Fourier Toxical Chem 16(10): 2009-2019  Mount D.R., D.D. Gulley J.R. Hockett and Control of the Control of Fourier Toxical Chem 16(10): 2009-2019  Mount D.R., D.D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D.D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D.D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D.D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Control of the Control of Toxical Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D. Gulley J.R. Hockett and Chem 16(10): 2009-2019  Mount D.R., D.R.	Statistical Mode	Mount D.R.: D.D. Gulley, J.R. Hockett	18272	160319
100	Decreases, Univ., 200 rep., 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 631-660  Decreases, Univ., 210. Cooling, 611. Maj Variability in the Deformance of the 7 D. Cention of Taxicol Chem 11(10): 611-660  Decreases, Univ., 210. Cooling, 611-660  Decreases, Coolin	Variability in the	DeGraeve, G. IVI., J.D. Cooney, B.H. Mai	11157	120823
100	Decrease, CM 1D Cooper, Data Walleshills, in the retrollmente of the 2-D CE Emitted 1. This LC DET 200	Variability in the	Defração CM ID Coppos BII Mai	11157	120020
100	a Parformance of the 7-D Calenviron Toxicol Cham 11(6): 851-866	Variability in the	DeGraeve G M ID Coopey B H Mai	11152	120828
199	115.5 DeGraver, S. M., J. D. Conown, J. Common Variability in the Deformance of the 7.0 Celement. Toxical Chem 1161; 851.866	Variability in the	DeGraeve G M ID Coopey B H Mai	11152	120839
199	to Performance of the 7-D Celenviron Toxicol Chem 11(6): 851-866	Variability in the	11152 DeGraeve G.M. I.D. Cooney, B.H. MadVariability in the Performance of the 7-D Ce	11152	120824
199	e Performance of the 7-D Ce Environ Toxicol Chem 11/6): 851-866	Variability in the	DeGraeve G.M. I.D. Cooney B.H. Ma	11152	120841
100	1112] Dectraces, Mr. Jud. Cooley, B. H. Mally anadming in the rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. H. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. M. Mally anadming in the Rendmander of the 7-D Celeminal. Tokical. Chem. 114(5): 531-566 1112] Dectraces, G.M. In Cooley, B. M. Mally anadming in the Rendmander of the 7-D Celeminal of the 7-D C	Variability in the	DeGraeve G. M. I.D. Cooney, B.H. Ma	11152	120837
100	Variability in the Performance of the 7-D Ce Environ Toxicol Chem 11(6): 851-866	Variability in the	11152 DeGraeve, G. M. I.D. Cooney, B.H. Mar	11152	120837
100	Postormance of the 7-D Celenviron, Toxicol, Chem 11(6), 851-866	Variability in the	DeGracy & M. I.D. Cooney, B.H. Ma	11152	120925
100	11152 DeGraeve G. M. D. Connex R. H. Marl Variability in the Deformance of the Z. T. De Environ. Toxicol. Chem. 11(6): 951-952.	Wariability in the	DeGraeve G M I D Coopey B H Ma	11152	120829
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1992	111.52 Decideve, S.y.m., J.D.: Cooniey, B.F.: Mail variability in the Performance of the 7-D Celebriton. Toxicoi. Chem.1.1(b): 851-866	Wariability in the	DeGraeve G.M. I.D. Cooney, B.H. Ma	11152	120840
195	Declines (2, 10), 3-12, county, but it will president to the 2-12 cell resident, 10, 10, 10, 10, 10, 10, 10, 10, 10, 10	Variability in the	Decline CM ID Cooner, Bill Mo	1115	170071
1992	to Performance of the 7-D Ce Environ Toxicol Chem 11/6: 851-866	r Variability in the	DeGraeve G.M. I.D. Cooney, B.H. Ma	11152	120826
100	1153 DeGraevie G M I D Connex R H MalVariability in the Deformance and order in Toxicol Chem 11/6: 851-866	Nariability in the	DeGraeve GM ID Coppey R H Ma	11152	120843
2011	Environ Toxical Chem 30/4): 930-938	Influence of Wa	158924 Soucek D.L. T.K. Linton, C.D. Tarr, A. D.	158924	2075418
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	Project No.20-22235A, Report Prepared for	Chloride Toxicity Test Results		112901	2060190
	Project No.20-22235A, Report Prepared for	Chloride Toxicity Test Results	ENVIRON International Corp.	112901	2060191
	Project No.20-22235A, Report Prepared for	Chloride Toxicity Test Results	ENVIRON International Corp.	112901	2060198
	Project No.20-22235A, Report Prepared for	Chloride Toxicity Test Results	ENVIRON International Corp.	112901	2060161
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	Project No.20-22235A, Report Prepared for	Chloride Toxicity Test Results	ENVIRON International Corp.	112901	2060148
	Mosq. News33(1): 78-83	Effect of Various Sodium Chloride Concentr	Lee,F.C.	111908	2152592
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2011	Aquaculture319(1/2): 307-310	Magondu, E.W., J. Rasowo, E. Oyoo-Ok Evaluation of Sodium Chloride (NaCl) for Po Aquaculture319(1/2): 307-310	Magondu,E.W., J. Rasowo, E. Oyoo-Ol	163945	2162290
2011	o Aquaculture319(1/2): 307-310	Magondu, E.W., J. Rasowo, E. Oyoo-Ok Evaluation of Sodium Chloride (NaCl) for Po	Magondu,E.W., J. Rasowo, E. Oyoo-Ok	163945	2162289
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2009	Project No.20-22235A, Report Prepared for	Chloride Toxicity Test Results	112901 ENVIRON International Corp.	112901	2060212
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1976	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	Standard Test Fish Development. Part I. Fat EPA-600/3-76-061A, U.S.EPA, Duluth,	Adelman,I.R., and L.L.,Jr. Smith	2145	196749
1976	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	Standard Test Fish Development. Part I. Fat EPA-600/3-76-061A, U.S.EPA, Duluth,	Adelman,I.R., and L.L.,Jr. Smith	2145	196757
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1976	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	Standard Test Fish Development. Part I. Fat	Adelman,I.R., and L.L.,Jr. Smith	2145	196761
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1976	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	Standard Test Fish Development. Part I. Fat EPA-600/3-76-061A, U.S.EPA, Duluth,	Adelman,I.R., and L.L.,Jr. Smith	2145	196765
1976	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	Standard Test Fish Development. Part I. Fat EPA-600/3-76-061A, U.S.EPA, Duluth,	Adelman,I.R., and L.L.,Jr. Smith	2145	196769
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1985	University of Kentucky, Lexington, KY:73 p.	Recommendations on Numerical Values for	6 Birge,W.J., J.A. Black, A.G. Westerman	45826	201537
1997	Environ. Toxicol. Chem.16(10): 2009-2019	Statistical Models to Predict the Toxicity of	Mount,D.R., D.D. Gulley, J.R. Hockett,	18272	160323
1976	J. Fish. Res. Board Can.33(2): 203-208	SAcute Toxicity of Sodium Chloride, Pentach J. Fish. Res. Board Can.33(2): 203-208	5230 Adelman, I.R., L.L., Jr. Smith, and G.D. S	5230	167861
1997	Environ. Toxicol. Chem.16(10): 2009-2019		Mount,D.R., D.D. Gulley, J.R. Hockett,	18272	160324
1997	Environ. Toxicol. Chem.16(10): 2009-2019		Mount,D.R., D.D. Gulley, J.R. Hockett,	18272	160325
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197	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	Standard Test Fish Development. Part I. Fat EPA-600/3-76-061A, U.S.EPA, Duluth,	2145 Adelman,I.R., and L.L.,Jr. Smith	2145	196762
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1989	Ph.D.Thesis, University of North Texas, Deni	Response of Freshwater and Saltwater Toxid Ph.D. Thesis, University of North Texas,	19549 Price,E.E.	19549	742070
1989	Ph.D.Thesis, University of North Texas, Deni	Response of Freshwater and Saltwater Toxi Ph.D.Thesis, University of North Texas	19549 Price, E.E.	19549	742057
1989	Ph.D.Thesis, University of North Texas, Den	Response of Freshwater and Saltwater Toxi Ph.D.Thesis, University of North Texas,	Price,E.E.	19549	742029
1989	orth Texas	Response of Freshwater and Saltwater Toxi Ph.D.Thesis, University of North Texas,	19549 Price,E.E.	19549	742059
1996	Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and Prog. Fish-Cult. 58(2): 77-84	Waller,D.L., S.W. Fisher, and H. Dabro	76054	740696
1996	Prog. Fish-Cult.58(2): 77-84	Waller, D.L., S.W. Fisher, and H. Dabro Prevention of Zebra Mussel Infestation and	Waller, D.L., S.W. Fisher, and H. Dabrov	76054	740694
1992			Mount,D.R., and D.D. Gulley	14761	768558
1985	ngton, KY:	Recommendations on Numerical Values for University of Kentucky, Lexington, KY:	Birge,W.J., J.A. Black, A.G. Westerman	45826	201538
1996	Prog. Fish-Cult.58(2): 77-84		Waller, D.L., S.W. Fisher, and H. Dabrov	76054	740697
2005	Arch. Environ. Contam. Toxicol.49(4): 511-5	Effects of Pulsed Contaminant Exposures or Arch. Environ. Contam. Toxicol.49(4):	Diamond,J., M. Bowersox, H. Latimer,	102785	742666
2005	Arch. Environ. Contam. Toxicol.49(4): 511-5	Diamond, J., M. Bowersox, H. Latimer, Effects of Pulsed Contaminant Exposures or Arch. Environ. Contam. Toxicol.49(4): 511-5	Diamond,J., M. Bowersox, H. Latimer,	102785	741400
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders, D.F.	116830	768118
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	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders, D.F.	116830	768083
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1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders,D.F.	116830	768114
	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco		116830	768081
	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	768075
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	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	768164
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1993	Site Water Bioassays for Huff and Huff, IncolPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	768089
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1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders.D.F.	116830	768144
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	768128
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders,D.F.	116830	768097
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders, D.F.	116830	768059
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco		116830	768067
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1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff. Inco IPS Environmental and Analytical Services.	Sanders D.F.	116830	768073
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2005	Arch. Environ. Contam. Toxicol.49(4): 511-5	Effects of Pulsed Contaminant Exposures on Arch. Environ. Contam. Toxicol.49(4):	102785 Diamond, J., M. Bowersox, H. Latimer,	102785	741744
1998	Report 2900, EA Engineering, Science	EA Engineering Science and Technolog Results of Acute and Chronic Toxicity Testin Report 2900, EA Engineering, Science,	8 EA Engineering Science and Technolog	165478	2210509
1985	University of Kentucky, Lexington, KY	45826 Birge, W.J., J.A. Black, A.G. Westerman Recommendations on Numerical Values for	6 Birge,W.J., J.A. Black, A.G. Westerman	45826	201561
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders,D.F.	116830	768117
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1997	Statistical Models to Predict the Toxicity of Environ. Toxicol. Chem.16(10): 2009-2019		18272 Mount, D.R., D.D. Gulley, J.R. Hockett,	18272	160287
1997	Environ. Toxicol. Chem.16(10): 2009-2019		18272 Mount, D.R., D.D. Gulley, J.R. Hockett,	18272	160288
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2007	Environ. Toxicol. Chem.26(10): 2086-2093	.IAcute and Chronic Toxicity of Technical-Gra	Bringolf,R.B., W.G. Cope, C.B. Eads, P	100597	803850
2007	Environ. Toxicol. Chem.26(10): 2086-2093	.IAcute and Chronic Toxicity of Technical-Gra	Bringolf,R.B., W.G. Cope, C.B. Eads, P	100597	741583
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2007	Environ. Toxicol. Chem.26(10): 2086-2093	. Acute and Chronic Toxicity of Technical-Gra	Bringolf,R.B., W.G. Cope, C.B. Eads, P	100597	741562
2007	Environ. Toxicol. Chem.26(10): 2086-2093	. Acute and Chronic Toxicity of Technical-Gra	Bringolf,R.B., W.G. Cope, C.B. Eads, P	100597	803853
2011	Environ. Toxicol. Chem.30(1): 239-246	Elphick, J. R. F., K.D. Bergh, and H.C. Bai Chronic Toxicity of Chloride to Freshwater \$\infty\$Environ. Toxicol. Chem. 30(1): 239-246	Elphick,J.R.F., K.D. Bergh, and H.C. Ba	158449	2152833
2011	Environ. Toxicol. Chem.30(4): 930-938	Influence of Water Hardness and Sulfate on	Soucek,D.J., T.K. Linton, C.D. Tarr, A. D	158924	2075477
2011	Environ. Toxicol. Chem.30(4): 930-938	Influence of Water Hardness and Sulfate on	Soucek,D.J., T.K. Linton, C.D. Tarr, A. D	158924	2075476
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the Environ. Pollut.120(2): 219-231	68238 Blasius, B.J., and R.W. Merritt	68238	741742
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the Environ. Pollut.120(2): 219-231	Blasius,B.J., and R.W. Merritt	68238	741743
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the E	Blasius,B.J., and R.W. Merritt	68238	741788
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the Environ. Pollut.120(2): 219-231	68238 Blasius, B.J., and R.W. Merritt	68238	741781
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra Proc. Pa. Acad. Sci.35:51-56	Wurtz,C.B., and C.H. Bridges	2050	231555
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1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra	Wurtz,C.B., and C.H. Bridges	2050	231557
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra Proc. Pa. Acad. Sci.35:51-56	Wurtz,C.B., and C.H. Bridges	2050	231559
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra	Wurtz,C.B., and C.H. Bridges	2050	231561
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2011	Environ. Toxicol. Chem.30(4): 930-938	Soucek,D.J., T.K. Linton, C.D. Tarr, A. Dinfluence of Water Hardness and Sulfate on Environ. Toxicol. Chem.30(4): 930-938	Soucek,D.J., T.K. Linton, C.D. Tarr, A.	158924	2075473
1996	Prog. Fish-Cult.58(2): 77-84	76054 Waller DT S W. Fisher and H. Dahro Prevention of Jehra Mussel Infestration and Prog. Fish-Cult.58(2): 77-84	Waller D.I. S.W. Fisher and H. Dabro	76054	740685
1996	Prog. Fish-Cult 58(2): 77-84	Waller D. S.W. Fisher and H. Dahroj Prevention of Zehra Mussel Infestation and Prog. Fish-Cult 58(2): 77-84	Waller D.I. S.W. Fisher and H. Dahr	76054	740686
2011	J. Appl. Aquacult.23(4): 351-357	Use of lodized Versus Non-lodized Sodium (J. Appl. Aquacult.23(4): 351-357	164438 McDermott A.L. and B.D. Petty	164438	2170552
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the EEnviron. Pollut.120(2): 219-231	Blasius.B.J., and R.W. Merritt	68238	741751
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the Elenviron, Pollut. 120(2): 219-231	Blasius.B.J., and R.W. Merritt	68238	741783
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the EEnviron. Pollut.120(2): 219-231	68238 Blasius B.J., and R.W. Merritt	68238	741756
2002	Environ. Pollut.120(2): 219-231	Field and Laboratory Investigations on the EEnviron. Pollut.120(2): 219-231	Blasius B.L. and B.W. Merritt	68738	742664
2002	Environ. Pollut 120(2): 219-231	Field and Laboratory Investigations on the #Environ. Pollut 120(2): 219-231	68738 Blacing B. L. and B.W. Merritt	8238	741/93
2002	Environ Pollut 120/21: 219-231	ations on the F	Blacing Bland BW/ Marritt	50000	741705
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2002	Project No. 20-22233A, Report Prepared for			112001	2060321
2000	loxicity of selected chemicals to certain Anj. Water Politic Control Feb. 3/19/: 1506-151	loxicity of Selected Chemicals to Certain An	Dowden,B.F., and H.J. Bennett	915	26116
1965	nJ. Water Pollut. Control Fed. 37(9): 1308-131	Toxicity of Selected Chemicals to Certain An		915	26115
1961	1Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebral Proc. Pa. Acad. Sci. 35:51-56	2050 Wurtz,C.B., and C.H. Bridges	2050	231529
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra Proc. Pa. Acad. Sci.35:51-56	Wurtz,C.B., and C.H. Bridges	2050	231530
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra Proc. Pa. Acad. Sci.35:51-56	2050 Wurtz,C.B., and C.H. Bridges	2050	231531
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra Proc. Pa. Acad. Sci.35:51-56	2050 Wurtz,C.B., and C.H. Bridges	2050	231528
1961	Proc. Pa. Acad. Sci.35:51-56	Preliminary Results From Macro-Invertebra Proc. Pa. Acad. Sci.35:51-56	2050 Wurtz,C.B., and C.H. Bridges	2050	231532
1989	Ph.D.Thesis, University of North Texas, Den	Response of Freshwater and Saltwater Toxi Ph.D.Thesis, University of North Texas,	19549 Price, E.E.	19549	742026
1989	Ph.D.Thesis, University of North Texas, Den	Response of Freshwater and Saltwater Toxi Ph.D.Thesis, University of North Texas,	19549 Price,E.E.	19549	742030
1989	Ph.D.Thesis, University of North Texas, Den	Response of Freshwater and Saltwater Toxic Ph.D.Thesis, University of North Texas,	19549 Price,E.E.	19549	742027
1992	Final Rep.GRI-92/0301, ENSR Contract No.5	Development of a Salinity/Toxicity Relation Final Rep. GRI-92/0301, ENSR Contract	14761 Mount,D.R., and D.D. Gulley	14761	768559
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1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, I	Sanders,D.F.	116830	767958
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767884
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders,D.F.	116830	767972
	Arch. Hydrobiol.122(1): 33-56	Demographic Effects of Salinity, Water Hard Arch. Hydrobiol.122(1): 33-56		119414	2152727
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	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	Aragao,M.A., and E.V. Pereira	71919	741945
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differen	Aragao, M.A., and E.V. Pereira	71919	742565
	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	-	71919	742563
	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer		71919	741949
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	Aragao,M.A., and E.V. Pereira	71919	741957
	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	Aragao,M.A., and E.V. Pereira	71919	742892
	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer		71919	741944
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2003	Sensitivity of Ceriodaphnia dubia of Differer Bull. Environ. Contam. Toxicol. 70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer		71919	741952
	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer Bull. Environ. Contam. Toxicol.70(6): 1247-		71919	741938
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differed Bull. Environ. Contam. Toxicol.70(6): 1247-1	71919 Aragao, M.A., and E.V. Pereira	71919	741947
1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve, G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem. 11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251012
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1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve, G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem. 11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251016
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1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve, G.M., J.D. Cooney, B.H. Marl Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem. 11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251014
1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve, G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Celenviron. Toxicol. Chem.11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251024
1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve, G.M., J.D. Cooney, B.H. Marl Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem.11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251022
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders D.F.	116830	767973
1993	IPS Fnyironmental and Analytical Services. A	Site Water Bioassays for Huff and Huff Inco IPS Environmental and Analytical Services.	116830 Sanders D.F.	116830	767965
1993	IDS Environmental and Analytical Services A	Site Water Biogsays for Huff and Huff Incollos Environmental and Analytical Services	116830 Sandar D.E.	116030	767900
1993	IDS Environmental and Analytical Services A	Site Water Bioassays for Huff and Huff Inco	116830 Sandors D.E.	116030	767063
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1993	IPS Environmental and Analytical Services. A	Site Water Bioassays for Huff and Huff Inco	Sanders D.F.	116830	767891
1993	IPS Environmental and Analytical Services. A	Site Water Bioassays for Huff and Huff Inco	Sanders D.F.	116830	766870
1993	IPS Environmental and Analytical Services. A	Site Water Bioassays for Huff and Huff Inco	116830 Sanders D.F.	116830	767866
1993	IPS Environmental and Analytical Services. A	Site Water Bioassays for Huff and Huff Incol IPS Environmental and Analytical Services.	Sanders D.F.	116830	766867
1993	IDS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff Inco	Sanders, U.F.	116830	767850
1993	IDS Environmental and Analytical Services	Cite Water Dioassays for Huff and Huff Inco	Salluels, p.r.	116830	707057
1993	IPS Environmental and Analytical Services. A			116930	75757
2003	Arch Fnyiron Contam. Toxicol.45(1): 79-85		Harmon S M	71674	747568
1993	Site Water Bioassays for Huff and Huff Incol IPS Environmental and Analytical Services. A	Site Water Bioassays for Hiff and Hiff Inco	Sanders D.E.	116830	767050
1993	IDS Environmental and Analytical Services A	Site Water Bioassays for Huff and Huff Inco	Sanders, D. F.	116830	767876
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Incolp's Environmental and Analytical Services	Sanders, D.F.	116830	767874
1993	AIPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D. F.	116830	766868
1993	VIPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	767868
1993	JIPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders, D.F.	116830	767860
1993	JIPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders, D.F.	116830	767901
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	766860
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	767957
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	Sanders, D.F.	116830	767883
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767971
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2000	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer Bull. Environ. Contam. Toxicol.70(6): 1247-	/1919 Aragao, M.A., and E.V. Pereira	/1919	/41946
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differed Bull. Environ. Contam. Toxicol.70(6): 1247-1	71919 Aragao, M.A., and E.V. Pereira	71919	741649
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	71919 Aragao, M.A., and E.V. Pereira	71919	741648
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differen	Aragao,M.A., and E.V. Pereira	71919	742901
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	71919 Aragao, M.A., and E.V. Pereira	71919	742893
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	71919 Aragao, M.A., and E.V. Pereira	71919	742891
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer Bull. Environ. Contam. Toxicol.70(6): 1247-1	Aragao,M.A., and E.V. Pereira	71919	741937
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	71919 Aragao,M.A., and E.V. Pereira	71919	742898
2005	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer	Aragao, M.A., and E.V. Pereira	71919	741953
2003	Bull. Environ. Contam. Toxicol.70(6): 1247-1	Sensitivity of Ceriodaphnia dubia of Differer Bull. Environ. Contam. Toxicol.70(6): 1247-1	71919 Aragao, M.A., and E.V. Pereira	71919	742896
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con	Stark,J.	116754 Stark,J.	761581
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Stark,J.	116754	761593
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Stark,J.	116754 Stark,J.	761573
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Stark,J.	116754 Stark,J.	761589
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Stark,J.	116754	761577
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Stark,J.	116754	761585
1997	Environ. Toxicol. Chem.11(6): 851-866	J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem.11(6): 851-866	DeGraeve,G.M.,	11152	251021
1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251011
1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve, G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce Environ. Toxicol. Chem. 11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251015
1992	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve,G.M., J.D. Cooney, B.H. Mar Variability in the Performance of the 7-D Ce	DeGraeve,G.M., J.D. Cooney, B.H. Ma	11152	251013
1992	Environ, Toxicol, Chem.11(6): 851-866	DeGraeve.G.M J.D. Cooney. B.H. Mar Variability in the Performance of the 7-D Ce	DeGraeve.G.M J.D. Coonev. B.H. Ma	11152	120856
1992	Environ, Toxicol, Chem.11(6): 851-866	DeGraeve G.M. J.D. Cooney B.H. Marl Variability in the Performance of the 7-D Celenviron Toxicol Chem. 11(6): 881-866	DeGraeve.G.M J.D. Cooney. B.H. Ma	11152	251017
1993	Environ. Toxicol. Chem.11(6): 851-866	DeGraeve G.M., J.D. Cooney, B.H. MarVariability in the Performance of the 7-D Ce Environ, Toxicol, Chem.11(6): 851-866	DeGraeve.G.M., J.D. Cooney, B.H. Ma		251023
1997	Environ Toxicol Chem 11/6): 851-866	DeGraeve G. M. I.D. Coopey, B.H. Marlyariability in the Performance of the 7-D Ce Environ. Toxicol. Chem. 11(6): 851-866		11152	251019
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff Inco IDS Environmental and Analytical Services,	Sanders D.E.	116830	767974
1993	NIPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,		116830	767066
199	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767964
199:	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767886
199:	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	116830 Sanders, D.F.	116830	767892
199:	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	766871
199	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	767867
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	766863
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders,D.F.	116830	767851
199	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767956
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	Sanders, D.F.	116830	767853
1999	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con-	Stark,J.	116754	761590
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con-	116754 Stark,J.	116754	761574
199	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con	116754 Stark,J.	116754	761586
1999	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con-	116754 Stark,J.	116754	761578
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1993	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767859
1993	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services,	116830 Sanders, D.F.	116830	767877
1993	Site Water Bioassays for Huff and Huff, Inco $ $ IPS Environmental and Analytical Services, $ \!\! $	Site Water Bioassays for Huff and Huff, Inco	Sanders,D.F.	116830	767894
199	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767875
1993	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	766869
1993	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, $A$	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders,D.F.	116830	767869
1993	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco	116830 Sanders, D.F.	116830	767861
1999	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con	Stark,J.	116754	761582
Publication Year	Source P	Title	Author	Number	Result Number
				Reference	

1004	Hydrohiologia100:25-31	The Influence of Mineral Salts on Feetingity Hydrobiologia108:25-31	10/318 Leblanc G A and D.C. Surprenant	107318	759463
1984	TENANCE OF A TORREST OF A TORRE	The intluence of Mineral Salts on Escundity			
1979	Res.Rep.No.BN-/9-10-546, Submitted to Sm	The Chronic Toxicity of Sulfate to the Water Res. Rep. No. BN-79-10-546, Submitted	115903 E.G. and G. Bionomics	115903	/59022
1984	Hydrobiologia108:25-31	The Influence of Mineral Salts on Fecundity	Surprenant	107318	759464
2011	Environ. Toxicol. Chem.30(1): 247-253	(An Aquatic Toxicological Evaluation of Sulfa Environ. Toxicol. Chem.30(1): 247-253	0	153684	2017150
2011	Environ. Toxicol. Chem.30(1): 247-253	(An Aquatic Toxicological Evaluation of Sulfa Environ. Toxicol. Chem.30(1): 247-253	153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017151
2011	Environ. Toxicol. Chem.30(1): 247-253	1 -		153684	2017149
201:	Environ. Toxicol. Chem.30(1): 247-253	An Aquatic Toxicological Evaluation of Sulfa		153684	2017148
201	Environ. Toxicol. Chem.30(1): 247-253	An Aquatic Toxicological Evaluation of Sulfa		153684	2017147
1988	Prepared for Presentation at American Insti	Toxicity of Common Salts to Three Biotoxici Prepared for Presentation at American Insti	Taylor,P.A., A.J. Stewart, and L. Holt	116816	761751
198	Prepared for Presentation at American Insti	Toxicity of Common Salts to Three Biotoxici	Taylor,P.A., A.J. Stewart, and L. Holt	116816	761750
1999	ln: W.A.Price, B.Hurt and C.Howell (Eds.), Pr		Pickard,J., P. McKee, and J. Stroiazzo	116829	759075
2011	Environ. Toxicol. Chem.30(1): 247-253	$\sim 1$	Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017061
201	Environ. Toxicol. Chem.30(1): 247-253	$\sim$	153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017052
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325	Soucek,D.J.	114662	766233
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325	Soucek,D.J.	114662	766231
199	IPS Environmental and Analytical Services, A	Site Water Bioassays for Huff and Huff, Inco IPS Environmental and Analytical Servi	Sanders, D.F.	116830	766855
200	~ 1	Effects of Water Quality on Acute and Chrod Third Quarterly Report, Submitted to	Illinois National History Survey	116831	766738
200	Third Quarterly Report, Submitted to U.S.EF		Illinois National History Survey	116831	766735
201	Environ. Toxicol. Chem.30(1): 247-253		153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017044
201	Environ. Toxicol. Chem.30(1): 247-253	$\sim$	153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017033
199	In: W.A.Price, B.Hurt and C.Howell (Eds.), Pr		Pickard, J., P. McKee, and J. Stroiazzo	116829	759074
201	Environ. Toxicol. Chem.30(1): 247-253		153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017060
201	Environ. Toxicol. Chem.30(1): 247-253	$\overline{}$		153684	2017051
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325		114662	766232
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325		114662	766356
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325		114662	766351
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325		114662	766230
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on the Ecotoxicology 16(3): 317-325		114662	766355
200	Ecotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on the Ecotoxicology 16(3): 317-325		114662	766344
200		Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology 16(3): 317-325		114662	766354
200		Effects of Water Quality on Acute and Chrod Third Quarterly Report Submitted to	1 Illinois National History Survey	116831	766737
200	Third Quarterly Report Submitted to U.S.ER	- 1	116831 Illinois National History Survey	116831	766734
203	Environ. Toxicol. Chem.30(1): 247-253	$\sim$	153684 Elphick J.R., M. Davies, G. Gilron, E.C.	153684	2017043
20:	Environ. Toxicol. Chem.30(1): 247-253		153684 Elphick.J.R., M. Davies, G. Gilron, E.C.	153684	2017032
190	Site Specific Multi-Species Toxicity Testing aln: W.A. Price, B. Hurt and C. Howell (Eds.). Pr	Site Specific Multi-Species Toxicity Testing of		116829	759077
201	Fnviron. Toxicol. Chem.29(2): 347-358	Observed and Predicted Reproduction of Celenviron, Toxicol, Chem. 29(2): 347-358		158925	2148467
207	Environ Toxicol Chem 29(2): 347-358	Observed and Predicted Reproduction of Celenviron Toxicol. Chem 29(2): 347-358	Clasier P Land I R Hardin	158925	2148469
)0C	Site Specific Multi-Species Toxicity Testing qin: w.A.Price, B.Hurt dila C.Howell (Eds.), Pi	Observed and Bradist of Boardwest on of Co		158275	71/08/75
1999	SF Analytical Laboratories Inc., Milwaukee,	Bioassay Report Chronic Toxicity Tests. Con SF Analytical Laboratories Inc., Milwau	4 Stark,J.	116754	761597
201	Environ. Toxicol. Chem.29(2): 347-358	Observed and Predicted Reproduction of Ce Environ. Toxicol. Chem.29(2): 347-358		158925	2148470
2010	Environ. Toxicol. Chem.29(2): 347-358	Observed and Predicted Reproduction of Ce Environ. Toxicol. Chem.29(2): 347-358	158925 Lasier,P.J., and I.R. Hardin	158925	2148476
201	Environ. Toxicol. Chem.29(2): 347-358	Observed and Predicted Reproduction of Ce Environ. Toxicol. Chem.29(2): 347-358	5 Lasier,P.J., and I.R. Hardin	158925	2148468
2011	Environ. Toxicol. Chem.30(1): 247-253	1_	4 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017045
201	Environ. Toxicol. Chem.30(1): 247-253	حا	153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017062
20:	Environ. Toxicol. Chem.30(1): 247-253	I ~	Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017053
2007	HEcotoxicology16(3): 317-325	Bioenergetic Effects of Sodium Sulfate on th Ecotoxicology16(3): 317-325	2 Soucek,D.J.	114662	766234
2007	HEcotoxicology16(3): 317-325		Soucek,D.J.	114662	766229
201	Environ. Toxicol. Chem.30(1): 247-253		Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017034
2011	Environ. Toxicol. Chem.30(1): 247-253	. ∮An Aquatic Toxicological Evaluation of Sulfa Environ. Toxicol. Chem.30(1): 247-253	153684 Elphick,J.R., M. Davies, G. Gilron, E.C.	153684	2017063
Publication Year	Source Pu	Title	Author	Number	Result Number
				Doforono	

Filter Criteria Used to Filter Raw Data from ECOTOX

- - - 2 Rename tab as "as recd from ECOTOX"

Removed species that are not found in the United States Removed additional studies that included confounding factors

> Add reagent grade chemicals to deionized water. Approximate equilibrium pH after 24 h of acration. Taken in part from Marking and Dawson (1973).

> > 0.0FZ

120.0

30.0

Rengent Added (mg/L)2

Caso, 2H, O Mgso,

values

values

senjey

sanje

senjev

Sort On

•

X Delete Level

15 Filled in hardness values as applicable (see table below)

After separating out into sublethal and lethal:

0.488

192.0

0.06

0.84

15.0

NaHCO

Water Type (Hardness) per EPA 2002:

Endpoint

Effect (HLF)

Lethal or Sublethal

Expressed as mg CaCO<sub>3</sub>/L.

Very hard

PleH

Hos

Lype

Water

Lyeu pl

Lyeu ph

Then by

Yd hos

Column evel ≜t Add Level

breH Moderately

Very soft

- - I Performed search using criteria in "Tolerance Value Search Criteria"
- 7 Under "Observed Duration Mean" column, filtered by "NR." Then filtered "Observed Duration Min" for values less than 1 and "NR." Deleted associated rows. 6 Under "Observed Duration Mean" column, filtered by all values less than 1 and deleted all associated rows
- 8 Filtered "Conc 2 Mean" and deselected "(Blanks)" and deleted all associated rows (to remove any study results that measured more than one chemical).

13 Filtered "Chemical Name" column and deselected all chemicals not containing potassium; deleted associated rows. (Potassium associated with high toxicity.)

552-542

110-150

t9-LS

SE-0E

Alka-

Cancel

-

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\_

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Wy data has headers

160-180

80-100

84-04

10-13

Hardness (mg/L CaCO3)

Hard

1102

Very Soft

Moderately Hard

8.0-8.4 280-320

0.8-9.7

8.7-4.7

9.7-2.7

Hd

081-091

001-08

84-04

Hardness

Approximate Final Water Quality

OK

Z of A

ZOIY

ZOIA

Order

• Z OJ Y

\_ ZOIA

0.01

0.4

2.0

50

KCI

TABLE 3. PREPARATION OF SYNTHETIC FRESHWATER USING REAGENT GRADE CHEMICALS!

240.0

130.0

0.08

- 9 Deleted columns related to Conc 2 and Conc 3 (now completely blank after filtering out those with values).

14 Removed entries that contained confounding factors in the "Experimental Design" column (e.g., use of K-medium, parasite infection).

11 Filtered "Conc 1 Type" and deselected "Total"; deleted associated rows ("Dissolved" and "Formulation"). 10 Filtered "Conc 1 Units" to select all but ug/L and removed associated rows to another worksheet labeled "diff units"

12 Filtered "Species Group" column and deselected all but Amphibians and related groups; deleted associated rows.

- - - 5 Deleted species taxonomic information columns (columns G through P in "as recd" worksheet) 4 Under "Species Family" column, filtered by "Salmonidae" and deleted all associated rows
      - 2 Made copy of tab for filtering and sorting and named "sorted values"

Riomacc	L	100000	ZZ	7 NR	Not reported		Fresh water Lab	Unmeasured Fres	Renewal	7647145 Sodium chloride (NaCl)	761560
Biomass		Day(s)	NK	4NK	Whole organism	PARTIAL RENEWAL AT 48 HOUR	h water Lab	Unmeasured Fres		Chica	LEGAL VALLE
Biomass	NOEC	Day(s)	NR.	7 NR	Not reported		h water Lab	7		Sodium chlor	761567
Biomass		Day(s)	NR	7 NR	Not reported		resh water Lab	77		7647145 Sodium chloride (NaCl)	761548
Biomass		Day(s)	NR :	7 NR	Not reported		resh water Lab		Renewal	2017	761549
Weight		Day(s)	2 2 2	4 NR	Whole organism	2-A REPLICATES	Fresh water Lab	Inmeasured Fres	Renewal	7547145 Sodium chloride (NaCl)	163091
Weight		Day(s)	N.R	56 NR	Whole organism		resh water Lab	Measured Fres	+	L'u	202202
Biomass		Day(s)	NR	54 NR	Whole organism		Fresh water Lab			7647145 Sodium chloride (NaCl)	152750
Biomass		Day(s)	NR	54 NR	Whole organism		h water Lab			Sodium	152747
Biomass	_	Dav(s)	200	54 NR	Whole organism		resh water Lab		Renewal	37.1.3	157748
Biomass	NOEC	Davis	28	54 NR	Whole organism		Fresh water Lab	Measured Free	Renewal	7647145 Sodium chloride (Naci)	157746
Weight		Until nato	NK	NR.	Whole organism NK		resh water Lab			7647145 Sodium chloride (NaCl)	251464
Weight		Day(s)	NR.	56 NR	Whole organism		h water Lab		Flow-through	7647145 Sodium chloride (NaCl)	251447
Weight	NOEC	Day(s)	NR.	90 NR	Whole organism	2 REPLICATES	Fresh water Lab	Measured Free	ough	7647145 Sodium chloride (NaCl)	251448
Weight		Day(s)	NR	14 NR	Whole organism		h water Lab		Renewal	300	741685
Weight		Day(s)	NR	56 NR	Whole organism		Fresh water Lab		Flow-through	7647145 Sodium chloride (NaCl)	202204
Weight	NOEL	Dav(s)	NR.	4 NR	Whole organism	A SEMI-STATIC SYSTEM WITH 50% OF DAILY	resh water Lab	Unmeasured Fres	Renewal	Sol	741673
Weight		Day(s)	N N	4 NR	Whole organism	DE DAIL	resh water Lab	77		7647145 Sodium chloride (NaCi)	743277
Leight	1	Day(s)	200	TOOLNA	Whole organism	CACLEVY WILL EVOY OF DAIL	n Water Lab	1		/64/145 Sodium chloride (NaCi)	165524
Weight	lă	Until met	N N	N. N.	Whole organism NR	10 REPLICATES, CHRONIC TEST	Fresh water Lab	Unmeasured Free	Renewal	7647145 Sodium chloride (NaCl)	154068
Length	NOEC	Day(s)	NR.	100 NR	Whole organism		resh water Lab			7647145 Sodium chloride (NaCl)	165525
Length		Day(s)	NR		Whole organism	D IN THE LAB, LARVAE MO	esh water Lab	27		Sodium chlor	060023
Length		Day(s)	NR	300 NR	Whole organism		Fresh water Lab	Unmeasured Fres	Renewal	7647145 Sodium chloride (NaCl)	2165533
Length	LOEC	Day(s)	NR	300 NR	Whole organism		h water Lab			2	165532
Weight gain	st-hatdLOEC	Day(s) po:	NR	5 NR	Not reported	EGGS	Fresh water Lab	Unmeasured Free	Flow-through	7647145 Sodium chloride (NaCl)	741651
Weight gain	t-hatdNOEC	Day(s) po:	NR.	5 NR	Not reported	EGGS	h water Lab			7647145 Sodium chloride (NaCl)	741652
Weight	t-hatdNOEC	Day(s) po:	NR	5 NR	Not reported	GGS			ngh	5	741641
Length	NOEC	Day(s)	NR.	15 NR	Whole organism	3 REPLICATES	resh water Lab	Unmeasured Free	Renewal	7647145 Sodium chloride (NaCl)	2162413
Weight	EC25	Day(s)	NR.	28 NR	Whole organism	TES, CONDUCTED IN STAND	resh water Lab	ed	Renewal	7647145 Sodium chloride (NaCl)	2164559
Biomass	NOEC	Dav(s)	NR.	28 NR	Not reported	0.000		Massired Fra		2 4	313030
Riomass	200	Day(s)	NR	SIN SC	Not reported	A REPUCATES	esh water Lab	7 2	nellewal	7647145 Sodium chloride (NaCi)	/19701
DIGITIOSS	FOEC	Dayls)	NN	70 NN	Not reported		resh water Lab	Measured		0	61875
Specific growth	NOEC	Day(s)	N.	28 NR	Not reported		resh water Lab	d	Renewal	7647145 Sodium chloride (NaCl)	73638
Weight gain	NOEC	Day(s)	NR	28 NR	Not reported	SREPLICATES	h water Lab			7647145 Sodium chloride (NaCl)	73639
Length	NOEC	Day(s)	N.R	7 NR	Whole organism	TEST WATER CHEMISTRY SHOWS NITRITE 5.	Fresh water Lab		Static	7647145 Sodium chloride (NaCl)	748411
Length	MATC	Day(s)	NR	21 NR	Not reported		resh water Lab	Measured Free		7647145 Sodium chloride (NaCl)	01557
Length	NOEC	Day(s)	NR.	21 NR	Not reported		sh water Lab	Measured Fre		7647145 Sodium chloride (NaCi)	201555
Size	NOEC	Day(s) N	NR	21 NR	Whole organism	10 REPLICATES, SALINITY ACCLIMATION, CO	sh water Lab	b	Renewal	7647145 Sodium chloride (NaCl)	60499
Size	LOEC	Day(s)	NR	21 NR	Whole organism	10 REPLICATES, SALINITY ACCLIMATION, CO	resh water Lab	TD		7647145 Sodium chloride (NaCl)	60501
Length	LOEC	Day(s)	NR	21 NR	Not reported		resh water Lab	Measured Fre	Renewal	7647145 Sodium chloride (NaCl)	01556
Biomass	NOEC	Day(s)	NR.	20 NR	Not reported	4 REPLICATES	sh water Lab	T	-	S	52814
Biomass	ICIO.	Day(s)	NR	20 NR	Not reported	4 REPLICATES		m		7647145 Sodium chloride (NaCl)	2152811
Biomass	LOEC	Day(s)	NR	20 NR	Not reported		resh water Lab			7647145 Sodium chloride (NaCl)	52815
Biomass	IC50	Day(s)	NR	20 NR	Not reported		Fresh water Lab	Measured Fre	Renewal	7647145 Sodium chloride (NaCl)	2152813
Biomass	1025	Day(s)	NR	20 NR	Not reported		sh water Lab	V		7647145 Sodium chloride (NaCl)	52812
Dry weight (AC	LOEC	Day(s)	N <sub>R</sub>	80 NR	Not reported		resh water Lab		Renewal		759436
Length	LOEC	Day(s)	NR	80 NR	Carapace		Fresh water Lab	Unmeasured Fre	Renewal	7647145 Sodium chloride (NaCl)	759444
Dry weight (AQ	NOEC	Day(s)	NR	80 NR	Not reported		resh water Lab		Renewal		759437
Dry weight (AQUI	NOEC	Day(s)	NR :	80 NR	Not reported		sh water Lab	2 0	Renewal	7647145 Sodium chloride (NaCl)	150/01
Growth rate	NOFC	Davis	Z Z	80 NR	Not reported		sh water Lab	Uninedsured Fre	Renewal	7647145 Sodium chloride (Naci)	STAGG
Length	NOFO	Davis	2 20 2	div 08	Carapace		Fresh Water Lab	eo	Renewal	764/145 Sodium chloride (NaCi)	59438
rengin	NOEC	Daylst	NA.	SO NR	Carapace		sn water Lab			7647145 Sodium chloride (NaCl)	759439
Lengin	NOEC	Day(s)	N.	80 NR	Carapace		Fresh water Lab	Unmeasured Fre	Renewal	7647145 Sodium chloride (NaCl)	759445
Weight	amorpi NOEC	Until met	NK	N.K	Whole organism NK	10 REPLICATES, CHRONIC TEST	Fresh water Lab	Unmeasured Fre	Renewal	7647145 Sodium chloride (NaCl)	154065
Weight	amorpitoec	Until met	NR.	N.R	Whole organism NR	10 REPLICATES, CHRONIC TEST	sh water Lab		Renewal	7647145 Sodium chloride (NaCl)	154061
Weight	IC25	Day(s)	NR.	7 NR		GROWTH AND MORTALITY	Fresh water Lab		Renewal	-	718612
Weight	NOEC	Day(s)	NR	7 NR	Whole organism	AUTHOR TESTS GROWTH AND MORTALITY,	sh water Lab	Unmeasured Fre	Renewal	7447407 Potassium chloride (KCI)	718611
Weight	TOEC	Day(s)	NR	7 NR	Whole organism	AUTHOR TESTS GROWTH AND MORTALITY,	sh water Lab	Unmeasured Fre	Renewal	7447407 Potassium chloride (KCI)	729236
Growth, genera	NOEC	Day(s)	NR	7 NR	Not reported		Fresh water Lab	Unmeasured Fre	Renewal	7447407 Potassium chloride (KCI)	43166
Growth, general	NOEC	Day(s)	NR	7 NR	Not reported		sh water Lab			77	96902
Growth, genera	MATC	Day(s)	NR	7 NR	Not reported			-	Renewal		96894
Growth, genera	NOEC	Day(s)	NR	7 NR	Not reported		resh water Lab	ed	Renewal	7447407 Potassium chloride (KCI)	96897
Weight	LOEC	Day(s)	NR	7 NR	Not reported		Fresh water Lab		Renewal		761770
Weight	NOEC	Day(s)	NR	7 NR	Not reported		sh water Lab	ed	Renewal	7447407 Potassium chloride (KCI)	761771
Growth, genera	LOEC	Day(s)	NR	7 NR	Not reported			Unmeasured Fre	Renewal	7447407 Potassium chloride (KCI)	96893
Growth, genera	NOEC	Day(s)	NR	7 NR	Not reported		Fresh water Lab		Renewal	7447407 Potassium chloride (KCI)	96891
Growth, genera	NOEC	Day(s)	NR.	7 NR	Not reported					7447407 Potassium chloride (KCI)	96886
Growth, general	NOEC	Day(s)	NR.	7 NR	Not reported		Fresh water Lab	Unmeasured Fre	Renewal	7447407 Potassium chloride (KCI)	43171
Growth, genera	MATC	Dav(s)	NR :	7 NR	Not reported	ACTOCK TO SECURITY AND MICHIGAN	oll Water Lab			744 7407 Potassium chloride (KCI)	/28144
Weight	DEC	Day(s)	N. S.	7 NR	Whole organism	GROWTH AND MORTALITY	SI Water Lab		ŀ	7 7	128596
Weight	1025	Dav(s)	NR :	7 NR	whole organism	ORTALITY	resh water Lab		Renewal	이 구	/18595
Weight	NOEC	Day(s)	NR	7 NR	Whole organism	ALITHOR TESTS GROWTH AND MORTALITY,	Fresh water Lab	Unmeasured fre		7447407 Porassium chloride (KCI)	70000
Weight	1025	Day(s)	Z <sub>D</sub>	7 NR	Whole organism	AUTHOR TESTS GROWTH AND MORTALITY,	ch water Lab			7447407 FOLGSSIGHT CHIORIGE (KCI)	710507
Weight	NOEC	Day(s)	NR.	4 NR	Whole organism	A DEPLICATES, 13 TESTS	resh water Lab	Office and Fre	Danawai	744/407 POISSIBIL CHICKER (NCI)	241090
Waight	105	Davis)	NR S	4 4	Whole organism		sh water Lab	Unmeasured	Renewal	7447407 Potassium chloride (KCI)	060110
VVeign:	IL)K	Day(s)	UNI	A A A	Whole organism		Fresh water Lab	Unmeasured	Renewal	7447407 Potassium chloride (KCI)	060109
Length	NOEL		NK	12 NK	Not reported	10 REPLICATES	sh water Lab	Unmeasured	Kenewai	12	793901
Length	LOEL	ш	- 1		The state of the s				Inc. word		
Marine II	13001			12 NK		10 REPUCATES	sh water Lab	Unmeasured	Renewal		793900
THE PROPERTY AND INCHES	Con all and	-	NR.	12 NR	11	10 REPUCATES	Fresh water Lab	Unmeasured	Renewal Renewal	7447407 Potassium chloride (KCI)	93900

768061	768100	768121	768122	768132	768131	768102	768103	2163908	2163912	2163911	761765	761764	761559	741814	742694	741815	741780	741831	768110	768109	768120	768119	769069	768069	768071	2152755	2152754	2152752	2152753	768139	768140	768076	768077	768085	768084	768159	768160	768078	768079	768150	768149	768095	768094	760667	191997	768162	768141	768142	768112	768111	768093	768092	768130	768170	769157	768151	96882	42867	14380	42862	14424	761570	761554	761571	761567	155197	761557	761565	761561	25156	761566	761556	755197	76155	761565	761558	78478	382762	761564	RESUIT NUMBER
7647145 So	7547145 50	7647145 So	7647145 50	7647145 So	7647145 So	7647145 50	7647145 So		7647145 Sc		7647145 50	7647145	7647145 56	7647145 Sc	7647145 Sc	7647145 50	S	7647145 50	7647145 50	7647145 50	7647145	7647145 50	7677175	7647145 50	7647145 50	7647145 Sc	10		7647145 50	7647145 50	7647145 50	7647145 50	7647145 50	7647145 50	7647145 50		7647145 50	7647145 50	7647145 5		7647145 5		7647145	7647145	7647145	7647145	7647145	7647145		7647145 S		7647145	7647145	7647145	7647145	7647145 5	7647145 5	7647145	7647145	7647145	7647145 S	7647145 S	7647145 5	7647145	76/71/45	7647145	7647145		7647145	T	7647145	7647145	76/71/45	7647145 5		764714	764714	764714	764714	CAS Numbe
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Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Survival	Survival	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	LOEC Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Survival	Mortality	Mortality	Mortality	Mortality	Mortality

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Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh Water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	FIGSH WOLES LOD	Fresh water Lab	_		Fresh water Lab		Fresh water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab			Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh Water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab			Fresh water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab	-	Fresh water Lab			Fresh water Lab	Greek water Lab	Fresh water Lab	Fresh water Lab	Fresh Water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Marit San and
							4 REPUCATES	S REPI ICATES										INFECTED BY PSEUDODACTILOGIRUS SP	Laurent Marchine	DARASITICALLY INSECTED	A BERLICATES	S BEBLICATES BOILED BOND WATER	4 REPLICATES BOILED BOND WATER		ACUTE STUDY	TRIALA	TRIALB																						ACUTE STUDY	TRIALA	TRIALB	3 REPUICATES				FED, MODERATELY HARD RECONSTITUTED								IWO-FOOR REPUCALES, SWIGHT CARACIC	TWO-FOUR REPLICATES, 9 MIGH CA RATIO		THREE REPLICATES											
Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	(Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	The sale of the last of the la
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NR	NR.	NR.	NR	NR	NR	NR	NR S	NR 3	NR NR	NR	Z.	Z.	NN	NA NA	N.	S.	NA NA	NO NA	NID NI	NR NR	NR S	NO	NA ON	NR.	NR	NR	NR	NR.	NR	NR.	NR	NR	NR	NR	NR	NR.	Z Z	NB NB	NR R	NR	NR.	NR	NR	NR	N.	NR II	2 2	E N	NR.	NR	NR	N Z	N X	2 2	Z Z	NR	NR.	NR.	NR	NR :	Z :	NR II	NR NA	No	NE	NA NA	NR.	NR	NR .	NR	NR	NR	NR RN	Z Z	Z Z	Z Z	ND I	
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LC50	М			LC50		NOEL	RO	NOEC	1050	LC50			LCSC	Ų.	12		רכטט	20	NIB-7EBO	NR-ZERO	NR-J FTH	NOEC	NOEC	NR-LETH	LC50	NR-ZERO	NR-ZERO	LD50	LD50	NR-LETH	LD50	NR-LETH	LD50	NR-LETH	LD50	NR-LETH	NR-LETH	1050	NR-LETH	NR-LETH	NR-ZERO	LD50	NR-LETH	LD50	NR-LETH Mortality	NR-LETH	NR-I FTH	LUSU LETT	LC50	NR-ZERO	NR-ZERO	ZERO	NR-LETH	LC50	LC50	NR-ZERO						1050			FCOD	1000	LCOS			LCS0		LC50	NOEC	LOEC	NOEC	NOEC	NOFC	SOFC
Mortality	Mortality	Mortality	Viortality	Mortality	Viortality	Time to 100% mortalit	Mortality	Hatch	Mortality	Mortality	Mortality	Mortality	WICHTARLY	Mortality	Mortality	Mortality	MOLIGITA	Mortality	Mortality	Viortality	Mortality	Survival	Hatch	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	Mortality	MAINT FRIED

			of reported	2	Grach water lah	Unmeasured	Renewal		
		5 NR	Not reported	N	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	196813 7
	2 2	1 NR	ot reported	2 2	Fresh water Lab	ŀ	Renewal	7647145 Sodium chloride (NaCl)	196810 7
		2 NR	Not reported	N	Fresh water Lab		Renewal	47145 Sodium chloride (NaCl)	
		1 NR	ot reported		Fresh water Lab		Renewal	47145 Sodium chloride (NaCl)	À
		4 NR	Not reported	UNHEALTHY	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCi)	196844 7
		1 NR	ot reported	Z		Unmeasured	Renewal	7647145 Sodium chloride (NaCl)	
		4 NR	ot reported	Z	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	
		7 NR	Not reported	2 2	Fresh water Lab	Unmeasured	Renewal	7647145 Sodium chloride (NaCl)	196825 7
		Z ZR	ot reported		Fresh water Lab	Unmeasured	Renewal	4/145 sodium chloride (NaCl)	
		5 NR	ot reported	2	-		Renewal	47145 Sodium chloride (NaCl)	
150		4 NR	ot reported	2	Fresh water Lab		Renewal	47145 Sodium chloride (NaCl)	196804 7
-		1 NR	Not reported	N		Unmeasured	Renewal	7647145 Sodium chloride (NaCl)	
× :		4 NR	ot reported	Z			Renewal	47145 Sodium chloride (NaCl)	196780 7
Y		3 C	Not reported	2 2	Fresh water Lab		Renewal	A7145 Sodium chloride (NaCl)	T
SIVE		4 NR	Not reported	UNHEALIHY			Renewal	4/145 Sodium chloride (NaCl)	198845
S)AE		A NR	ot reported	Z	Fresh water Lab		Renewal	47145 Sodium chloride (NaCl)	П
SIVE		2 NR	Not reported	Z	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	196827 7
34(5)		4 NR	at reported	2	Fresh water Lab		Renewal	47145 Sodium chloride (NaCl)	
(s)AE		1 NR	ot reported	Z	Fresh water Lab		Renewal	47145 Sodium chloride (NaCl)	
ay(s)		2 NR	Not reported	Z	Fresh water Lab		Renewal	7647145 Sodium chloride (NaCl)	196775 7
ay(s)			Not reported NR		Fresh water Lab	Unmeasured	Renewal		
ay(s)		1 NR	Not reported	UNHEALTHY	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	
ay(s)			ot reported		Fresh water Lab	Unmeasured	Renewal		Ť
ay(s)		US NR	Not reported	Z	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	
ay(s)		4 NR	Not reported	Z	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	Ī
ay(s)			ot reported	Z	Fresh water Lab	Unmeasured	Renewal	Sodium chlo	
ay(s)	L	N.R	Not reported	UNHEALTHY	Fresh water Lab	Unmeasured	Renewal	764/145 Sodium chloride (NaCl)	196841 7
ay(s)			Not reported			Unmeasured	Renewal	Sodium chio	Τ
dy(s)			ot reported		-	Unmeasured	Renewal		Ī
(c) kp			Not reported	2 2	TIEST WATER LAD	Missonen	Static	ATTAE CALL CHIOTIGE (NaCI)	10407
(c) Ap		d NN	Not reported		+	Not vebotted	Kenewai	4/145 Sodium chioride (Naci)	10/000
avis)			of reported			Not Reported	Renewal	7647145 Sodium chloride (Naci)	167963
aute)	П	AND	Not reported		Create March Char	Not Benorted	Denewal	AZIAS CONTINUED (NOCI)	12967
av(s)			ot reported	Z	Fresh water Lab	Unmeasured	Renewal	50	196838
av(s)			Not reported	2		Unmeasured	Renewal	47145 Sodium chloride (NaCl)	1
av(s)		4 NR	Not reported	2	Fresh water Lab	Unmeasured	Renewal	7647145 Sodium chloride (NaCl)	196828
av(s)	П		ot reported	2	Fresh water Lab	Unmeasured	Renewal	8	1
ay(s)			ot reported	2		Unmeasured	Renewal	Sodium chlo	
ay(s)		4 NR	Not reported	2	Fresh water Lab	Unmeasured	Renewal	7647145 Sodium chloride (NaCl)	196776
ay(s)	- 1	n NR	Not reported	2	Fresh water Lab	Unmeasured	Renewal	0	T
ay(s)	П	2 NR	ot reported	2	Fresh water Lab	Unmeasured	Renewal	ŝ	
ay(s)	М	1 NR	Not reported	N	Fresh water Lab	Unmeasured	Renewal	647145 Sodium chloride (NaCl)	196814
ay(s)		2 NR	Not reported	N	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	196831
ay(s)		2 NR	lot reported	N	Fresh water Lab	Unmeasured	Renewal	S	
Day(s)			lot reported	N	Fresh water Lab	Unmeasured	Renewal	7647145 Sodium chloride (NaCl)	196817
ay(s)		6 NR	lot reported	7	Fresh water Lab	Unmeasured	Renewal	47145 Sodium chloride (NaCl)	196801
ay(s)			Not reported		Fresh water Lab	Unmeasured	Renewal	S	196818
ay(s)			ot reported		ab	Unmeasured	Static	S	742016
ay(s)		4 NR	Not reported	TITUTED WATER (N		Unmeasured	Static	647145 Sodium chloride (NaCl)	742038
ay(s)			lot reported	MINERAL WATER N	Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	187904
ay(s)			Not reported			Unmeasured	Static	547145 Sodium chloride (NaCl)	187906
ay(s)			lot reported		Fresh water Lab	Unmeasured	Static	647145 Sodium chloride (NaCl)	187901
ay(s)			lot reported		Lab	Unmeasured	Static	547145 Sodium chloride (NaCl)	
ay(s)		NR	Not reported		Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	187902
(c) Ap	1		ortebotted		Lan	Unmedsured	STATIC	04/145 Sodium chioride (Naci)	Γ
Telko			ocieboiten		Lab	Unimedaured	Signic	Sodiam chio	T
(c) Ap		1 IND	Not reported	WINCESCO WATER, TEX	Flesh water Lab	Officedsured	Sidile	/64/140 sodiom chloride (Naci)	T07903
Tallan	1	1	ot reported		Fresh water Lab	Cilileasured	Single	200	
avis)	1	ZNR	Not reported	the second second second second	Fresh water lah	Unmeasured	Static	7647145 Sodium chloride (Nect)	107000
av(s)	1	1 NR	Not reported	MEDIUM HARD RECONSTITUTED WATER INV	Frech water lah	Unmeasured	Static	2 3	247077
av(s)	1	4 NR	ot reported	ONSTITUTED WATER IN	Greek water Lab	Unmeasured	Charic	7647145 Sodium chloride (NaCl)	900247
ay(e)	1		or reported	N-WEDILINA N	FIRST Water Lab	Unmeasured	Static	64/145 South chorde (Naci)	
20/6	١		otreported	WIEDLOW DESCRIPTION OF THE WHOLE IN	Fresh mater Lab	Unmeasured	Static	04/140 SOCIOTA CHIOCIDE (Maci)	
au(s)		1 NR	Not reported	MEDITIN HARD RECONSTITUTED WATER (MA	Fresh water Lab	Unmasured	Static	04/140 Socioni chloride (Naci)	742910
av(s)	1	1	lot reported		Fresh water   sh	Inmeseured	Cratic		
av(s)			Not reported			Unmeasured	Static	7647145 Sodium chloride (NaCl)	7060300
av(s)	-	1 NR	ot reported	7	Fresh water lah	Unmassured	Statio		
av(s)			Not reported	Þ	Fresh water Lab	-4	Flow-through		1
av(s)			Not reported	4 REPLICATES	Fresh water Lab	Measured	Static	Sodium chlor	7
ay(s)	ш	NR	ot reported	7	Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCi)	2060983
ay(s)			lot reported	7	Fresh water Lab	Unmeasured	Static	547145 Sodium chloride (NaCl)	
ay(s)			lot reported	7	Fresh water Lab	Unmeasured	Static	647145 Sodium chloride (NaCl)	
ay(s)		1 NR	Not reported	7		Unmeasured	Static	7647145 Sodium chloride (NaCl)	764326
ay(s)			lot reported	7	Fresh water Lab	Unmeasured	Static	647145 Sodium chloride (NaCl)	
ay(s)	NR.		Not reported	7	Fresh water Lab	Unmeasured	Static	547145 Sodium chloride (NaCl)	764325
ay(s)			lot reported	77		Unmeasured	Static	647145 Sodium chloride (NaCl)	764333
ay(s)	П		Not reported	3 REPLICATES	Fresh water Lab	Unmeasured	Static	547145 Sodium chloride (NaCl)	2164637
Tollan		0.0417 NB	tot reported	3 DEDUCATED		Cililicopiico	Signe	04/145 Sodium Cilioride (Naci)	70560907
laline.	1		tot reported		THE WASTER DAY	Officespied	Static	04/143 Sociality Ciliotide (Maci)	HQCTC7
av(s)			Not reported		Fresh water Lab	Unmeasured	Static	SA7145 Sodium chloride (NaCl)	I
ay(s)		3 NR	lot reported	2	Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	231565
ay(s)			Not reported		Fresh water	Unmeasured	Static	547145 Sodium chloride (NaCl)	
ay(s)			Not reported		LIESH MOTE!	Commence	100000000000000000000000000000000000000		
A Lacon	1				Panala vivatan	heritseamni	Static	7647145 Sodium chloride (NaCl)	231566

767890	766867	766866	766872	766873	2153774	2075410	767967	767968	767870	767871	767960	767959	2075468	767872	767873	2075434	2075442	767895	767896	767880	767879	2153268	767962	767961	767898	767897	2153271	766864	766865	207546	767903	76790	2152825	76785	20771	741699	741550	214681	214681	2146816	74169	20771	20771	741686	20771	742567	741687	206099	75944	75939	75944	759417	75939	75944	759442	2611	19680	196787	18301	196781	19683	19680	19677	19682	19684	19682	196835	196815	19679	19677	19861	19680	19679	19678	19682	19678	19679	19679	Kesuit Mulling
	7647145 Sodium chloride (NaCl)	Sodiun	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (N	7647145 Sodium chloride (N	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (N	7647145 Sodium chloride (N		764714	764714	764714		764714	764714	764714	764714	764714	7647145 Sodium chloride (NaCl)	764714	7647145 Sodium chloride (N	Sodium chlor	7647145 Sodium chloride (NaCl)	7647145 Sodium chlo	7647145 5	7647145	7647145 Sodium chloride (NaCl)		7647145	T	7647145 Sodium chloride (f	Sodium chlor	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride	Т	Т	/64/145 Sodium chloride (	7647145 Sodium chloride (NaCl)	13	7647145	7647145 Sodium chloride (		764714	764714		7647145	7647145 Sodium chloride	7647145 Sodium chloride (	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (	7647145 Sodium chloride (NaCl)	7647145	7647145 Sodium chloride (	7647145 Sc	7647145	7647145 50	7647145 Sodium chlor	7647145 Sodium chlor	764714	Civ.	7647145 Sodium chloride (	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (		7647145 Sodium chloride (NaCl)	T		7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride	5 7647145 Sodium chloride (NaCl)	2 7647145 Sodium chloride	1 7647145 Sodium chloride I	5 7647145 Sodium chloride	196798 7647145 Sodium chloride (NaCl)	4 7647145 Sodium chloride	CAS Northber Chermical Name
				aCI) Renewal			laCl) Renewal							IaCl) Renewal								laCl) Renewal			vaCI) Renewai		vaCI) Renewal				vaCi) Renewai				(NaCl) Renewal			VaCI) Renewal					VaCl) Renewal			NaCl) Renewal		NaCl) Static	(NaCl) Static	NaCl) Renewal		NaCl) Renewal				NaCl) Static		ide (NaCl) Renewal						NaCl) Renewal			NaCl) Renewal			NaCi) Renewal	NaCi) Renewal			NaCl) Renewal	(aCl)	NaCl) Renewal	JaCI)	NaCl) Renewal	2
Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Linmeasured	Unmeasured	Unmeasured	Measured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Measured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Measured	Unmeasured	Measured	Measured	Measured	Unmeasured	Unmeasured	Unmeasured	Measured	Measured	Measured	Measured	Measured	Measured	Measured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	+							
Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab				Fresh water lab		Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab			Fresh water Lab			Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab		Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	Fresh water Lab	
				HARDNESS OF 80 MIG/L CACOS																		HARDNESS OF 20 MG/L CACO3				4	HARDNESS OF 40 MG/L CACO3					1100	4 REPLICATES		CINCORC DE COCCUE	CHRONIC 3 BROOM STRV	CHRONIC -3 BROOD SIDY	10 REPUICATES	10 REPUCATES	10 REPLICATES	ACUTE			ACUTE		CHRONIC -3 BROOD STDY	C -3 BROOD																		UNHEALTHY														
Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	
7 NR	7 NR	7 NR	7 NR	7 NR	2 NR	2 NR	7 NR	7 NR	7 NR	7 NB	7 77	7 200	2 NB	7 NB	7 NR	22 NR	2 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	2 NB	7 NR	7 NR	2 NR	7 NR	8 NR	NO NA	7 NR	7 NR	7 NR	7 NR	2 NR	2 NR	8 NR	2 NR	2 NR	7 NR	7 NR	0.0417 NR	90 NR	80 NR	80 NR	80 NR	80 NR	80 NR	80 NR	1 NR	5 NR	2 NR	J N N	6 NR	A NR	1 NR	1 NR	4 NR	4 NR	6 NR	2 NR	2 NR	A NR	2 NB	SNA S	4 NR	4 NR	1 N8	6 NR	1 NR	1 NR	1 NR	
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NR.																																				567	NO NA	NO NA	N.R.	NR	NR	NR	NR	NR.	NR	NR	N.R.	NA .	NR :	NR S	Z Z	N N	20 2	NK NK	Z Z	NR NR	NR	NR	NR	NR	NR	NR S	NR	NR :	NR	NR	NR	NR.	NR	NR	NR	NR	NR.	NR	NR	NR	NR	NR	NR NA	Days Duration islan Inward	The same same same same same same same sam
Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Dav(s)	Davis	Dav(s)	Davis	Day(s)	Day(s)	Day(s)	Davis	Dav(s)	Day(s)	Dayle	Dayle	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Davis)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Dav(s)	Dav(s)	Dav(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s) E	Day(s)	Day(s)	Day(s)	ODSEL ACTION	Table Land Division
NR-LETH Mo	NR-LETH MA	LC50 Mc	NR-ZERO Mo	NR-ZERO MA	NR-ZERO MA	NR-LETH Mc	NR-LETH Mc	INV-SEVO MI	NR-ZERO MA	NR-ZERO MA	LCS0 Mi			LCS0 Mo						LCS0 Mi	NR-ZERO Ma	NR-LETH MO	EC50 Su	NR-ZERO Mo	LC50 Me	LC50 M	LCS0 M	1050 M	1050 M	1050 W	LOCK Se	DEC. ST	NOEC SI	1050 M	NR-ZERO MA	EC50 M	NOE!	INTERIOR IN	LCS0 M	NR-ZERO M	LC50 M	LC50 Mi	NR-ZERO M	LC50 M.	NR-LETH M	NR-LETH M	LD50 M	LD99 M	NR-LETH M	NR-ZERO M	050	CSO	NN-ZENC	NK-LEIN	CSO	CSO	C50	VR-LETH	VR-ZERC	C75	063	C15	NOEC	VR-ZERO	C25	663	OEC	C10	VR-LETH	VR-ZERC	C80	VR-LETH	.C50	C05	C40	C85	C95	MATC	C01	COO	ndngint
Mortality	ortality	ortality	ortality	ortality	ortality	ortality	rtality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	rvival	ortality	ortality	ortality	ortality	ortality	ortality	ortality	rtality	INVIVAL	Divide	ortality	ortality	ortality	accin	treb	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	ortality	Mortality	ortality	ortality	rvival	EVIVA	rvival	rvival	ortality	rvival	rvival	Irvival	rviva	ortality	ortality	irvival	ortality	Mortality	irvival	rvival	Survival	rvival	Survival	Political	Survival	- Address specials

No.	Mortality		NA.		Not reported	3 REPLICATES	Fresh water Lab		Renewal		2053814
Colora   C	A THE TANK	(S)AR	NID	Г	Morrichorica	L ACTOR DE LA COLONIA DE LA CO	The state of the s	Commence	THE ASSTRACT		
Coloradia protection   Coloradia   Color	Mortality	ay(s)	NO NA		Not reported	3 REPUCATES	Fresh water Lab	Unmeasured	Renewal	7647145 Sodiam chloride (Naci)	3053818
Company   Comp	Mortality	ay(s)	NR R	16	Not reported		Fresh water Lab	Unmeasured	Renewal	764/145 Sodium chloride (NaCl)	2053816
Coloration of Coloration   Co	ZERO Mortality	3y(s)	NR		Not reported		Fresh water Lab		Flow-through	7647145 Sodium chloride (NaCl)	742663
Colora   C	0 Mortality	ay(s)	NR		Not reported	STUDY	Fresh water Lab		Static	Sodium chlor	741789
Column   C	LETH Mortality	1V(S)	NR NR	NR.	Not reported		Fresh water Lab		Static		2061000
Coltamo   Colt	0 Mortality	ay(s)	2 2	NR NR	Not reported		Fresh water Lab		Flow-through		2061031
Column   C	Survival	ay(s)	NR	П	Not reported	5 REPLICATES	-	Unmeasured	Renewal	So	773637
Coltame   Colt	0 Mortality	ay(s)	NR.		Not reported			Unmeasured	Static	So	2064154
Coltable	0 Mortality	ay(s)	NR R		Not reported			Unmeasured	Static	7647145 Sodium chloride (NaCl)	2064156
Colora   Colora Colora Colora Colora   Colora Col	LEIH Mortality	ay(s)	NB NR		Not reported			Unmeasured	Static	7647145 Sodium chloride (NaCl)	2062417
Part	ZERO Mortality	ay(s)	NR		Not reported			Unmeasured	Static	7647145 Sodium chloride (NaCl)	2062416
Part	0 Mortality	ay(s)	NR		Not reported	CHANGED AT 192 HOURS			Renewal	7647145 Sodium chloride (NaCl)	741834
Part	LETH Mortality	ay(s)	NR		Not reported				Static	7647145 Sodium chloride (NaCl)	2060998
Part	LETH Mortality	ay(s)	NR		Not reported		Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	2060995
Mathematical process	LETH Mortality	dy(s)	NR S		Not reported		Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	2060994
Coltable	O Mortality	ay(s)	Z Z	1	Not reported		Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	251526
Part	0 Mortality	ay(s)	NR		Not reported		Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	231525
Part	0 Mortality	ay(s)	NR.		Not reported			Unmeasured	Static	Sodium chlo	231527
Mathematical Mat	0 Mortality	ay(s)	NR.		Not reported			Unmeasured	Static	Sodium chlo	231523
Part	0 Mortality	ay(s)	NR.		Not reported			Unmeasured	Static		231524
Part	O Survival	ay(s)	NR		Not reported	RECONSTITUTED WATER		Unmeasured	Static	So	2052775
MATCH Singles relotes   MACH   Contact   Con	O Survival	av(s)	NR	7	Not reported	3 REPLICATES		Measured	Static	So	803856
Participation of the color   Participation	O Survival	av(s)	NR I		Not reported			Measured	Static	Sodium chlori	741563
Part   Distant Control   Part   Par	LEIH Mortality	dy(s)	ND NO	UN C7:7	Not reported			Unmeasured	Static	Sodium chlor	70004/
Patrolis	CELL MOLITAINA	dy(s)	NW	1.25 NA	Not reported		-	Uninedsured	Static	פחומות כוווטו	740090
Part	LETH Mortality	ay(s)	Z Z		Not reported		4	Unmeasured	Static	Sodium chlor	2061001
Part	LETH Mortality	ay(s)	NR	1	Not reported			Unmeasured	Static	Sodium chlor	2060988
Part	Mortality	ay(s)	NR		Not reported	4 REPLICATES, WITHOUT FISH KAIROMONE		Unmeasured	Static		2160468
Part	EC Survival	ay(s)	NR		Not reported	T FISH KAIROMONE		Unmeasured	Static	un	2160460
Part	C Survival	ay(s)	NR.		Not reported	SH KAIROMONE	Fresh water Lab	Unmeasured	Static	Sodium chlo	2160464
Part	0 Mortality	ay(s)	NR.		Not reported	SHKAIROMONE	Fresh water Lab	Unmeasured	Static	Sodium chlo	2160465
Patricul policies   Manager   Mana	C Survival	av(s)	NR S		Not reported	KAIROMONE	Fresh water Lab	Unmeasured	Renewal	0 3	2160490
Part	EC SURVIVAI	aylsi	NO S		Not reported	TOTAL NATIONS	Fresh Water Lab	Unmeasured	nellewal	2 8	2160/461
Part	C Survival	ay(s)	2 2		Not reported	ISH KAIROMON	Fresh water Lab	Unmeasured	Renewal	Sodium chlor	2160484
Part	C Survival	ay(s)	NA NA		Not reported	WITH FISH KAIROMONE	Fresh water Lab	Unmeasured	Static	Sodium chlor	2160471
Part	Mortality	ay(s)	NR		Not reported	WITH FISH KAIROMONE	Fresh water Lab	Unmeasured	Static	w	2160473
Part	EC Survival	ay(s)	NR	2 NR	Not reported		Fresh water Lab	Unmeasured	Static	Sodium chlor	2160470
Part	ш	ay(s)	NR	ш	Not reported		Fresh water Lab	Unmeasured	Static	Sc	2160472
Part		ay(s)	N.R	- 1	Not reported		Fresh water Lab	Unmeasured	Static	S	2160475
Part		av(s)	NR S		Not reported	1	Fresh water Lab	Unmeasured	Static	Sodium	2160469
Part	- 1	av(s)	NR :	2 NR	Not reported	4 REPLICATES WITHOUT FISH KAIROMONE	Fresh water Lab	Unmeasured	Static		2160450
Part 245   Soldinm mitoride ( NRC)   Static   Measured Fresh water ( Jab.   Eth. MOCERATE V HADD ECCONSTITUTE) (Not separed   2   NR   NR   20   MR   NR   20   MR		av(s)	200		Not reported	10 REPLICATES SALINITY ACCUMATION CO	Fresh water Lab	Unmeasured	Renewal		2160493
	- 1	ay(s)	NB NA		Not reported	4-10 REPLICATES ADULT EATHEAD MINNOY		Inmestired	Denewal	7647145 Sodium chloride (Naci)	2005000
Part 245 Studim mitotinicis (Nati)		dy(s)	NA NA		Not reported	CATES, LARGEMOOTH BASS PREL		Unmeasured	Kenewai	7647145 Sodium chloride (Naci)	2055020
Part 145 Sedim michiode Navi)		ay(s)	NR R		Not reported	CATES	Fresh water Lab	Unmeasured	Renewal	Sodi	2055812
Partial Stadium chloride Navil	П	ay(s)	NR.	2 NR	Not reported	DILUTION WATER	Fresh water Lab	Measured	Static	Sodium chlor	201566
Part		ay(s)	NR	70	Not reported	WATER	Fresh water Lab	Unmeasured	Static	Sodium chlor	741434
75621348 (Sodium chloride (NaSCI)         Static         Measured (Assert Lab Sodium chloride (NaSCI)         Static         Measured (Assert Merchan)         Static (Assert Merchan)         Sta	RO	ay(s)	NR		Not reported		Fresh water Lab	Measured	Renewal		201558
Tright   Signatum nithoride   Nacil   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Most reported   2 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Not reported   1 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Not reported   1 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Not reported   1 NR   NR   Cayl   NR-ZERO   Mortality   Tright   Static   Measured   Fresh water   Jab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2		av(s)	NR		Not reported	4-10 REPLICATES, 24 HOUR ADULT FATHEAU	Fresh water Lab	Unmeasured	Renewal		2055828
		ay(s)	NR :		Not reported			Measured	Static	S	201545
Page		av(s)	NR S	Y [1]	Not reported			Unmeasured	Static	S L	09481
75647145 Sedium chloride (NaCl)         Static         Measured         Fresh water (ab)         ED MODERATELY HARD RECONSTITUTED (Not reported)         2 NR         MR         Day(s)         MR-2ERO (Mortality)           75647145 Sedium chloride (NaCl)         Static         Measured         Fresh water (ab)         FED MODERATELY HARD RECONSTITUTED (Not reported)         2 NR         NR         Day(s)         MR-2ERO (Mortality)           75647145 Sedium chloride (NaCl)         Static         Measured         Fresh water (ab)         FED MODERATELY HARD RECONSTITUTED (Not reported)         2 NR         NR         Day(s)         MR-2ERO (Mortality)           75647145 Sedium chloride (NaCl)         Static         Measured         Fresh water (ab)         FED MODERATELY HARD RECONSTITUTED (Not reported)         2 NR         NR         Day(s)         MR-2ERO (Mortality)           75647145 Sedium chloride (NaCl)         Static         Measured         Fresh water (ab)         FED MODERATELY HARD RECONSTITUTED (Not reported)         1 NR         NR         Day(s)         MR-2ERO (Mortality)           75647145 Sedium chloride (NaCl)         Static         Measured         Fresh water (ab)         FED MODERATELY HARD RECONSTITUTED (Not reported)         1 NR         NR         Day(s)         NR-2ERO (Mortality)           75647145 Sedium chloride (NaCl)         Static         Measured <td< td=""><td>1</td><td>ay(s)</td><td>N N</td><td></td><td>Not reported</td><td></td><td></td><td>Unmeasured</td><td>Static</td><td>318</td><td>742018</td></td<>	1	ay(s)	N N		Not reported			Unmeasured	Static	318	742018
7567145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         2 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         2 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         2 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         2 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         2 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         2 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         Static         Measured Fresh water Lab         EED, MODERATELY HARD RECONSTITUTED Voict reported         1 NR         NR         Day(s)         NR-ZERO Modrality         7667145 Sodium chloride (NaCl)         N	50 Mortality	ay(s)	Z R	150	Not reported			Measured	Static	7647145 Sodium chloride (NaCl)	767308
75627145 [Sodium chloride (NaCl)         Static         Measured         Fresh water lab         EED, MODERATELY HARD RECONSTITUTED Voic reported         2 NR         NR         Day(s)         NR-2826 Mordality           7667145 [Sodium chloride (NaCl)         Static         Measured         Fresh water lab         FED, MODERATELY HARD RECONSTITUTED Voic reported         2 NR         NR         Day(s)         NR-2826 Mordality           7667145 [Sodium chloride (NaCl)         Static         Measured         Fresh water lab         FED, MODERATELY HARD RECONSTITUTED Voic reported         2 NR         NR         Day(s)         NR-2826 Mordality           7667145 [Sodium chloride (NaCl)         Static         Measured         Fresh water lab         DEFINITIVE ACUTE*, A REPLACITE*, A REPLA	-ZERO Mortality	ay(s)	NR	2 NR	Not reported			Unmeasured	Static	7647145 Sodium chloride (NaCl)	742927
Tock1745  Sodium chloride (NaCl)   Static   Measured Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   2   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   2   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   2   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   2   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   NR reported   1   NR   NR   Day(s)   NR-ZERO   Morsility   NR REPORTED	-ZERO Mortality	ay(s)	NR		Not reported		Fresh water Lab	Unmeasured	Static		742068
Tock1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (NaCl)   Static   Measured   Fresh water   ab   ED_ MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Toch1745  Sodium chloride (N	50 Mortality	aγ(s)	NR	~ [	Not reported		Fresh water Lab	Measured	Static		772103
Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   1 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   1 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   1 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   NNt reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   Static   Measured   Fresh water   Lab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   Total   NR   Day(s)   NR-ZERO   Mortality	50 Montality	ay(s)	NR.	-	Not reported		Fresh water Lab	Measured	Static	Sodium	767302
Tock1745  Sodium chloride (Naci)	-ZERO Mortality	lav(s)	NR.	2 NR	Not reported		Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (NaCl)	742926
7567145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   EED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(s)   NR-ZERO   Mortality   767145  Sodium chloride (Nacl)   Static	-ZERO Mortality	)av(s)	N. Z	Z NR	Not reported		Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (Naci)	742931
Technical Sedium chloride (Naci)   Static   Measured Fresh water   Ab   EED, MODERATELY HARD RECONSTITUTED Valor reported   2 NR   NR   Day(s)   NR-ZERO   Mortality	-ZERO Mortality	Jay(s)	200	S NB	Not reported	RANGEFINDING ACOIC	Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (Naci)	SOCOTES
Todatas  Sedium chloride (Naci)   Static   Measured   Fresh water   Ab   ED, MODERATELY HARD RECONSTITUTED   Not reported   1   NR   Day(s)   NR-ZERO   Mortality   Todatas  Sodium chloride (Naci)   Static   Measured   Fresh water   Ab   ED, MODERATELY HARD RECONSTITUTED   Not reported   2   NR   NR   Day(s)   NR-ZERO   Mortality   Todatas  Sodium chloride (Naci)   Static   Measured   Fresh water   Ab   ED, MODERATELY HARD RECONSTITUTED   Not reported   2   NR   NR   Day(s)   NR-ZERO   Mortality   Todatas  Sodium chloride (Naci)   Static   Measured   Fresh water   Ab   DEFINITY   NACITE	-LEIH MORTALITY	Pay(s)	Z Z		Not reported	RECONSTITUTED	iei	Measured	Static	/64/145 Sodium chloride (Naci)	768486
7567145   Sodium chloride (Nacl)   Static   Measured Fresh water   ab   ED_MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(5)   NR-ZERO   Mortality   NR-ZERO	-LETH Mortality	Jay(s)	NR		Not reported	RECONSTITUTED	Fresh water Lab	Measured	Static		768521
Todatas  Sodium chloride (Naci)   Static   Measured Fresh water   Ab ED, MODERATELY HARD RECONSTITUTED   Not reported   1 NR   NR   Day(s)   NR-ZERO   Mortality	50 Mortality	ay(s)	NR	1 NR	Not reported		Fresh water Lab	Unmeasured	Static		160321
	-ZERO Mortality	Jay(s)	NR	6-	Not reported	MODERATELY HARD RECONSTITUTED	Fresh water Lab	Measured	Static		768526
7647145   Sodium chloride (NaCl)   Static   Measured   Fresh water   Lab   FED_MODERATELY HARD RECONSTITUTED   Nat reported   2   NR	-LETH Mortality	ay(s)	NR	100	Not reported	FED, MODERATELY HARD RECONSTITUTED V	Fresh water Lab	Measured	Static	7647145 Sodium chloride (NaCl)	768467
7647145   Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   FED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(5)   NR-ZERO   Mortality   7647145   Sodium chloride (Nacl)   Static   Measured   Fresh water   ab   FED, MODERATELY HARD RECONSTITUTED   Not reported   2 NR   NR   Day(5)   NR-ZERO   Mortality   NR-ZERO   MO	-ZERO Mortality	Jay(s)	NR.	2 NR	Not reported	MODERATELY HARD RECONSTITUTED	Fresh water Lab	Measured	Static	7647145 Sodium chloride (NaCl)	768488
	-LETH Mortality	Day(s)	NR II	- 1	Not reported	FED. MODERATELY HARD RECONSTITUTED V		Measured	Static	7647145 Sodium chloride (Naci)	768499
7647145 Sodium chloride (NaCl)  Static  Measured  Fresh water Lab  Fresh w	-I FTH Mortality	)av(s)	NR Z	913	Not reported	CED MODERATELY HARD RECONSTITUTED A	Fresh Water Lab	Measured	Static	7647145 Sodium chloride (NaCl)	768475
7647145 Sodium chloride (NaCl.)  Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED (Not reported 2) NR NR-ZERO Mortality 7647145 Sodium chloride (NaCl.)  Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED (Not reported 2) NR NR-ZERO Mortality 7647145 Sodium chloride (NaCl.)  Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED (Not reported 2) NR Day(s) NR-ZERO (Mortality 7647145 Sodium chloride (NaCl.)  Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED (Not reported 2) NR Day(s) NR-ZERO (Mortality NR-ZERO Mortality NR-ZERO Mortality NR-ZERO MOTALITY NR-ZERO MOTALI	ZERO Mortality	Jay(s)	N N		Not reported	ACUIE, 4 REPUCATES	Fresh water Lab	Unmeasured	Static	7647145 Sodium chloride (Naci)	2210504
7647145 Sedium chloride (Nacl)         Static         Measured         Fresh water Lab         FED, MODERATELY HARD RECONSTITUTED (Not reported)         2 (NR         NR         Dayls         MR-ZBRO Modrality           7647145 Sedium chloride (Nacl)         Static         Measured         Fresh water Lab         FED, MODERATELY HARD RECONSTITUTED (Not reported)         1 (NR         DAYLS         NR         Dayls         NR-ZBRO Modrality           7647145 Sedium chloride (Nacl)         Static         Measured         Fresh water Lab         FED, MODERATELY HARD RECONSTITUTED (Not reported)         1 (NR         DAYLS         NR         Dayls         NR-ZBRO Modrality           7647145 Sedium chloride (Nacl)         Static         Measured         Fresh water Lab         FED, MODERATELY HARD RECONSTITUTED (Not reported)         2 (NR         NR         Dayls         NR-ZBRO Modrality           7647145 Sedium chloride (Nacl)         Static         Measured         Fresh water Lab         FED, MODERATELY HARD RECONSTITUTED (Not reported)         2 (NR         NR         Dayls         NR-ZBRO Modrality	-ZERO Mortality	Jay(s)	NR.		Not reported	FED, MODERATELY HARD RECONSTITUTED		Measured	Static	7647145 Sodium chloride (NaCl)	768530
7647145 Sodium chloride (NaCl) Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED Not reported 2 NR NR Day(s) NR-ZERO Mortality 7647145 Sodium chloride (NaCl) Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED Not reported 1 NR NR Day(s) NR-LETH Motrality	-ZERO Mortality	Day(s)	NR	100	Not reported	FED, MODERATELY HARD RECONSTITUTED		Measured	Static	7647145 Sodium chloride (NaCl)	768523
7647145 Sodium chloride (NaCl) Static Measured Fresh water Lab FED, MODERATELY HARD RECONSTITUTED Not reported 2 NR NR Dayls) NR-ZERO Mortality	-LETH Mortality	ay(s)	NR	1 NR	Not reported	FED, MODERATELY HARD RECONSTITUTED	Fresh Water Lab	Measured	Digit	7647145 Sodium chloride (NaCi)	768496
	-ZERO Mortality	Pay(s)	NR.				7		7545	The rate of the last of the la	0.00

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ffect Measurement	Aortality	Mortality	Mortality	Antality	Acrtality	Mortality	Mortality	Mortality	Mortality	Aortality	Aortality	Mortality	Mortality	Mortality	Jortality	Mortality	Anriality	Acrtality	Aprilative	Mortality	Mortality	Mortality	Mortality	Mortality	Aortality	Mortality	Mortality	Acception	Mortality	Mortality	nortality	Mortality	Mortality	Aortality	Mortality	urvival	Mortality	Hatch	Hatch	flortality	Mortality	Mortality	Mortality	fortality	Mortality	Mortality	Mortality	urvival	fortality	Mortality	fortality	fortality	Mortality	urvivorship	urvivorship	Survivorship	urvivorship	urvivorship	Survivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvivorship	urvival	urvival	invival	invival	i para i	or vival	survival	Survival	Irvival	urvival	Invival	urvivai	Invivat	Survival	J.Viva.
Endpoint E	LC50 N	NR-LETH N	NR-LETH N	NR-I FTH	NO-1 ETH A	NR-LEIT N	T	T			N		10	NOEC	l.	LOEC				000		0121	Ī	1610		200		T	1005 N	10 1571		NK-ZEKU IN	LCSOT N		U	NOEC	JR-LETH	OEC	OEC	NR-ZERO N	NR-ZERO N	NR-ZERO N	NR-ZERO N	NR-ZERO N	NR-ZERO IN	NR-ZERO N	NR-ZERO N		NR-ZERO N	NR-ZERO N	NR-LETH N	NR-LETH N	NR-LETH N	LOEC			IOEC Si	0		NOEC S	NOEC S	LOEC S	LOEC	NOEC	LOEC	NOEC SI	LOEC St	NOEC SI	NOEC St	NOEC St		0,	FC20 Si	FC50 S	0003	2000	7 6		,,,,	ECSO St	0110	ברפת פייבע	ECSU SE		EC50 St	1
erved Dural	(S	(5	(8	2	6	6	(2)	6 3	(5)	(s	(5	(s	(5	(5	s)	(8)	1	100	(6)	6 6	10	100	100	100	100	1	0 0		(s)		(8)	(8)	(5)	er stage	2)	Day(s) post-hate	(5	Day(s) post-hatdn	Day(s) post-hato	(5	(5	(9					(9	s) post-hato						(1)			(0)	()	(2)			()	~					)																		
on Max (Days) Obse	Day	Day	Dav(s)	David	Dayle	Day	Day	Dayle	Davis	Day	Day(s)	(Day)	)APQ	Day(s	Dav(s)	Dav	Davis	live O	Dayle	Davie	Dayle	land.	Dav(s)	Dayle	Daylel	) And	Day(s)	Van C	Day(s)	)APO	Day	Day(s)	Day(s)	Gosr	Day(s)	Day	Day	Day	Day	Day(s)	Day(	Day(s)	Day(s)	)Ap(	Day(s)	Day(s)	Day(s)	Day(	Day(s)	Day(s)	Day(s)	Day(	Day(s)	Day	Day	Day(s)	Day(s)	Day(	Day(	Day(s)	Day(s)	Day(s	Day(s)	Day(s)	Dayle	Day(s	Day(s)	Day(s	Day(s)	Dav(s)	Day(s	Dav(s	Dav(s)	Dav(c)	Jan J	Cay(a)	Day	Day(s)	Day(s)	Day	Day(s)	Dank	Day(s)	Claylo Complete	Dank	1120
ion Min (DaysDurati	NR	NR	NR	N	No	NN	NR	No No	NB	NR	NR	NR	N.	NR	NR	NR	a N	GN C	ON ON	ON ON	NA	NB	an	an an	ON	W W	S ON	ON ON	NR G	AN CA	NR.	N. S.	NK	NR	NR	NR	NR	N.	NR.	NR	NR	N.	NR	NR	NR	NR	NR	Z.	N.S.	NR	NR	NR.	N.	N. S.	Z S	S S	NR.	N.	NR	NR	NR	N	N.S.	N.	NR	NR	SS	NR	S	NR	NR	N.	N.	N.	NEW MIN	VIVI	CAL CON	NR	N.	NR.	N N	NN ON	2 0 2 2	Z Z	NA GA	oxi
Duration Mean   Duration Min (Day4Duration Max (Days)   Observed Durat Endpoint   Effect Measuremen   0.125 NR   Nortality   NR   1.55 NR   Mortality   0.125 NR   1.55 NR   1.5	3 NR	1 NR	2 NR	N C	N N SCE O	0.125 NR	SNR	N O N	A A	N N	4 NR	4 NR	INR	1 NR	1 NR	N L	4 NR	NA NE		ON PE	NN SC	ZIND	AN TC	ON IC	ON C	UNI N	AN SC	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	14 NR	NA TOWN	0.0417 INR	4 NR	ZNR		15 NR		1.25 NR	0.25 NR	0.25 NR	52 NR	25 NR		32 NR	38 NR	37 NR	28 NR	20 NR	7 NR	2 NR	2 NR	0.0833 NR	1.0417 NR	0.0417 NR	14 NR	14 NR	14 NR	14 NR		14 NR	14 NR	14 NR	14 NR	14 NR	14 NR	14 NR	14 NR	14 NB	INR	NR.	aN I	ON T	UNIT T	T IN	ZINK	N. I	INK	I NR	TIND	A NR	N N	NN T	wilt.				
Response Site		Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not renorted	Not reported	Motroported	Motroported	Mot reported	Mor reported	Motreported	Morroportod	Mot reported	Morroported	Mot reported	and reported	Not reported	Not reported			Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Notroported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported												
ocation Experimental Design	3 REPLICATES				2010000	4 KEPLICATES											A REPLICATES	4 TO 6 BEDLICATES CONDITCTED IN STAND	CONDICTED	A TO 6 PERIODIES, CONDUCTED IN STAND	A TO 6 BERLICATES CONDUCTED IN STAND	A TO G DEDILICATES CONDUCTED IN STAND	A TO & BEBLICATES CONDUCTED IN STAND	4 TO 6 DEDUCATES CONDUCTED IN STAND	A TO 6 PERICATES CONDUCTED IN STAND	4 TO C DEDITIONES, CONDUCTED IN STAND	4 TO 6 PEDICATES CONDUCTED IN STAND	4 TO B REFLICATES, CONDOCTED IN STAND	4 TO B REPLICATES, CONDUCTED IN STANDANG	4 I U & REPUCATES, CONDUCTED IN STAND	NES.	POSITIVE CONTROLS OF BIOASSAY BATTERY		3 REPLICATES	3 REPLICATES	UNDISSOCIATED EGGS		DISSOCIATED EGGS	DISSOCIATED EGGS	FISH IN TANKS, EXPERIMENT 1	FISH IN TANKS, EXPERIMENT 2	FISH IN TANKS, EXPERIMENT 10	FISH IN TANKS, EXPERIMENT 7	FISH IN TANKS, EXPERIMENT 6	FISH IN TANKS, EXPERIMENT 11	FISH IN TANKS, EXPERIMENT 3	FISH IN TANKS, EXPERIMENT 8	UNDISSOCIATED EGGS			3 REPLICATES, BUCKEYE LAKE WATER Not reported	BALLASTED TANKS ON TRANSOCEANIC CON	TANKS WITH RESIDUAL BALLAST ON TRANS	FALL	WINTER	SPRING	SPRING	WINTER	SUMMER	SUMMER	SPRING	SPRING	WINTER	FALL	FALL	WINTER	WINTER	WINTER	FALL	WINTER	WINTER	RECONSTITUTED WATER	GRAND RIVER WATER	SYDENIA DIMED WATED	STOCKHAMI RIVEN WATER	STUDENTIAN RIVER WATER	MAIILAND RIVER WAIER		RECONSTITUTED WATER, 2008	RECONSTITUTED WATER, 2009	THAMES RIVER WATER	THAMES RIVER WATER		Disapped married with the	GRAND RIVER WATER	MAITLAND KIVEK WATER
31	Lab	Lab	Lab	Lab As I	Cab	rap	Lab	Lab	lah	Lab	Lab	Lab	Lab	Lab	Lab	(ap	de l	2 2	Lab.	ran	Lab	200	Lab	7 7 7	Can	Lan	OPT -		rap	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	lab	lah.	907		Tag	Lan	Lab	rap	Lab	rap	Lab	Lab	Lab	rab	rap
sis Media Type Test	Fresh water	Fresh water	Fresh water	Frach water Lab	Fresh Water	Fresh water	Fresh water	rresh water	Frach wate	Fresh water Lab	Fresh water	Less water	Liesti water	Fresh water	Centh water	Fresh water	Freeh mater	riesti water	riesh water	Fresh Water	LIESU WALE	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water Lab	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water L	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water L	Fresh water	Fresh water	Fresh water	Frach water	Proch woter	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water L	Fresh water	Fresh water	Fresh water							
Chemical Analy	Unmeasured	Unmeasured	Unmeasured	Unmassured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmassured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Immeacured	Massirad	Nicasolea Nicasolea	Contreasored	Offittedsured	Unmedsured	Office and ed	Officeaured	Universities	OfficedSured	numedsured	unmeasured	Ontheasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Measured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Inmeasured	Immeaning	o constant	Onmeasured	unmeasured	Unmeasured	Measured	i		Unmeasured			Measured		
Exposure Type	Renewal	Static	Static	Charle	Static	Renewal	Static	Static	Charic	Static	Verlewal	Renewal	Renewal	Delewal	Periewal	Veriewal	Reflewal	Kenewai	Kenewal	Renewal	Renewa	Kenewal	Renewa	Static	Static	Static	Renewal	Flow-through	Static	Flow-through	Flow-through	Static	Static	Static	Static	Static	Static	Static	Static	Flow-through	Static	Static	Static	Static	Static	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Static	Charle	Cereta	Static	Static	Static	Static	Static	Static	Static	Static	Renewal	Static	Static	Static								
er Chemical Name	15 Sodium chloride (NaCl)	45 Sodium chloride (NaCl)	45 Sodium chloride (NaCl)	se continue objection (Naci)	45 Sodium chloride (Naci)	45 Sodium chloride (NaCl)	45 Sodium chloride (NaCl)	45 Sodium chloride (NaCl)	15 Sodium chloride (Naci)	45 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	35 Sodium chloride (NaCl)	15 Codium chloride (NaCl)	15 Codium chlorida (NaCl)	43 Souldin Chilofide (Naci)	do sodium chloride (Naci)	45 Sodium chloride (NaCl)	50 Sodium chloride (NaCl)	20 Souldin Chichiae (Mach)	45 Sogium chloride (NaCl)	to sodium chioride (Naci)	45 Sodium chibride (Naci)	5 Sodium chioride (NaCi)	45 Sodium chloride (NaCl)	45 Sodium chloride (NaCi)	45 Sodium chloride (NaCI)	45 Sodium chloride (NaCl)	45 Sodium chloride (NaCl)	45 Sodium chloride (NaCI)	45 Sodium chloride (NaCI)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCI)	(Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	(5 Sodium chloride (NaCl)	15 Sodium chloride (NaCI)	(S Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	(Sissing chloride (NaCl)	(5) Sodium chloride (NaCI)	(5 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCl)	15 Sodium chloride (NaCI)	15 Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	(5 Sodium chloride (NaCl)	(5 Sodium chloride (NaCl)	S Sodium chlorida (NaCl)	o codium chloride (Naci)	Southern Chicago	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCI)	5 Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	5 Sodium chloride (NaCI)	(5 Sodium chloride (NaCI)	(S Sodium chloride (NaCI)	5 Sodium chloride (NaCl)	5 Sodium chioriae (NaLI)										
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Measured Malysis	Measured	Measured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Denissemul	Unmeasured	Inmascired	Unmeasured	Unmostricad	Outriedsored	Unineasured	Unmeasured	Unmeasured	Unmeasured	Conneasored	Unmeasured	Measured	Unmeasured	Measured	Measured	Measured	Measured	Measured	Measured	Measured	Measured	Measured	Measured	Measured	Massurad	Unmeasured	Deniseamil	Unmeasured	heritzeamil	I'm pagerad	Unineasured (	Unmeasured	Unmeasured	Unmeasured	Measured	Measured	Measured	Unmeasured	Measured	Unmeasured	Measured	Unmeasured	Measured	Measured	Measured	Unmeasured	Measured	Measured	Unmeasured	Unmeasured	Measured	Unmeasured	Measured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Measured	Unmeasured	Morringd	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured	Unmeasured		Unmeasured		
Exposure Type Static	Renewal	Static	Static	Static	Static	Flow-through	Static	Static	Static	Static	Static	Static	Cratic	Pagettal	Renewal	Renewal	Denemal	Kenewai	Renewal	Charle	Static	Static	Static	Static	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Denamal	Renewal	Ranawal	Renewal	Danatal	Danawal	Designal	Kenewal	Renewal	Renewal	Static	Static	Static	Renewal	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static	Charle	Static	Static	Static	Static	Static	Static	Static	Static	Static	Static
Chemical Name Sodium chloride (NaCl)	Sodium chloride (NaCI)	Sodium chloride (NaCl)	Sulfuric acid	Sulfuric acid	Sulfuric acid	Sulfuric acid	8	Sulfuric acid, Monosodium salt	5 Sulfunc acid sodium salt (1:2)	Suituric acid sodium salt (1.2)	influring acid codium calt (1-2)	Sufferic acid sodium salt (1.2)	Sulfarir acid codium salt (1.2)	Mercinal codium cale (1.2)	Sulfuric acid sodium said (1:2)	Sulfuric acid sodium sait (1.2)	Sulful Cacid Southin Salt (1:2)	Sulfuric acid sodium sait (1:2)	b suiture acid sodium salt (1.2)	Sulfuric acid sodium san (1:2)	Sullunc acid sodium salk (1.2)	Ufferit acid sodium salt (1.2)	Sulfuric acid codium calt (1:2)	ulfuric acid sodium salt (1:2)	fulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	6 Sulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	official acid sodium salt (1.2)	official acid sodium salt (1.2)	official acid acidim salt (1:2)	ulfuric acid sodium salt (1:2)	ufunic acid codium calt (1:2)	Sulfinic acid codium calt (1.5)	uferring and confirm cale (1.2)	Sulfationarid and immediate (1.2)	Sulfuric acid socium salt (1:2)	sulturic acid sodium sait (1:2)	fulfuric acid sodium salt (1:2)	sulfuric acid sodium salt (1:2)	sulfuric acid sodium salt (1:2)	sulfuric acid sodi	Sulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium sait (1:2)	7757826 Sulfuric acid sodium salt (1:2)	utilities acid sodium salt (1:2)	aufuric acid sodium salt (1.2)	ulfuric acid sodium salt (1:2)	26 Sulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfunc acid sodium salt (1:2)	ulfunc acid sodium salt (1:2)	6 Sulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)	Sulfuric acid sodium salt (1:2)	Sulfuric acid sodium salt (1:2)	Sulfuric acid sodium salt (1:2)	ulfuric acid sodium saft (1-2)	7757876 Sufficie acid sodium salt (1:2)	Sulfation and and in the rate (1995)	ulfuric acid sodium salt (1.2)	Sulfuric acid sodium salt (1-2)	ulfuric acid codium salt (1.2)	Sulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1-2)	Sulfuric acid sodium salt (1:2)	ulturic acid sodium sait (1:2)	Sulfuric acid sodium salt (1:2)	7757826 Sulfuric acid sodium salt (1:2)	ulfuric acid sodium salt (1:2)							
Result Number CAS Number (741561 7647145)	764714	764714	766493	766493	766493	7664939	67863 7681381	768138	1/5/82	225257	775793	7757876	7757075	735707	7375707	791911	70/5//	1/5/82	159266 1767876	79/5//	79/5//	735757	775787	775782	775782	775787	775787	775782	775782	775782	775782	775782	2017041 7757876	775787	775787	775707	766278 7757826	775727	766735 7757836	775703	77570	0/0//	77578	77578	77578.	759078 77578265	77578.							150330 //5/8255		757375 7757876	77578	77578	767353 77578265	77578	77578	775782	775782	767411 7757826 5	775782	775782	775782	775782	7757876	766429 7757826 5	7757876	7757876	1	מספונון סונספון	7757276	7757876	7757876	7757876	7757876	7757826	7757826	7757826	7757826	766720 7757826 \$1

7757826 Suffuric acid sodium sait (1.2) Static 7757826 Suffuric acid sodium sait (1.2) Flow-through 7757826 Suffuric acid sodium sait (1.2) Flow-through 7757826 Suffuric acid sodium sait (1.2) Flow-through		Fresh water Lab	FED, MODERATELY HARD RECONSTITUTED V	Not reported 2 NR	N N		NR-LETH IN	Mortality
Sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab	FED, MODERATELY HARD RECONSTITUTED V	2	NR	Day(s)	NR-ZERO N	fortality
Sulfuric acid sodium salt (1:2)		Fresh water Lab		- 0	N N N		LCSO* N	fortality
sulfuric acid sodium salt (1:2)		Fresh water Lab		1	NR		LC50 N	lortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	E S	Day(s)	LCSO N	Mortality
Sulfuric acid sodium salt (1:2) Sulfuric acid sodium salt (1:2)	hrough Measured	Fresh water Lab		4	NR		NR-ZERO N	lortality
Sulfuric acid sodium salt (1:2)	hrough Measured	Fresh water Lab		4	NR		NR-ZERO N	Tortality
10-14 the mails on the call (19-2)	hrough Measured	Fresh water Lab		Not reported A NR	N N	Day(s)	NR-ZERO N	ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab	CA:MG RATIO 2.33	4	NR			Tortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab	CA:MG RATIO 2.33	4	NR			lortality
	Unmeasured	Fresh water Lab		4	NR	Day(s)		Mortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab	CA:MG BATIO 2:33	Not reported	N N			ortality
	Unmeasured	Fresh water Lab		4	NR			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	NR			lortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	NR			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	NR			Mortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab		Not reported 4 NR	NR		Į.	tortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab	CA:MG RATIO 2.33	Not reported 4 NR	N. S.	Day(s)	LC50 M	ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	rresh water Lab		t e	NA NA			Mortality
7757976 Sulfuric acid sodium salt (1.2) Static	Unmeasured	Fresh water Lab	CA-MG RATIO 2 33	Not reported	S SN	Dav(s)	1C50 M	Mortality
	Unmeasured	Fresh water Lab		4	NB			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	NR			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	NR			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		4	NR			Mortality
	Unmeasured	Fresh water Lab		Not reported 4 NR	NR		-	ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		Not reported 4 NR	NR		LCSO N	ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		Not reported	N N	Day(s)	V 050	ortality
	Onmeasured	Fresh water Lab		Not reported	N N		T	Mortality
	Measured	Fresh water Lab		Not reported	W. W.			rivival
	Manuscal	Frach woter Lab		Not reported	2			rvival
7757826 Sulfuric acid sodium salt (1:2) Static	Measured	Fresh water Lab	4 REPLICATES	Not reported 4 NR	N	Day(s)		Survival
	Measured	Fresh water Lab		Not reported 4 NR	NR		1	ırvival
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab	2.33	Not reported 4 NR	NR			ortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab		Not reported 4 NR	NR			ortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab	2.33	Not reported 4 NR	NR		LCS0 M	Mortality
T	Unmeasured	Fresh water Lab		Not reported 4 NR	N. N.			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		Not reported	N. S.	Day(s)	ICSO IN	Mortality
7757626 Sulfarit and sodium salt (1:2) Static	Massinad	Grach water Lab	TREDANIER CREEK WATER	Not reported	NR			origal
7757806 Sulfane acid sodium salt (1.2) Static	Unmeacured	Fresh water lah	97.0	Not reported	N.			Mortality
Sulfure acid sodium salt (1:2)	Unmeasured	ab		Not reported 4 NR	NR		LCS0 M	ortality
	Unmeasured	Lab		Not reported 4 NR	NR	Day(s)	LCS0 M	Mortality
	Unmeasured	Tab.		Not reported 4 NR	NR		LCS0 M	Mortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	del		Not reported 4 NR	N. N.		LCS0 M	Mortality
	Unmeasured	.ab		Not reported 4 NR	NR.		LCS0 M	ortality
Sulfuric acid sodium salt (1:2)	Unmeasured		CA:MG RATIO 2.33	Not reported 4 NR	N S	Day(s)	MOEC M	Mortality
775/826 Sulfuric acid sodium salt (1:2) Static	Measured	de.		Not reported	NN		NR.ZERO M	Mortalito
Т		Fresh water liab		Not reported	N. S.		NR-ZERO M	Mortality
7757826 Sulfuric acid sodium salt (1:2) Static	Measured	Fresh water Lab		Not reported 4 NR	NR	Day(s)		ortality
7757826 Sulfuric acid sodium salt (1;2) Static	Measured	Fresh water Lab	CA:MG MOLAR RATIO 7.0	4	NR	Day(s)	LC50 M	Mortality
	Measured	Fresh water Lab		4	NR		6	ortality
Sulfuric acid sodium	Measured	Fresh water Lab		4	NR	Day(s)	LC50 M	ortality
Sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab	CA:MG MOLAR RATIO 1.8	4	NR			ortality
sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab		4	N.	1		ortality
7757826 Sulfuric acid sodium salt (1:2) Static	Measured	Fresh water Lab		Not reported A NR	N N		J	Mortality
Sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab	TREPANIER CREEK WATER	1 4	N ON	Dav(s)	NOFC Su	rvival
7757876 Sulfunc acid codium salt (1:2) Static	Measured	Fresh water ligh		Vot reported	N. N.	Dav(s)	LCSO M.	ortality
ľ	Measured	Fresh water Lab		Not reported 4 NR	NR	Day(s)	LOEC M	ortality
7757826 Sulfuric acid sodium salt (1:2) Static	Measured	Fresh water Lab		Not reported 4 NR	NR	Day(s)		Mortality
7757826 Sulfunc acid sodium salt (1:2) Static	Measured	Fresh water Lab		4	NR			ortality
Sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab		4	N.		NR-LETH M	ortality
7757826 Sulfuric acid sodium salt (1:2) Static	Measured	Fresh water Lab		Not reported 4 NR	NR	Day(s)		Survival
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab	REFORMULATED MODERATELY HARD WATER	4	N N			ortality
Sulfuric acid sodium salt (1:2)	Unmeasured	Fresh water Lab		Not reported	N N	Day(s)	LCIU IN	Mortality
775/826 Suituric acid sodium sait (1:2) Static	ineasured in	Fresh water Lab		Not reported	W W	Day(s)	IOFC Su	rvival
Suituric acid sodium salt (1.2)	Unimedanie	Fresh water Lab		4 0	N N			ortality
Sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab		r	N. N.		LOEC Su	Survival
Т	Measured	Fresh water lab		2	N.S.			rvival
Sufficio acid sodium salt (1:2)	Measured	Fresh water Lab		4	NR			rvival
Sulfuric acid sodium salt (1:2)	Measured	Fresh water Lab		m	NR		F	rvival
-	Unmeasured	Fresh water Lab	REFORMULATED MODERATELY HARD WATEN	4	NR			Mortality
1 1	Measured	gen		S	NR			rvival
	Unmeasured	de.		4	NR			ortality
7757826 Sulfuric acid sodium salt (1:2) Static	Unmeasured	Fresh water Lab	CA:MG RATIO 1.46	Not reported 2 NR	NR		LCSO Me	Mortality
1	Unmpasured	ab		4 4	NR	Dav(s)		ortality

T66668         775728.6 Sulfurie and sodium salt (1.2)           766667         775728.5 Sulfurie and sodium salt (1.2)           766669         775728.5 Sulfurie and sodium salt (1.2)           766667         7757326 Sulfurie and sodium salt (1.2)           766668         7757328 Sulfurie and sodium salt (1.2)           766666         7757328 Sulfurie and sodium salt (1.2)           766667         7757328 Sulfurie and sodium salt (1.2)           8009         7757328 Sulfurie and sodium salt (1.2)           40509         7757328 Sulfurie and sodium salt (1.2)           13819         7757328 Sulfurie and sodium salt (1.2)           12607         7757328 Sulfurie and sodium salt (1.2)           126070         7757328 Sulfurie and sodium salt (1.2)           126070         7757328 Sulfurie and sodium salt (1.2)           126070         7757328 Sulfurie and sodium salt (1.2)           7757328 Sulfurie and sodium salt (1.2)<	Static Static Static	Unmeasured Unmeasured	Fresh water Lab	CAMOG RATIO 2.33	Not reported Not reported Not reported	2 NR NR Day(s) (CSO 4 NR NR DAY(s) (CSO 4 NR NR NR NR NR NR CSO 4 NR	NR NR NB	Day(s) Day(s) Day(s) Day(s)		Mortality Mortality
77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578	Static	Unmeasured	resh water Lab	CA:MG RATIO 1.46 CA:MG RATIO 2.33 CA:MG RATIO 2.33		4 A NR	NR NR	Day(s) Day(s)		Mortality
77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578			resh water Lab	CA:MG RATIO 2.33	Not reported	AINR	NR NB	Day(s) Day(s)	1050	
77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578			200	CA:MG RATIO 2.33		(1A) E	- N	Day(s)	1000	Mortality
77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578 77578			resh water Lau	CA:MG RATIO 2 33	Not reported	4 NR	N N	Daviel	1050	Mortality
77578 77578 77578 77578 77578 77578 77578 77578 77578 77578		Unmeasured	Fresh water Lab	CA:MG RATIO 1.46	Not reported	2 NR	Z.	Dav(s)		Mortality
7578 7578 77578 77578 77578 77578 77578 77578 77578 77578 77578	Static		resh water Lab	CA:MG RATIO 2.33	Not reported	2 NR	NR	Day(s)	1050	Mortality
77578 77578 77578 77578 77578 77578 77578 77578 77578	Ï		Fresh water Lab		Not reported	4 NR	NR	Day(s)	1050	Survival
77578 77578 77578 77578 77578 77578 77578 77578	Static		resh water Lab	COMPOSITION OF WATER REPORTED	Not reported	A NR	NR	Day(s)		Mortality
77578 77578 77578 77578 77578 77578 77578 77578	l		resh water Lab	COMPOSITION OF WATER REPORTED	Not reported	A NR	N. S.	Day(s)		Mortality
77578 77578 77578 77578 77578 77578 77577		Т	resh water Lab	o papaharreps see papen	Not reported	T NE	N N	Day(s)		Mortality
27578 27578 27578 27577 27578 27577 27577		Т	resh water Lab	2) SEL FAFEN	Not reported	N N	NB	Day(s)		Mortality
77578 27577 27578 27577 27577 27577	Static	Inmeasured	resh water lah		Not reported	4 4 4 A	NR	Davis		Mortality
287277 287277 287277 287277	Cratic		rach water lab		Notreported	A A	N.B.	Davie	0501	Mortality
77578 77578 77578 77578 77578	Static	Unmeasured	resh water lab		Not reported	- NB	NB	Davisi	NR.I FTH Mortality	Mortality
775782	Static	T	Fresh water Lab		Not reported	2 NB	NB	Daviel		Mortality
775782	Charie		rest water lab		Not reported	N N N	WD	Dayle)		Adortality
775782		T	Tiesii water Lab		Not reported	NAME OF STREET	NN NN	Day(s)		MOT LATELY
113/87	Static	Unmedsured	Fresh water Lab		Not reported	# INR	N. N.	Day(s)	000	Mortality
COLICE		T	rest water Lab		Not reported	2 0 2	ND	Dawlel		Mortality
6211 (/3/626) Sulfulle acid southill Sale (1.2)		T	rest water tab		Mot reported	NAME OF THE PARTY	ND ND	Davies		Modelity
1/2/87	Static	Unmeasured	resh water Lab	4 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	Datiodal ton	N. S.	N	Day(s)		Mortality
1/3/87		Unmeasured	resh water Lab	2 REPUCALES	Not reported	t in	INE	Day(s)		Mortality
0			resh water Lab	2 NEPLICATES	Not reported	SINK	N. N.	Day(s)		Mortality
- 1	Static	Unmeasured	Fresh Water Lab		Not reported	N. C.	NH.	Day(s)	1000	Mortality
8026/0 //5/826 Surfuric acid sodium sait (1:2)			resh water Lab	2 REPLICATES	Not reported	1 INK	NA.	Day(s)		Mortality
_ 1		1	resn water Lab		Not reported	INK	NH.	Day(s)		Mortality
- 1		Unmeasured	Fresh water Lab		Not reported	SINK	N. N.	Day(s)	1	Mortality
- 1	Static		resh water Lab		Not reported	4 NK	NR	Day(s)	1	Mortality
8025// //5/826 Sulfuric acid sodium salt (1:2)			resh water Lab		Not reported	Z NR	HN.	Day(s)	t	Mortality
			resn water Lab	Z KEPLICATES	Not reported	TN T	NH.	Day(s)	T	Mortality
41640 7757826 Sulturic acid sodium salt (1:2)	Static	Unmeasured	resh water Lab		Not reported	ZNK	N. S	Day(s)		Mortality
- 1			resh water Lab	2 REPLICATES	Not reported	2 NR	NR	Day(s)	П	Mortality
- 1			resh water Lab		Not reported	3 NR	N.	Day(s)	П	Mortality
	Static	Unmeasured	resh water Lab		Not reported	A NR	NA	Day(s)	Т	Mortality
802678 7757826 Sulfuric acid sodium salt (1:2)	Static		Fresh water Lab	2 REPLICATES	Not reported	1 NR	NR	Day(s)	007	Mortality
			resh water Lab		Not reported	2 NR	NR	Day(s)		Mortality
	Static	Unmeasured	resh water Lab	2 REPLICATES	Not reported	2 NR	NR	Day(s)	LC100	Mortality
			resh water Lab		Not reported	1 NR	N.S.	Day(s)		Mortality
			resh water Lab		Not reported	1 NR	NR	Day(s)		Mortality
	Static		resh water Lab		Not reported	2 NR	NR	Day(s)	LC50	Mortality
802672 7757826 Sulfuric acid sodium salt (1:2)		Unmeasured	Fresh water Lab	2 REPLICATES	Not reported	2 NR	NR	Day(s)	LC100	Mortality
	Static		resh water Lab		Not reported	3 NR	NR	Day(s)		Mortality
	Static	Ш	resh water Lab	2 REPLICATES	Not reported	T N T	NR	Day(s)		Mortality
	Static		resh water Lab		Not reported	3 NR	NS NS	Day(s)	i,	Mortality
802674 7757826 Sulfuric acid sodium salt (1:2)	Static	Unmeasured	resh water Lab	2 REPLICATES	Not reported	4 NR	NR	Day(s)	100101	Mortality
	Static		resh water Lab	2 REPLICATES	Not reported	3 NR	NR	Day(s)		Mortality
			resh water Lab		Not reported	A NR	NR	Day(s)	1050	Mortality
775782		Unmeasured	resh water Lab	2 REPLICATES	Not reported	3 NR	N.	Dav(s)	LC100	Mortality
775782	Static		resh water Lab		Not reported	TNR NR	N.	Dav(s)	1,050	Mortality
775787			rech water Lab		Not reported	4 NB	N.S.	Dav(s)		Mortality
775782		L	resh water Lab	2 REPLICATES	Not reported	4 NR	NR	Dav(s)	007	Mortality
775782	Renewal	1	resh water Lab		Not reported	14 NR	NR	Day(s)	NR-LETH	Mortality
775787			resh water Lab	3 REPLICATES	Not reported	7 NR	NR	Dav(s)	NOEC	Survival
775782			resh water Lab		Not reported	7 NB	NB.	Dav(s)	NOEC	Survival
761509 7757906 Guifuric acid codium sait (1.5)	Penewal		Fresh water lab		Not reported	7 NB	NB NB	Day(s)	NOFC	Mortality
775783			resh water lah	3 REPLICATES	Not reported	7 NB	2	Davis		Survival
201511	1		Fresh water Lab		Not reported	AND T	NID	Daviel	2000	Survival
113/82/			rest water Lab		Not reported	ANI /	NIO	Day(s)	2000	Cuminal
201/210 //5/826 Sulfund acid sodium salf (1:2)	Kenewal		resh water Lab	3 REPUCATES	Not reported	A NR	NR.	Day(s)	5773	Survival
1757826			resh water Lab		Not reported	N. I	NR.	Day(s)	NOEL	Survival
			resh water Lab	3 REPLICATES	Not reported	NR	NK	Day(s)	ECSO	Survival
775782	ugh	1	resh water Lab		Not reported	AN AN	NK	Day(s)	NK-ZEKO	Mortairty
775782	ugno		resh water Lab		Not reported	ANR ANR	NR	Day(s)	NR-ZERO	Mortality
775782	Flow-through		Fresh water Lab		Not reported	A NR	NR	Day(s)	NR-ZERO	Mortality
775782			resh water Lab		Not reported	7 NR	NR	Day(s)	LOEC	Survival
775782	Renewal		resh water Lab	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC10	Survival
775782			resh water Lab		Not reported	7 NR	NR	Dav(s)	NOEC	Survival
775797			rach water lah		Not reported	2 NB	NB	Daviel	EC50	Survival
201011	neilewal		207		7		NIO.	Davie)	2000	Completed
17.5 77.526 Sulfuric acid souldin salt (1.2)	Reliewal		LIESH MAIEL LAD		national los	UNI C	NA.	Dayls)	0	on vival
115/82	ongn		resn water Lab		Not reported	AN 4	NK	Day(s)	2	Mortality
7757826			Fresh water Lab		Not reported	7 NR	NR	Day(s)		Survival
2017188 7757826 Sulfuric acid sodium salt (1:2)	Renewal		Fresh water Lab	3 REPLICATES	Not reported	7 NR	NR	Day(s)	EC10	Survival
7757826			resh water Lab		Not reported	7 NR	NR	Day(s)		Survival
775782	,		resh water Lab	TREPANIER CREEK WATER	Not reported	7 NR	NR	Day(s)		Survival
775782			resh water Lab		Not reported	4 NR	NR	Dav(s)		Mortality
767598 7757826 Sulfuric acid sodium salt (1:2)		Г	esh water Lab	TREPANIER CREEK WATER	Not reported	7 NR	NR	Dav(s)		Survival
(c.t) starting the print of the	1	1	Frach water Lab		Notreported	a N	NB	(a) (a)		Survival
7.17 Jupe Highland acid south (1.17)		1	esti water Lab		Not reported	200	UNI.	Day(s)	T	SUIVING
115/82			esh water Lab		Not reported	/ NK	NK	Day(s)		survival
775782		Measured	esh water Lab	3 REPLICATES	Not reported	ZINB	NR.	Day(s)	NOEC	Survival
775782		П	esh water Lab		Not reported	7 NR	NR	Day(s)		Survival
- 1	Renewal		esh water Lab	3 REPLICATES	Not reported	7 NR	NR	Day(s)		Survival
7757826 Sulfuric acid sodium salt (1:2)	ugno	Measured F	Fresh water Lab		Not reported	4 NR	NR	Day(s)	ERO	Mortality
7757826 Sulfuric acid sodium salt (1:2)	Flow-through I		esh water Lab	3 REPLICATES	Not reported	21 NR	NR	Day(s)		Hatch
7757826 Sulfuric acid sodium salt (1:2)	Flow-through 7		esh water Lab		Not reported	21 NR	NR	Day(s)		Survival
sulfuric acid sodium salt (1:2)	Static	p	esh water Lab		Not reported	2 NR	NR	Day(s)		Mortality

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Observed Durat Endpoint Day(s) NR-ZERO Day(s) LC50	NR-LETH	UC50	NR-ZERO	1050	1050	1050	LCSO	1050	0521	1050	1050	NR-LETH	NR-ZERO	NR-ZERO	NR-ZERO	LC50	1050	LC30	NR-I FTH	NR-LETH	NR-LETH	NR-LETH	NR-LETH	NR-LETH	NR-LETH	LCSO	1050	1050	NR-LETH	NR-LETH	NR-ZERO	1C50*	NR-ZEKO	1,050	NR-7FRO	1050	LCSO	1050	1050	LC50	1050	CSO	NOEC	LOEC	1050	1C50	1050	1050	NR-ZERO	NR-ZERO		NR-ZERO	NOEC	NOEC	1050	1050	1,050			LC50	NR-LETH	NR-LETH	NR-LETH	NR-LETH	NR-LETH	1050	1050	NR-LETH	NR-LETH	NR-LETH	NR-LETH	NR-LEID	ioer	1	NOEC	
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Experimental Design CA, MG, NA+K, SO4, CL2	FED, MODERATELY HARD RECONSTITUTED	EED MODERATELY HARD RECONSTITUTED	FED, MODERATELY HARD RECONSTITUTED									IODERATELY HARD RECONSTITUTED	FED, MODERATELY HARD RECONSTITUTED V	IODERATELY HARD RECONSTITUTED	ODERATELY HARD RECONSTITUTED																	WATER CHEM PARAMETERS RPTD		OTOG MADE OBTOWN MONTHS	COLUMN WALLEN CHICKING AND COLUMN					CATES	CATES	CATES	CATES	CATES							RECIRCULATING		TWO REPLICATES, NO CONTROL DATA AFTE	NO CONTROL DATA AFTE										ERATELY HARD RECONSTITUTED	ERATELY HARD RECONSTITUTED											
ocation	FED, N	S CPR	FED, N		1							FED, N	FED, N	FED, N	FED, M		1															WATER		CONADO	- Carlon					4 REPLI	4 REPLI	4 REPLI	A DEDI-	4 REPLICATES							RECIRC		TWOR	TWOR										FED, M	FED, MOD				REP 3	REP 7		REP 10		-		
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Calcium	Calcium	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCI2)	4 Calcium chloride (CaCl2)	Calcium chloride (CaCI2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (Cacio)	Calcium chloride (CaCl2)	4 Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	4 Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	calcium chioride (Laciz)	Calcium chloride (CaCio)	Palcium chloride (CaCI2)	Calcium chloride (CaCl2)	Salcium chloride (CaCl2)	Calcium chloride (CaCl2)	Salcium chloride (CaCl2)	Salcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Laicium chioride (Caciz)	Calcium chloride (Caciz)	Jalcium chloride (CaCi2)	alcium chloride (CaCI2)	algium chloride (CaCl2)	4 Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Algina chloride (Caciz)	d Calcium chloride (CaCl2)	alcium chloride (CaCl2)	algium chloride (CaCl2)	Calcium chloride (CaCI2)	ि	Calcium chloride (CaCI2)	Salcium chloride (CaCI2)	Jaicium chloride (CaCl2)	Jaicium chloride (CaCl2)	4 Calcium chloride (CaCl2)	Calcium chloride (CaCl2)	Salcium chloride (CaCI2)	Social chloride (CaCl2)	alcium chloride (CaCI2)	alcium chloride (CaCl2)	alcium chloride (CaCl2)	10043524 Calcium chloride (CaCl2)	alcium chloride (CaCl2)	alcium chloride (CaCl2)	alcium chloride (CaCl2)	alcium chloride (CaCl2)	10043524 Calcium chloride (CaCl2)	alcium chloride (CaCIZ)	4 Calcium chloride (CaCl2)	alcium chloride (caciz)	aloum chorde (cacia)	alcium chioride (caciz)											
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LOEC Hatch	ERO Mortality	TH Mortality	ERO Mortality	ERO Mortality	ERO Mortality	ETH Mortality	Proseny counts/number	Propeny counts/number	Fecundity	ď		* Reproduction, general	* Reproduction, general	-	riogeny counts/numb			Beat/Cross frequency			Motility		Beat/Cross frequency		Linearity		Motinity Maximum amplifude o	Velocity	Motility	Motility			۵	T	Progeny counts/numb                      Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Reproduction, general	Reproduction, general	Reproduction, general	Reproduction, general	Pregnant, Paris or Grav	Gamete production	Progeny counts/numbi	Progeny counts/numbe	Progeny counts/numbe	Progeny counts/numbr	Progeny counts/number	Progeny counts/number	Progeny counts/number	Progeny counts/numbe	Progeny counts/numbe	Progeny counts/numb	Reproducing organism	Reproducing organism	Reproducing organism	Reproducing organism	Progeny counts/numbi	Progeny counts/numbi	Progeny counts/numbi	Progeny counts/numbi	Progeny counts/numb	Progeny counts/numbi	Reproduction, general	Parameter appear								
Ved Durat Endpo	NR-ZE	NR-LE	NR-ZE	NR-ZE	NR-ZE	NR-LE	NOEC	1028	TOET	LOEC	NOEC	ECS0.	ECSO.	LOEC	NOF	LOEC	LOEC	NOEC	NOEC	NOEC	NOEC	NOFC	NOEC	NOEC	NOEC	NOEC	NOEC	LOEC	LOEC	NOEC	NOEC	NOEC	NOEC	NOEC	050	IC20	1010	NOEC	NOEC	0770	1750	1010	LOEC	NOEC	NOEL	1025	IC10	NOEC	LOEC	TOEC	NOEC	NOEC	LOEC	DEC	NOEC	TOEC	NOEC	LOEC	LOEC	NOEC	1050	NOEC	LOEC	NOEC NOEC	LOEC	LOEC	NOEC	LOEC	NOEC	NOEC	LOEC	1025	
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1.5 NR	NR.	0.0138 NR	1.5 NR	1.5 NR	1 NR		7 NB	7 NR	12 NR	7 NR		21 NR	21 NR	ZI NK	O DOOD INR	0.0002 NR	0.0002 NR	0.0001 NR	0.0002 NR	0.0001 NR	0.0001 NR	O DODOS NR	0.0002 NR	0.0001 NR	0.0003 NR	0.0002 NR	D.OOOT NB	0.0001 NB	0.0001 NR	0.0002 NR	0.0002 NR	0.0003 NR	98 NR	98 NR	A A A	4 NR	4 NR	21 NR	21 NR				98 NR	98 NR	4.5417 NR	2 NR	2 NR	2 NR	2 NR	80 NR	80 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NB	7 NR	7 NR	7 NR	3 NR	7 NR	7 NR	ZINK	7 NB	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	
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experimental pesign							C WALLE		IOREPLICATES			SEE PAPER																					BIOMPHALARIA ALEXANDRINA PRE-EXPOSE	BIOMPHALARIA ALEXANDRINA PRE-EXPOSE	THREE REPLICATES	THREE REPLICATES	TWO-FOUR REPLICATES, 9 MG:1 CA RATIO			TWO-FOUR REPLICATES, 9 MG:1 CA RATIO	MIG.1 CA RATIO		ALEXANDRINA PRE-EXPOSE	BIOMPHALARIA ALEXANDRINA PRE-EXPOSE		8 REPLICATES			8 REPLICATES													TEST 4	TEST 4		TEST E							HARDNESS OF 80 MG/L CACO3	
Fresh water Lab	esh water Lab	esh water Lab	esh water Lab	esh water Lab	esh water Lab	sh water Lab	Frach water Lab	sh water Lab	esh water Lab	esh water Lab	esh water Lab	ssh water Lab	Fresh water Lab	ssh water Lab	Fresh water Lab	sh water Lab	sh water Lab	Fresh water Lab	sh water Lab	Fresh water Lab	sh water Lab	sh water Lab	sh water Lab	Fresh water Lab	Fresh water Lab	sh water Lab	Fresh water Lab	Sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	Fresh water Lab	sh water Lab	Fresh water Lab	sh water Lab	sh water Lab	Fresh water Lab	qe-	Fresh water Lab	OP.		sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	Fresh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	sh water Lab	Fresh water lab	sh water Lab	sh water Lab	sh water Lab	Fresh water Lab			sh water Lab	sh water Lab	resh water Lab	sh water Lab	sh water Lab	Fresh water Lab			
Measured	Unmeasured	Unmeasured	Unmeasured Fre	Unmeasured	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Measured Fre	Measured Fre	Unmeasured	Unmeasured	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Measured Fre	Measured Fre	Measured Fre	Unmeasured Fre	Unmeasured Fre	Measured hre	Measured Fre	Measured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Measured Fre	Measured Fre	Measured Fre	Measured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unimpasiired Fre	Unmeasured Fre	Unmeasured Fre	Measured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Fre	Unmeasured Free	Unmeasured Free	Unmeasured Free	Measured Free																					
Static	Static	Static	Static				-	Renewa			Renewal	- 1	Renewal		-1		1				- 1	-		1			- 1				1	Static	ш	Renewal	(I) Renewal	(1) Renewal	(11) Renewal	(1) Renewal	:1) Renewal	T) Renewal	1) Renewal	11 Renewal	:1) Renewal	:1) Renewal	Static	Static	Static	Static	Static	Renewal		Ш		П	Renewal			Renewal				Static		П	П			Renewal			П		
Number Chemical Name 0043524 Calcium chloride (CaCl2)	0043524 Calcium chloride (CaCl2)	10043524 Calcium chloride (CaCl2)	0043524 Calcium chloride (CaCl2)	0043524 Calcium chloride (CaCI2)	0043524 Calcium chloride (CaCl2)	0043524 Calcium chloride (CaCl2)	2447407 Deserving chloride (cenz)	7447407 Potassium chloride (KCI)	744/40/ Potassium chloride (KC)	7447407 Potassium chloride (KCI)	744/407 Potassium chloride (KCI)	744/407 Potassium chloride (KCI)	7447407 Potassium chloride (KCI)	7447407 Potassium chloride (KCI)	7447407 Potassium chloride (KCI)	9 7447407 Potassium chloride (KCI)	7447407 Potassium chloride (KCI)	7447407 Potassium chloride (KCI)	748/889 Sulfuric acid magnesium sait (1	7487889 Sulfuric acid magnesium salt (1	748/889 Sulfuric acid magnesium sait (1	7487889 Sulfurc acid magnesium salt (1	7487889 Sulfuric acid magnesium salt (1	7487889 Sulfuric acid magnesium salt (1	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	1647145  Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl) Static	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	'647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	764/145 Sodium chloride (NaCl)	647.145 Sodium chloride (NaCl)	(647145 Sodium chloride (NaCl)	(647145) Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	'647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	7647145 Sodium chloride (NaCl)	SA7145 Sodium chloride (NaCl)	647145 Sodium chloride (NaCl)	647145 Sodium chloride (NaCl)	647145 Sodium chloride (NaCl)	'647145 Sodium chloride (NaCI)	7647145 Sodium chloride (NaCl)	647145 Sodium chloride (NaCl)	647145 Sodium chloride (NaCI)																								
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Effect Measurement Progeny counts/numb	Progeny counts/num	Progeny counts/num	Progeny counts/num	Progeny counts/num	Progeny counts/num	Reproduction, genera	Reproduction, general	Progeny counts/num	Progeny counts/num	Progeny counts/num	Reproduction, genera	Reproduction, genera	Progeny counts/num	Reproduction, genera	Reproduction, genera	Reproduction, genera	Reproduction, genera	Progeny counts/num	Reproducing organisr	Progeny counts/num	Reproducing organism	Progeny counts/num	Progeny counts/num	Reproducing organisr	Progeny counts/num	Reproducing organism	Reproducing organist	Reproducing organisr	Reproducing organisr	Reproducing organism	Reproducing organism	Reproducing organish	Progeny counts/numi	Progeny counts/numi	Progeny counts/numi	Reproducing organism	Reproducing organism	Reproducing organism	Reproducing organism	Reproducing organism	Reproducing organism	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numk	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Reproduction, genera	Reproduction, genera	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Progeny counts/numb	Reproduction, general	Reproduction, general	rogeny counts/numb	Togethy counts/memb					
Endpoint	NOEL	EC50	TOET	LOEC	NOEC	1025	NOED	10FC	LOEC	NOEC	1025	1050	ICSO	(CS0	1025	1C25	1050	NOEC	LOEC	ECSO	ECSO	NOEC		LOEC	NOEC	1050	NOEC	NOEC	1050	1025	1050	NOEC	LOEC	LOEC	NOEC	1050	1050	1025			Ü		ICSO			NOEC	1025	NOEC	(C2S	1050	ICS0			NOFC	LOFC	NOF	IOFC	LOEC	NOEC	LOEC	NOEC	LOEC	NOEC	LOEC	NOEC	NOEC					NOEC	1025	1050	LOEC		LOEC			NOEC I	П	
Observed Durat Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Davie)	Daviel	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Dav(s)	Dav(s)	Day(s)	Day(s)	Day(s)	Day(s)	Jay(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Day(s)	Dav(s)	Day(s)	Day(s)	Oay(s)	Day(s)	Day(s)	Day(s)	Jay(s)	Day(s)	Jay(s)	Day(s)	Say(s)	(s) (a)	(s)\ec	Jay(s)	Day(s)	Jay(s)	Day(s)	Day(s)	Davie	lavíc)	Dav(s)	Dav(s)	Day(s)	(s)(a)	Day(s)	Day(s)	(s) (a)	Day(s)	Day(s)	(s) (a)	Day(s)	Day(s)	ay(s)	ay(s)	Day(s)	Day(s)	Day(s)	ay(s)	Day(s)	Day(s)	ay(s)	ay(s)	Day(s)	Day(s)	ayls)
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Days Duration Min (Days Duration R	П	R NR	Z			N S			2	2	Z	N	N	Z		NR NR	N	NR	1	Z	Z	N	N N	S.	N N	Z.	N.	N.	Z.	Z	Z	Z		N. N.	N N	ž	Z	N	N	N.	Z	ž		N.		NB			ا و	9	0	ں م	0 4	o a	2 2		N.		NR	NR	NR	NR	NR		NR	NR	NR	NR	NR			NR	RN			S. S.			S S		
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Response Site Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported	Not reported
Experimental Design						10 REPLICATES	10 her dichies				HARDNESS OF 10 MG/L CACO3	HARDNESS OF 10 MG/L CACO3		HARDNESS OF 160 MG/L CACO3	HARDNESS OF 160 MG/L CACO3	HARDNESS OF 320 MG/L CACO3	HARDNESS OF 320 MG/L CACO3	CHRONIC -3 BROOD STDY	CHRONIC -3 BROOD STDY	CHRONIC -3 BROOD STDY		10.REPLICATES	10 REPLICATES	TEST 7		TEST 5			TEST 12		TEST 1	TEST 11	TEST 11	TEST 8	TEST 8	EST 4	TEST 2				TEST 5	TEST 9	TEST 6	TEST 10	TEST 2	FEST 2			10 REPLICATES	10 REPLICATES	10 REPLICATES	IOREPLICATES	10 REPLICATES	IO REPLICATES																		HARDNESS OF 40 MG/L CACO3	HARDNESS OF 40 MG/L CACO3					HARDNESS OF 20 MG/L CACO3	ARDNESS OF 20 MG/L CACUS		TEST MATERIAL 3
Test Location	er Lab	er Lab	er Lab	er Lab	er Lab	er Lab	r Lab	r lah	er Lab	r Lab	er Lab	er Lab	Lab	de.	de.	de.	de.	.ab	r Lab	r Lab	r Lab	Lab	Lab	de.	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	r Lab	Lab	Lab	Lab	Lab	r Lab	r Lab	r Lab	Lab	Lab	Tab	de	ab	de de	Lab	Lab	Lab	Lab	Lab	Lab	Lab	Lab	r Lab	- Lab	- Lab	Lab	Lab	Lab	gen	Lab	de.	ge.	.ab	rab	. Lab	Lab	de.	de.		Lab					
Fresh water I	Fresh wate	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Grach wate	Frach water	Fresh water	Fresh wate	Fresh wate	Fresh water	Fresh water	Fresh water	Fresh water	Fresh wate	Fresh wate	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh wate	Fresh water	Fresh wate	Fresh water	Fresh water	Fresh water	Frach water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water	Fresh water Lab	Fresh water Lab	Fresh water
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Exposure Type Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Static	Renewal	Static	Renewal	Renewal	Static	Renewal	Static	Static	Static	Static	Static	Static	Static	Renewal	Renewal	Renewal	Static	Static	Static	Static	Static	Static	Renewal	Renewal	Static	Static	Static	Static	Static	Static	Reciewal	Denemal	Ranawai	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Nenewai
Chemical Name Sodium chloride	Sodium chloride (NaCl)	Sodium	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCi)	Sodium chloride (NaCI)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	poo	Sodium chloride (NaCl)	Sodium chloride (NaCl)	pos	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCi)	Sodium chloride (NaCl)			Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCI)	Sodium chloride (NaCI)	5 Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodiun	Sodium chloride (NaCI)	Sodium	Sodium chloride (NaCI)	5 Sodium chloride (NaCl)	Sodium chloride (NaCI)	5 Sodium chloride (NaCl)	Sodium chloride (NaCI)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	odium chloride (NaCI)	odium chloride (NaCI)	Sodium chloride (NaCl)	5 Sodium chloride (NaCl)	odium chloride (NaCl)	Sodium chloride (NaCl)	odium chloride (NaCl)	odium chloride (NaCl)	Sodium chloride (NaCI)	S Sodium chloride (NaCl)	odium chloride (NaCl)	Sodium chloride (NaCl)	sodium chloride (NaCl)	sodium chloride (NaCl)	sodium chloride (NaCl)	Sodium chloride (NaCl)	Sodium chloride (NaCl)	odium chloride (NaCI)	sodium chloride (NaCI)	7647145 Sodium chloride (NaCl)	odfum chloride (NaCI)	odium chloride (Naci)										
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Result Number 0	2152726	2152725	2152727	767893	767894	2152721	7575731	767876	767858	767859	2152855	2152873	767552	2153276	2153275	2153278	2153279	741688	742568	741661	207720	2146818	2146819	741952	761594	741936	761578	761586	741950	761580	742564	741953	742900	741944	742898	741951	741929	761575	761574	761587	741937	742892	742894	742895	741957	742891	761576	761590	2148464	2148463	2148479	2148480	2148473	21484/4	10/0/2	250/0/	767955	767850	767851	766862	766863	767866	767867	766870	756871	767892	767891	767885	767886	767963	767964	2153269	2153270	767899	767900	767965	767966	2153267	2152881	767974	10/2/07

Int Effect Measurement Progeny counts/numbr	Progeny counts/numbi	Progeny counts/number	Progeny counts/numb	Progeny counts/numbe	Progeny counts/number	Progeny counts/numbe	Progeny counts/number	Progeny counts/numbe	Progeny counts/number	Progeny counts/numbe	Progeny counts/numb	Progeny counts/numb	Progeny counts/numbe	Progeny counts/numb	Progeny counts/numbe	Progeny counts/number	Progeny counts/numbe	Progeny counts/numb	Progeny counts/numb	Progeny counts/numbe	Progeny counts/numbe	Progeny counts/numbr	Progeny counts/numbi	Reproducing organism	Progeny counts/number	Progeny counts/numbe	Reproducing organism Progeny counts/numbe	Progeny counts/numbe	Progeny counts/numbe	Progeny counts/number	Reproducing organism	Reproducing organism	Progeny counts/numbr	Progeny counts/numbe	Reproducing organism	Reproducing organism	Progeny counts/number	Progeny counts/number	Progeny counts/numbe	Reproducing organism	Reproducing organism	Reproducing organism	Reproducing organism	Reproducing organism	Fecundity	Progeny counts/numbs	Progeny counts/numb	Progeny counts/number	Progeny counts/numbi	Fecundity	Fecundity	Fecundity	Fecundity	Fecundity	Fecundity	Fecundity	Progeny counts/numbe	Progeny counts/numbe	Progeny counts/number	Reproduction, Series
20	ICSO	050	NOEC	LOEC	NOEL	1050	1050	1050	LOEC	1050	NOEC	NOEC	1050	NOEC	1050	1050	LOEC	NOEC	NOEC	1050	1050	LOEC	LOEC	NOEC	NOEC	1025	NOEC	1025	NOEC	NOEC	NOEC	LOEC	1025	1050	NOEC	LOEC	IC25	NOEC	1025	CSD	NOEC	LOEC	LOEC	NOEC	NOEC	LOEC	ECS0	NOEC	ECSO	NOEC	NOEC	NOEC	NOEC	LOEC	OEC	LOEC	NOEL		J.	V
y(s)	y(s)	V(s)	Day(s)	Day(s)	Day(s)	y(s)	y(s) y(s)	y(s)	Day(s)	y(s)	Day(s)	Day(s)	y(s)	(s) (s)	V(s)	y(s)	V(s)	Day(s)	V(S)	Day(s)	Day(s)	Dav(s)	Day(s)	(5)/	Day(s)	/(s)	/(s) /(s)	(s)	/(s)	(s)	(s)/	(s)	(s)	(s)/	/(s) /(s)	(s)	(s)	(s) (s)	(s)	(s) (s)	(s)	(8)	(s)	(5)	(s)	(s)	(s)	(s)	(5)	od or litter	od or litter	od or litter	od or litter	Brood or litter	od or litter	Brood or litter	(s)	(s)	(s)	(5)
an Duration Min (Days Duration Max (Days)   Observed Duration 2   NR   NR   NR   Day(s)   LOE	Da	Da	Da	Da	EQ	Da	Da	Da	Da	Da	Da	Da	Da	Da	Da	B C	Da	Da	P C	Da	Da	Da	Da	Da	Da	Da	Davis)	Da	Day(s)	Day	Day	(e)	Day	Dai	Day	Day	Day(s)	Day	Day(s)	Day	Day(s)	Day(s)	Day(s)	Day	(eQ	Day(s)	Day(s)	Day(s)	Day(s)	Brood	Bro	Bro	Bro	Bro	Bro	Bro	Day	Day(s)	Day(s)	
(Days Dura	NR.	Z Z	S	NR ON	N. S.	S S	N N	N S	N N	a z	Z Z	NR	N. S.	NR	N.	Z Z	N.	N.	Z Z	N.	NR.	N N	NR	N N	N.	N S	N N	N.	N ON	N. N.	NR	N N	N.	N.	N N	N.	N N	N. S.	N S	NR NR	NR	N N	N.	N N	S.	N N	S.	S S	S S	N.	N N	N.	Z 2	Z Z	N.	Z Z	NR	NR.	N N	- CAN
Duration Mean Duration Mir	7 NR	7 NR	7 NR	7 NR	9 NR	7 NR	7 NR	7 NB	7 NR	7 NR	N N N	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	10 NR	10 NR	10 NR	10 NR	SINR	O NR	AN A	N S S S S S S S S S S S S S S S S S S S	4 NR	3 NR	N N N	10 NR	10 NR	10 NR	
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TEST MATERIAL 3	TEST 2, TEST MATERIAL 1	TEST 2, TEST MATERIAL 3	ATERIAL 3	ATERIAL 1	SROOD TEST	TEST MATERIAL 1	rest 1, test intalerial 1	FEST 2, TEST MATERIAL 3	TEST MATERIAL 3	EST MATERIAL 1	TEST MATERIAL 3	TEST MATERIAL 1	EST MATERIAL 3	TEST MATERIAL 1	LAB B, TEST MATERIAL 3	TEST 1, TEST MATERIAL 1	ATERIAL 1	TEST MATERIAL 1	ATERIAL 1	LAB D, TEST 1, TEST MATERIAL 3	EST MATERIAL 3	LAB B, TEST MATERIAL 1	TEST MATERIAL 1																							C -3 BROOD STDY	C -3 BROOD STDY	CHRONIC -3 BROOD STDY	ROOD TEST	CATES	CATES	CATES	CATES	CATES	CATES	CATES			- Carrier	
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	7 NR	NN V	21 NR	14 NR	14 NR	14 NR	14 NR	7 NB	7 NR	7 NR	7 NR	7 NR	7 NR	7 NR	GZ.	21 NR	21 NR	21 NR	21 NR	21 NR	21 NR	98 NR	80 4 RN RN	21 NR	0 NR	ONR	9 NR	9 NR	2 N N N N N N N N N N N N N N N N N N N	7 NR		21 NR	10 NR	10 NR	0.0417 NR	O.O417 NR	1	0.0417 NR	O OA12 NR		0.0001 NR	0.0002 NR	0.0003 NR	0.0001 NR	0.0002 NR	0.0001 NR	0.0001 NR	0.0001 NR														
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MODERATELY HARD WATER	MODERATELY HARD WATER											THOUSE AND STREET OF STREET OF STREET	FED ALGAE, ANKISTRODESMUS CONVOLUTI	FED ALGAE, ANKISTRODESMUS CONVOLU	FED ALGAE, ANKISTRODESMUS CONVOLUTI	RODESMUS	FED ALGAE, ANKISTRODESMUS CONVOLUTE	4 REPLICATES, SEDIMENT	4 REPLICATES, SEDIMENT	4 REPLICATES, SEDIMENT	4 REPLICATES, SEDIMENT	4 REPLICATES, SEDIMENT		TREPANIER CREEK WATER	TREPANIER CREEK DILUTION WATER				3 REPLICATES	3 REPLICATES	3 REPLICATES	3 REPLICATES	CATACO SPANOAL GOS CIOAC	CACLE TON DANDINESS CONTROL		BIOMPHALAREA ALEXANDRINA PRE-EXPOS	BIOMPHALAREA ALEXANDRINA PRE-EXPORTANO TWO REPLICATES	SEE PAPER							3 REPLICATES OF DOSED, 2 REPLICATES OF						BUFFERED TAP WATER	BUFFERED TAP WATER										
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Renewal		П	Renewal							П							Henewal	Renewal		1	Renewal		1	Renewal	Renewal	Renewal	Renewal	Renewal	Flow-through	Flow-through	III.		Renewal				Renewal	Renewal	Static	Static	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Renewal	Static	Static	Static	Static										
826 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	826 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	826 Sulfuric acid sodium salt (1:2)	826 Sulfuric acid sodium salt (1:2)	256 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1;2)	326 Sulfuric acid sodium salt (1:2)	S26 Sulfuric acid sodium salt (1:2)	326 Sulturic acid sodium salt (1:2)	256 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	126 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	S26 Sulfuric acid sodium salt (1:2)	26 Sulfuric acid sodium selt (1:2)	326 Sulfuric acid sodium salt (1:2)	126 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	326 Sulfuric acid sodium salt (1:2)	26 Sulfuric acid sodium salt (1:2)	226 Sulfuric acid sodium salt (1:2)	(26 Sulfuric acid sodium salt (1:2)	7826 Sulfuric acid sodium salt (1:2)	26 Sulfunc acid sodium salt (1:2)	26 Sulfuric acid sodium salt (1:2)	89 Sulfuric acid, Calcium salt (1:1)	89 Sulfuric acid, Calcium sait (1:1)	89 Sulfuric acid, Calcium salt (1:1)	89 Sulfuric acid, Calcium salt (1:1)	89 Sulfuric acid, Calcium sait (1:1)	03 Magnesium chloride	03 Magnesium chloride	Os Magnesium chloride	24 Calcium chloride (CaCI2)	24 Calcium chloride (CaCI2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCI2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	24 Calcium chloride (CaCl2)	34 Calcium chloride (CaCl2)	M Calcium chlorida (Cacta)				
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Raw Data from ECOTOX (Unfiltered)

mment Conc 1	Total	Total	Total	Total	Total	Total	Total	Total							T/AVE FINAL DRY WT/// Total	I		Total				l					768094 768095 Total						Total	Total	Total	Total	Total	Total	Total	Total	Total	Total	Total	Total	Total		ISMT/CONC/ AVERAGE 7 DAY CONCENTRATOTAL	Total	Total	/FORK LENGTH/ Dissolve	Dissolve	Total	Total	Total	Total	Total
ypeConc 1 Mean Conc 1 Mean (State				1090000	1000000	340000	1470000	1000000	1430000	1090000	1000000	2070000	4000000	4000000	4000000	4000000	1250000	2500000	1250000	1250000	2500000	1250000	2500000	2500000	250000	5000000	2500000	2500030	2500000	2000000	7500000	250000	1250000	2500000	1250000	958000	558000	1058000	1250000									1000000					2500000		2500000	2500000
Ann 1 (Stank	NR	N N	NR	N. N.	NR	N. S.N.	S. O.	NR	N N	NR	Z Z	NR	N N	NR	a a	NR	Z Z	NR	N N N	NR	N.S.	Z Z	NR	NR	NR.	N. C.	NR NR	NR.	N. N.	S. S.	N N	NR	N. N.	NR		000007		N. N.	NR	SN S	N N	N.	X XX	NR	W 22	NR	NR NR	NR NR	N N	NR	NR	N. N.	NB	NR NR	NR	NR
	NR.	N N	NR	N N	NR	N. N.	NR	NR	N N	NR	Z Z	NR	Z Z	NR	N N	N.	as as	N.	N N N	N. N.	NR	N N	NR	N. S.	W. W.	NR.	NR	N S	NR NR	N.	N. N.	N.	N N	N. S.				NR SE	N N	N. I.	N N	W I	X X	N.	N N	N.	or o	NR	N S	NR	NR NR	NR	NR	NR NR	NR	NR
Juits (Conc. 2	1/8n	% na/L	ng/L	ug/L	ug/L	ug/L	Ug/L	ng/L	ug/L	1/8n	ug/L	1/8n	Ug/L	1/8n	ug/L	ng/L	ug/L	ug/L	ng/L	1/8n	7/8n	ug/L	ng/L	1/8n	ng/L	ug/L	1/Sn	ug/L	1/8n	1/8n	1/8n	ug/L	7/80	ng/L	ug/L	1582000 ug/L	ug/L	ug/L	1/8/1	ug/L	1/8n	ng/L	ug/L	ng/L	1/8n	ng/L	Ug/L	ug/t	ug/L	ng/L	ng/L	ng/r	1/80	ug/t.	ng/L	1/8n
Type (Stan Conc 2 Mean (StaConc 2 Units (Standa Application Rate Application Rate Units																							7																																	
(Standa Application																																																								

10.000   1.0	68060 68062	Total	2500000	N N		NR NR	ng/L			
	68063	Total	2000000	N S			1/Bn			
Triangle    47196	Total	284000	Z Z			1/80				
Triple   No.   N	47204	Total	284000	N. S.			1/Bn			
Total   Not   No	54834 hly)	Total	NR	UZ.	2130000	854000	1/2n 0 ng/r			
Total   1970	54835	Total		3	2130000	854000	0 ug/L			
Total   Fire	34264 WSW1/LENGTH///	Total	2700	2 2			ng/r ng/r			
Total	50460 MSMT/WIDTH///	Total	20007	NR			ng/L			
Total   Tota	10560 MSMT/LENGTH///	Total >	7000	N S			ug/L			
Total   1970	33065 MSMT/WIDTH///	Total	4000	N. N.			ug/L			
Triangle    .0559 MSMT/WIDTH///	Total	3500	NR			ng/L				
Total	33066 MSMT/LENGTH///	Total >	2000	N. N.			ug/L			
Total	9972	Total	1400000	2 2			1/8/1			
Total	in.	Total	0000009	NR			1/8n			
Total   1900000	9386 MSMT/REPLICATE 2///	Total	2530000	NR			T/8n			
Grad         SECONDO         IAR         NR           Getal         SECONDO         IAR         NR           Getal         SECONDO         IAR         NR           Getal         SECONDO         IAR         NR           Getal         1,20000         IAR         NR           Distand         1,20000         IAR	9387 MSMT/REPLICATE 2///	Total	1900000	N S			T/Sn			
Total	9373	Total	0000009	N. N.			ng/r			
Total	9388 (v)	Total	0000009	NR			1/Bn			
Total   1500000		Total	0000009	NR			ng/L			
Total	IGHT TO SHELL	Total	1500000	N. S.			ng/L			
Total	5901	Total	1500000	Z Z			ug/L			
Discolated   101370	5902 liy)	Total	150000	N N			ug/L			
Coloration of the coloration		Dissolved	101370	NR			ug/L			
Total   1224900		Dissolved	101370	NR			1/8n			
No.   Discoved   Control   Control		Total	2254900		2010720	267782	lug/L			
NY)         Total of the control o		Dissolved	101370	NR			ug/L			
Vy)         Total         > 1381000         NR         NR           Ny)         Total         344500         NR         NR         NR           Ny)         Total         344500         NR         NR         NR           Ny)         Total         1350000         NR         NR         NR           Total         1350000         NR         NR         NR           Total         1350000         NR         NR         NR           Total         1320000         NR         NR         NR           Total         132000         NR		Total <	220000	NR			ng/L			
WHY         10441         9482500         NR         PR         PR           WHY         10441         3595000         NR         NR         NR           HY         10441         1200000         NR         NR         NR           10441         1200000         NR         1640         NR         NR           10441         1200000         NR         1640         NR         NR           10441         1230000         NR         1640         NR         202000           10441         1230000         NR         1640         NR         1640         NR           10441         1230000         NR         1640         NR         NR         164000         NR         164000         NR         164000         N		Total >	1381000	N S			ug/L			
VA)         CORD         NA         NA           VI)         (1041)         1,200000         NR         NR           VI)         (1044)         1,200000         NR         NR           VI)         (1044)         1,200000         NR         NR           VI         (1044)         1,250000         NR         NR	9.5	otal	3449810	N S			1/80			
Total   1200000   NR   164000   NR   1640000   NR   164000   NR   164000   NR   1640000   NR   16400000   NR   164000000   NR   16400000   NR   164000000   NR   16400000   NR   16400000   NR   164000000   NR   1640000000   NR   164000000   NR   164000000   NR   164000000   NR   164000000   NR   1640000000   NR   1640000000   NR   164000000   NR   1640000000   NR   164000000000000000000000000000000000000		Total	1060000	C N			7/80			
Total   135,0000   NR   146,000		Total	220000	Z			7/8n			
Total   1553000   NR   1545000   NR   15450000   NR   NR   15450000   NR   15450000   NR   NR   154500000   NR   NR   NR   NR   154500000   NR   NR   NR   154500000   NR   NR   NR   NR   NR   154500000   NR   NR   NR   NR   NR   154500000   NR   NR   NR   NR   NR   NR   NR		Total	1300900	N. N.			ug/L			
Total         18950000         NR         1046000         NR         202000           Total         1342000         NR         NR         NR         NR           Total         1342000         NR         126700         NR         NR         NR         NR         NR         126700         NR         NR         NR         NR         NR         126700         NR         NR <td< td=""><td></td><td>Total</td><td>1560000</td><td>N.</td><td></td><td></td><td>ng/L</td><td></td><td></td><td></td></td<>		Total	1560000	N.			ng/L			
Total   1350000   NR   NR   NR     Total   13500000   NR   NR     Total   135000000   NR   NR     Total   135000000   NR   NR     Total   13500000   NR   NR     Total   135000000   NR   NR     Total   135000000   NR   NR     Total   135000000   NR   NR     Total   135000000   NR   NR     Total   13500000		Total	1853000		1646000	208200	ng/L			
Total		Total	1950000	N S			ug/L			
Total		Cotal	1352000	e a			1/8n			
Total		Total	978000	N.			ng/r			
Total		Total	1348000	NR			ug/L			
Total         1075000         NR         NR         NR           Total         343000         NR         NR         NR           Total         350000         NR         NR         NR           Total         2500000         NR         NR         NR           T		Total	1510000		1217000	216700	1/gn(			
Total         343000         NR         NR         NR           Total         343000         NR         NR         NR           Total         532000         NR         NR         NR           Total         55000         NR         NR         NR           Total         543000         NR         NR         NR           Total         543000         NR         NR         NR           Total         54000         NR         NR         NR           Total         355000         NR         NR         NR           Total         355000         NR         NR         NR           Total         355000         NR         NR         NR           Total         350000         NR         NR         NR           Total         350000         NR         NR         NR           Total         350000         NR         NR         NR           Total         250000         NR         NR         NR           Total         250000         NR         NR         NR           Total         250000         NR         NR         NR           Total <td></td> <td>Fotal</td> <td>1925000</td> <td>RN</td> <td></td> <td>NR.</td> <td>ng/L</td> <td></td> <td></td> <td></td>		Fotal	1925000	RN		NR.	ng/L			
Total		lotal	1075000	NR		NR	ug/L			
Total		Total	343000	N.		NR	ng/L			
Total		Total	343000	S. S.		N. S.	ug/t			
Total   85,0000   NR		otal	732000	E :		N.	ug/L		İ	
Total		otal	97000	NR.		N. N.	T/Iomn			
Total   1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1,		otal	850000	N. C.		N S	ng/r			
Total		otal	1410000	X Z		N. N.	1/8n			
Total		otal	1410000	XX			1/80			
Total   154000   140000   154000   154000   1540000   1540000   1540000   1540000   1540000   1540000   156000000   156000000   156000000   156000000   156000000000000000000000000000000000000		otal	000799		000/85	740000	ug/L			
Total		otal	1534000		1400000	1661000	1/8/1			
Total   189000**   NR		otal	76000		24000		ug/L			
Total   1830000		otal	355000	N S		N. Y.	1/80			
Total		otal	390000	N N		N C	ng/c		l	
Total		010	190000*	g a		NB	7/20			
Total		otal	251000*	a a		N. N.	7.60			
Total		Otal	210000	a N		NR	1/60			
Total   2500000   NR   NR   NR     Total   12000000   NR   NR   NR     Total   120000000   NR   NR   NR     Total   1200000000000000000000000000000000000		oral	280000	Z Z		NB	700			
Total		Otal	0000000	GN C		NB	100/1			
Total		otal	2000000	N N		N. N.	760			
Total 1000000 NR NR NR NR NR NR NR NR NR NR NR NR NR		otal	1800000	S. N.		NR	l/an			
Total		otal	1000000	N. N.		Z.	ng/t			
Total 2000000   NR   NR   NR     Total 229500000   NR   NR   NR     Total 229590000   NR   NR   NR     Total 229590000   NR   NR   NR     Total 40850000   NR   NR   NR     Total 41200000   NR   NR   NR     Total 41200000   NR   NR   NR     Total 189500000   NR   NR   NR     Total 189500000   NR   NR   NR     Total 189500000   NR   NR   NR     Total 189500000   NR   NR   NR   NR   NR     Total 189500000   NR   NR   NR   NR   NR   NR   NR		otal	1200000	N. N.		NR	1/411			
Total		Otal	2000000	S. S.		NR	1/0/1			
Total		Oldi.	1200000	N N		NE	ug/r			
Total 28839000 NR NR NR NR NR NR NR NR NR NR NR NR NR		0.0	300000	UN N		NB	UB/1			
Total         4350000         NR         NR           Total         44850000         NR         NR           Total         4480000         NR         NR           Total         4320000         NR         NR           Total         4320000         NR         NR           Total         1385000         NR         NR           Total         1385000         NR         NR           Total         13850000         NR         NR		Oral Oral	000000000	NN NN		N N	7/80			
Total		otal	73033000	N. C.		NA	ug/r			
Total		otal	41560000	Z Z		NR	1/8n			
Total		otal	47600000	NA S		NN	ng/r			
Total		otal	43609000	HN CA		NK.	ng/r	-		
Total		otal	4120000	EN S		NK:	1/8n			
105a  1.8850000		otal	18950000	N N		NK	ng/L			
Total   18900000   NR   NR			1005001	1000		411				
		otal	19200000	INR		NR	ng/r			

Column	2148461	Total	11510000	sel do mar t a	Islan conc. I was op isla	NR ug/L	out 2 Type (Staff Cott, 2 Mean (Staff	one a orins (standa Application Rate	ation rate. Application rate of
1500         1315,000         104         1	2148458	Total	18900000						
The continue of the continue	2148459	Total	11560000						
1971   1975	2148460	Total	11560000						
1988   1988	18/928	Total	39130000						
Total	768170	Total	299090						
1979   1979	160280	Total	630000		30000	63000			
Total	793907	Total	690000						
Constant	768254	Total	150490						
Total   1989   1980	2060131	Total				707000			
Cotal   1,5000   1,6000   1,	2060128	Total		2	20000	S00000 ug/L			
1985   1985	160281	Total	630000	io.	30000	920000			
Total	798.61	Total	150490	X X					
Total	769361	Total	76080	N ON					
COME         STREAM         NOR         NOR           COME         255250         NOR         NOR           COME         255250         NOR         NOR           COME         255250         NOR         NOR           COME         251100         NOR         NOR           COME         251100         NOR         NOR           COMB         2500000         NOR         ALSENDO           COMB         2500000         NOR         NOR           COMB         2500000         NOR         ALSENDO           COMB         2500000	768315	Total	150490	E W					
Total	768353	Total	241540	NR					
Contain         SSS 55-00         NR         NR           Total         2512-00         NR         NR           Total         2511/00         NR         NR           Total         2511/00         NR         NR           Total         2511/00         NR         NR           Total         2500000         NR         NR           Total         2500000         NR         NR           Total         2500000         NR         4150000         NR           Total         2500000         NR         450000	768312	Total	596280	NR					
Total	768357	Total	251540	NR					
Treat   21110	768251	Total	596280	NR					
Total	768257	Total	596280	NR					
Total	727030	Total	21110	NR					
Total   2000000	762358	Total	21100	NR					
Total	2056433	Total	3394000	NR					
Total	165611	Total	2000000	П		100000000000000000000000000000000000000			
Total	165609	Total	2000000	416	0000	6010000 ug/L			
Total   55,0000   43,0000   1,0000	165610	Total	3170000	226	0000	4400000 ug/L			
Total	165622	Total	2300000	43	00000	6520000 ug/l			
Closal         28,90000         43,90000         64,90000           Closal         28,90000         5,20000         64,90000           Total         28,00000         5,20000         7,20000           Total         28,00000         8,20000         7,20000           Total         28,00000         4,20000         7,20000           Total         28,00000         1,20000         7,20000           Total         28,00000         1,20000         1,20000           Total         28,0000         1,20000         1,20000           Total	165615	Total	9000000	109	0000	7240000 ug/L			
Clear         5250000         A43000         649000         750000<	165614	Total	2890000	238	0000	3200000 nB/L			
Total	165621	Total	2360000	44	0000	6490000 ug/L			
Total	165613	Total	6280000	526	0000	7500000 ug/L			
Total   1250000   NR   2550000   NR   1250000   NR   12500000   NR   1250000   NR   12500000   NR   1250000   NR   12500000   NR   12500000   NR   12500000   NR   12500000   NR   12500000   NR   12500000   NR   125000000   NR   1	165616	Total	1770000	30	0000	\$260000 ug/t			
Total   1250000   NR   1250000   NR   1250000   NR   1250000   NR   1250000   NR   1250000   NR   12500000   12500000   125000000   12500000   12500000   12500000   125000000   125000000   125000000   125000000   125000000   125000000   125000000   125000000   125000000   12500000   125000000   1250	165620	Total	3510000	100	0000	4430000 08/1			
Total   1,000   1,00	200001	Total	1750000			****			
Total   1,000   1,00	20000	Total	0000031	No.					
Total   SESTONO   SESTONO   SESTONO   TOTAL    719091	10191	000000			-				
Total	165608	lotal	9830000	953	0000	/310000 ug/L			
Total   E200000   A500000   Total   E200000   Total   E200000   A500000   A500000   E200000   A500000   E200000   A500000   E200000   E200000   A500000   E200000   A500000   E200000   E200000   A500000   E200000   E200000   A500000   E200000   A500000   A500000   A500000   A500000   A500000   A500000   A500000   A50000   A500000   A5000000   A500000   A5000000   A5000000   A5000000   A5000000   A5000000   A5000000   A5000000   A5000000   A500000   A5	165619	lotal	5110000	418	0000	6240000 ug/L			
Total   5200000   4380000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   758000000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   75800000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   75800000   7580000   7580000   758000   758000   758000   758000   75800000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   7580000   75800000	16560/	lotal	6190000	53/	0000	/130000 ug/L			
Total	165623	lotal	970000	486	0000	/830000 ug/L			
Total	10001	lotal	2300000	4	0000	especial ug/r.			
Total	7152543	lotal	4260		3830	4/30 ug/L			
Total	2152542	lotal	3820		3390	4380 ug/L			
Total	2152541	Total	4160	1		4780			
Total 14405007*   NR   NR   NR     Total 15570007   76500   NR   NR   NR     Total 15570007   76500   NR   NR   NR     Total 15570007   NR   NR   NR     Total 2590500   NR   3321400   NR   NR     Total 415000   NR   3321400   NR   NR     Total 415000   NR   3321400   NR   NR     Total 415000   NR   345000   NR     Total NR   357000   NR   NR   NR     Total 579000   NR   NR     Total 579000   NR   NR     Total 579000   NR   NR     Total 5790000   NR   NR     Total 579000   NR     Total	2146823	Total	4440000	NR					
Total	15436	Total	1406000*	NR					
Total	16498	Total	1567000*	N					
Total	2146826	Total	76500	NR					
Total   192100	768484	Total	596280	NR					
Total	768515	Total	192100	NR					
Total   41500   NR   32140   NR   492000   NR   34600   NR   482000   NR   NR   482000   NR   4820	768422	Total	299090	ZZ					
Total	768419	Total	596280	NR					
Total	233426	Total	415000			497100			
Total   11870   NR	233425	Total	405000	34		480700			
Total   NR	707031	Total	418870						
Total   NR	10072	1,000	000000	0.00					
Total   NR   517000   7070000   7070000   70700000   707000000   70700000   70700000   70700000   70700000   70700000   707000000   70700000   70700000   70700000   70700000   70700000   7070000   7070000   7070000   7070000   7070000   7070000   7070000   7070000   7070000   7070000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   707000   7070000   707000   707000   707000   707000   707000   707000   7070000   707000   707000   707000   707000   707000   707000   7070000   707000   707000   707000   707000   707000   707000   7070000   7070000   7070000   7070000   7070000   7070000   7070000   70700000   70700000   70700000   70700000   70700000   707000000   70700000   707000000   70700000   707000000   707000000   707000000   707000000   707000000   707000000   707000000   70700000   707000000   707000000   707000000   707000000   707000000   707000000   7070000000   707000000   707000000   7070000000   7070000000   70700000000	2060111	Total	1			000000			
Total   NR   73,7000   NR   73,0000   NR   70,0000     Total   34,3000   NR   75,000   NR   NR   NR   NR   NR   NR   NR	2000111	Total	N. C.	90	2000	/0/000 ug/L			
Total NR 357000 NR 250000 NR 250000	Z080143	Iptal	XX	10	7000	/0/000 ug/t			
Total	2060140	lotal				200000			
Total	119482	lotal	35/000	NK					
Total	318/8	lotal	343000	NA					
Total	18170 MSMT/TLM///	Total	29000	Z X					
Total	18166 MSMT/TLM///	Total	923000	NR					
Total   17000   NR	18167 MSMT/TLM///	Total	343000	S					
Total	18169 MSMT/TLM///	Total	117000	N.					
Total	18168 MSMT/TLM///	Total	337000	NR					
E A/// E A/// Total         Total 299200         NR         NR           E B/// Total         Total 290200         NR         NR           I Total Total         990200         NR         NR           I Total Total         990200         NR         NR           I Total Total         290200         NR         NR           I Total Total         295200         NR         A40000           I Total Total         295200         NR         NR	768482	Total	596280	NR					
E A/// E B/// Total         Total 596200         NR NR         NR NR         NR NR         NR NR         NR NR         NR NR         NR NR         NR S80000         NR NR         NR S80000         NR NR         S80000         NR S80000         NR NR         S80000         NR NR         N	768483	Total	299090	NR		1			
E B/// TOTAL         TOTAL         968000 24000         NR         NR         NR           TOTAL         70431         24000         NR         NR         NR           TOTAL         7984000         NR         NR         NR           TOTAL         2954000         NR         NR         NR           TOTAL         2557100         NR         NR         NR           TOTAL         295280         NR         NR         NR           TOTAL         2950200         NR         NR         NR           TOTAL         382300         NR         44000         NR           TOTAL         1278500         NR         NR         NR           TOTAL         1278500         NR         NR         NR           TOTAL         10781         1285000         NR         NR	759474 MSMT/REPLICATE A///	Total	24000	NR					
E B/// Total         Total 101al         24000 286000         NR         NR         NR           Total         994000         NR         NR         NR         880000           Total         994000         NR         880000         NR         880000           Total         25557100         NR         NR         NR         NR         880000           Total         529590         NR         440000         NR         880000           Total         3823000         NR         NR         NR         80000           Total         1275500         NR         NR         NR         NR           Total         1275500         NR         NR         NR         NR           Total         1280000         NR         NR         NR         NR	767301	Total	000896	NR					
Total   986000   NR	759477 MSMT/REPLICATE B///	Total	24000	m Z					
Total   2994000   NR 580000   NR 280000   NR 2800000   NR	767309	Total	986000	aN					
Total   740000   NR 580000   NR 5800000   NR 580000   NR 5800000   NR 5800000   NR 5800000   NR 5800000   NR 580000   NR 5800000   NR 5800000   NR 580000   NR 580000   NR 5	757300	Total	994000	N N					
Total   2557200   NR 200000   NR 200000   NR 2000000	Cachar	Total	740000	ľ		000000			
Total	190202	Total	/40000	1		asonon			
Total	768506	iotai	255/100	NX					
Total	768494	Total	596280	NA					
Total	768485	Total	299090						
Total 382300 NR NR NR NR NR NR   Total 1279500 NR NR NR   NR NR NR NR NR NR NR NR NR NR NR NR NR	160283	Total	000099	1	000	880000			
Total	768513	Total	382300	N.S.	-				
Total 596280 NR 10000000 11600000 11600000	768508	Total	1279500	N.S.	_				
Total 1000000 1160000	768421	Total	596280	DIN DIN DIN DIN DIN DIN DIN DIN DIN DIN	4				
000001	200000		Table 1	ling	AT				

ug/t ug/t	ug/t ug/t	2/L	3/F	7/8n 7/8n	1/5	8/r 3/r	2/L	7/2 //	17 T	7/7	27/	2/1	7/7	2/L	3/L	27	7/1	y/L	**	2/1	(/r	3/1	1/r	V/L	ug/L	ug/L	VL	//r	ν/L	7/r	1/1	1/4	1/1	1//	ug/L	1/1	J.	1/F	1/1	1/r	7/	/L	7/1	7.	// //	77		J.C.
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Raw Data from ECOTOX (Unfiltered)

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120844	Total	910000	NN		NR	ng/L			
251003	Total	200000	N.S.		NR	ng/L			
120840	Total	280000	N. S		NR.	1/Bn			
251001	Total	300000	NR S		NK .	1/8n			
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251009	lotal	2500000	N W		N N	1/8n			
170842	lotal	1940000	N N		N N	7/20			
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251004 MSMT/FOUND IN *10% OF LABS///	Total	200000	a N		e e	1/80			
251002 MSMT/FOUND IN *10% OF LABS///	Total	300000	N.		N.	ng/L			
120845	Total	1430000	NR		N.	ng/L			
120837	Total <	330000	NN		NR	T/Bn			
251008 MSMT/FOUND BETWEEN 50 AND 60% OF L/Tota	Total	1500000	NR		NR	ng/L			
120827	Total	1830000	NR		NR	ng/L			
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   | 20   | 2200000 ug/  | /Bn  | /Bn  | 5 mS/   
   | /Sin   | /Sn  | /Sn  | 4154000 ug/l   
           | 6670000 ug/l   |                 | /Sn  | /Bn  | /8n  | 8000000 ug/l  
  | 1/Sn              | 1/8n   | mm   | 1/3n 00000001 | 1000000 ug/  | 1/8n 00000001 | 10000000118/1  | /An  
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Raw Data from ECOTOX (Unfiltered)

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Result Number	766740	766739	756717	1/999/	756560	766575	766581	766777	766573	766730	766731	766578	766595	766586	766610	766745	799/42	766555	766566	766718	766570	766723	766605	766591	766724	766572	766548	700000	(86299)	/66588	766608	766551	766563	767585	767586	767587	766854	766592	766726	766748	766587	755512	766561	766590	766593	1,000,00	766571	766584	/1799/	18144	18145	84497	767473	759175	767574	119467	767794	767307	767306	767796	767303	200707	10/293	967/9/	9/9/9/	18452	18147	18453	18454	18451	184501	750001	759090	759097	767297	767305	767299	767496	727294	101101	767475

766612 76612 MSMT/MEAN OF 3 TESTS/ 76612 MSMT/MEAN OF 3 TESTS/ 76613 MSMT/MEAN OF 3 TESTS/ 76613 MSMT/MEAN OF 3 TESTS/ 76662 7 766614 7 76614 MSMT/MEAN OF 3 TESTS/ 76617 MSMT/MEAN OF 3 TESTS/ 76611 MSMT/MEAN OF 3 TESTS/ 76611 MSMT/MEAN OF 3 TESTS/ 76611 MSMT/MEAN OF 3 TESTS/ 76611 MSMT/MEAN OF 3 TESTS/ 76613	Total     Tota	34000000 134000000 15500000 15500000 15500000 250000 250000 14750000 117500000 117500000 1175000000 1175000000 11750000000000	N	A         B	1/8n 1/8n 1/8n 1/8n 1/8n 1/8n 1/8n 1/8n	
AN OF 3 TESTS/ AN OF 3 TESTS/	Total   Tota	4945000 1901000 29550000 1562000 3462000 2002000 1287000 1438000 1438000 2121000 2022000	NR NR NR NR 1118210 NR NR NR NR		1/8n   1/	
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	Total   Tota	2240000 2101000 569000 3650000 1126130 3650000 1126130 3650000 3144000	NR NR NR NR 1118210 NR NR NR		98/L 98/L 98/L 98/L 98/L 98/L 98/L 98/L	
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4///	Total	5401000	N.			ng/t				
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	Total	220000*	AN.			ug/L				
	Total	100000	RN	2		ug/L				
	Total	4500000	N	3		ug/L				
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	Total	000906	N			ng/L				
	Total	160000	NR			T/Bn				
	Total	2850000	NR			ng/L		*		
	Total	1555000		869000	2032000	ug/L				
	Total	1300000	NR		NR	7/8n				
	Total	2938000			3385000	1/80			2000	
	Total	2183000		1499000	2634000	1/0/1				
	Takal	24000	a z		NR	1/20				
	Total	4553000			5157000	1/2/1				
	Total	3231000		COUNTY	3840000	1/80				
	Total	3801000		2817000	4356000	ua/i				
	- Cuai	1355400	02		(D) *	1/90				
	lotal	D00000011	N N	00000444	NA TENADOOD	ug/r				
	Total	15200000			15940000	1/8n	1			
	Total	210000	NR		NR	ng/L				
	Total	229000		293000	805000	1/Bn				
	Total	1250000	NR		NR	1/Bn				
	Total	000565	NR			1/811				
	Total	1649000		1255,000	OOGZOOC	1/00/1				
	lotal	1643000		1232000	2027200	ug/t.				
	Total	933000			1230000	1/Br				
	Total	474000	NR			ia/l				
	lotal	DOUBLE OF THE PERSON OF THE PE	N N		NR	JB/L				
	Dissolved	1013/0	NN			ug/L				
			-							
	Dissolved	101370	NR			J/Sr				

160334   100	^	8080000 7960000 24000	7070000 > 6800000 > NR		10000000 10000000			
ASMT/TUM!//		7960000	12.0		10000000 ug/L			
ISMITIUM!//		33	N.					
ASMT/TUM!//		5500000			NR ug/L			
SSMTTUM!//		1060000	NR					
///w////		15996000	NR					
		20040000	NR					
		1075000	N.R.					
		719000	234000	000	1041000			
		1850000						
	^	1850000	N.S.					
		1205000	363000	00	1510000			
		1925000	NR					
		1507000	907000	000	1963000 ug/L			
		082000	1460	000	1302000 ug/L			
		343000	NR		NR ug/L			
		1190000	750000	20	2002000 ug/L			
		978000	l					
		0000000	gN					
		0000505	gN					
		2023020	Q.Z					
		2018000	UNI ON					
		7700007	UNI.					
		1202000	NN C		INK UE/L			
		2186000	NY.					
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	tion	7340000	N. N.					
	tion	000099	N.					
		300000					The second second second	
	^	1970000 >	1970000	> 00	1970000			
		0750771						
		1447270	ND					
		0/5/547	NO.		WD NB			
		859960	NY					
		764840	N.S.					
	^	1910000	1910000	< 00	1970000			
		754840						
		Ototo:	000		1/20			
		144/3/0	NY.					
	^	1940000 >	19400	> 00	1990000			
		233000	NR		NR ug/L			
		000886	N. N.					
		000000						
		224000	un.					
		254000	NE					
		1600000	NR					
		1470240	NR		NR ug/L			
		1470270	NR					
	^	1970000 >	1970000	> 00	1970000			
		25,0960	an					
		00000000	N.D.					
		14/0240	NX					
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		00000000	0.4					
	^	Secucion	NK					
	^	26000000	NR					
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	^	< 000001	1970000	< 00	1970000			
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		13/0000			200			
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		1470200	ú		1			
	^	< 000001	1970000	> 00	1970000			
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Total		220400	NR		NR ug/L			
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lotal		//0400	INK					
Total		425900)	NR					
Total		643620	N.N.					
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		030030	0					
lotal		753630	INN					
Total		253650						
Total	V	680000 <	620000	00				
Let Co		270000		O	780000			
1372		OCTOBE	Aib					
Total		770400	NX					
Total		253650	NR					
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Total		770400	NK					
Total		600000	NR					
1 1 1		340000	ND					
lotal		340000	NK					
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letoT	-	Masan	NE					
10101		775C76	INK				+	
Total		253650	NR					
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Colore		7500000	N.		
10000000   NR   NR   NR   NR   NR   NR	Total Total Total Total Total Total Total Total Total		2000	NR ug/L	
10000000   NR   NR   NR   NR   NR   NR	Total   Tota	10000000	N.S.	NR ug/L	
No.   Colored   No.	Total Total Total Total Total Total Total	10000000	NR	NR ug/L	
No.   No.	Total Total Total Total	100000000	NR	NR ug/L	
Mail	Total	20000000	NR	NR ug/L	
Mail	Total	0.001	NR		
1800000   N	TO THE STATE OF TH	NR	125000	15000	
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10	Tayot T	7.	021		
N	iorai	*	400		
N		4	Z,		
NR   NR   NR   NR   NR   NR   NR   NR		. 2	NR		
NE	Total	1	N.S.		
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NR		50	av		
No.   No.			9		
NR   NR   NR   NR   NR	lotal	र्ग ।	NX		
NR	Total	. 1	N.S.		
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NR	Total	4	an		
National Color					
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NR   NR   NR   NR   NR   NR   NR   NR		4	N.S.		
NR   NR   NR   NR   NR		2	e Z		
1000000	1 1				
1250000	rotal	- 3	NR		
10.155   NR   NR   NR     0.12500000	Total	0.25	N.S.		
1250000	Total	35.0	NB		
1250000		200			
1250000		0.125	N.		
1250000	TION DAYS/	1250000	NR		
150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   15000000   1500000   15000	TION PER SNAIL F	1250000	a Z		
1000   1000		00000	0000	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	
14500   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   15000000   1500000   1500000   15000000   1500000   1500000   1500000   15000000   1500000   150000		36000	91000	1/2n000 ug/L	
15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   15000   1500000   15000000   15000000   15000000   1500000   1500000   15000000   1500000   1500000   1500000   15000000   1		4500	2200	6200 ug/L	
1,000   1,000    1,		13000	11000	1/2000 112000	
350,000		2500	003	77000	
12000   NR   24000   NR   NR   24000   NR   24000   NR   NR   NR   NR   24000   NR   NR   NR   NR   24000   NR   NR   NR   NR   NR   NR   NR		0000		1400	
12000   NR   24000   NR   24000   24000   24000   24000   24000   24000   24000   24000   24000   24000   24000   24000   24000   240000		360000	NR		
17000   1710000   171000   171000   171000   171000   171000   171000   1710000   171000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   1710000   171000		360000	NR		
120000   12000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   1510000   15100000   15100000   15100		apoon	17000	1/21/00073	
1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000   1,2000	T	00000	00000	7/30 00000	
1000000   100000   100000   100000   100000   10000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   10000000   1000000   1000000   1000000   1000000   1000000   1000000   1000000   10000000   10000000   10000000   100000000		122000	00086	121000 ug/L	
150000   NR		63000	26000	7/2000 lug/L	
12000000   NR		36000	27000	4200011971	
1000000   NR	The state of the s	00000			
1500000   NR 50000   NR 1570	CARIA PRODUCTION PER SINAL RIOTAL	100000	N.		
150000   NR   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   1500000   150000   150000   150000   150000   150000   150000   15000000   150	S CERCARIA PRODUCED (SHEDD Total	2000000	NR		
150000	Total Daylor Deconnect Total	00000	alle alle		
135000   163000   155000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   108   1550000   1550000   108   1550000   1550000   108   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   15500000   1550000   1550000   1550000   1550000   1550000   15500000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   15500000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   1550000   15500000   15500000   1550000   1550000   1550000   1550000   1550000   1550000   15	VEN.	nnnnne			
1345000   1631000   1345000   1345000   1345000   1345000   1345000   1345000   1345000   1345000	Total	1505000	240000	1670000 ug/L	
1144000   NR   1144000   NR   1144000   NR   NR   1144000   NR   NR   NR   NR   NR   NR   NR	Total	1945000	1631000	2263000 ug/L	
1,200000   NR	100	2000	000110	7/95 000000	
1100000   NR	lotal	1241000	١	1345000	
1250000   NR	Total	1120000	NR		
1200000   NR	Total	2230000	NR		
1500000	'crc'	0000002	0.2		
1500000   NR		7,000,00	NR		
15000000		1900000	NR		
1250000		1600000	82		
1250000		000000	0.2		
1250000	lotal	820000	INK		
1250000   NR	Total	1250000	NR		
125000   NR	Total	2500000	NR		
1350000	Total	0000361	O.V.		
1250000		000000000000000000000000000000000000000			
S20000   NR	lotal	1250000	ZX		
Section   NR	Total	620000	NR		
1250000	Total	620000	NR		
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140250   NR	Pioral	970079	IVA		
1000000   NR	AN OF ALL LABORATORY REFERENTOTAL	1450250	NR		
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1000000   NR	THOR REPORTS REPRODUCTION a Total	750000	av		
1250000	1.00	000003	NI NI NI NI NI NI NI NI NI NI NI NI NI N		
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1250000	THOR REPORTS REPRODUCTION 4 JOTAI	1000000	NR		
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1,250.000	e c	DOODEL	Oly City		
1250000   NR	IOTAI	1250000	NK		
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697400   540000   901000	IDIGI	454000	000157	1/Bnlnnning.r	
	Total	697400	540000	1/8n 000106	

1971   1971	Rate Units									T						I														Ì																			i i													T		Ī		Ī	I		1	T					
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1,000,000   1,00	Conc 2 Type (Star																																																																										
1970   1970	++1			1/80	П	7/8n 000	1/80 000	1/Sn	ng/L	ug/L	1/8n 000	610 ug/l	000 ng/L	000 ug/L	000 ug/L	1/8/1	1/8/1	000 ng/L	000 ng/L	ng/L	ng/L	UB/L	1/80	ne/L	ug/L	ng/L	ng/t	ng/L	1/Bn	ng/L	ng/L	ng/L	ng/L	ng/r	7/Sn	ug/L	1/80	1/811	1/80	ug/L	ng/L	ng/L	ng/r	ng/L	ng/r	1/8/L	na/L	1/8n	ug/L	ug/L	ng/L	ug/L	ug/L	1/8/1	1/8n	ug/L	ng/L	ng/L	ng/L	ug/L	UB/L	ug/L	UB/L	ng/r	7/B/L	ng/r	OD US/L	DO UB/L	ng/L	1/Bn	1/8/1			1/011100	00 ug/L
1530000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   150000000   NR   150000000   NR   150000000   NR   1500000000   NR   1500000000   NR   15000000000   NR   1500000000000000000000000000000000000	tai Conc 1 Max (	1 1		N.		815		NR.	NR	1	211	1565	1177	733	700	NR NR	N.	1640	2000		Z S	N N	a N	Z.	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	N S	X Z	N N	N. N.	N N	Z.	NR	NR	NR	N S	N N	N N	NB NB	NR	NR	NR	NR	N.S.	Z S	N N	a.	NR	NR	NR	N. S.	NR.	X C	N S	N N	NR	N N	2550			NR.	N S	N N	NK	3950	2800	2800 NR
1530000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   1500000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   15000000   NR   150000000   NR   150000000   NR   150000000   NR   1500000000   NR   1500000000   NR   15000000000   NR   1500000000000000000000000000000000000	Conc 1 Max Op (S																																																																										
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	onc 1 Mean (St.C	1296000	7160000	2500000	1250000	454000	620000	1250000	1250000	97,000	161000	1245610	895000	580000	221000	440000	850000	1350000		630	1250	220000	1100000	1500000	1000000	670000	1060000	1130000	250000	200000	200000	250000	000096	920000	1160000	1170000	200000	1000000	0000086	610000	1000000	200000	1200000	200000	347000	563000	340000	653000	456000	620000	1250000	130000	620000	620000	1250000	620000	1250000	620000	1250000	620000	310000	4250000	1230000	020000	1250000	146000	761000	481000	1250000	620000	1250000	201000	201000	264000	264000
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Tristance   Et   Tris	Result Number EE	21527	21527.	7678	7678	2152721	76787	7678	767858ers	301311	21528	76755	2153276	21552.	21232	74168	74256	74166	2077.	214681	21468	76150	74193	76157	76158	74195	76158	74256	74195	74290	74194	74285	74195	74192	76157	76150	74193	74289	74289	74289	74195	74289	76157	76159	214845	214840	214848	214847.	214847	767853	76785.	76795.	76795	76785	76686.	76686	76786.	76786	76687	76687	76789.	76789	767002	75705	76796	96/97	7153269	215327	767899	767900	76796	75536	213326.	2152887	2152881

1	1500000		NR UB/			
1,10000   Mai	0000001					
1,000,000   MS   MS   MS   MS   MS   MS   MS	12100001	NR				
1,000000	1760000	NR				
1,000,000   M   M   M   M   M   M   M   M   M	300000	NR				
No.   No.	1500000	N. S.				
1860000   NR   NR   NR   NR   NR   NR   NR	467000	NR				
No.   1,19,000   N.   N.   N.   N.   N.   N.   N.	467000	N. S.				
1,000,000   No.	1300000	N. C.				
1500000   NR   NR   NR   NR   NR   NR   NR	740000	N N				
1980000   Note	000039	2				
1300000   Not	1340000	NR				
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1,00000   NR   NR   NR   NR   NR   NR   NR	330000	S. S.				
1380000   NR	720000	a Z				
1500000   NR	1390000	N.				
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Colored   No.   No.	1500000	NR				
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1,00000	000006	N				
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1140000   NR	2,0000	ZZ				
1000000   NR   NR   NR     1070000   NR   NR     1070000   NR   NR   NR     1070000   NR   NR	200000	NA				
1500000   NR	1140000	NR				
1000000   NR	720000	NR				
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1000000   NR	200000	NR				
SOCOCO   NR	1000000	NR				
NR   NR   NR   NR   NR   NR   NR   NR	220000	NR				
1900000   NR	200000	NR			44	
NR   NR   NR   NR   NR   NR   NR   NR	1500000	NR				
SEGONO   NR	710000	N.				
SECONDO   NR	000089	œz				
Colored   NR	500000					
Color	250000	2 Z				
Color	350000	0.00				
11550000   NR	000000	2.0				
NR   NR   NR   NR   NR   NR   NR   NR	750000	N. S.				
Company   Comp	2000005	NR				
NE   NE   NE   NE   NE   NE   NE   NE	250000	XX				
1155000   NR	250000	NR				
1500000   NR 530000   NR 530000   NR 530000   NR 530000   NR 530000   NR 530000   NR 530000   NR 530000   NR 530000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 5300000   NR 530000000   R 530000000      NR 53000000000000000000000000000000000000	1155000	NR				
SECONDO   NR   NR   NR   NR   NR   NR   NR   N	1650000	NR				
650000	820000	NR				
440000   NR	000059		790000			
A	440000					
5	467000	000				
1000000   NR	000000	0.2				
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1000000   NR   NR   NR   NR   NR   NR   N	200000	Z.				
1000000   NR   NR   NR   NR   NR   NR   N	TOOOOOT	NX				
1000000   NR   NR   NR   NR   NR   NR   N	1000000	NK				
1000000   NR   NR   NR   NR   NR   NR   N	1000000	N. N.				
2000000 NR NR NR 2000000 NR NR NR 1200000 NR NR NR 1250000 NR 4000000 NR NR NR NR NR NR NR NR NR NR NR 1250000 NR 40000000 NR NR NR NR NR NR NR NR NR NR NR NR NR NR NR NR NR NR N	1000000	NR				
NR   NR   NR   NR   NR   NR   NR   NR	2000000	NR				
1200000   NR   NR   NR   NR   NR   NR   NR	2000000	NR	1			
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NR   1296.000   NR   450.000	1000000	S N				
NR	1296000	NID				1
NR 2160000 NR NR NR NR AND NR NR ADDROX NR NR ADDROX NR NR NR NR NR NR NR NR NR NR NR NR NR	7700677	INK	9500000			
S160000 NR NR S05000 NR	NK V	400000018	4500000			
Total C	2160000	NR				
TOTAL SCHOOLS INK	206000	NR				
Total 1037000 684000 1491000	1037000	684000	1491000			
2152734 (10tal < 2152733 (Total )		900000   900000   300000   300000   300000   330000   300   33000   33000   33000   33000   3300   3300   3300   3300   3300   3300   3300	4 1 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	NR	NR	NR

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1000   1000	3153735		ant Conc 1 Mean (St. Conc 1	Min Op (StaConc Min 1 (Sta	onc 1 Max Op (Sta	nc 1 Max (Stan Co	-	Conc 2 Mean	anda	Rate Application Rate Units
Total   1900000   No.	2152732	Total	421000	0009C	NH.	90,00000	yr.			
Total   1000000   NR   NR   NR   NR   NR   NR   N	2164729	Total	200000	NR	NR	2000	/L			
Fired   1914	2164728	Total	1000000	NR	NR		/t			
Colore   C	201550 MSMT/CHRONIC VALUE///	Total	314000	NR	NR		/L			
COLOR DE CARDON   CALON   CA	201552 MSMT/CHRONIC VALUE///	Total	372000	N. S.	N. S.		7.1			
COOL OF CASE   COOL	201549 MSMT/CHKONIC VALUE///	Total	372000	N N	N N		/L			
Fige bit   Fige bit	201547	Total	314000	a N	NR		// //			
Color of Markey   Color of M	201551 prs	Total	441000	NR	NR		V			
Colore   C		- 13	1200000	S.S.	NR		1/1			
Total		Formul	1000001	Z Z	Z Z		/L			
Total   Section   No.   Sect	2152784	Total	1366000			1541000	1/4			
Coloral   Colorad   Coloral   Colorad   Coloral   Colorad   Coloral   Colo	2152783	Total	825000	54900		1256000	1/1			
Total   C   Section   NR	2152786	Total	366000				Λ.			
Page 25420   NR   NR   NR   NR	2152785	Total <	366000	NR	S. S.		Λ.			
Page 1989   Page 298	2163917 MSMT/MEASURED AS TOTAL EGGS PER FE	MDissolved	25430	N.R.	N.N.		Λ.			
Part   Part	2163920 MSMT/MEASURED AS PERCENTAGE OF CH	ADissolved	25430	NR	E I		η,			
Cotal         35520         NR         NR           Foral         1280         NR         NR           Foral         1280         NR         NR           Foral         1210         NR         NR           Foral         1280         NR         NR           Foral         1280         NR         NR           Foral         1280         NR         NR           Foral         1280         NR         NR           Foral         1290         NR         NR           Foral         1200         N	2163919 MSMT/MEASURED AS CONDUCATIVE EGGS 2163919 MSMT/MEASURED AS MEAN EGGS PER FEI	Missolved	05450	NAN	2 2		// //			
Total   120	2163916	Dissolved	25430	N N	N N		7/			
Total   1,20	2254781	Total	180	NN NN	ON		7			
Total   120	2254792	Total	180	NB	an an					
Total   120	2254801	Total	120	NR	NR					
Total   120	2254784	Total	120	NR	NR					
Total   Total   Total   NR	2254788	Total	180	NR	NB		-			
Total   NR	2254793	Total	180	NR	NR					
Total   180	2254797	Total	09	NR	NR		V			
Total   120	2254804 Flateral head displacement	Total	180	NR	NR		V			
Total   120   NR	2254803 f lateral head displacement	Total	180	NR	NR		V			
Total   120	2254798	Total	30	NR	NR		V			
Total   NR	2254785	Total	120	NR	RN		1		1	
Total   120	2254805 f lateral head displacement	Total	180	NR	NR		V			
Total         NR         NR           Total         120         NR         NR           Total         120         NR         NR           Total         120         NR         NR           Total         2000000         NR         252000           Total         252000         NR         252000           Total         252000         NR         252000           Total         252000         NR         35000           Total         252000         NR         35000           Total         252000         NR         NR           Total	2254789	Total	180	NR	NR		V			
Total   180	2254802 f lateral head displacement	Total	120	NR	NR		V			
Total   180	2254800	Total	180	NR	NR		4			
Total   NR	2254810	Total	180	NR	NR		4			
Total   1000000	2254813	Total	180	NR	NR		-			
Total   1,000,000	767224 MSMT/CERCARIA PRODUCTION DAYS/	Total	2000000	NR.	N. N.					
Total   Signoo	767225 MSMT/CERCARIA PRODUCTION PER SNAIL	PTotal	1000000							
Total   Tota	152810	Total	000000	1						
Total   Colored   Colore	200000000000000000000000000000000000000	Total	752000							
Total   Section   NR	152807	Total	60500			Suppopular/				
Total	152809	Total	462000			000760				
Total   6500000	153137 MSMT/MEAN PER SNAIL PAIR/	Total >	338000	NR	NR					
Total   G52000	759367 MSMT/COMBINED THREE TESTS, EGGS PER	Total	9000009	NR	NR					
Total   1,495,000	148468 prs	Total	625000	NR	NR					
Total         NR         NR           Total         1,55000         NR         1,56809           Total         1,252000         NR         1,56809           Total         2,041000         NR         1,56809           Total         2,051000         NR         NR           Total         1,50000         NR         NR           Total         455,000         NR         NR           Total         2,50000         NR         NR           <	148476 prs	Total	496000	NR.	NR					
Total   1260000	148475ers	Total	715000	NR	NR					
Total   1252000	148470prs	Total	1060000	NR	N.					
Total   1266/90	(48469Prs	Total	1252000							
Total   3650000   NR   1535210   NR   2782460     Total   1500000   NR   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   17000   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   1500000   NR   NR   NR     Total   15000000   NR   NR   NR     Total   10000000   NR   NR     Total   100000	59076general)	Total	1266760	325610		8				
Total   1,000,000	SSU//general)	otal	2061180	1		2262460 ug/				
Total   1200000	590/4 general)	Total	3650000	N.S.	NR	/Bn				
Total   1,9000	59U/5general)	iotal	1060000	Z.	NR	/Bn				
Total	117032 brs	Total	150000	NB	NR	/Bn				
Total         NR         NR           Total         130000         NR         204000           Total         137000         132000         232000           Total         246500         172000         NR           Total         246500         NR         172000         NR           Total         645000         NR         NR         235000           Total         652000         NR         NR         NR           Total         855000         NR         NR         NR           Total         265000         NR         NR         NR           Total         39600         NR         NR         NR           Total         130900         NR         NR         NR           Total         1000000         NR         NR         NR           Total         1000000         NR         NR         NR           Total         10000000         NR         NR         <	61597 prs	Total >	1381000	NR	NR	/Bn				
Total   C   130000	48467 ers	Total	766000	NR	RN	/an				
Total   137000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   350000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   350000   350000   3500000   350000   350000   350000   350000   35000000   3500000   3500000   3500000   3500000   35000000   35000000   3500000   3500000   3500000   3500000   35000000   35000000   35000000   35000000   35000000   35000000   35000000   35000000   35000000   35000000   35000000   350000000   35000000   35000000   350000000   35000000   35000000   35000000   35000000   35	17033 prs	Total <	150000	NR	NR	l/gu				
Total         465000         338000         358000           Total         246000         NR         17200         NR           Total         625000         NR         NR         NR           Total         852000         NR         601000         NR         1036000           Total         780000         NR         NR         NR         NR         NR           Total         394000         NR         NR         NR         NR         NR           Total         1360200         NR         NR         NR         NR         NR           Total         1360200         NR         1558200         NR         NR         156820           Total         265000         NR         1558200         NR         NR         156820           Total         355000         NR         NR         NR         NR         156820           Total         1000000         NR         NR         NR         NR         156820           Total         1000000         NR         NR         NR         NR         156820           Total         1000000         NR         NR         NR         NR         NR	117036prs	Total	137000			204000 ug/l				
Total         246000         NR         172000         NR           Iotal         625000         NR         NR         NR           Iotal         625000         NR         601000         NR         1036000           Iotal         850000         NR         601000         NR         1036000           Iotal         984000         NR         NR         NR         1036000           Iotal         1360900         NR         NR         NR         NR           Iotal         1360900         NR         155510         NR         156600           Iotal         1000000         NR         NR         NR         NR           Iotal         10000000         NR         NR         NR         NR           Iotal         10000000         NR         NR         NR         NR	117034 prs	Total	465000	358000		\$92000 ug/l				
Total   1250000	117035 prs	Total	246000	172000		326000 ug/l				
Total   654500   NR   NR   NR   NR     Total   855000   NR   601000   NR   1036000     Total   855000   NR   601000   NR   NR     Total   856000   NR   NR   NR     Total   135000   NR   NR   NR     Total   1360300   NR   NR   NR     Total   2061180   NR   NR   NR     Total   2061180   NR   NR   NR     Total   2060000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR   NR     Total   10000000   NR   NR     Total   1	117043 prs	Total	1250000			l/an				
Total	)17044 prs	Total	645000	NB	N.	/80				
Total   285,500   NR 601000   NR 10360000   NR 1036000   NR 1036000   NR 1036000   NR 1036000   NR 1036000   NR 1036000   NR 1036000   NR 103600000   NR 10360000   NR 10360000   NR 10360000   NR 10360000   NR 10360000   NR 10360000   NR 10360000   NR 103600000   NR 1036000000   NR 103600000   NR 103600000   NR 103600000   NR 103600000   NR 103600000   NR 103600000   NR 103600000   NR 103600000   NR 103600000   NR 1036000000   NR 103600000000   NR 103600000000000000000000000000000000000	11 7047 215	Total	622000	2 Z	NBN	/61				
October   700,000	17046575	Total	965000		CN	Jan Cooper				
1014    200000	CCCCC MACANI OF STREETS	Total	0000002			TOSPONO NEVI				
rotal         1949         NR         NR           Total         1965/00         NR         NR           Total         1369/20         NR         NR           Total         2061/20         NR         NR           Total         2061/20         NR         NR           Total         355000         NR         NR           Total         1000000         NR         NR	SCHOOL MICHIGAN OF STEETS	Total	024000	22	CON CON	1/8n				
Total   135000	SCOOL MACHATINES STEELS	Total	224000	NN.	NA C	US/L				
Total         13,9000         NR         NR           Total         13,00300         NR         15,5610           Total         2661380         15,3510         NR           Total         266100         NR         NR           Total         1000000         NR         NR           Total         1000000         NR         NR           Total         1000000         NR         NR           Total         1000000         NR         NR           Iotal         NR         NR         NR           Iotal         NR         NR         NR           Ictal         NR         NR         NR           Ictal         NR         NR         NR	DOLOG INDIVITING AN OF 3 ESTS!	Total	nongoe	N.Y.	NX.	1/Bn				
Total         1.266/200         NR         325510         VR         1568/200           Total         2.061180         1.55510         2.262/460         2.262/460           Total         1.0660000         NR         NR         NR           Total         1.000000         NR         NR         NR           Total         1.000000         NR         NR         NR           Total         1.000000         NR         NR         NR           Total         1.256000         NR         NR         NR           Total         1.000000         NR         NR         NR	SOCIAL MISINITY MERINGE STEELS	Total	1300000	NA NA	N N	1/Bn				
Total   1269.00   1558.0	2130000	Total	7370300	NK	NK					
Total   250,000   NR   NR   NR   NR   NR   NR   NR	27.003.63	Potal	1799/90	325610		920				
Total         300000         NR         NR           Total         1000000         NR         NR           Total         1000000         NR         NR           Total         1000000         NR         NR           Total         123000         NR         NR           Total         NR         NR         NR	27.000.13	Total	201190	1555210		77P7 08/1				
Total   1,000.000	32501 55	Total	3550000	N. S.	NK.	Ug/L				
Total   1000000	7591 Brs	Total	1060000	N.S.	a. N	1/8n				
Total   128200   NR   NR   NR   NR   NR   NR   NR	56354 MSMT/MEASURED IN F3 GENERATION/	Total	1000000	NR	NR	1/Bn				
Total   1000000   NR   NR   NR   NR   Lotal	56229brs	Total	1458000	NR	NR	1/Bn				
Total   1000000   NR   NR   NR   Total   Total   NR   NR   NR   Total   NR   NR   Total   NR   NR   Total   NR   NR   NR   Total   NR   NR   NR   NR   Total   NR   NR   NR   Total   Total   NR   NR   NR   NR   Total   To	56344 Brs	Total	1000000	NR	SS	1/Bn				
Total   NR 1286000	66355 MSMT/MEASURED IN F4 GENERATION/	Total	1000000	NR	NR	1/Bn				
Total   NR   >   0   1236000   1246000   1236000   NR   NR   1236000   NR   Teats   1400000   NR   1236000   NR   1236000   NR   1236000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   1236000000   NR   1236000000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   123600000   NR   1236000000   NR   1236000000   NR   1236000000   NR   1236000000   NR   12360000000   NR   1236000000000   NR   12360000000000000   NR   1236000000000000000000000000000000000000	56230 prs	Total	1236000	NR	NR	1/Bn				
Total 1000000 NR NR	66231 Frs	Total	NR ×	o		1236000 ug/L				
T > + > 1	66351 MSMT/MEASURED IN F2 GENERATION/	Total	1000000	Z	NR	l/an				
1000000 NR	66356 MSMT/MEASURED IN PS GENERATION/	Total	1000000	NR	NR	I/an				

USC   USC
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mg/L CaCO3	Mod Hard	Mod Hard	Mod Hard	Mod Hard	N. S. S. S. S. S. S. S. S. S. S. S. S. S.	NR	N.	NR	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	N. N.	an an	TANK CACOS	mg/L cacus	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	N.S.	NR	NR	NR	NR	NR	NR	N.S.	Z.	NR	N.S.	NR	NR	NR	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	ZZ	Z.	mg/L CaCO3	mg/L cacos	NA CA	NB	ma/l CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	NR	NR	N.S.	NR	NR.	NR.	N N	NAN	Na Na	NR	NR	NR	mg/L	mg/L CaCO3	mg/L	mg/L	mg/L	mg/L	mg/LCaCO3	mg/L CaCO3	mg/Lcacos	mg/L cacos	mg/L	NR	NR.	NR	VR	me/L CaCO3	IIB/L Cacoo	N. N.
									94	94	28	28	8			00	7	94	\$6	36															100	100	100	100	100								100	100	100	100																															
mg/L CaCO3																																																																		4															
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2000	0,05			50.0						0.05	0.05	0.05	50.0			90.0	000	200	0.05	0.05				0.01	0.3	0.005	0.001	0.05	0.84	0.34	0.065	0	0.05	0.05			0.01		0.01	0.05	0.002	0.1064	0.03	30.0	0.00	0.0	0.01			0,01		0.113	0.05	0.05	0.05	0.066	0.824	0.477	0.824	0.874	0.05	0.05	0.05	0.05	0.01	0.05	0.05	0.05	0.05	0.01		100	1000	0.05	0.657				0.01	1000	
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793900 Significant	9 No significance	O Not applicable	2 Not applicable	1 No significance	7 Not applicable	5 No significanc	6 Not applicable	4 Significant	7 Not applicable	1 No significano	6 No significano	1 No significano	3 Significant	1 Not applicable	3 Not applicable	7 No significance	A NO SIGNIFICATION	Not applicable	2 No significano	o No significano	6 Significant	1 No significano.	2 Not applicable	1 Significant at a	5 Not significant	S No significano.	9 No significano	759438 Significant	Not significant	/ Not significan	Not significant	/ No significance	4 Significant	6 Significant	2 Not applicable	2152813 Not applicable	5 Significant	I Not applicable	4 No significance	5 Significant	Significant	No significance	o No significance	Not applicable	Not significant	Not significant	Significant	Not applicable	Not applicable	No significance	Not applicable	2162413 Not significant at all concentrations	Not significant	No significance	Significant	Significant at a	Not significant	Not significant	2163525 Not significant at all concentrations	Not significant	Significant	No significance	Not significant	Significant	Not significant	Not significant	Significant	Not significant	No significance	No significance	Not applicable	Not applicable	Not applicable	No significance	No significance	Not applicable	761548 Not applicable	Not applicable	Not significant	Not significable	Not approach
793900	206010	206011	206014	206014	71859	71859	718596	72814	8896	4317	9688	9689	9689	76177	76177	9689	0000	2000	9690	4316	729236	71861.	71861.	215406.	215406	75944	75943	75943	75941	15839	75942	75943	75944	759436	215281.	2152813	215281	215281.	2152814	201554	216050	215049	201555	20155.	74041.	773638	2152819	2152817	2152816	2152818	2164559	2162413	741641	741651	741651	2165532	2165533	2060023	2154068	2165524	741597	741576	741623	202204	741685	251448	251447	251464	251446	2152749	2152/48	14/7577	2132/30	202203	2163091	761549	761548	761567	784790	761560	20100

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		0.05	NR		NR			86		94 mg
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18/922 Not applicable 2148459 Significant	NA 0.05		NR NR	ZZZ					80	06	ng/L CaCO
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200	NA		NA	42					0	8	IR Call
768170 Not applicable	NA		NA	d'N						R	IR.
160280 Not applicable	NA		NA	AN.							R
3899 Not applicable	NA PO		NA ND	AN IN			44				mg/L CaCO3
Not applicable	AN AN		NA	AN			177				ng/L caco
0131 Not applicable	4N		AN.	NA.							fod Hard
0128 No significance	0.05		NR	NR							1od Hard
0281 Not applicable	NA		NA	NA	(A)	- C				-	В
768261 Not applicable	NA		NA	NA						-	ж
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SSES Not applicable	t e		AN AN								2 0
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8357 Not applicable	AN		NA NA	AN							e a
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768257 Not applicable	NA		NA	NA		1 1 2				2	В
7030 Not applicable	NA		NA	NA			93	l l			1/8:
2358 Not applicable	NA		NA	AN		93*				2	1/8t
5611 Not applicable	NA		NA NA	AN AN					00	2 000	R COOL
Sold Not applicable	NA NA		W. C.	AN AN				-	07	130	ig/L cacc
5610 Not applicable	AN		A A	AM				4 12	120	130 0	a/l Caco
5652 Not applicable	AN		NA NA	AN					200	130 1	שלו השכנ
5615 Not applicable	4Z		NA	AN				1	20	130 n	e/L CaCC
6514 Not applicable	AN		NA	NA				17	20	130 n	a/L Cacc
621 Not applicable	AN		NA	NA				11	20	130 п	e/L CaCo
613 Not applicable	AN		NA	AN				1.	20	130 m	g/L CaCO
616 Not applicable	N.A.		NA	AN				1	20	130 п	R/L CaCO
620 Not applicable	NA		NA	NA				1.	50	130 n	g/L CaCO
612 Not applicable	NA		NA	NA				1.	20	130 п	g/L CaCO
617 Not applicable	NA		AN	NA				1.	20	130 n	g/L CaCO
608 Not applicable	NA		NA	AN				T	50	130 n	g/L CaCO
165619 Not applicable	NA		NA	INA				1	120	130 n	g/L CaCO
607 Not applicable	NA		NA	AN	1			4	50	130 m	g/L CaCU
623 Not applicable	NA		NA	AN A				1	50	130 m	g/L CaCu
518 Not applicable	A .		NA NA	AN .:				1	50	13	g/L Caco
543 Not applicable	AN		A P	NA		X TO		16*			g/L CaCO
542 Not applicable	AA		AA	NA				16*			g/L CaCO
541 Not applicable	NA		AA.	AN				16*		20*	g/L Cacc
323 Not applicable	NA		NA	NA				40*			g/L CaCO
136 Not applicable	AN		NA	NA			124			E	g/L Cacc
198 Not applicable	NA.		NA.	NA			124				g/L CaCo
326 Not applicable	A S		AA .	AN AN				40.		18.	g/L caco
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108 No significance	90'0			NB						2	od Hard
111 Not applicable	NA		A.A.	AN						2	od Hard
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878 Not applicable	NA		A.A.	AN						Z	~
170 Not applicable	NA	_	AA.	NA						Z	~
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167 Not applicable	NA	4	JA.	NA			100			Z	2
169 Not applicable	NA	4	JA	NA						Z	~
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474 Not applicable	NA	2	IA	NA			170			ш	3/L CaCO3
801 Not applicable	AN	~	IA.	NA			106			E	3/L CaCO
477 Not applicable	NA	2	NA.	AN			170			8	S/L CaCO
809 Not applicable	AN	6	Ą	AN			106			E	Z/L CaCO3
300 Not applicable	AN	~	IA.	AN :			106			E	3/L CaCO3
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194 Not applicable	NA	4	IA.	NA						Z	
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COTOX (Unfiltered) Statistical Significance Level Med Significance Level Med Significance		Not applicable Not applicable	Not applicable			2153706 Not applicable NA		181627 Not applicable NA							181621 Not applicable NA				8825 Not reported															165598 Not applicable						165605 Not applicable NA			221428 Not applicable NA			221429 Not applicable NA											40518 Not applicable NA					Not applicable							2153894 Not applicable         NA           18172 Not applicable         NA           18173 Not applicable         NA           18173 Not applicable         NA

ppm CaCO3	ppm CaCO3	nom CaCO3	ppm CaCO3	ppm cacus	ppm CaCO3	mg/L	NA NA	N N	NR	NR	N.	NR	NR	NR	ZZ.	NK	NR.	NA NA	NA CAN	NN	NR	ZR	NR	NR	NR.	NR	NR	NR	NR.	N.S.	Z S	Z L	Z Z	X C	N N	ND	NB	NB	N. N.	N.	NR	NR	N.	NY.	N N	2 N	N. N.	NR	N.S.	Z.	NR	NR	N.S.	Z Z	NK NK	N. N. N. N. N. N. N. N. N. N. N. N. N. N	NR	NB	NR	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	mg/L CaCO3	NR	N.S.	mg/L CaCO3	N.S.	NR	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	NR	NR	NR
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tapplicable	NA	NA		AN			2.7	35		76 m	Mg/L CaCO3
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it applicable	NA	AN		NA				42		m 06	g/L CaCO3
t applicable	NA	NA		NA				40		70 m	g/L CaCO3
t applicable	NA	NA		NA				18		58 m	g/L CaCO3
nificant at all concentrations	0.0193	N.		NR						Z	- 2
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	474 Not applicable	AN		NA	Z Z Z	207	56			mg/L CaCC
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No. 10.   No.	842 Not applicable 556 Not applicable	A AN		A N	A A			OS		100 mg/L CaCC
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0.05   NR	953 Not applicable	NA		NA	NA					NR
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0.02	513 Not significant at all concentrations		0.05	N. N.	N. N.					Z Z
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0.055   NR   NR   NR     0.055   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.056   NR   NR   NR     0.057   NR   NR   NR     0.058   NR   NR   NR     0.059   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   NR   NR   NR     0.050   N	692 No significance		0.05	NR	Z.					S.S.
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NA   NA   NA   NA   NA   NA   NA   NA	82 Not applicable	NA		NA	NA		292			mg/L CaCO
NA   NA   NA   NA   NA   NA   NA   NA	83 Not applicable	NA		NA	NA		322			mg/L CaCO
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	79 Not applicable	NA		AN.	AN		278			mg/L CaCO
NA NA	84 Not applicable	AN		Z Z Z	NA NA		322			mg/L CaCO3

815 Not applicable	NA NA	NA NA	NA NA	NA NA	naturess Weart Op Datum	naturess iviedit naturess ivin	5	nardness Min Hardness Max Up Inardness Max Hardness Unit 160* mg/L CaCO3	Op Hardness May	mg/L CaCO3
2052778 Not applicable	NA	Z Z Z		NA NA		322				mg/L CaCO3 mg/L CaCO3
052770 Not applicable	AN A	A A		NA NA		47				NR.
6	NA	NA		NA		66				mg/L CaCO3
741558 Not applicable	NA NA	A AN		AN				170	192	mg/L CaCO3
41594 Not applicable	NA	NA		NA				170	19.	mg/L CaCO3
24806 Not applicable 64655 Not applicable	AN N	NA NA		NA			160*		180*	mg/L CaCO3
Not applicable		NA		NA						N. N.
2164654 Significant at all concentrations	0.001	N.		NR						NR
64656 Not applicable	A A	A A A		AN AN						NR
96203 Not applicable	NA	NA		NA		10				mg/L CaCO3
10845 Not applicable	NA	NA		NA			140*		160*	mg/L
18184 Not applicable	4 4 7	A N		NA				000		NR NF
01540 Not applicable	NA	AN		NA		101.7		23.5	70	mg/L CaCO3
13417 Not applicable	NA	NA		NA						NR
740684 Not applicable	NA NA	AN A		NA		140				mg/L CaCO3
20765 Not applicable	T de Z	NA		AN		140				mg/L CaCO3
24820 Not applicable	NA	NA		NA			160*		180*	mg/L CaCO3
24821 Not applicable	NA	NA		NA			160*		180*	mg/L CaCO3
31519 Not applicable	NA	NA		NA	100*					mdd
31583 Not applicable	AZ .	NA		NA	100-					ppm
31522 Not applicable	NA NA	4 2 2		A NA	100-					mdd
31 582 Not applicable	AN AN	42		NA NA	100*					mdd
31521 Not applicable	I d	2 2		AN AN	1001					mdd
31518 Not applicable	AZ	AN		AN	1001					midd
31520 Not applicable	NA	NA		NA	100*					mdd
03677 Not applicable	NA	NA		NA		100.8				mg/L CaCO3
03678 Not applicable		NA		NA		100.8				mg/L CaCO3
54076 Not significant at all concentrations	0.05	Z Z		Z Z						NR.
60018 Stenificant	0.05	N. N.		NB						NA NA
60020 Not significant at all concentrations	0.382	Z.		NR						NR
2059995 No significance	0.05	N.		NR						NR
2059533 Not significant at all concentrations	50'0	NR		NR						NR
76801 Not applicable	NA	NA		NA			160*		180*	mg/L
11963 Not applicable	NA	NA		NA						NR
2155523 Not significant at all concentrations	0.274	C an		a a						NA
	NA	A.N		NA				80	1001	me/l CaCO3
0317 Not applicable	NA	AN		NA	296*					mg/L CaCO3
0288 Not applicable	NA	NA		NA	29€*					mg/L CaCO3
0318 Not applicable	NA	AN		NA	\$36¢					mg/L caco3
0291 No significance	0.01	N. S.		NR	296*					mg/L CaCO3
0315 Not applicable	1 4 2	AN AN		AM AM	.962					mg/L cacos
0319 Not applicable	T d	42		42	*300					mg/L Cacos
0292 Significant	0.01	NR		NR	296*					mg/L CaCO3
0307 Not applicable	NA	NA		AN	296*					mg/L CaCO3
0302 Not applicable	NA	NA		NA	₹36₹					mg/L CaCO3
0309 Not applicable	NA	NA		NA	₹36*					mg/L CaCO3
0290 Not applicable	AN	N.A.		AN	296*					me/I CaCO3
0276 Not applicable	NA	NA		NA	296*					mg/L CaCO3
0293 Not applicable	AN	NA		NA	₹36₹					mg/L CaCO3
0306 Not applicable	NA	NA		NA	296*					ng/L CaCO3
8185 Not applicable	NA	NA		NA						Z.S.
4505 Not applicable	NA	NA		NA						Z.E.
8186 Not applicable	NA	NA		NA						VR
1739 Not reported		NR		NR						Z.R.
4605 No significance	0,05	NR		NR						4R
4606 Significant	0.05	NR		NR						48
4588 Significant	0,05	NR		NR		1				Z.R.
2164587 No significance	0.05	NR		NR						NR
4592 Significant	50:0	NR		NR						4R
4591 No significance	0.05	NR		NR						A.R.
1600) Significant	0.05	NR		NR						IR.
2164599 No significance	0.05	N. S.		NR						J.B.
tous no significance	0.05	NR		NA.						IR.
1609 Significant	0.05	NR		NR						IR
4603 Significant	0.05	NR		NR						IR.
1602 No significance	0.05	N.S.		NR						4R
1594 No significance	0.05	Z Z		N.S.						J. N.
See Significant	50.0	N. A.		NK						H H
557 No significance	0.05	N N		Z Z					I	X G
741624 Significant	0.05	NR		NR			-			2 2
SSO Significant	200	2		(41)						
0	0.00	INK		N.S.						IR

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2163112 Significant 2163110 Not applicable 2163109 Not applicable 2163101 No significance	740693 Not	741709 Not	201543 Not	231546 Not	231545 Not	231549 Not	231554 Not	231544 Not	231555 NOT	231542 Not	231551 Not	231548 Not.	231540 Not applicable	231539 Not	231538 Not appli	231552 Not.	231541 Not	196750 Not	196772 Not	196755 Not	196726 Not	196768 Not	196767 Not	196713 Not	196744 Not	196769 Not.	196724 Not	196763 Not a	740694 Not	759180 Not	740697 Not applicable	196745 Not	196714 Not	196728 Not	196741 Not	196756 Not	196734 Not	196760 Not a	196759 Not a	196761 Not a	96738 Not a	96839 Not a	96770 Not	96720 Not a	196753 Not a	196766 Not a	196757 Not a	196732 Not a	196740 Not a	768104 Not r	201560 No si	768072 Not r	768073 Not reported	768158 Not r	768157 Not r	201562 Not a	768066 Not n	768059 Not r.	168058 Not r	768097 Not n	201537 Not a	768128 Not reported

196716 Not applicable	NA	NA NA	ברבאבו ואווון פולווורפוורפ רבאבו ואופץ כול	NA NA	naturiess intean Op	narghess iviedin nan	diress with Op hard	raness Min Hardness Max Op	Hardness Max	Hardness Units SOFT
7749 Not applicable 7722 Not applicable	AN	A A		NA NA					S	SOFT
751 Not applicable	NA	NA		NA	-				SO	H
711 Not applicable	NA	A A A		NA NA					S	E 1
712 Not applicable	NA	NA		NA					S	E
764 Not applicable Not applicable	4 4 4 A	A A A		A A A					8 8	E
746 Not applicable	N.A.	NA		NA					SOS	t
733 Not applicable	NA	AN		AN					OS (S	11
773 Not applicable	AN	AN AN		Z Z Z					SS	E
725 Not applicable	NA	NA		AN					SOS	FT
196752 Not applicable	NA	NA		NA					SO	L.
730 Not applicable	AZ Z	Z Z		A A A					SOS	
727 Not applicable	NA	AN		NA					OS OS	t
771 Not applicable	NA	NA		AN.					OS	13
739 Not applicable	4 4 4 4 A	A A		NA					S	tal ta
754 Not applicable	NA	NA		NA					OS OS	, t-
717 Not applicable	NA	NA		NA					SOI	15
747 Not applicable	NA	NA S		NA.					SOS	1
745 Not applicable	42 2	A S		NA					SOS	
12 Not applicable	AN AN	Z Z		NA NA					SOS	
144 Not reported	NR	NR		NR					S. N	
143 Not reported	NR	NR		NR					NR	
090 Not reported	NR	NR S		Z Z					NR.	
091 Not reported	XX XX	X X		X X				- G	NR AZ	( CaCo
549 Not applicable	AN AN	4N		AN				46	70 mg	/L CaCO
550 Not applicable	NA	NA		NA				44	68 mg	/L CaCO
542 Not applicable		AN		NA				52	96 mg	/L CaCO3
400 Not significant at all concentrations	0.05	N S		S S				90	110 mg	7
666 No significance	0.05	NR		NR				06	110 mg	1 2
907 Not significant at all concentrati>	0.05	NR		NR			4 1		NR	
914 Not significant at all concentrat >	0.05	NR		NR					NR	
089 Not reported	XX GX	N N		N N					Z Z	
124 Not reported	Z.S.	NR		NR					N. N.	
123 Not reported	NR.	NR		NR					NR	
.63 Not reported	NR	NR		NR					N N	
64 Not reported	NR.	N G		N G					N S	
54 Not reported	2 2	N N		E 22					N. N.	
998 Not reported	NR	N.		N.					NR	
999 Not reported	NR	NR		NR					N.	
74 Not reported	NR	N.		NR					NR	
75 Not reported	N. C.	N S		N.S.					NA.	0.0
55 Not applicable	A a a	A an		AN W		120			Mg/	L CaCO
81 Not reported	N.S.	NR		NR				l	N N	
59 Not applicable	NA	AN		NA					N N	
29 Not applicable	NA	NA		NA					NR	
Not applicable	NA.	AN		NA					NB	
070 Not applicable	NA AN	A A		NA					2 2	
277 Not applicable	C d	AN		NA					2 2	
332 Not applicable	NA	AN		NA					A.S.	
58 Not applicable	NA	NA		NA					N.	
325 Not applicable	NA	NA		N.A.					NR	
324 Not applicable	NA	A S		NA					N. N.	
999 No significance	0.05	N S		N N				98	94 mg/	L CaCO3
65 Not applicable	AN SOCIO	NA AN		NA NA		210-		900	100 E	L Cacos
361 Not applicable	NA	NA		NA		210*			/Bw	L CaCO3
83 No significance		NR		NR				36	94 mg/	L CaCO3
85 No significance	0.05	NR		NR				98	94 mg/	L CaCO3
56 Significant	0.05	N S		NR.				86	94 mg/	L CaCO3
13 Not reported	Z Z	2 2		2 2					E E	
08 Not reported	Z	Z.		NR					NR	
07 Not reported	NR	NR		NR					NR	
38 Not applicable	NA	NA		NA		96.3			/Bm	L CaCO3
33 Not reported	N. C.	N S		N S					N S	
82 Not reported	N. N. N. N. N. N. N. N. N. N. N. N. N. N	N N		N N N					N ON	
7 Not reported	2 2	Z Z		S N					N N	
768147 Not reported	NR	NR		NR			4		NR	
48 Not reported	NR	N.R.		NR					NR	
064 Not reported	NR	N.		NR					NR	

1   1   1   1   1   1   1   1   1   1	120			NR NR NR
NA   NA   NA   NA   NA   NA   NA   NA				mg/L CaCO3
NA			98	94 mg/L CaCO3
NA			98	94 mg/L CaCO3
NA			98	94 mg/L CaCO3
NA			98	94 mg/L CaCO3
NA			90 00	94 mg/L CaCO3
NA			0	NR
NA				NR
Contractions			86	94 mg/L CaCO3
NA			06	110 mg/L
NA			06	110 mg/L
NA   NA   NA   NA   NA   NA   NA   NA				NR
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NA				
NA	290*	288*	.292	mg/L CaCO3
NA		288*	292	
NA		288*	292	
NA				
NA	75.0*			mg/l CaCO3
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NA				mg/L CaCO3
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NA NA				mg/r cacoo
42			80	100 mg/L CaCO3
En!			200	103-011
NA		700	170	192 mg/L cacO3
4Z		0	170	192 mg/L CaCO3

741561 Not applicable NA	NA NA	NA NA	Significance Level Max Op Significan	CE CEVEL IVIAN MAINTENS IVIEGIL OP	Hardiess iviedii II iai siir	Op naturess wear naturess with Op naturess with hardin	ess Max Op natone	192 mg/L CaCO3
	NA	NA	NA			170		92 mg/L CaCO3
	4 4 X	4 X X	A A			170		92 mg/L CaCO3
	AN	AN	AN					N. N.
064223 Not applicable	NA	NA	NA					NR
211034 Not applicable	NA.	NA	NA					NR
67863 Not applicable	4 2 2	AN .	AN					N. S.
5/502 Not applicable	¥ 22	T S N	422					N N
18176 Not applicable	d Z	d Z	42					S S S
18177 Not applicable	NA	NA	NA					NR
18179 Not applicable	NA	NA	NA					NR
64340 Not applicable	NA	NA	NA					NR
64585 Not applicable	NA	NA	AN					NR
59286 No significance	0 900	XX XX	N N					NR.
SOST Not applicable	4N	4 N	d					N ON
59267 Not applicable	( a	( a	( d Z					N N
59266 Not applicable	AN	AN	AN					Z Z
59265 Not applicable	NA	NA	AN					N.
56604 Not applicable	NA	NA	NA		288			mg/L CaCO3
56435 Not applicable	NA	NA	NA		578			mg/L CaCO3
56426 Not applicable	NA	NA	NA		194			mg/L CaCO3
57351 Not applicable	NA	NA.	AN :					NR
pod 36 Not applicable	NA D	NA	4 2 2 C					mg/L cacos
7040 No significant	NA DA	2 02	N. N.		160			mg/L
2048 Not applicable	4 2	d d	d d		160			me/l
7057 Not applicable	ev.	AN	AN		320			1/aw
7059 Significant	C Z	a.v.	Z Z		320			T/au
7058 No significance	22	NR	Z		320			T/Sul
7040 Not applicable	AN	AN	4N		80			mg/L
2042 Significant	2	<u>a</u> 2	2		08			mo/l
7041 No cignificance	2	. Z	2		80			ma/l
2020 Not applicable	4 2	4 2	্ব ব		40			ma/l
2030 No cignificance	0 m	2 2	2		40			ma/i
2021 Significant	2	2	2		40			mo/!
6228 Not applicable	4 2	d: 2	4 2					ma/l CaCO3
OCCOUNT AND ADDRESS OF THE PROPERTY OF THE PRO	0.00	S. V.			-00			mg/c cacos
SOCIAL NOT Applicable		0.2	2 2		200			mg/cacco
6223 Similifering	50.0	02 Z	2 2		92*			mg/L cacos
5224 Digillication	600	× × ×	C 22		200			mg/L Cacos
SAZI Not applicable		T.	T. 22		76			mg/L Cacos
5222 Significant	SO'O	N. C.	N. C.		76			mg/L cacos
5225 No significance		22	XX.		.76			mg/L cacus
5226 Not applicable	NA.	AN	AN ::		-26			mg/L CaCO3
9078 Not applicable	AA	AZ.	NA.					NR
9072 Not reported	NR	NR	N. N.					NR
9073 Not reported	NR	NR	N.S.					NR
1596 Not applicable	NA	AA	A A					Z.
7459 Not applicable	NA	NA	NA					NR
5432 Not applicable	AA	NA	NA		484			mg/L CaCO3
7457 Not applicable	NA	AA	NA					NR
5427 Not applicable	4X	NA	A N		194			mg/L CaCO3
3330 Not applicable	NA.	NA	AA					N.
7456 Not reported	NR	Z.S.	NR					NR
7412 Not applicable	NA	NA	AN					Z Z
7375 Not applicable	NA.	AN	AS					NR
5430 Not applicable	NA	NA	AN		390			mg/L CaCO3
7380 Not applicable	AN	AN	AN					NR
7353 Not applicable	AN	AN	AN					N.
5418 Not applicable	d Z	AN	AN.		107			mg/L CaCO3
5433 Not applicable	AN	AN	AN		484			mg/L CaCO3
7350 Not applicable	d'X	4N	42					NR
2428 Not applicable	d N	4N	42		288			mø/l. CaCO3
2411 Not soulicable		9	2		2			NR NR
art increphicane	4.4	(	C V		501			100 COCO
and applicable	T.N.	471	T TO		707			mg/c cacos
5437 Not applicable	NA	NA.	42.		- 250			mg/L cacos
5431 Not applicable	NA	NA	AN		390			mg/L CaCO3
5215 Not applicable	NA	NA	INA	31	94*			mg/L CaCO3
0331 Not applicable	NA	NA	NA					NR
5429 Not applicable	NA	NA	NA		288			mg/L CaCO3
5434 Not applicable	NA	NA	NA		578			mg/L CaCO3
7454 Not applicable	NA	NA	NA					NR
766216 Not applicable	NA	NA	AN	6	****			mg/L CaCO3
TOTAL SAMILARIA	MA	NA	4Z					NR
766576 Not applicable	DIN DIN	Q. V	d		886			mp/l CaCO3
20/6 NOT applicable	EN.	WA!	CAL VIN		2 00			118/1 Carres
opos Not applicable	T.V.	NA	T .		5 1			mg/L cacus
5554 Not applicable	NA	NA	NA		102			mg/L CaCO3
5567 Not applicable	NA	NA	NA		96			mg/L CaCO3
580 Not applicable	AN	NA	NA		96	11		mg/L CaCO3
5565 Not applicable	NA	NA	NA		8			mg/L CaCO3
766607 Not applicable	NA	AN	NA		390			mg/L CaCO3
SSENOt applicable	NA	NA	AN		9%			mg/l CaCO3
COOR INCLUDIOSES	1				-			100000000000000000000000000000000000000
CC730 Mot anniicable	ΔM	ΔN	AN.	į	100			mg/L CaCO3

Hardness Max Op Hardness Max Hardness Uni	NR	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	X X	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR ,	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	mg/L CaCO3	NR	mg/L CaCO3 -	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	N N	694* mg/L CaCO3	mg/L CaCO3	NR	mg/L CaCO3	mg/L caco3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	N. N.	N.S.	N N	N.S.	N N N	mg/L caco3	mg/L CaCO3	NR NB	N. N.	unil
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766574 Not applicable	39 5	766717 Not applicable	766560 Not app	766603 Not app	766581 Not app.	766722 Not app	766730 Not rept	766731 Not rept	766578 Not app 766595 Not appl	766586 Not appl	766610 Not app.	766555 Not appl	766566 Not appl	766570 Not apply	766723 Not appl	766591 Not appli	766724 Not appl	766572 Not appl	766606 Not appli	766585 Not appli	766608 Not appli	766551 Not appli	766563 Not appli	767586 No signifi	767587 Not appli	766592 Not applic	766726 Not appli	766587 Not applic	766612 Not appli	766561 Not appli	766593 Not applic	766594 Not appli	756584 Not applic	766217 Not applic	18144 Not applic	18145 Not applie	767473 Not applic	759175 Not applic 767575 No signific	767574 Significant	119467 Not applic 767294 Not applic	767307 Not applic	767296 Not applic	767303 Not applic	767295 Not applic	767576 Not applic	18452 Not applic	18147 Not applic	18454 Not applic.	18451 Not applicable	18146 Not applica	759090 Not reported 759090 Not reported	759092 Not report	767305 Not applica	767299 Not applica	767494 Not applica	767475 Not applica	constalator soulies

NR NR NR	X X	NR	NR ON	144 mg/L CaCO3	140 mg/L CaCO3	144 mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L	mg/L	mg/L	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	116 mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3		142 mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	116 mg/L CaCO3	116 mg/L CaCO3	NR.	N N	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3
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				139	134	139	292	392	100	100	98 00	396	96	100	175	396	482	100	96	86	100	100	1001	86	80	80	08	482	100	100	292	100	00+	100	86	296	192	100	137	141	20	102	75	102	102	62				123	100	107	94*	200	94*	100	300	200	107	300	100	100
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Not applicable Not applicable Not applicable	ot applicable	ot applicable	Not applicable Not applicable	ot applicable	764943 Not applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	of applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	gnificant	or applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	ot applicable	o significance it applicable	ot applicable	it applicable	ot applicable	t applicable	767311 Not applicable	t applicable	nificant	767600 No significance	treported	T reported	t applicable	rt applicable	t applicable	t applicable	766441 Significant at all concentrations	Not applicable	nificant at all concentration	Not significant at all concentrations	nificant at all concentrations	Not applicable	it all conc	3 Not applicable	t applicable
160333 NG 767505 NG 767507 NG	2064207 Nc	2064204 N.	2064206 No	764940 N	764943 N.	764936 No	766638 Ni	766639 NC	766712 NG	766716 N.	765513 N	766641 No	766616 Nc	766625 No	765631 No	766640 Nc	766644 No	766627 Nr	766624 No	766619 NC	766714 NC	766715 NC	766710 No	766623 No	2017143 Nc	2017146 Sig	2017113 NC	766643 No	766630 No	766711 NO	766637 No	766622 No	767601 NC	766628 No	766626 No	766636 No	766634 No	766621 No	2017145 No	764947 No	767290 No.	767310 No	767291 NO	767311 No.	767312 No	767599 Sign	767600 No	759093 No	159095 No.	767292 No.	766440 No.	766416 Not	766367 NO	766441 Sign	766365 Not	766439 Sign	766444 No.	766442 Sign	766415 Not	766659 Not	756663 Not	766664 Not

N	766658 Not applicable				NA NA		-	Tal calcala	initiess intex op param	ESS IVI da
	766666 Not applicable	AN AN		NA	N N		100			mg/L CaCO3
	766657 Not applicable	AN AN		TZ V	A A		100			mg/L C
	766656 Not applicable	AN		NA	NA		100			mg/LC
	766665 Not applicable	NA		NA	NA		100			mg/L C
	200196 Not applicable	4 4 2		AN AN	A A		200			mg/L C
No. 10.   No.	40509 Not applicable	NA		NA	NA		2	35		62 mg/L C
	40508 Not applicable	NA		NA	AN			35		62 mg/L C
March   Marc	13180 Not applicable	4 2 Z		AN ON O	4 < Z					N S
No.   No.	126071 Not reported	NR		NB	α Z					S S
Mail	126072 Not reported	NR		NR	NR					Z.
No. of the control	126070 Not reported	NR		NR	NR					NR
No.   No.	764588 Not applicable	NA A		NA	A S					Z :
100   101	18151 Not applicable	T d		T AZ	1 4					Z Z
Mail	18152 Not applicable	NA		NA	AN					N.S.
Mathematical Control	18149 Not applicable	4N		NA	NA					NR
	6211 Not applicable	NA		NA	NA					NR
No.   No.	41536 Not applicable	4 × 3		d d	AN NA					Z :
No. 10.   No.	802668 Not applicable	C V		AN AN	र व					Z Z
No. 10.   No.	41641 Not applicable	NA		NA	NA					N.
No.   No.	802670 Not applicable	NA		NA	AN					NR
No. 10.   No.	802671 Not applicable	4Z		NA	NA					NR
Mail	A1642 Not applicable	4 × 2		AZ	42.2					æ č
No.   No.	802677 Not applicable	AN AN		AZ AZ	C d					e a
Mail	802667 Not applicable	4Z		NA	AN					N. N.
NA	41640 Not applicable	NA		NA.	AN					NR
NA	802664 Not applicable	NA		NA	NA					NR
NA	802665 Not applicable	NA		NA.	NA					NR
Mathematical Colored	6214 Not applicable	A		AN	NA					N. N.
14	8026/8 Not applicable	ď «2		47.2	d'A					X I
10, 10, 10, 10, 10, 10, 10, 10, 10, 10,	803669 Not applicable	T TN		T.V.	AN ON					X Z
Marked   M	6215 Not applicable	d'a		AN AN	AZ.					N N
NA	41635 Not applicable	AN		Ą.V.	AN					NR
NA   NA   NA   NA   NA   NA   NA   NA	6212 Not applicable	NA		AA	AN					NN
NA	802672 Not applicable	AN		NA	NA					NR
NA	6213 Not applicable	NA		NA	NA NA					NR
NA   NA   NA   NA   NA   NA   NA   NA	802663 Not applicable	A		A S	A Z					N. S.
MA	802674 Not applicable	2 2		C di	C 4					N N
NA	802676 Not applicable	d'A		42	d Z					2
NA	41638 Not applicable	AN		NA	AN					N.N.
NA	802673 Not applicable	NA		NA	AN					NR
NA   NA   NA   NA   NA   NA   NA   NA	41639 Not applicable	AN		NA	NA					NR
NA	6218 Not applicable	NA		NA NA	AZ Z					N. S.
NR         NR         NR           NR         NR         NR           NR         NR         NR           NA         NA         NA	759009 Not applicable	C dd Z		42	X Z					N N
NR   NR   NR   NR   NR     NR   NR   N	2017191 No significance	0 00		C 22	C cc		160			INDA/I
NA         NA         NA           NA         NA         NA	766856 Not reported	NR		NR	Z.S.					NR
NR   NR   NR   NR   NR   NR   NR   NR	761598 Not applicable	NA		NA	NA					NR
NA   NA   NA   NA   NA   NA   NA   NA	2017213 Significant	NR		NR	NR		320			mg/L
NA   NA   NA   NA   NA   NA   NA   NA	2017209 Not applicable	AN		NA	NA		320			mg/L
NA	2017210 Not applicable	NA		NA	AN.		320			mg/L
NA   NA   NA   NA   NA   NA   NA   NA	201/212 No significance	EN N		N.X.	NR		370			mg/L
MA         NA         NA           NA         NA         NA	764930 Not applicable	II 4		11 4	t d		145	140		148 mg/L
NA	764935 Not applicable	NA		NA	N. A.		147	140		152 mg/L Ca
NR   NR   NR   NR   NR   NR   NR   NR	764929 Not applicable	NA		NA	NA		131	128		132 mg/L Ca
NR	2017182 Significant	NR		NR	NR		80			mg/L
NA	2017178 Not applicable	A .		NA .	NA		80			mg/L
NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NB         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA           NA         NA         NA	2017180 Not applicable	2 Z		4 2	X X		80			1/8w
NA   NA   NA   NA   NA   NA   NA   NA	2017179 Not applicable	AZ.		NA	NA		80			mg/L
NA	764932 Not applicable	AN		NA	NA		136	124		144 mg/L Ca
NA   NA   NA   NA   NA   NA   NA   NA	2017190 Not applicable	N. A.		NA	NA		160			1/gm
NA   NA   NA   NA   NA   NA   NA   NA	2017188 Not applicable	AN		NA	NA		160			T/Bu
NA   NA   NA   NA   NA   NA   NA   NA	201/105 Not applicable	t a N		NA NA	NA	105*	Ibu		I	mg/L mg/l Ca
NA   NA   NA   NA   NA   NA   NA   NA	759176 Not applicable	AN.		NA NA	NA	563*		404*	* 469	mg/L Ca
NA   NA   NA   NA   NA   NA   NA   NA	767593 No significance		0.05	NR	NR	105*				mg/L Ca
NR NR NR NR NR NA NA NA NA NA NA NA NA NA NA NA NA NA	2017168 Not applicable	NA		NA	NA.		40			mg/L
NR NR NR NR NA NA NA NA NA NA NA NA NA NA NA NA NA	2017172 Significant	RN.		N.S.	NR		40			mg/L
NA         NA         NA           NA straightealne         NA         NA           NA straightealne         NA         NA	2017171 No significance	ZZ.		N. N. N. N. N. N. N. N. N. N. N. N. N. N	NR		40			mg/L
NA AND EXPENSION NA NA NA NA NA NA NA NA NA NA NA NA NA	2017170 Not applicable	NA		NA	NA		40			mg/L
THE PROPERTY IN THE PROPERTY I	764934 Not applicable	AN AN		NA	NA		131	128		132 mg/L Ca
Not significant at all concentral(>	2163930 Not significant at all concentration			NR.	NR		177	1		N. O.
	2163923 Not significant at all concentrat			NA	6.2					2000
Not applicable NA NA	160335 Not applicable	2		100	NA.					NR
		42		NA	N A					NR N

Statistical Significance	Difficance Level Mea Sporific	cance Level Mea Signi	ificance Level Min On	Significance Level Min	Significance Level Max On Sign	ificance Level Max Hardness	Mean On Hardness Mean Hardness Min On	se Min On Hardness Mir	Min Hardness Max On H	Sardness Max Hardne	ce I Inite
Vot applicable	NA			NA	AN			-		N.	
Not applicable	NA.			NA	NA					NR	
	N N			NR N	Z Z		131		128	136 mg/L Ca	9003
767592 Significant		0.05		NR	NN					mg/L Ca	BCO3
18153 Not applicable	AN			NA	AN					N. I.	Ī
2017315 No significance	Z Z			NR	N N					MR/L	T
2017301 Not applicable	NA			NA	NA					mg/L	
2017306 Significant	N. S.			NR	AN S	NR.	15			mg/L	
2017313 Not applicable	AN			NA	AN AN					mg/L	
2017316 Significant	NR			NR	NR		i			mg/L	
2017314 Not applicable	AN			NA.	AN					T/BW	
747008 Not applicable	AN			T 42 Z	AN		**			mg/L	603
2017303 Not applicable	AN			NA	NA					mg/L	
2017305 No significance	NR			NR	NR		15			mg/L	
766706 Not applicable	NA			NA	AN					mg/L Ca	CO3
766705 Not applicable	A AN			NA	A N					mg/L Ca	503
766707 Not applicable	AN			र द	AN AN		93			mg/L Ca	503
766364 Not applicable	AN			NA	AN					mg/L Ca	CO3
766703 Not applicable	NA			NA	NA					mg/L Ca	503
766704 Not applicable	AN			NA	NA					mg/L Ca	5003
766708 Not applicable	AN			NA	AN		06			mg/L Ca	5003
161/40 Not reported	ZZ ZZ			N.Y.	Z Z					mg/L Ca	503
2052622 Not applicable	42			T d	AN AN					mg/L Ca	503
160500 Not applicable				NA	NA					NR NR	
767397 Not reported				NR	NB					NR	
767360 Not applicable				NA	AN					N.N.	
767399 Not applicable				NA	NA					N N	
767463 Not applicable				AN CO	NA .					N S	Ī
160498 Not applicable				NA	A S					Z Z	1
767317 Not applicable				NA da	Z Z					N N	T
160497 Not applicable				C d	AN N					z a	
2148118 Not significant at all concentration		0.05		NR	S					Z.	
2148117 Not significant at all concentrat <		0.05		NR	NR					N. N.	
2148116 Not significant at all concentrali		0.05		N.S.	NR					NR	
2148115 Not significant at all concentrat <		0.05		NR	NR					NR	
759021 Not significant at all concentrations		0.05		82	N. S.			2	99	1468 mg/L Ca	503
767463 Not applicable	2 2			Z Z	4 2 2					N N	T
160499 Not applicable				AN	A N					S S	I
767469 Not applicable				NA	NA					NR	
767484 Not applicable				NA	NA					NR	
759462 Not applicable				NA	NA		170			mg/L Ca(	CO3
759465 Not applicable				AN	NA		170			mg/L Ca	503
2063748 Not applicable				AZ.	AN .					Z S	T
2063749 Not applicable				42.2	AN A					S S	1
2053/20 Not applicable				C N	C Z		0.			INK MANUAL COL	200
26240 Not applicable	AN AN			1 4 2	AN AN		07			MB/L Cal	500
759073 Not significant at all concentrations		0.05		C C C	Z Z				73	763 mg/l Ca(	800
759024 Not significant at all concentrations		0:05		α :: α :: α :: α :: α :: α :: α :: α :	N N				73	763 mg/L Ca	503
160503 Not applicable				AA	AN					NR	
160502 Not applicable				NA	AN					NR	
767512 Not applicable				NA	NA					NR	
767515 Not applicable				NA	NA					NR	
160501 Not applicable				NA	NA					NR	1
749018 Not applicable				NA.	d .					N S	T
748389 Not applicable				NA NA	42					NA NA	1
767446 Not applicable				( d	AN AN					N N	Ī
767354 Not applicable	AN.			4v	A Z					NR	
767421 Not applicable	NA			NA	AN					NR	
767385 Not applicable	AN			NA	AN					NR	
767443 Not applicable	AN			NA	NA					NR	
767357 Not applicable	NA			NA	AN					NR	
767427 Not applicable	AN			NA	NA					N. S.	
767390 Not reported	NK			Z	NA			1		NA	Ī
160505 Not applicable	K KZ			NA NA	AN AN					X Z	T
767387 Not applicable	AN W			4 2 2	AN AN					N N	T
767437 Not reported	N N			( m27	Z Z					2 2	T
767432 Not applicable	NA			NA.	AN					S. S.	
767382 Not applicable	NA			AA	NA					NR	
2147175 Not applicable	NA			NA	NA					mg/L CaC	203
759765 Not applicable	AN			NA.	NA		40.5			mg/L CaC	:03
2147174 Not applicable	N.A.			NA	NA NA		1			mg/L Cac	503
767482 Not applicable	AN AN			JA JA	AN					N N	Ť
767477 Not applicable	AZ.			42	AN					. K	Ī
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AAAAAA		NA NA NA NA NA NA	NA NA NA NA NA NA			NN NN NN NN NN NN NN NN NN NN NN NN NN
AAAAAA		NA NA NA NA NA	NA NA NA NA	40.5	42	NR NR mg/L CaCO3 48 mg/L CaCO3 NR
A A A A A		NA NA NA NA	NA NA NA NA NA NA NA NA NA NA NA NA NA N			8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8
		NA NA NA NA NA NA	NA NA NA NA			N N N N N N N N N N N N N N N N N N N
2 Z Z Z Z Z		N A A A A A A A A A A A A A A A A A A A	NA NA NA NA			N N N N N N N N N N N N N N N N N N N
AAAA		NA NA NA	NA NA NA NA			N N N N S
2 2 2 7		NA NA NA	NA NA NA			N N N N N N N N N N N N N N N N N N N
		NA NA NA NA	N N N N N N N N N N N N N N N N N N N			N N N N
		NA NA NA	N N N			N N N
		2	Z Z Z Z Z Z Z Z Z Z Z Z Z Z Z Z Z Z Z			2 Z Z Z
	0.05	NA NR NA	NA NA			N N N N N N N
	0.05	NA NA NA NA	N N N N N N N N N N N N N N N N N N N			N N N
		NA NA	NA NA		76	NR NR
A A A		AN AN AN	NA NA			2 2 Z
	50:0	NA NR	NR NR	270		NR mg/L CaCO3
	0.05	X X X	Z Z Z	150		mg/L CaCO3 mg/L CaCO3
Z Z		NA NA	A A A			NR NR
	0.05	NR NA	NA AN			NR NR
		NA NA	NA AN			NR NR
3 3		NA NA	A A A			N N
3 3		NA NA	NA A			N. N.
A A		NA	NA AN			NR
3 3		NA	NA NA			NR
1313		NA	NA A			N.R.
N N		NA NA	NA			N. N.
žž		NA NA	NA			N N
1313		NA NA	4 d d			Z Z
2 2		NA NA	NA			N N N
						22

mg/L CaCO3	Z Z	NR	NR	NR.	X Q	NR	NR	N.N.	NR	N. N.	N. 1	200	N C	NA WA	NY S	NR.	N. C.	N. N.	N. I	NK NA	mg/L Cacos	mp/l CaCO3	mg/l CaCO3	10000 July 10000	mg/l CaCO2	mg/i CaCO3	ma/l CaCO3	an an	NR	a N	ma/l CaCO3	ma/i CaCO3	mg/l CaCO3	NB COCO	mg/L CaCO3	mg/L CaCO3	0 mg/L CaCO3	mg/L CaCO3	NR	NB	N.S.	N. C.	NK.	NB	NB NB	- C	NR	NR	NR	N.S.	NR	mg/L CaCO3	mg/L CaCO3	Z.	NR	mg/Lacos	NR NR	N N	Z.	NR	NR	NR	N.S.	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR	NR	NR	NR	NR	NR	NR	mg/L CaCO3	NR	NR	mg/L CaCO3	mg/L CaCO3	ma/l CaCO3
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Result Number 2057502	768300	160622	768396	768462	2152738	2152736	18686	18685	18688	18684	18683	18687	742079	768457	768402	768460	768398	160621	32466	119478	2153763	2133576	740750	740700	740732	2153/00	740764	10/01/01	3053736	302325	740725	CB/O+/	11/04/	10577	747747	196145	40507	2153801	126054	126052	126053	18689	2054808	2054805	2054806	2024807	2054804	18693	18691	18690	18692	2153798	740767	184730	2152710	759027	759028	994/2	90478	61681	777466	61680	160623	741718	2153836	2153835	768541	768539	160624	160625	742032	742060	742031	740738	742933	742078	742890	741711	744,000

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Hardness Mean Hardness Miles   98   98   98   98   98   98   98   9	44 44 44 44 44 44 44 44 44 44 44 44 44	4 4 4 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
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Result Number Statistical Significance 251024 Significant	gnificance Level Mea Signific	ance Level Meal Signif	cance Level Min Op	ifficance Level Min	Significance Level Max Op	Significance Level Max. Hardness NR. NA.	Hardness Mean Up Hardness Mean		Hardness Min Op Hardness Min Hardnes	Hardness Max Op Hardness Max	NR NR
120854 Not applicable 120851 Not applicable	AN			NA		T A					N N
120833 Not applicable	AN			AN		A a					e e
251014 Significant	NR	ll a		NR		R					α Z
741538 Not applicable 741566 Not significant at all concentration	NA ns	0,05		NA NR		4 8		101	100		103 mg/L 103 mg/L
120830 Not applicable	NA			NA		А					N.
120832 Not applicable	AN AN			NA		4 4					S S
120853 Not applicable	AN			NA		A					N.
120850 Not applicable	AN			AN		A					N.
120836 Not applicable	N N			AN AN		4					NR N
251023 No significance	NR			NR	1	æ					NR
251017 No significance	œ 2			N N		ac a					N N
120859 Not applicable	N N			NA	-	× ×					NR
251016 Significant	NR			NR.		Я					N.
251013 No significance	N N			NR NA		R 4					a a
120835 Not applicable	Z Z			AN AN							N.
120848 Not applicable	NA			NA	2	A		-			Z Z
251010 Significant	NR			NR		62					NR.
251015 No significance	Y W			X 82		× 02					N N
251021 No significance	Z Z			NR.							N. N.
120847 Not applicable	NA			NA	_	A					NR
120854 Not applicable	NA			NA	_	A					N.
120831 Not applicable	A CZ			NA	2 2	Α 0					N ON
251013 Significant	N N			× × ×		e 00					N N
742893 No significance		0.5		NR					40*	48*	mg/LC
741958 Not applicable	NA			NA	2	A			40*	* 83	mg/LCs
761585 Not applicable	NA			NA	2 .	A					N.
761579 Not applicable	A AN			NA NA		4 4			40+	*****	mg/LC
761577 Not applicable	NA			NA		A					N.
761588 Not applicable	NA			NA	~	A					NR
761589 Not applicable	AN			NA	2 .	Α.					NR.
761572 Not applicable	AN AN			a a	2	£ 4					N N
742901 No significance		0.02		NR	N	R			40*	48*	mg/L Ca
741949 Significant		0.05		NR	4	oc oc			40*	48*	mg/L Ca
761584 Not applicable	NA			NA	2	A					N.
761591 Not applicable	AN S			NA NA	2 2	A					Z Z
741545 Not applicable	A A			4 4 4		r 4			40*	48*	mo// Ca
741648 No significance	NR			NR	2	oc.			40*	48*	mg/L Ca
742563 Significant	NR			NR	2	oc.		1 1 1 1 1	40*	48*	mg/L Ca
761583 Not applicable	NA			NA	2	A		31			N.
761593 Not applicable	d sa			NA		4					N N
761581 Not applicable	A A			NA NA		1 41					N N
742899 Not applicable	C 40			AN AN	2	4			40*	48*	mg/L Ca
742565 Significant <		0.05		NR	4	25			40*	48*	mg/L Ca
741649 No significance		0.05		NR.	Z	œ			40*	48*	mg/L Ca
741945 Significant at all concentrations<		0.05		VR	2	m			40*	48*	mg/L Ca
741946 No significance		0.05		220	2 2	~ ~			40*	488	mg/L Ca
741942 No significance		50:0		N. N.	2				40*	48*	mg/L Ca
742342 No significance		0.05		VR.	Z				40*	48*	mg/L Ca
761753 Not applicable	NA			NA.	2	4					NR
761752 Not applicable	NA			NA	2	ν					N.
741696 Significant		0.05		A.R.	2	er.			54*	72*	mg/L Ca
741654 Not applicable	AN			NA	2	d .			54*	72*	mg/L Ca(
741559 No significance		0.05		X 2	2 2	~ 6		150		1,57	72 mg/L Cal
741540 Not significant at an concentration	42	2000		47	2,2			691	166		72 mg/L Cal
2164764 No significance		0.05		4R	Z		14	9.6			mg/L
2164766 Significant <		0.05		27	Z	3	14	9.61			mg/L
2164761 No significance		0.05		VR.	Z	3	14	19.6			mg/L
2164763 No significance <		0.05		4R	Z	~	14	9.6			mg/L
2164765 No significance		0.05		Z.B.	Z		14	9.60			mg/L
2164767 Significant		0.05		8 9	2 2	~ .	14	9,51			mg/L
2164768 Significant		0.05		E .	2 2		14	9.50			mg/L
2164720 Stanificant		0.05		2 22	2 2		14	900			mg/L
2164762 No significance		0.05		# H	Z		14	149.6			mg/L
2152714 No significance	NR			VR.	Z			170			mg/L Ca
2152713 Not applicable	AN			4A	Z	+		170			mg/L Ca(
2152715 Significant	NR			4R	Z	~		170			mg/L CaC
2152734 No significance		.00			-						Mino/ Cal
		0.01		X.	N				OR S		1

mg/L CaCO3	mg/L CaCO3	mg/L	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	NR NR	NR	NR Maril Carons	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	N. N.	NR	NR	N N	NR	N.S.	NR	NR	N.S.	NA NA	NR	NR	Z C	N. N.	NR	NR.	VR	NR 	mg/L CaCO3	mg/L CaCO3	mg/L CaCO3	VR.	VR.	mg/L	mg/L ne/L	ng/L	ng/t	N N	18	VR	mg/L	ng/L	ng/L	ng/L	ng/L	ng/L	ng/L	ng/L	IR.	E 0	NR	IR.	ng/L CaCO3	mg/L CaCO3	ng/L CaCO3	ng/L CaCO3	ng/L CaCO3	mg/L CaCO3	1g/L CaCO3	mo/l CaCO3
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NR	NA	NR	NR.	NA NA	NR	NR NR	N.	NN.	N AN	N. A.	NR	N N	NR	NR	NR.	N N	NR	Z Z	Z.	NR	Z Z	NR	NR	NR	N N	N. N.	NR	N N	NR	N. N.	NR	NA	NA NB	NA	NR	NA	NA	NA	NA	NA NA	NR	NR	NR NA	NA	N.	A A A	NA	N. N.	N.S.	NAN	NR	NR No	NR	NR.	NA NA	NR	NR	NA	N.S.	N. N.	NR	
0.01	20.0	50.0	0.05		0.05	0.05	0.05	0,3	0.3		0.01	0.01	0.05	0.05	0.05	0.05	0.05	0.05	50:0	50.0	0.05	0.05	0.05	0.05	0.05	0.05	50.0	0.05	0.05	0.05	0.01		9.01	1000	0.618																			7		0.05	0.05	0.05	0.05	0.05	0.05	2000
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Significant	applicable	Significant	ignificance	Not applicable	18 Significant	ignificance	significant at all	significant at all	significant at all	Not applicable	ificant	ignificance	Not significant at all co	2163918 Not significant at all concentrat	significant at all	significant at all ificant	ignificance	No significance	ficant	ignificance	ficant	ficant	ignificance	5 No significance	ignificance	No significance	ficant	No significance	significant at all o	Significant at all concentrations	ficant	applicable	applicable	applicable	ignificant at all c	applicable	applicable	spplicable	applicable	pplicable	eported	eported	ficant	pplicable	gnificance	pplicable	pplicable	icant	gnificance	pplicable	eported	eported	eported	eported	pplicable	icant	gnificance	pplicable	icant at all conce	766230 Significant at all concentrations <	gnificance	
2152735 Sign	2152732 Not	2164728 Sign	201550 No	201552 NOT	201548 Sign	201547 No s	2160498 Not	773634 Not	773635 Not	2152783 Not	2152786 Sign	2152785 Nos	2163920 Not	2163918 Not	2163919 Not	2254781 Sign.	2254792 No sign	2254801 No s	2254788 Sign	2254793 No significance	2254797 Sign	2254803 Sign	2254798 No s	2254785 No s	2254805 NO S	2254802 No s	2254800 Signi	2254810 No s	767224 Not:	767225 Sign	2152810 Signi	2152808 Not.	2152807 Not	2153137 Not	759367 Not:	2148468 Not	2148475 Not	2148470 Not	2148469 Not	759077 Not	759074 Not r	759075 Not	2017032 Signi	2148467 Not a	2017033 No si	2017036 Not a	2017035 Not a	2017043 Signi	2017044 No s	2017046 Not a	766735 Not r	766734 Not a	766737 Not r	766855 Not r	767589 Not a	767590 Signit	767591 No si	766229 Not a	766344 Signif	766230 Signif	766231 No sig	The same of the sa

766223 Significant < 766233 No significance <	0.05	N.R.	Z Z Z		92*		mg/L CaCO3 mg/L CaCO3
766234 Not applicable	d'A	A 2	N.				E) 1
51 Significant 54 Not applicable	NA NA	NA NA	N N		160		
3 Not applicable	NA	NA	ĄN		160		c
S Not applicable	NA NB	A W	N N		160		c   6
1 No significance	NR	N.	N.		320		E
2017063 Not applicable	NA	NA	N		320		5
Not applicable	42	A N	42		320		
Significant	NR	NR	N.		320		E
Not applicable	AN	NA	AN.		80		c
Not applicable	NA	NA	an :				2 3
Not applicable	NA NA	a a	82				2
Not significant at all concentrats	0.05	NR	a z				Z
Not significant at all concentration	0.05	NR	NR				Z
Not significant at all concentrat	0.05	NR	NR				2
Not significant at all concentrat <	0,05	NR	NR				2
Not significant at all concentrati	0.05	NR	NR	7.1			2
Not significant at all concentration	0.05	NR	NR				Z
Not applicable	NA	NA	AN		08		E
No significance	N.N.	NR	N		8		E
Significant	NR.	N. N.	NR		80		-
Not applicable	NA	AN	AN		80		-
Not applicable	NA	A N	N		1		
Significant	0.05	X Z	AN S		105		
No significance	0.05	NA	NA STATE OF THE ST		105		
Not applicable	NA.	AN	AN STATE		105		
	200	AN A	AN CONTRACTOR		COT		
	22	ZV.	EN S				
	7 7	2 2	Z Z				
759082 Not reported	C 000	N. N.	N. N.				
Concentration	0.05	NR	Z				
Not significant at all concentration	0.05	a z	Z				
Not significant at all concentration	0.05	a Z	Z				
Not significant at all concentrat>	0.05	NR	N.				3 =
Vot significant at all concentrati>	0.05	NR	S.N.				
759464 Not significant at all concentrations	0.05	NR	NR		170		Y-10-1
Not significant at all concentrations	50'0	NR	RN			256	1468
Not significant at all concentrations	0.05	NR	NR		170		
Not significant at all concentrations	0.05	NR	RN			256	1468
Not significant at all concentrati>	50:0	NR	N				_
significant at all concentrations <	0.01	N.S.	S.S.				
	NA	4Z	AN				
Not applicable	4A	NA	d Z		45.3		
Significant <	0.05	NR.	W.Z.				
2148186 Not significant at all concentrati	0.05	N. N.	S. S.				
No significance <		NR	S. S.				
	NR	N.S.	N.				
	ZR.	Z	N.				
	NA.	NA	N				_
Not applicable	VA.	NA	NA				Z
Not applicable	NA	NA	NA				Z
2148354 Significant at all concentrations <	0.01	N.R.	N				Z
Not applicable	4A	AN	d Z		45.3		E
Significant	NR	az.	a Z				Z
No significance	22	æ :	Z :				Z
Not applicable	NA NA	YZ.	d'A				Z
2148178 Significant at all concentrations	0.05	Z Z	X OZ				2 2
No significance	9.05	Z Z	NN				2 2
Significant at all concentrations <	0.05	X C	Z Z				2 2
No significance	0.05	N. C.	NN GIA				2 2
Significant	0.05	X C	XX.				2 2
2148139 Significant <	9,05	Y Y	N. N.				-
2148141 Significant <	0.05	α Z	N. S.				
No significance <	0.05	N.N.	AN AN AN AN AN AN AN AN AN AN AN AN AN A				2
Not significant at all concentrat>	0.05	a Z	N. N.				2 .
Not significant at all concentrat >	0.05	N.R.	NR				2
Not significant at all concentrati>	0.05	NR	NR				Z
2254826 No significance	50:0	NR	NR				Z
No significance >>	0.05	NR	NR				
No significance	0.05	S. S.	NR				4
No citation of	0.05	a z	82				2
No significance	500	200	2				2
Significant	0.05	NR	NA.				2 2
2254814 No significance >	0.05	NR	NR				Ż
Significant >	0.05	NR	NR				Z
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1316	87386 2786	2145	d L.L.	Standard Test Fish Development, Part I. Fat Standard Test Fish Development, Part I. Fat	EPA-600/3-76-061A, U.S.EPA EPA-600/3-76-061A, U.S.EPA	1
13.154   Selentin II. 2015   Selentin II. 2016   Selentin III. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin II. 2016   Selentin III. 2016   Selen	5821	2145	d L.L.	Standard Test Fish Development. Part I. Fat	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	÷.
2.125  delicional, 1, and LL, 1, Similho   Standard Test to Decemberate Herit II.	5782 5796	2145	d L.L.Jr.	Standard Test Fish Development, Part I. Fat Standard Test Fish Development, Part I. Fat	REPA-600/3-76-061A, U.S.EPA, Duluth, MN:7 REPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	r r
2.254   Journal, N. 2012   Journal, J. 2012   Journal, N. 2012   Journal, J. 2012   Jou	6800	2145	nith	Standard Test Fish Development, Part I. Fat	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	15
235   Adentino, M. and L. L. Sorths   Control, M. Control   Control of First Discussioners   Part L. Sorths	5812	2145	nith	Standard Test Fish Development, Part I, Fat Standard Test Eich Development, Bort I Est	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	# 1
2.25  Selembar, M. and L.L. Smith   Standard Test fits Decentionment that L.L. Smith   Standard Test fits Decention that Decention   Standard Test fits Decention that Decention   Standard Test fits Decention that Decention   Standard Test fits Decention   Standard Test fits Decention   Standard Test fits Dece	6279	2145	nith	Standard Test Fish Development, Part I. Fat	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	1,
23.5   Section M. J. B. C. B. P. B. B. B. B. B. B. B. B. B. B. B. B. B.	26792	2145	nith	Standard Test Fish Development. Part I. Fat	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	-
225 Johnson, M. J. 2015 Johnson, M. J. 2015 Johnson, M. J. 2014 Johnson, M. J. 2015 Jo	5815	2145	nith	Standard Test Fish Development, Part I, Fat Standard Test Fish Development, Part I, Fat	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	1 5
12.25   Johanne J. J. 2005   Johanne J. 2005   Johanne J. J. 2005   Johanne J. J. 2005   Johanne J. 200	6285	2145	nith	Standard Test Fish Development, Part I. Fati	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	15
2155   Selemon 1, 1, 1, 1971   Standard 1, 1, 1, 1971   Standard 1 test for Development Part 1	5843	2145	Adelman, I.R., and L.L., Jr. Smith	andard Test Fish Development, Part I. F.	REPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	11 12
2255   Jedemino L. J., and L. L. J., Sinth   Standard Test find Development. Part I. S. Month Microbial Control of the Contr	55774	2145	R., and	andard Test Fish Development. Part I. F	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	151
1212   Sammari, A. 2014   Sammari, A. 2014   LLD. Solino   Statistical field in Development Part 1.     1212   Sammari, A. 2014   LLD. Solino   Statistical field in Development Part 1.     1213   Sammari, A. 2014   LLD. Solino   Statistical field in Development Part 1.     1214   Sammari, A. 2014   LLD. Solino   Statistical field in Development Part 1.     1215   Sammari, A. 2014   Sammari, A. 2014   LLD. Solino   Statistical field in Development Part 1.     1216   Sammari, A. 2014   Sammari, A. 2014   LLD. Solino   Statistical field in Development Part 1.     1216   Sammari, A. 2014   Sammari	5806	2145	R., and	Fish Development, Part I. F	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	18
2315   Geldman, La, and L. Senth   Stroke Reference Part   Stroke Reference	6832	2145	R., and	Standard Test Fish Development, Part 1. Fat Standard Test Fish Development Part   Fat	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	1
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13.13   Powerbook N. 19.	6819	2145	man,I.R., and L.L.,Jr.	Part I. F	EPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	1.
MANY	6787	2145	man, i.R., and L.L., Jr.	Part	FPA-600/3-76-061A, U.S.EPA, Duluth, MN:7	100
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1317    1317	/L/WOND	71674	Harmon, S.M., W.L. Specht, and G.T. Chandl	A Comparison of the Daphnids Ceriodaphnia	rch. Environ. Contam.	20
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State   Checker   Checke	7718 CONC/FROM GRAPH//	45168	Milazzo	The Response of the Three Brood Ceriodap	ch. Environ. Contam	21.00
1860   March 1, 2005   March 2, 2005   March	1686 DNUM/6//DOSES/0:4/9/0:8/9/1:2//1:/9/2	45168		The Response of the Three Brood Ceriodaph	Arch, Environ, Contam, Toxicol,21(1): 35-40	15
151079    151074    1510	7716 CONC/FROM GRAPH//	45168		The Response of the Three Brood Ceriodap	ch. Environ. Contam	15
18.00   Janes A. J. L. Cole Neals, and J. F. Bartinial   Toxicty of Chincide Under Winter Low-Flowing Bills Environ.	1695 DNUM/6//DOSES/0.479/0.879/1.27/1.79/2	71674	T. Char	A Comparison of the Daphnids Ceriodaphnia	Arch. Environ. Contam. Toxicol.45(1): 79-85	20
15005    Allert AL., C.L. Cole Alsal, and J.F. Farchild   Toncity of Chloride Under Winter Low-Flow Bull Environ. CONK/FROM (8APH)   A 518   Anter A.L., C.L. Cole Alsal, and J.F. Farchild   Toncity of Chloride Under Winter Low-Flow Bull Environ. CONK/FROM (8APH)   A 518   Control Alsa   A 518   A 51	6816	161067	Fairchi	Toxicity of Chloride Under Winter Low-Flow Toxicity of Chloride Under Winter Low-Flow	Bull, Environ, Contam, Toxicol,89(2); 296-30. Bull, Environ, Contam, Toxicol,89(2); 296-30.	20
Processors   Pro	6815	161067	irchi	Toxicity of Chloride Under Winter Low-Flow	Bull. Environ, Contam. Toxicol.89(2): 296-30	20
1553   Coorgil JUM., and D.P. Milazon   The Response of the Three Brood Geriodaphida. Funtion. DONE/STAIN   2553   Coorgil JUM., and D.P. Milazon   The Response of the Three Brood Geriodaphida. Funtion. DONE/STAIN   2554   Service. Str. M. Beeth, and G.T. Charlif A. Comparison of the Daphwids Ceriodaphida. Funtion. DONE/STAIN   2554   Service. Str. M. Beeth, and G.T. Charlif A. Comparison of the Daphwids Ceriodaphida. Funtion. DONE/STAIN gi/J/TESTID/20/1   15532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Strutton. Done/SENA gi/J/TESTID/20/1   11532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Strutton. Done/SENA gi/J/TESTID/20/1   11532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Strutton. Done/SENA gi/J/TESTID/20/1   11532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Strutton. Done/SENA gi/J/TESTID/20/1   11532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Strutton. Done/SENA gi/J/TESTID/20/1   11532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Struttonner DOSES/NA gi/J/TESTID/20/1   11532   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Cilolotide to Freshwater   Struttonner DOSES/NA gi/J/TESTID/24/1   11533   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Circuit and Hulf, Incell PS Environmen   DOSES/NA gi/J/TESTID/24/1   11533   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Circuit de Freshwater   Struttonner OOSES/NA gi/J/TESTID/24/1   11533   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Circuit de Hulf and Hulf, Incell PS Environmen   DOSES/NA gi/J/TESTID/24/1   11533   Sanders, D.F. S. G. Beegh, and H.C. Bailey   Circuit Coxidity of Circuit de Hulf and Hulf, Incell PS Environmen   DOSES/NA gi/J/TESTID/24/1   11533   Sanders, D.F. S. G. S		71674	Char	A Comparison of the Daphnids Ceriodaphnia	Arch. Environ. Contam. Toxicol.45(1): 79-85	20
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158349  Ephick.J.R.F. K.D. Bergh, and H.C. Bailey   Chronic Toxicity of Chloride to Freshwaters	DOSFS/	116830	2	Site Water Bioassays for Huff and Huff, Inco	PS Environme	15
Size Water Boassays for Huff and Huff, Inco   1982/2/ Standers, D.F.		158449	Iphick, J.R.F., K.D. Bergh, and H.C. Bailey	Chronic Toxicity of Chloride to Freshwater S	Environ. Toxical. Chem.30(1): 239-246	2C
118530   Sameta, D.F.   1185	7904 DOSES/NA g/L//TESTID/20//	116830	anders,D.F.	Site Water Bioassays for Huff and Huff, Inco		15
1585243   Sander, D.F.	7903 DOSES/NA g/L//TESTID/20//			Site Water Bioassays for Huff and Huff, Inco		15
185820  Sanders,D.F.   18680  Sanders,D.F.	0		T.K. Linton, C.D.	Influence of Water Hardness and Sulfate on	Environ, Toxicol, Chem.30(4): 930-938	35
158409   Enthe Lance   158409   Enthe Lance   158409   Enthe Lance   158409   Enthe Lance   158409   Enthe Lance   188400   Enthe Lance   188400   Enthe Lance   188400   Enthe Lance   188400   Enthe Lance   188400   Enthe Lance   188400   Enthe Enthe Enter	000		anders.D.F.	Site Water Bioassays for Huff and Huff, Inco		13
116820  Sander,D.F.   116820  Sander,D.F.			K.D. Bergh,	Chronic Toxicity of Chloride to Freshwater S	Environ, Toxicol, Chem.30(1): 239-246	20
1868.05/SIANA gL/LTESTID/23/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/23/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/23/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/14/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/14/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/13/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/12/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/12/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/12/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/12/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco DOSES/NA gL/LTESTID/24/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco SOSES/NA gL/LTESTID/24/  1188.05/Siander, D.F.   Site Water Bloassays for Huff and Huff, Inco SOSES/NA gL/LTESTID/24/  Site Water Bloassays for Huff and Huff,	7897 DOSES/NA g/L//TESTID/19//	116830		Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
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1956/1976   1956	7961 DOSES/NA.g/L//TESTID/23//	116830	anders,D.F.	Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	15
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116830   Sander, D.F.   1168	7879 DOSES/NA e/L//TESTID/14//	116830		Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
116820   Sanders, D.F.   Site Water Biolosays for fruit and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/18/    116820   Sanders, D.F.   Site Water Biolosays for fruit and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/18/    116820   Sanders, D.F.   Site Water Biolosays for fruit and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/13/    116820   Sanders, D.F.   Site Water Biolosays for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/13/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/12/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/24/    116820   Sanders, D.F.   Site Water Biolosasys for Huff and Huff, Incol [PE Etw. DOSES/NA g/L/TESTID/	7880 DOSES/NA g/L//TESTID/14//	116830		Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
158924 SOURCE, D. T. A. DICKING IMPLEASE OF WATER TRACES AND STATE OF BOSSES/NA BUL/TESTID/18/1   158924 SOURCE, D. J. T. K. LINDO, C.D. Tarr, A. DICKING IMPLEASE OF WATER TRACES and Suitate of Environ DOSES/NA BUL/TESTID/13/1   158924 SOURCE, D. J. T. K. LINDO, C.D. Tarr, A. DICKING IMPLEASE OF WATER TRACES and Suitate of Environ DOSES/NA BUL/TESTID/13/1   118800 SAURCE, D. T. K. LINDO, C.D. Tarr, A. DICKING IMPLEASE OF THAIT AND HAIT, INCO IPS ENVIRONMENT OF STATE BIOASSANS for HAIT AND HAIT, INCO IPS ENVIRONMENT OF STATE TRACES AND SAURCE OF THAIT AND HAIT, INCO IPS ENVIRONMENT OF THAIT AND HAIT	7896 DOSES/NA g/L//TESTID/18//	116830 8	F.	Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
158242   SOUCES, DA. A. LINDO, C.D. Tart, A. Dicking illustuace of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. A. LINDO, C.D. Tart, A. Dicking illustuace of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. A. LINDO, C.D. Tart, A. Dicking illustuace of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking illustuace of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking illustuace of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking illustuace of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking influence of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking influence of Water Handress and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrates and Sulfate on Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrate Bloassays for Huff and Huff, Incol IPS Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrate Bloassays for Huff and Huff, Incol IPS Environ   158242   SOUCES, D.J. T. K. LINDO, C.D. Tart, A. Dicking illustrate Bloassays for Huff and Huff, Incol IPS En	a.	116830 5		Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	15
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116820   Sanders D.F.     116820   Sanders	24.54 78.73 DOSES/NA #/1 //TESTID/13//	116830	A. Littory C.D. 1811, A. Dick	Site Water Bioassavs for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
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COSES/NA g/L/TESTID/22/f         118830   Sanders, D.F.         Site Water Bloassays for hulf and Hulf, Incol IPS Envi.           DOSES/NA g/L/TESTID/22/f         118830   Sanders, D.F.         Site Water Bloassays for hulf and Hulf, Incol IPS Env.           DOSES/NA g/L/TESTID/22/f         118830   Sanders, D.F.         Site Water Bloassays for Hulf and Hulf, Incol IPS Env.           DOSES/NA g/L/TESTID/24/f         118830   Sanders, D.F.         Site Water Bloassays for Hulf and Hulf, Incol IPS Env.           DOSES/NA g/L/TESTID/24/f         118830   Sanders, D.F.         Site Water Bloassays for Hulf and Hulf, Incol IPS Env.           DOSES/NA g/L/TESTID/24/f         118830   Sanders, D.F.         Site Water Bloassays for Hulf and Hulf, Incol IPS Env.           DOSES/NA g/L/TESTID/24/f         118830   Sanders, D.F.         Site Water Bloassays for Hulf and Hulf, Incol IPS Env.           158224   Sanders, D.F.         Sanders, D.F.         Site Water Bloassays for Hulf and Hulf, Incol IPS Env.           158224   Sanders, D.F.         Sanders, D.F.         Sanders, D.F.           158224   Sanders, D.F.         Sanders, D.F.         Sanders, D.F.           158224   Sanders, D.F.         Sanders, D.F.         Sanders, D.F.           158224   Sanders, D.F.         Sanders, D.F.         Sanders, D.F.           158224   Sanders, D.F.         Sanders, D.F.         Sanders, D.F.           158224   Sanders, D.F.         Sand	5468		, T.K. Linton, C.D. Tarr, A. Dicki	Influence of Water Hardness and Sulfate on	Environ. Toxical. Chem.30(4): 930-938	20
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116830   Sanders, D.F.   116840   Sanders, D.F.	OCSES/NA B/L/	116830 5	anders, D. F.	Site Water Bloassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
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15849 [Elpick, JR.F., KD. Bergh, and H.C. Bailey   Chronic Toxicity of Chloride to Freshwater-Struvion   11830 [Sanders, JR.F., KD. Bergh, and H.C. Bailey   Chronic Toxicity of Chloride to Freshwater-Struvion   11830 [Sanders, JR.F. Bergh, and Holf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   ODGES/NA gl/JTESTID/Si/   11830 [Sanders, JR.F. Bargh, and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and Hulf, Incolle Struvion   Site Water Biosassays for Hulf and	471	158924 S	K. Linton, C.D. Tarr, A. Dicki	Influence of Water Hardness and Sulfate on	Environ, Toxical, Chem.30(4): 930-938	20
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DOSES/NA BLI//TESTID/5// 11683a Sanders, D.F. Site Water Bioassays for Huff and Huff, IncolPS Envi	872 DOSES/NA g/L//TESTID/6//			Site Water Bioassays for Huff and Huff, Inco	IPS Environmental and Analytical Services, A	19
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Raw Data from ECOTOX (Unfiltered)

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1 TIME/OTHER DURATIONS REPORTED//DNU	IIME/OTHER DORATIONS REPORTED//DING			2271252			742574 DNUM/5//DOSES/6.4/12.6/15/25.6 mS/cm/	CONTRACTOR MONTH	DNUM/11//DOSES/NA ppm//TESTID/27//O	6170 CONTR/< 10 % MORT///	CONTR/< 10 % MORT//	802493 DNUM/11//DOSES/NA ppm//TESTID/27//O	6169 CONTR/< 10 % MORT//	CONTR/< 10 % MORT// DONIM/11//DOSES/NA pom//TESTID/78//O	DNUM/11//DOSES/NA ppm//TESTID/28//O	DNUM/11//DC	CONTR/< 10 % MORT//	DNUM/11//DOSES/NA ppm//TESTID/28//O	DNUM/11/DOSES/NA ppm//TESTID/28//O	802490 DNUM/11//DOSES/NA ppm//TESTID/27//O	DNUM/11//DDSES/NA ppm//TESTID/27//O	CONTR/< 10 % MORI//	802496 DNUM/11//DOSES/NA ppm//TESTID/27//O	CONTR/< 1	DNUM/11//DOSES/NA ppm//TESTID/28//O	DNUM/11//DOSES/NA ppm//1ESTID/28//O		16494 CONTR/11 % MORT//		2061033	CONTR/B,C//	205035 CONTR/B,C//	CONTR/B,C//	CONTR/B,C//	2060327 CONTR/B,C//	CONTR/B.C//	2060325 CONTR/B,C//	CONTR/B,C//	2060326 CONTR/B,C//	2060359 CONTR/B.C//		WTAT/101 MG//COMPEP/1//ETIME/90 D//	ETIME/56 D//	449 TESTID/3//DOSES/117000/306000/643000/	DNUM/6//TESTID/2//OEF/MIXTURE-ROAD	DNUM/6//TESTID/2//OEF/MIXTURE-ROAD I	205757 COMPEP/2//WTAT/551 MG//ETIME/90 D//	WTAT/553 MG//COMPEP/2//ETIME/90 D//	ETIME(30 D/)	DNUM/5//DOSES/6500/7500/8500/10000 g		CONC1/ONLY CONC TESTED//	740663 CONC1/ONLY CONC TESTED//									CONCI/ONLY CONC TESTED//	CONCI/ONLY CONC TESTED//	CONC.1/ONLY CONC TESTED//	CONC1/ONLY CONC TESTED//			2163090

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Raw Data from ECOTOX (Unfiltered)

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TODGESSAM PRIMITESTID(ASV)         2002 Hughes J.S.         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         2002 Hughes J.S.         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         2002 Hughes J.S.         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         2002 Hughes J.S.         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         2002 Hughes J.S.         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         2002 Hughes J.S.         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (DOSESANA PRIMITESTID(ASV)         Acute Caretory of Thirty Chemicals bis 5th (Associated Primites Desired Primites Desi	DNUM/11//E	0 201.		Acute Toxicity of Thirty Chemicals to Striped	oc. Annu. Conf. Western Assoc	19]
No. Moork.ALT.VI)   2012   Indigned, 3.5.   Acute Totaloth of Thirty Chemicate District Discussion of Thirty Chemicate District	668 DNUM/11//DOSES/NA ppm//TESTID/49//	.00 201.		Acute Toxicity of Thirty Chemicals to Striped	oc. Annu. Conf. Western Assoc	19,
2005/51/Methol. 3.0012   Images. 1.5.   Acute Total Control Chinary Chemiste 19.20   Montal Control Chinary Chemiste 19.20   Montal Chinary	641 CONTR/< 10 % MORTALITY//	201	Hughes, J.S.	Acute Toxicity of Thirty Chemicals to Striped	oc. Annu, Conf. Western Assoc.	19
Policy State (Principles)         2013 [magnes 1.5.         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thirty Chemicals to STID (Magnes 1.5.)         Acute Toxicity of Thi	671 DNI IM/11//DOSES/NA DBm//TESTID/49//	201	Hughes I.S.	Acute Toxicity of Thirty Chemicals to Stribed	oc. Annu. Conf. Western Assoc	197
ODSSENAR pont/TESTID/SQI/O         2023 [haghed.1.5]         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O         Acute Tools of Pinty Chemicals to Strip (DOSSS/MA pont/TESTID/SQI/O	217 CONTR/< 10 % MORTALITY//	201	Hughes, J.S.	Acute Toxicity of Thirty Chemicals to Striped	oc. Annu. Conf. Western Assoc	197
DOSESNA pgm/TESTID/95/10  2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2022 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2022 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   Acute Towickly of Thirty Chemicate bottom (NORTALLY)   2021 Hughes J.S.   A	CONTR/< 10 % MORT	2013	Hughes, J.S.	Acute Toxicity of Thirty Chemicals to Striped F	oc. Annu. Conf. Western Assoc	197
Acute Tools of Thirty Chemists to Strip   2021 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2024 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2022 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2023 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2024 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2024 Hughres   3.   Acute Tools of Thirty Chemists to Strip   2024 Hughres   3.	M/11//DOSES	0 201	Hughes,J.S.	7	Proc. Annu. Conf. Western Assoc. State Gan	19
2012   Hughres J.S.   Acute Touching of Thirty Chemicals to Strip (2005SENA pom/TESTID/49)  2012   Hughres J.S.   Acute Touching value and the Strip (2015   Hughres J.S.   Acute Touching value to Strip (2015   Hughres J.S.   Acute Touching value at the Strip (2015   Hughres J.	M/11//DOSES	0 201	Hughes,J.S.	Thirty Chemicals to Striped	Proc. Annu. Conf. Western Assoc. State Gan	19.
Acute Toucking of March 1971   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of Thing Chemists to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking of the Toucking Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking toucking to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking toucking to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking toucking to Stip (2005/SIAA pom/TESTID/SIA)   2021 Hugghes J.S.   Acute Toucking Stip (2005/SIAA)   2021 Hugghes J.S.   Acute Toucking Stip (2005/SIAA)   2021 Hugghes J.S.   2021 Hugghes J.S.   2021 Hu	640 CONTR/< 10 % MORTALITY//	2017	Hughes, 1.5.	0	Proc. Annu. Conf. Western Assoc. State Gan	19.
AMORFALITY    2012   Highes 1.5.   Acute Toxicity of Thirty Chemicals to Sign	664 DNUM/11//DOSES/NA ppm//TESTID/49//	201.	Hughes 15.	0 0	Proc. Annu. Conf. Western Assoc. State Gam Proc. Annu. Conf. Western Assoc. State Gam	197
Acute Totology   2012   Hughes 1.5   Acute Totology   2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughes 1.5   Acute Totology of Theiry Chemical to Sirgo CONTR-1.0 % MORFALTH// 2012   Hughe	5214 CONTR/< 10 % MORTALITY//	2013	Hughes, J. S.	Acute Toxicity of Thirty Chemicals to Striped	Proc. Annu. Conf. Western Assoc. State Gan	197
98 MORTALITY()         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         2002 Hugbers J.S.         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm/(TSSTD/94)/O         Actual Toxicity of Thirty Chemicals to Strip POSSESNA ppm	DNUM/11//DOSES/N	2012	Hughes, J. S.	Acute Toxicity of Thirty Chemicals to Striped	Proc. Annu. Conf. Western Assoc. State Gam	197
DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip S MONTALITY// 2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip DODGE/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip Dodge/NA ppm//TESTID/29/10   2012 Hughres, 15.   Acute Toxicity of Thirty Chemicals to Strip Dodge/NA ppm/	CONTR/< 10 % MORTALITY//	2012	Hughes,J.S.	Acute Toxicity of Thirty Chemicals to Striped	Proc. Annu. Conf. Western Assoc. State Gan	197
Na MONTALITY  2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA pom/TESTID/SQ/) 2012   Higgles, 1.5.   Acute toxicity of Thirty Chemicals to Strip (DOSES/NA	DNUM/11//DOSES/NA ppm//	0 2012	Hughes,J.S.	Acute Toxicity of Thirty Chemicals to Striped	Proc. Annu. Conf. Western Assoc. State Gan	197
	CONTR/< 10	2012	Hughes,J.S.	Acute Toxicity of Thirty Chemicals to Striped	Proc. Annu. Conf. Western Assoc. State Gan	197
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116526  Sanders, D.F.	191 (high poses) / 300/ 1000 ppm// res	153684	Elphick J.B.: M. Davies, G. Gilron, E.C. Canar	An Aquatic Toxicological Evaluation of Sulfa	Sovieton Toxicol, Chem. 30(1): 247-253	201
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61 Mount, D.R.,	61 Mount, E	72 Mount,C	'61 Mount, D	72 Mount,D	11 FISHER, S.W. 65 Camargo, J.	37 Trama, F.	72 Mount,D	72 Mount, D.R.	19 Ragab, F.	21 Boutet,C	21 Boutet,C	OS Tatara, C.	72 Mount,D	51 Mount,D	51 Mount,D	51 Mount,D	89 Baudouit	5 Dowden,	S Dowden,	51 Mount,D	51 Mount,D	Mount,D	Mount D	Mount,D	18272 Mount, D.R	Mount,D	Mount,D	8 Wallen I	8 Wallen,I.	8 Harless, N	8 Harless, N	8 Harless N	8 Harless, N		1 Boutet	1 Boutet,C.	1 Boutet,C.	2 Mount, D.	2 Mount,D.	8 Silva,L.V.R	8 Silva,L.V.F	8 Silva,L.V.F	8 Wooldrid	0 Luo,S.Q.,	U Luo,S.Q., 1 Kardatzke	1 Kardatzke	1 Kardatzke, J	1 Kardatzke,J 1 Kardatzke,J	1 Kardatzke,J	6 Olasehind	Cowgill, U	1 Mount, D.	Mount, D.	1 Mount, D.	Mount, D.	Cowgill	Mount,D.	Mount,D.	Mount, D.
14761	14751	182	147	182	1162	80:	182	182	1130	547	5421	1860	18272	1476	1476	1476	533	91	91	1476	1476	1476	1476			l			205	154088	15408	15408	154088	15408	1183	5421	542	18272	18272	8486	8486	84868	286	83190	10704	107041	107041	107041	107041	110666	11941	14761	1827.	1476	14761	11941	1476	1476	14761
mg/L/	mg/L//NA/26.3*	1	mg/L//NA/26.3* mg/L//NA/26.3*	1	/TESTID/1//OEF.			1	8% SULFUR//Dd				1 2010111111111111111111111111111111111	mg/L//NA/26.31	mg/L//NA/26.3*	mg/L//NA/26.31	ALCONC//CONT			mg/L//NA/26.3*	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3*	CA/14.0" mg/L//MG/12.0" mg/L//NA/26.3"	mg/L//NA/26.3	mg/L//NA/26.31	15 HARD/MODERATELYHARD//	mg/L//NA/26.31	mg/L//NA/26.3*								D AS LETHAL//				and the same and the same	II//TESTID/3//O	1//TESTID/4//O	II//TESTID/3//O	CONC1/ONLY	1,1	//DNUM/11//	1//DNUM/11//	1//DNUM/11//	1//DNUM/11//	//II/MNNQ//1		I/FROM GRAPH	ng/L//NA/26.31		ng/L//NA/26.3*	ng/L//NA/26.3*	TE/L//NA/26.3	12.0° mg/L//NA/26.3°	mg/L//NA/26.3*	mg/L//NA/26.31
//MG/12.	/L//MG/12.0*	RATELYHARD/	/L//MG/12.0* /L//MG/12.0*	RATELYHARD/	35ES/95 mg/L//7		J/MODERATELYHARD/	RATELYHARD/	SSIUM OXIDE, 1				HARD/MODERATELYHARD/	/L//MG/12.0*	/L//MG/12.0*	/L//MG/12.0*	O EFCT CONT/MEDIAN LETH	TO THE PERSON OF		/L//MG/12.0*	/L//MG/12.0*	/L//MG/12.0*	1//WG/12.0*	///MG/12.0*	RATELYHARD/	1//MG/12.0*	L//MG/12.0"								GENS COUNTE		// UDANA DA	HARD/MODERATELYHARD//	RATELYHARD//	SES/14.12 mg/ SFS/32.71 mg/	/DOSES/32.71 mg/l//TESTID/4/	SES/14.12 mg/	O=TAP WATER//	CONC1/ONLY CONC TESTED,	CONC TESTED,	NCE TO ADUL	NCE TO ADUL	TIME/EMERGENCE TO ADULT//DNUM/11/	ENCE TO ADUL		TMETH/APHA, 1989// CONC1/FROM GRAF	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.	ATELYHARD//	L//MG/12.0* n	CA/14.0* mg/L//MG/12.0* mg/L//NA/26.3	1989// CONC.	//WG/	//WG/	//MG/12.0*
	V14.0* mg/L	ARD/MODERAL	1/14.0* mg	ARD/MODE	NUM/2//D		ARD/MODE	HARD/MODE	HAR/POTAS				ARD/MODE	V14.0* mg	1/14.0* mg	1/14.0* mg	EFCT CONT	TO MAN TO		/14.0* mg	/14.0* mg	/14.0* mg	/14.0° mg	/14.0* mg	RD/MODE	/14.0* mg	/14.0° mg/								-I/IEKAIO		20044/00	RD/MODER	RD/MODE	UM/2//DO	UM/2//DC	UM/2//DO	NTR/C,0,0	NC1/ONLY	NCI/ONLY	1E/EMERGE	1E/EMERGENCE	IE/EMERGE	E/EMERGE		ЕТН/АРНА	14.0" mg/	RD/MODER	14.0* mg/	14.0* mg/	TMETH/APHA	14.0° mg/	/14,0* mg/	CA/14.0* mg/L

Raw Data from ECOTOX (Unfiltered)

Year	1974	1997	1992	1992	1991	1991	1991	1965	1965	1965	1965	1965	1989	1992	1992	1992	1992	1997	1961	1961	1996	1996	1996	1996	1996	1996	1996	1996	1957	1957	1957	1996	1995	1000	1000	1000	0000	Cast	1994	1996	1959	1959	1959	1965	2011	2011	2011	2011	2011	2011	1965	1965	1965	1965	1996	1996	1994	1999	1978	1978	1986	1986	1986	1986	1986	1986	1997	1996	1996	1996	0001	4000	7661	1881	1661	1989	1989	1989	9661	6861	6861	2003	2003	2003
Publication																																																														) -																				Ĭ		
5-2162	.12(6): 745-75	- 2004-2019	ontract No.5	ntract No.5				9): 1308-131	11.1308-131	1 1308-131	3): 1308-131	9): 1308-131	Texas. Dent	ntract No.5	ntract No.5	3 Contract No.5	ontract No.5	009-2019											11	11	11						Designation of the	S.P.UDIIC FIE	105:185-20		Series No.95,	Series No.95,	Series No.95,	9): 1308-131	37-1641	37-1641	37-1641	1637-1641	37-1641	1637-1641	1308-131	:1308-131	1308-131	: 1308-131					, Submitted to Sm	tted to Sm	1): 46-53	1): 46-53	1): 46-53	1): 46-53	1): 46-53	1): 46-53	909-2019		Ī		JOSOT ENSP Contract No Se	CONTRACTOR	LEACT NO.54	6102-60	6107-60	exas, Deni	exas, Deni	exas, Deni		exas, Deni	exas, Dent			
echnol.44(6): 215		Chem 16/10)		ũ	1): 33-56	1): 33-56	1): 33-56	J. Water Pollut. Control Fed.37(9): 1308-	rol Fed 37(9):	to Fed 37/6	Water Pollut, Control Fed. 37(9): 1308-	rol Fed.37(9	sity of North Texas.	11, ENSR Co	11, ENSR Co	11, ENSR Co	/0301, ENSR Co	m.16(10): 2	:77-85	:77-85	77-84	77-84	77-84	77-84	77-84	77-84	77-84	77-84	9(6): 695-7	9(6): 695-7	9(6): 695-7	77-84	77-84	77 04	127 140	77.64	10.11	0(LZK1), U.S	niladelphia	1.2	te Cont., Se	Conf.,	Conf.	ol Fed.37(9	m.30(7): 16	m.30(7): 16	m.30(7): 16	n.30(7): 16.	m.30(7): 16	m.30(7): 16:	ol Fed.37(9	ol Fed.37(9	ol Fed.37(9	ol Fed.37(9	77-84	77-84	269-272	15-1047	006, Submi	006, Submi	Environ. Health32(1); 46-53	Environ. Health32(1); 46-53	Environ. Health32(1): 46-53	7. Health32	ron. Health32(1): 46-53	h. Health32	Chem. 16(10): 2009-2019	77-84	77-84	77-84	ENISP Con	ENCD CON	L, ENSK LON	n.15(10): 20	17: (AT)01: V	University of North Texas,	University of North Texas, Den	University of North Texas,	77-84	y of North	y of North T	279-287	279-287	279-287
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Source in Environ.	wa Bull. Environ.	f Frying	ns Final Rep.	nsFinal Rep	rd Arch. Hydrob					In Water	in J. Water F	13	10				107	f Environ. Tox	Proc. La.	Proc. La.	d Prog. Fish	d Prog. Fish	d Prog. Fish	d Prog. Fish	d Prog. Fish-Cult.58(2): 77-84	d Prog. Fish	1 Prog. Fish	1 Prog. Fish	Sewage Ind.	e Sewage In				Drog Cirb	Prog Cich Cult 30(3)	Drog Lich	100 100	erinal Rep.No.R	Drae Cat	Prog. FISh	ZIN: Proc.1		z In: Proc.13	n. Water Pol	d Environ, T	d Environ. T	d Environ. T	d Environ. T	dEnviron, T	dEnviron, T	17	1	J. Water P	J. Water P.	ď.		Prog. Fish-C	Ü	Res.Rep.No	Res.Rep.No.BW	Jpn. J. Tox	Ipn. J. Toxicol.	t Jpn. J. Toxi	Jpn. J. Toxi	Jpn. J. Toxicol	-		Prog. Fish-	Prog. Fish-	Prop Fish-	Einal Ban GB	Flord Don	rinal kep.	Environ. 10	Environ. Ioxid		E	Ph.D. I hesis,	Prog. Fish-	Ph.D.Thesi	Ph.D.Thesi	Aquacultur	Aquacultur	Aquacultur
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Iters Gene	arious Meta	o Predict th	Salinity/Tox	salinity/Toxi	ts of Salinity	ts of Salinity	ts of Salinity	Chemicals	Chemicals	Chemicals	Chemicals	Chemicals	vater and Sa	salinity/Toxi	alinity/Toxi	of a Salinity/Toxi	alinity/Toxi	o Predict th	es of Some	es of Some	ebra Mussel Infestation	Mussel Inf	Mussel Inf	Mussel Inf	Mussel Inf	Mussel Inf	Mussel Inf	Mussel Infi	a affinis of	Gambusia affinis of Certai	a affinis of	Mussel Inf	Mussel inf	Adviced Inf	with of Disto	Zahra Mussel Infestation	in in a second	וואול או אלחמוור רווב וא רבו ומוו	African lef	Mussel Infe	Bluegill Sun	Bluegill Sun	Bluegill Sun	Chemicals t	cal Deicers	cal Deicers	cal Deicers	cal Deicers	cal Deicers on	al Deicers	Chemicals	Chemicals t	Chemicals t	ed Chemicals to Certain	Mussel Infe	Mussel Infe	um Does no	Calcium Against	Sulfate on Eggs and Fry of	e on Eggs at	iring Tempe	iring Tempe	ring Tempe	ring Tempe	iring Tempe	ring Tempe	Predict the	Mussel Infe	Missel Infe		linity/Toxic	Taite /Toule	alluity/ loxic	Predict the	Predict the	ater and Sal	ater and Sal	and	Mussel Infe	ater and Sai	ster and Sal	Catfish, Rha	atfish, Rha	atfish, Rha
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//Q31	200	SD//	.0° mg/L//h	.0* mg/L//h	CONC1/FROM GRAP	1989// CONC1/FROM GRAF							70/580/660	1G/12.0" mg/L//NA/26.3"	0* mg/L//NA/26.3*	12.0* mg/L//N	0* mg/L//NA/26.3	//q	)		//GBJ	//Q3J	LED//	TED//	LED//	TED//	//Q3	LED//				/FD//	/LU1/	1/03	1100	////5	1100		1/43	EU//															//Q3	//Q3		//Q3	//DOSES/85	//DOSES/85							1/10	//Q3	1/03	ED//	I mail line	The man I I I I I I I I	MENT/IN	110	111	5	IG/L//TESTID	IE/L// ES !!!	//d=	g/L//TESTIL	000/2260/2	ng/I//TESTI	ng/I//TESTIC	DE/I//TESTIC
ments CONC TES	200	RATEI YHA!	/L//MG/12			A, 1989// CI	4, 1989//						DSES/410/4	/L//MG/12.	mg/L//MG/12.0*	//WG/	MG/	RATELYHARD			CONC TEST	ONLY CONC TEST	CONC TEST	CONC TEST	CONC TEST	CONC TEST	CONC TESTED/	CONC TEST				CONC TEST	CONC TEST	COMIC TEST	200	ONI V CONC TESTED	2000		CONOTESTED	CONCIESI															CONC TEST	CONC TEST		CONC TEST	//DNOM/2	//DNUM/2							ATELYHAR	CONC TEST	CONC TEST	CONC TEST	1/MG/13 C	1/2 and 1/2 and 1/2	-//WG/12.L	ALELTHAR	MIELYMAKI	SES/1760/2	DOSES/2840 mg/L	SES/2840 mg/L	CONC TEST	SES/2840 m	SES/1760/2	SES/62.94 n	SES/24.95 n	SES/24.95 n
General Comments		HARD/MODERATELY	A/14.0* mg/L/	A/14.0* mg	TMETH/APHA, 1989//	тметн/арна,	TMETH/APHA,						NUM/6//DC	A/14.0" mg	CA/14.0* mg.	A/14.0* mg/l	E	ARD/MODE			ONC1/ONLY	ONC1/ONLY	CONC1/ONLY CONC TESTED,	ONC1/ONLY	ONC1/ONLY	CONC1/ONLY CONC TESTED/	ONC1/ONLY	ONCI/ONLY				CONC. JONEY CONCITESTED/	VINO/TONC	CONCT (ONLY CONC TESTED)	1000	VINOLI /ONLY	CIACT/ CIAC		VINOT FORD	ONCI/ONLY															DNC1/ONLY CONC TESTED/	ONC1/ONLY		CONCI/CNLY CONC	74/10* mg/L	04/10° mg/L							HARD/MODERATELYHARD/	ONC1/ONLY CON	NC1/ONLY	NC1/ONLY	CA/14 OF ma// /	1911 OF THE	1,14.U mg/	HARD/MODER	ARD/MODER	2	DNUM/2//DO	OU//2//DO	NCI/ONLY	VUM/2//DO	NOM/e//DO	NUM/2//DO:	DNUM/2//DOS	JUM/2//DO:
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From: Sent: Kesler, Karen [Kesler.Karen@epa.gov] Thursday, September 29, 2016 11:34 AM

To:

Heather Ferguson

Subject:

RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

Hi Heather,

Thank you for sending this information over. Once I have a chance to review it, I will let you know if I have any questions or comments.

Thanks, Karen

From: Heather Ferguson [mailto:hlf@ftn-assoc.com]

Sent: Tuesday, September 27, 2016 2:19 PM

To: Kesler, Karen < Kesler.Karen@epa.gov >; Crocker, Philip < Crocker.philip@epa.gov >; Nelson, Russell

<nelson.russell@epa.gov>

**Cc:** 'Allan Gates' < <u>AGates@mwlaw.com</u>>; 'Frank Mills' < <u>flmills@aep.com</u>>; Patrick Downey < <u>pjd@ftn-assoc.com</u>>; James Malcolm < <u>jtm@ftn-assoc.com</u>>; 'Sarah Clem' < <u>clem@adeq.state.ar.us</u>>

**Subject:** Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in Arkansas

Good afternoon Karen,

On behalf of Southwestern Electric Power Company, please find attached the requested supplemental information for dissolved minerals tolerance values.

The attached information includes a memorandum that briefly describes our approach to obtaining the information you requested and a spreadsheet summarizing lethal and sublethal toxicity endpoints and associated hardness values. We believe this information satisfies your request and demonstrates through a weight-of-evidence approach the protection of aquatic life in the downstream reach of the Red River.

If you have any questions or comments regarding the attached information, please do not hesitate to call Jim Malcolm, Pat Downey, or me at (501) 225-7779.

Respectfully submitted,



Heather Ferguson FTN Associates, Ltd.

3 Innwood Circle, Suite 220 Se Little Rock, AR 72211

hlf@ftn-assoc.com

(501) 225-7779 🗫 fax (501) 225-6738

http://www.ftn-assoc.com

From: Heather Ferguson [hlf@ftn-assoc.com]
Sent: Heather Ferguson [hlf@ftn-assoc.com]
Thursday, September 29, 2016 11:46 AM

To: 'Kesler, Karen'

Cc: James Malcolm; Patrick Downey

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

#### Thanks Karen!



Heather Ferguson FTN Associates, Ltd. 3 Innwood Circle, Suite 220 Se Little Rock, AR 72211

3 Innwood Circle, Suite 220 So Little Rock, AR 72211 (501) 225-7779 So fax (501) 225-6738 hlf@ftn-assoc.com http://www.ftn-assoc.com

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]
Sent: Thursday, September 29, 2016 11:34 AM

To: Heather Ferguson

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

**Arkansas** 

Hi Heather,

Thank you for sending this information over. Once I have a chance to review it, I will let you know if I have any questions or comments.

Thanks, Karen

From: Heather Ferguson [mailto:hlf@ftn-assoc.com]

Sent: Tuesday, September 27, 2016 2:19 PM

**To:** Kesler, Karen < <a href="mailto:Kesler.Karen@epa.gov">Kesler.Karen@epa.gov">Kesler, Philip < <a href="mailto:crocker.philip@epa.gov">crocker.philip@epa.gov</a>>; Nelson, Russell <a href="mailto:nelson.russell@epa.gov">nelson.russell@epa.gov</a>>

**Cc:** 'Allan Gates' <<u>AGates@mwlaw.com</u>>; 'Frank Mills' <<u>flmills@aep.com</u>>; Patrick Downey <<u>pjd@ftn-assoc.com</u>>; James Malcolm <<u>jtm@ftn-assoc.com</u>>; 'Sarah Clem' <clem@adeg.state.ar.us>

**Subject:** Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in Arkansas

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Respectfully submitted,



Heather Ferguson FTN Associates, Ltd. 3 Innwood Circle, Suite 220 & Little Rock, AR 72211 hlf@ftn-assoc.com

From: Jim Malcolm [jtm@ftn-assoc.com]
Sent: Thursday, September 29, 2016 1:43 PM
To: 'Heather Ferguson'; 'Kesler, Karen'
Cc: 'Patrick Downey'; James Malcolm

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

Hey Karen, good to see you yesterday, from this reply I assume your flight back was OK. Really appreciate you taking the time to come to AR for the meeting but especially the site walkover yesterday – many thanks.

Jim

From: Heather Ferguson [mailto:hlf@ftn-assoc.com]
Sent: Thursday, September 29, 2016 11:46 AM

To: 'Kesler, Karen'

Cc: James Malcolm; Patrick Downey

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

#### Thanks Karen!



Heather Ferguson FTN Associates, Ltd.

3 Innwood Circle, Suite 220 Stittle Rock, AR 72211 hlf@ftn-assoc.com

(501) 225-7779 **%** fax (501) 225-6738 http://www.ftn-assoc.com

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]
Sent: Thursday, September 29, 2016 11:34 AM

To: Heather Ferguson

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

Hi Heather,

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Thanks, Karen

From: Heather Ferguson [mailto:hlf@ftn-assoc.com]

Sent: Tuesday, September 27, 2016 2:19 PM

To: Kesler, Karen < Kesler.Karen@epa.gov >; Crocker, Philip < Crocker.philip@epa.gov >; Nelson, Russell

<nelson.russell@epa.gov>

 $\textbf{Cc: 'Allan Gates'} < \underline{\textbf{AGates@mwlaw.com}} > ; \textbf{'Frank Mills'} < \underline{\textbf{flmills@aep.com}} > ; \textbf{Patrick Downey } < \underline{\textbf{pid@ftn-assoc.com}} > ; \textbf{James}$ 

Malcolm < itm@ftn-assoc.com >; 'Sarah Clem' < clem@adeq.state.ar.us >

**Subject:** Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in Arkansas

Good afternoon Karen,

On behalf of Southwestern Electric Power Company, please find attached the requested supplemental information for dissolved minerals tolerance values.

The attached information includes a memorandum that briefly describes our approach to obtaining the information you requested and a spreadsheet summarizing lethal and sublethal toxicity endpoints and associated hardness values. We believe this information satisfies your request and demonstrates through a weight-of-evidence approach the protection of aquatic life in the downstream reach of the Red River.

If you have any questions or comments regarding the attached information, please do not hesitate to call Jim Malcolm, Pat Downey, or me at (501) 225-7779.

Respectfully submitted,



Heather Ferguson FTN Associates, Ltd. 3 Innwood Circle, Suite 220 № Little Rock, AR 72211 hlf@ftn-assoc.com

From: Sent: Kesler, Karen [Kesler.Karen@epa.gov] Thursday, September 29, 2016 2:06 PM

To:

Jim Malcolm

Subject:

RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

Hi Jim,

Yes, the flight back went smoothly. Thank you for taking the time to show me around and really explain how the remediation process is going to work. It really helps me to have a greater understanding of how the whole system works. It was great to get to see it in person.

Thanks again,

Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]
Sent: Thursday, September 29, 2016 1:43 PM

**To:** 'Heather Ferguson' < <a href="mailto:hlf@ftn-assoc.com">hlf@ftn-assoc.com</a>; Kesler, Karen < <a href="mailto:Kesler.Karen@epa.gov">Kesler.Karen@epa.gov</a>> <a href="mailto:Cc:">Cc: 'Patrick Downey' < <a href="mailto:pid@ftn-assoc.com">pid@ftn-assoc.com</a>; James Malcolm < <a href="mailto:jtm@ftn-assoc.com">jtm@ftn-assoc.com</a>>

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

Hey Karen, good to see you yesterday, from this reply I assume your flight back was OK. Really appreciate you taking the time to come to AR for the meeting but especially the site walkover yesterday – many thanks.

Jim

From: Heather Ferguson [mailto:hlf@ftn-assoc.com] Sent: Thursday, September 29, 2016 11:46 AM

To: 'Kesler, Karen'

Cc: James Malcolm; Patrick Downey

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

**Arkansas** 

Thanks Karen!



Heather Ferguson FTN Associates, Ltd.

3 Innwood Circle, Suite 220 Se Little Rock, AR 72211 hlf@ftn-assoc.com

(501) 225-7779 **%** fax (501) 225-6738 http://www.ftn-assoc.com

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov] Sent: Thursday, September 29, 2016 11:34 AM

To: Heather Ferguson

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From: Heather Ferguson [mailto:hlf@ftn-assoc.com]

Sent: Tuesday, September 27, 2016 2:19 PM

**To:** Kesler, Karen < Kesler.Karen@epa.gov >; Crocker, Philip < Crocker.philip@epa.gov >; Nelson, Russell < nelson.russell@epa.gov >

 $\textbf{Cc: 'Allan Gates'} < \underline{\textbf{AGates@mwlaw.com'}}; \textbf{'Frank Mills'} < \underline{\textbf{flmills@aep.com'}}; \textbf{Patrick Downey } < \underline{\textbf{pid@ftn-assoc.com'}}; \textbf{James Malcolm } < \underline{\textbf{jtm@ftn-assoc.com'}}; \textbf{'Sarah Clem'} < \underline{\textbf{clem@adeq.state.ar.us'}}$ 

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From: Heather Ferguson [hlf@ftn-assoc.com]
Sent: Wednesday, November 02, 2016 2:15 PM

To: 'Kesler, Karen'

Cc: 'Sarah Clem'; James Malcolm; Patrick Downey

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Heather Ferguson
FTN Associates, Ltd.
3 Innwood Circle, Suite 220 Se Little Rock, AR 72211

From: Sent: Kesler, Karen [Kesler.Karen@epa.gov] Friday, November 04, 2016 2:03 PM

To:

Heather Ferguson

Cc:

'Sarah Clem'; James Malcolm; Patrick Downey; Crocker, Philip

Subject:

RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

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The one additional question that I have is that I noticed in your filtering of the data that you removed any tests that used more than one chemical. Did any of these tests with two chemicals involve both chlorides and sulfates? If so, those would probably be useful data to see as well.

Thanks, Karen

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Sent: Wednesday, November 02, 2016 2:15 PM
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Cc: 'Sarah Clem'; 'James Malcolm'; 'Crocker, Philip'

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

Attachments: Kunz\_et\_al-2013-Environmental\_Toxicology\_and\_Chemistry mussels.pdf; Lassier and

Hardin 2010.pdf; Mount et al 1997.pdf; Mount\_et\_al-2016-

Environmental\_Toxicology\_and\_Chemistry(1).pdf; Soucek nad Kennedy 2005-Environmental\_Toxicology\_and\_Chemistry several inverts.pdf; Soucek\_et\_al-2011-

Environmental\_Toxicology\_and\_Chemistry fingernail clams.pdf

#### Karen,

"More than one chemical" refers to a study (maybe 2) that looked at ions plus metals. In those studies the main toxicant was the metal so they were not relevant to this analysis. I have attached several studies that look at combinations of ions.

We hope that this provides sufficient information for you to complete your review.

 $\frac{1}{2} \frac{1}$ 

Pat Downey

Senior Project Manager FTN Associates, Ltd. 3 Innwood Circle Little Rock, AR 72211

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# USE OF RECONSTITUTED WATERS TO EVALUATE EFFECTS OF ELEVATED MAJOR IONS ASSOCIATED WITH MOUNTAINTOP COAL MINING ON FRESHWATER INVERTEBRATES

James L. Kunz,\*† Justin M. Conley,‡ David B. Buchwalter,‡ Teresa J. Norberg-King,§ Nile E. Kemble,† Ning Wang,† and Christopher G. Ingersoll†

†US Geological Survey, Columbia Environmental Research Center, Columbia, Missouri ‡North Carolina State University, Raleigh, North Carolina §US Environmental Protection Agency, Duluth, Minnesota

(Submitted 18 January 2013; Returned for Revision 2 April 2013; Accepted 21 August 2013)

Abstract: In previous laboratory chronic 7-d toxicity tests conducted with the cladoceran *Ceriodaphnia dubia*, surface waters collected from Appalachian sites impacted by coal mining have shown toxic effects associated with elevated total dissolved solids (TDS). The objective of the present study was to evaluate the effects of elevated major ions in chronic laboratory tests with *C. dubia* (7-d exposure), a unionid mussel (*Lampsilis siliquoidea*; 28-d exposure), an amphipod (*Hyalella azteca*; 28-d exposure), and a mayfly (*Centroptilum triangulifer*; 35-d exposure) in 3 reconstituted waters designed to be representative of 3 Appalachian sites impacted by coal mining. Two of the reconstituted waters had ionic compositions representative of alkaline mine drainage associated with mountaintop removal and valley fill-impacted streams (Winding Shoals and Boardtree, with elevated Mg, Ca, K, SO<sub>4</sub>, HCO<sub>3</sub>), and a third reconstituted water had an ionic composition representative of neutralized mine drainage (Upper Dempsey, with elevated Na, K, SO<sub>4</sub>, and HCO<sub>3</sub>). The waters with similar conductivities but, with different ionic compositions had different effects on the test organisms. The Winding Shoals and Boardtree reconstituted waters were consistently toxic to the mussel, the amphipod, and the mayfly. In contrast, the Upper Dempsey reconstituted water was toxic to the mussel, the amphipod, and the cladoceran but was not toxic to the mayfly. These results indicate that, although elevated TDS can be correlated with toxicity, the specific major ion composition of the water is important. Moreover, the choice of test organism is critical, particularly if a test species is to be used as a surrogate for a range of faunal groups. *Environ Toxicol Chem* 2013;32:2826–2835. © 2013 SETAC

Keywords: Invertebrate toxicology Mussels Mayfly Major ion toxicity

#### INTRODUCTION

Salinization of freshwater as a result of increased concentrations of major cations or major anions is a growing global concern that can result from a wide variety of human activities [1]. Major ions contributing to elevated total dissolved solids (TDS; often estimated using conductivity as a surrogate measurement) may include various combinations of Ca, Mg, Na, K, sulfate (SO<sub>4</sub>), chloride (Cl), and bicarbonate (HCO<sub>3</sub>), as well as other ions [2]. In North America, examples of activities that result in increased TDS include mountaintop mining of coal (and resultant valley filling [3]), coal processing [4,5], and the use of de-icing agents and road salts [6]. Although previous work has highlighted the potential for elevated TDS to be toxic to aquatic organisms [2,7-9], general understanding of TDS toxicity is currently inadequate, given the complexity of ionic matrices constituting high-TDS-impacted surface waters and the diversity of aquatic organisms inhabiting these waters.

Elevated concentrations of TDS in streams and rivers are widespread throughout the coal mining regions of Appalachia associated with mountaintop removal and valley fill [10,11]. During mountaintop coal mining and other surface mining, overburden layers are removed to mine the underlying coal seams. The overburden is returned to the mined areas or is deposited in headwater valleys, forming valley fills [12]. Natural

streams in this region are dilute, with conductivity generally below  $100\,\mu\text{S/cm}$ . However, conductivity is greater than  $1000\,\mu\text{S/cm}$  in streams with surface mining activities in the absence of other major TDS sources. Elevated conductivity found in surface mining streams can result from precipitation and ground water percolating through the pulverized overburden containing sulfate and carbonate materials, creating alkaline mine drainage [13].

Studies of macroinvertebrate communities from Kentucky, Virginia, West Virginia, and Pennsylvania impacted by mountaintop removal have shown that conductivity explains a substantial amount of the variance associated with various benthic assessment metrics [10]. Because macroinvertebrates inhabit dilute streams in the region, increased concentrations of major ions or ionic imbalance has been suggested to be contributing to this impairment of benthic communities. At other sites across North America, conductivity above 2000 µS/cm or TDS above about 1340 mg/L reportedly represents conditions that may adversely affect freshwater organisms [2,8,9]. In contrast, more than 90% of the sites in West Virginia and Kentucky receiving runoff water from coalfields exhibit impaired macroinvertebrate index of biotic integrity (IBI) scores when conductivity is greater than 500 µS/cm [10]. Mayflies (Ephemeroptera) are reportedly highly sensitive to TDS in these locations. Component major ions that elevate conductivity in alkaline mine drainage in these areas include Mg, Ca, K, HCO<sub>3</sub>,

In the absence of any other toxicant, major ions can be acutely or chronically toxic to aquatic life [9]. However, the correlation between increasing TDS or conductivity and toxicity varies with

\* Address correspondence to jkunz@usgs.gov Published online in Wiley Online Library

(wileyonlinelibrary.com).

DOI: 10.1002/etc.2391

All Supplemental Data may be found in the online version of this article.

ionic composition; therefore, TDS or conductivity measurements alone may not provide a basis to estimate reliably the toxicity of samples with differing ionic balances [9]. Moreover, ionic strength and ionic imbalance can both contribute to the toxicity of water to freshwater organisms [14,15].

The toxicity of major ions has been evaluated in 24-h to 96-h exposures with 3 commonly tested freshwater species (the cladocerans *Ceriodaphnia dubia* and *Daphnia magna* and fathead minnow *Pimephales promelas*) [7]. The relative ion toxicity was  $K > HCO_3 \approx Mg > Cl > SO_4$ , with *C. dubia* tending to be more sensitive compared with *D. magna* or *P. promelas* [7]. However, laboratory toxicity tests with reconstituted waters have not been conducted with test organisms more representative of taxa inhabiting freshwater Appalachian streams impacted by mountaintop removal and valley fill, including mayflies and mussels [13].

The objective of the present study was to evaluate the effects of elevated major ions in chronic laboratory tests with 4 aquatic invertebrates exposed to dilutions of 3 reconstituted waters designed to be representative of 3 Appalachian sites impacted by coal mining in southwestern West Virginia with high TDS downstream of valley fill: Winding Shoals Branch, Boardtree Branch, and Upper Dempsey Branch [13]. These 3 test sites were chosen from among multiple field-collected site waters previously demonstrated to be toxic to C. dubia in chronic 7d laboratory exposures and exhibit impaired populations of benthic invertebrates [13]. Two of these waters (Winding Shoals and Boardtree) had ionic signatures representative of alkaline mine drainage associated with streams affected by mountaintop removal and valley fill with elevated Mg, Ca, K, HCO<sub>3</sub>, and SO<sub>4</sub>), whereas Upper Dempsey had a somewhat different ionic composition, representing neutralized mine drainage or mine drainage that has experienced some cation exchange (elevated Na, K, SO<sub>4</sub>, and HCO<sub>3</sub>; Table 1 and Supplemental Data, Table S1 [13]).

The 3 reconstituted waters were used to conduct the chronic toxicity tests with a unionid mussel (*Lampsilis siliquoidea*; 28-d exposure), an amphipod (*Hyalella azteca*; 28-d exposure), and a cladoceran (*C. dubia*; 7-d exposure). Subsamples of the reconstituted waters were also provided to North Carolina State University, Raleigh, North Carolina, USA, for conducting exposures with a mayfly (*Centroptilum triangulifer*; 35-d exposure).

#### MATERIALS AND METHODS

Reconstituted test waters

The reconstituted waters were prepared at the Columbia Environmental Research Center in Columbia, Missouri, USA, by adding reagent-grade salts (K<sub>2</sub>SO<sub>4</sub>, CaSO<sub>4</sub>, NaHCO<sub>3</sub>, MgSO<sub>4</sub>, NaCl, CaCl<sub>2</sub>, or Na<sub>2</sub>SO<sub>4</sub>; Supplemental Data, Table S2) to well water or diluted well water to match the site water chemistry as closely as possible (Supplemental Data, Tables S1 and S3). The base water for Winding Shoals and Boardtree reconstituted waters was 25% well water and 75% deionized water, whereas the base water was 100% well water for Upper Dempsey reconstituted water (Supplemental Data, Table S2; the 100% well water was of approximately 300 mg/L hardness as CaCO<sub>3</sub>, with a pH of approximately 8). Salt addition recipes for the reconstituted waters were calculated to minimize differences among individual major cations or anions (i.e., attempting to match each major cation or major anion within about 20% of the site water), with no attempt to adjust alkalinity or pH (Supplemental Data, Table S1). The initial recipes were modified (e.g., by reducing NaHCO3 and not attempting to control alkalinity) to eliminate unwanted increases in pH and to ensure that the salts stayed in solution for the duration of the tests. Each salt was initially mixed individually in a 10-L jar with 8 L of the appropriate base water. The 10-L jars were placed on a stir plate and mixed for up to 24 h (depending on the solubility of individual salts). After the salts had been visibly dissolved, all salt solutions for a given reconstituted water were combined in 200-L plastic containers with a circulating pump and brought to a final volume of 130 with the appropriate base water. A similar process was used to prepare smaller 40-L samples of each reconstituted water for the second 2 wk of the exposures conducted at the Columbia Environmental Research Center or for use by North Carolina State University to conduct exposures with C. triangulifer.

Exposures with *L. siliquoidea, H. azteca*, and *C. dubia* were conducted in dilutions of Winding Shoals, Upper Dempsey, and Boardtree reconstituted waters (100%, 33%, and 10%, and a dilution water control). The *C. dubia* exposure with the Upper Dempsey reconstituted water had an additional dilution of 5% tested (in an attempt to define better the potential effects at the lower exposure concentration). All 3 reconstituted waters prepared at Columbia Environmental Research Center used

Table 1. Mean water quality characteristics of waters across all chronic toxicity tests in exposures conducted by the Columbia Environmental Research Center<sup>a</sup>

	Dissolved			Alkalinity	Hardness			Major	cations and	anions (	mg/L)	
Treatment	oxygen (mg/L)	pН	Conductivity (µS/cm)	(mg/L as CaCO <sub>3</sub> )	(mg/L as CaCO <sub>3</sub> )	Ammonia (mg N/L)	Ca	Mg	Na	K	Cl	SO <sub>4</sub>
Winding Sl	hoals											
0%	6.9 (0.8)	8.3 (0.2)	275 (25)	96 (9)	107 (10)	0.2(0.2)	30 (2)	10(1)	12(2)	2(1)	14 (2)	22 (3)
10%	7.2 (0.8)	8.2 (0.5)	504 (54)	93 (8)	223 (37)	0.3(0.1)	42 (4)	38 (6)	14 (2)	4(1)	13 (2)	141 (12)
33%	6.9 (1.1)	8.1 (0.1)	947 (48)	98 (3)	472 (13)	0.2(0.1)	65 (3)	93 (3)	18 (1)	10(1)	12 (1)	386 (33)
100%	7.2 (0.8)	8.1 (0.1)	1906 (162)	99 (6)	1154 (58)	0.3 (0.1)	109 (19)	216 (42)	24 (3)	21 (5)	12(2)	1023 (138)
Boardtree												
0%	6.9 (0.8)	8.3 (0.2)	275 (25)	96 (9)	107 (10)	0.2(0.2)	30 (2)	10(1)	12(2)	2(1)	14 (2)	22 (3)
10%	7.0 (0.8)	8.1 (0.1)	565 (36)	84 (8)	262 (26)	0.3 (0.2)	55 (6)	41 (7)	13 (2)	4(1)	12(1)	180 (37)
33%	6.9 (1.1)	8.1 (0.1)	1121 (127)	84 (11)	585 (42)	0.7 (0.6)	100 (4)	96 (7)	12(2)	8 (1)	12 (1)	489 (33)
100%	7.0 (0.8)	8.0 (0.2)	2367 (54)	72 (8)	1408 (37)	0.7 (0.1)	241 (4)	260 (6)	12(2)	21 (1)	11 (2)	1580 (12)
Upper Den	npsey											
0%	6.9 (0.8)	8.3 (0.2)	275 (25)	96 (9)	107 (10)	0.2(0.2)	30(2)	10(1)	12(2)	2(1)	14 (2)	22 (3)
10%	6.9 (0.9)	8.3 (0.2)	448 (40)	110 (15)	122 (18)	0.3 (0.1)	33 (2)	13 (1)	52 (5)	3(1)	15 (1)	78 (8)
33%	7.1 (0.8)	8.4 (0.1)	789 (55)	157 (14)	144 (13)	0.3 (0.1)	38 (2)	17 (1)	125 (6)	5 (1)	21 (3)	175 (17)
100%	6.8 (1.3)	8.4 (0.2)	1813 (157)	279 (20)	209 (36)	0.4 (0.1)	42 (9)	28 (1)	350 (16)	11 (1)	39 (2)	640 (77)

<sup>&</sup>lt;sup>a</sup> Values are means with standard deviation in parenthesis (n = 4 to 15).

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the same dilution water (control water) and were prepared by diluting well water with deionized water to a hardness of about 100 mg/L (as CaCO<sub>3</sub>), alkalinity of 85 mg/L (as CaCO<sub>3</sub>), and pH of approximately 8.3 (Supplemental Data, Table S3), Supplemental Data, Table S1, summarizes the target, nominal, and measured water-quality characteristics of the 3 test waters before dilution (i.e., at 100%). The C. triangulifer exposures were conducted in 2 rounds. Round 1 consisted of Winding Shoals, Upper Dempsey, and Boardtree waters at 100% and 50% and a dilution water control (50:50, well water:reconstituted soft water [16]; Supplemental Data, Table S3). Round 2 consisted of Winding Shoals water tested at 100%, 66%, and 33% and a dilution water control (30:70, well water:deionized water; Supplemental Data, Table S3). The first round of exposures with C. triangulifer was conducted to determine whether this organism would be responsive to the reconstituted waters. Given the results of round 1, the second round of C. triangulifer exposures was conducted to test the repeatability of results from round 1 (i.e., complete mortality at 100% strength) as well as to test additional dilutions of Winding Shoals water (i.e., 33% and 66%).

For exposures conducted at the Columbia Environmental Research Center, water-quality parameters (dissolved oxygen, pH, conductivity, hardness, alkalinity, ammonia) were determined in the control and all concentrations for each of the test waters at the beginning and the end of the tests, following standard methods [17]. In addition, dissolved oxygen and conductivity were measured weekly during the exposures. Concentrations of major ions (Ca, Mg, Na, K, Cl, SO<sub>4</sub>) were determined in the control, and all concentrations were measured at the beginning and end of the test (except for the C. dubia test, because of limited water volume). Major ions were also measured in the water samples collected from newly prepared batches of reconstituted test waters. The analyses of the major ions were conducted by Engineering Surveys and Services Testing Laboratories at Columbia, Missouri, using a Perkin Elmer AAnalyst 800 atomic absorption spectrometer and a Thermo Scientific Genesys 20 visible spectrophotometer in accordance with standard methods [17]. For exposures conducted with C. triangulifer at North Carolina State University, water-quality parameters (pH, conductivity) were measured weekly for the duration of the exposures. Dissolved oxygen was not measured, because past studies at North Carolina State University have indicated that dissolved oxygen consistently remains at approximately 6 mg/L to 8 mg/L over this period, and C. triangulifer do not display evidence of oxygen limitation until dissolved oxygen reaches approximately 1 mg/L to 2 mg/L (D.B. Buchwalter, unpublished data). At the end of round 2 for C. triangulifer, 20-mL water samples were collected from each exposure, filtered through a 0.22-µm syringe filter (Fisherbrand), and shipped on ice to the Columbia Environmental Research Center for analyses of major ions. Mean measured concentrations of these water-quality parameters of the 3 reconstituted waters used in all toxicity tests are summarized in Table 1. More details on water-quality measurements for each test with a test species are summarized in Supplemental Data, Tables S1, S3, and S4.

#### Test organisms

Lampsilis siliquoidea (approximately 2 mo old) were obtained from laboratory cultures at Missouri State University, Springfield (see Wang et al. [18] for mussel culture method). Hyalella azteca (approximately 7 d old) and C. dubia (>24 h old) used to start the exposures were obtained from laboratory

cultures at the Columbia Environmental Research Center (see Besser et al. [19] for methods used to culture H. azteca and Wang et al. [18] for methods used to culture C. dubia). These 3 species of test organisms were acclimated to the control dilution water and temperature for a minimum of 24 h before the start of the exposures [16,20–22]. Eggs of C. triangulifer (WCC-2 clone) were obtained from culture at Stroud Water Research Center, Avondale, Pennsylvania, USA. Exposures with C. triangulifer started with newly hatched larvae (first instar,  $\leq$ 48 h old) placed directly into test water at ambient laboratory temperature of 21 °C.

#### Mussel testing (Lampsilis siliquoidea)

Static-renewal exposures with mussels were conducted for 28 d in accordance with methods outlined ASTM International [20] (Supplemental Data, Table S5). Toxicity endpoints included survival and shell length. Ten juvenile mussels exhibiting foot movement were impartially transferred into each of 4 replicate 300-mL glass beakers containing 100%, 33%, and 10% test water or into 8 replicates of the dilution water control at 23 °C. Each beaker had a 2.5-cm hole in the side covered with 50-mesh (279-µm width opening) stainless steel screen and contained 200 mL water. Mussels were fed 1 mL of an algal mixture [23] twice daily on Monday through Friday and were fed 2 mL once daily on the weekends. About 2 volumes of test water were added to each beaker with flow splitters once on Mondays, Wednesdays, and Fridays [24]. Mussels were transferred into new test beakers each week. Survival of mussels was determined at the end of the exposures based on foot movement within a 5-min observation period using a dissecting microscope [20]. Surviving mussels were isolated on day 28 and preserved in 8% formalin for subsequent shell length measurement. The maximum shell length of surviving mussels was measured to the nearest 0.001 mm using a digitizing system with video micrometer software (Image Caliper, Resolution Technology).

## Amphipod testing (Hyalella azteca)

Static-renewal exposures with amphipods were conducted over 28 d in accordance with methods outlined by ASTM International [21] (Supplemental Data, Table S6). Toxicity endpoints included survival and biomass. At the beginning of the exposures, 10 amphipods were impartially transferred using a pipet into each of 4 replicate 300-mL glass beakers containing about 200 mL of 100%, 33%, and 10% test water or into 8 replicates of the dilution water control at 23 °C. A thin layer of substrate (5 mL fine sand) was added to each beaker. About 2 volumes of test water were added to each beaker with flow splitters once on Mondays, Wednesdays, and Fridays [24]. Amphipods were transferred into new test beakers each week. During exposure, amphipods in each beaker were fed 1.0 mL of yeast-cerophyll-trout chow (YCT; 1.7-1.9 g/L in a water suspension) daily. On day 28, the surviving amphipods were counted and preserved in 8% sugar formalin for subsequent length measurement. Lengths of surviving amphipods were measured from the base of the first antenna to the tip of the third uropod along the curve of the dorsal surface using a digitizing system with video micrometer software (Image Caliper, Resolution Technology) connected to a computer and a microscope. The biomass of surviving amphipods from each replicate was estimated as the sum of individual amphipod weights calculated from the empirical relationship: weight (mg) =  $([0.177 \times \text{length (mm)}] - 0.0292)^3$ [25,26].

#### Cladoceran testing (Ceriodaphnia dubia)

Static-renewal, 3-brood exposures with the cladoceran were conducted for 7-d in accordance with ASTM International [22] (Supplemental Data, Table S7). Toxicity endpoints included survival and reproduction. Test organisms were transferred to test chambers by placing 1 organism in each of 10 30-mL replicate cups containing 15 mL of 100%, 33%, and 10% test water or into 10 replicates of the dilution water control at 25 °C. The Upper Dempsey exposure had an additional dilution of 5%. Each day before water renewal, first-generation organisms were recorded as alive or dead, and live organisms were then transferred to a new test chamber containing fresh solution. Cladocerans were fed 0.1 mL of YCT (1.7-1.9 g/L in a water suspension) and 0.1 mL algal concentrate (Pseudokirchneriella subcapitata,  $3.0 \times 10^7$  cell/mL; Aquatic Biosystems) after daily water renewal. Exposures were conducted for 7 d or until >60% of the control organisms had produced 3 broods. The number of young released from firstgeneration females over each 24-h period was recorded daily.

#### Mayfly testing (Centroptilum triangulifer)

Static, nonrenewal exposures with C. triangulifer were conducted for a full life cycle (approximately 35 d; Supplemental Data, Table S8) [27-30]. The toxicity endpoints were adult survival and biomass. For each of the 2 rounds of experiments, nymphs were hatched from a single adult female clutch (clonal) to start all exposures. Two rounds of exposures were conducted. Round 1 consisted of controls and Boardtree, Upper Dempsey, and Winding Shoals waters at 100% and 50% strength. Round 2 consisted of controls and Winding Shoals water at 100%, 66%, and 33% strength. All treatments were conducted in triplicate, except in round 1, in which Upper Dempsey water exposures were conducted in duplicate because of a limited number of periphyton plates, and in round 2, in which 1 control and 1 33% Winding Shoals replicate developed a fungal growth on the periphyton surface and were omitted (resulting in n = 2 replicates). In round 1, each replicate received 20 nymphs; in round 2, each replicate received 15 nymphs. Mayflies were fed a natural periphyton diet grown at the Stroud Water Research Center by allowing fresh stream water from White Clay Creek, Pennsylvania, USA (39°51'47"N, 75°47'07"W) to flow continuously over acrylic plates  $(6.5 \times 23 \times 0.15 \text{ cm})$  in a greenhouse. Plates were cultivated until the periphyton had reached a thickness of about 1 to 2 mm and relatively uniform coverage, at which time the acrylic plates were shipped on ice overnight to North Carolina State University. Exposures were conducted in 2-L glass bottles with 1.8 L of exposure/control water, and nymphs were fed 2 sets of periphyton plates staggered 22 wk apart (i.e., days 0 and 14). Subimagos (i.e., subadults) emerged after hatching into meshlined collection lids during midafternoon to late afternoon. Subimagos were kept overnight in humid chambers containing moist paper towels to facilitate the final molt to adulthood. Gravid adults were stimulated to release eggs by wetting the abdomen in 3.5-cm Petri dishes containing 4.5 mL reconstituted soft water [16]. Adults (now postpartum) were stored in individual microcentrifuge tubes at -20 °C. Once all surviving organisms had emerged and the exposure was completed, the postpartum adults were removed from the freezer and dried at 65 °C for 48 h. Dried mayflies were individually weighed on a microbalance (Sartorius model CPA2P) to the nearest 0.001 mg.

## Data analysis

Mean survival, individual length, or biomass (total dry wt of surviving organisms per replicate) of *L. siliquoidea* and

H. azteca were calculated for each control and exposure replicate. Survival of C. dubia was calculated as number of individuals surviving in each treatment. Reproduction of C. dubia was calculated as the mean number of offspring across the control or exposure replicates.

Biomass of *C. triangulifer* was calculated based on the product of the mean body weight of surviving organisms and the number of survivors within a given replicate. Mean survival and mean biomass were calculated as the average across replicates for each control or treatment level, which had at least 3 replicates. Survival and biomass measures for controls in rounds 1 and 2 were pooled for comparison with Winding Shoals water because control performance was not significantly different between rounds. Mean survival and mean biomass were not calculated for exposures that had only 2 replicates (Upper Dempsey 50%, Upper Dempsey 100%, and Winding Shoals 33%); however, the biomass and survival of each of the 2 replicates are reported (Table 2).

The no-observed-effect concentration (NOEC) and lowestobserved-effect concentration (LOEC) for all endpoints were determined with TOXSTAT software (version 3.5; Western EcoSystems Technology) by one-way analysis of variance (ANOVA) with pairwise comparison performed using Williams' test. If the data were not normally distributed or did not have equal variances, Steel's many-one rank test or Wilcoxon rank sum test with Bonferroni adjustment was used for the determinations of NOEC and LOEC values [31]. An exception was that Wilcoxon rank sum test with Bonferroni adjustment was used to compare mean survival and biomass between the control and the Winding Shoals 50% or Boardtree 50% exposure treatments for C. triangulifer, because these tests had only 1 exposure treatment with partial mortality and at least 3 replicates (Table 2). The level of statistical significance was set at  $\alpha = 0.05$ . In most cases, no 20% effect concentration (EC20) could be estimated because the data from the toxicity test did not meet the conditions for any logistic regression analysis [32]. A 25% effect concentration (EC25) for C. dubia was estimated using the TOXSTAT software to compare responses of C. dubia in the present study and a previous study [13].

#### RESULTS

Water chemistry

Concentrations of major cation and major anions and relevant water-quality characteristics of site waters (target field chemistry reported by the US Environmental Protection Agency [USEPA] [13]) are presented in Supplemental Data, Table S1 as well as the nominal (predicted) and measured concentrations of the reconstituted waters. The quality of the reconstituted waters approximated the intended ionic compositions found in site waters (Supplemental Data, Table S1). The reconstituted Winding Shoals and Boardtree waters were largely similar to each other, with elevated concentrations of Ca, Mg, K, SO<sub>4</sub>, and HCO<sub>3</sub>, as opposed to Upper Dempsey water, which was high in K, Na, SO<sub>4</sub>, and HCO<sub>3</sub>. Relative to the dilution control water, the 100% Winding Shoals target water had proportionally higher concentrations of Ca (5.4×), Mg (24×), K (8.5×), SO<sub>4</sub> (44×), HCO<sub>3</sub> (19×), and conductivity (7.8×; Supplemental Data, Table S1). Relative to the dilution control water, the 100% Boardtree target water had proportionally higher concentrations of Ca (8.0×), Mg (25×), K (7.5×), SO<sub>4</sub> (69×), HCO<sub>3</sub> (6.0×), and conductivity (8.9×; Supplemental Data, Table S1). Relative to the dilution control water, the 100% Upper Dempsey target water had proportionally higher concentrations of Na (32×), K

Table 2. Responses of cladoceran (*Ceriodaphnia dubia*; 7-d exposures), mussel (*Lampsilis silquoidea*; 28-d exposures), amphipod (*Hyalella azteca*; 28-d exposure), and mayfly (*Centroptilum triangulifer*; ~35-d exposures) to 3 reconstituted water samples<sup>a</sup>

	Clad	oceran	Mu	ssel <sup>b</sup>	Amp	hipod <sup>b</sup>	Ma	ayfly
Treatment	Survival (%)	No. of young	Survival (%)	Length (mm)	Survival (%)	Biomass (mg)	Survival (%) <sup>c</sup>	Biomass (mg)
Winding She	oals							
Control	90	28 (10)	89 (6)	2.93 (0.6)	86 (13)	3.21 (0.8)	84 (11)	8.5 (2.0)
10%	100	31 (5)	18 (17)*	$2.79(0.3)^{d}$	83 (29)	3.25 (2.1)	- (11)	0.5 (2.0)
33%	90	30 (7)	8 (5)*	$2.49 (0.7)^{d}$	73 (17)	1.27 (0.2)*	27/33 <sup>e</sup>	1.9/2.2 <sup>e</sup>
50%	f	_	_	_	- 4.17	- (0.2)	12 (20)*	1.0 (1.7)*
66%	_	-	_	_			0*	0*
100%	100	33 (4)	3 (5)*	ND	65 (26)	$2.08 (0.9)^*$	0*	0*
NOEC	> 100%	>100%	<10%	>100%	>100%	10%	<33%g	<33% <sup>g</sup>
LOEC	>100%	>100%	10%	>100%	>100%	33%	33% <sup>g</sup>	33% <sup>g</sup>
Boardtree					7.75		5570	35 70
Control	90	28 (10)	89 (6)	2.93 (0.6)	86 (13)	3.21 (0.8)	80 (10)	8.9 (2.7)
10%	80	23 (13)	45 (29)*	3.01 (0.3)	80 (18)	2.67 (1.3)	-	0.2 (2.1)
33%	80	21 (12)	10 (8)*	$3.52 (0.5)^{d}$	73 (5)	2.53 (0.3)		
50%	-	_	_	_	_		37 (13)*	3.7 (1.8)*
100%	100	22 (7)*	30 (25)*	$2.91 (0.6)^{d}$	75 (10)	1.78 (0.2)*	0*	0*
NOEC	>100%	33%	<10%	>100%	>100%	33%	< 50%	<50%
LOEC	>100%	100%	10%	>100%	>100%	100%	50%	50%
Upper Demp	sey				7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7	2370 10	2070	3070
Control	100	40 (3)	89 (6)	2.93 (0.6)	86 (13)	3.21 (0.8)	80 (10)	8.9 (2.7)
5%	100	38 (6)				()	=	0.5 (2.7)
10%	100	36 (7)	25 (24)*	2.99 (0.5)	98 (5)	4.53 (0.3)	_	
33%	90	36 (13)	13 (10)*	$3.12 (0.4)^{d}$	95 (6)	3.55 (0.3)		
50%	_	_	_		-	-	95/70 <sup>e</sup>	11/7.9 <sup>e</sup>
100%	40*	18 (13)*	15 (6)*	2.86 (0.5)	95 (10)	$2.52(0.2)^*$	100/80 <sup>e</sup>	11/10 <sup>e</sup>
NOEC	33%	33%	<10%	>100%	>100%	33%	>100%	>100%
LOEC	100%	100%	10%	>100%	>100%	100%	>100%	>100%

<sup>a</sup> Values are means with standard deviation in parentheses (n = 10 for the cladoceran and n = 4 for other species unless otherwise noted).

<sup>b</sup> Mean starting sizes (and standard deviations) were: mussels, 1.9 mm/individual (0.3, n = 21); and amphipods, 1.89 mm/individual (0.25, n = 20).

 $^{c}$  n = 3, except for the control (n = 5), 33% (n = 2), and 100% (n = 6) treatments in the Winding Shoals water, the 50% and 100% treatments (n = 2) in the Upper Dempsey water.

<sup>d</sup>Based on 3 replicates as a result of 100% mortality in 1 replicate at these exposure concentrations.

<sup>e</sup> Two values were from the 2 replicates of this treatment and not used for statistical analysis.

<sup>f</sup>Species was not tested at this concentration.

g The 33% water was considered to be an effect concentration because the survival and biomass at this concentration was more than 60% less than those in the controls.

Significant reduction relative to control (p < 0.05).

NOEC = no-observed-effect concentration; LOEC = lowest-observed-effect concentration; ND = not determined (because of limited number of organisms for growth measurement).

(4.8×), SO<sub>4</sub> (21×), HCO<sub>3</sub> (40×), and conductivity (7.9×; Supplemental Data, Table S1).

The 3 reconstituted waters were designed to replicate the target waters, although in some cases an adjustment of the composition of reconstituted waters was necessary, as described in Materials and Methods. With limited exceptions, the nominal concentrations of major cations and major anions in all 3 reconstituted waters were within 30% of field target waters. Differences between nominal concentrations and final measured concentrations were generally within 20% for all 3 reconstituted waters (Supplemental Data, Table S1). The most notable difference between the reconstituted waters and the 100% target waters was in alkalinity (no attempt was made to control alkalinity), the reconstituted waters being below target levels of alkalinity by 72% for Winding Shoals water, 38% for Boardtree water, and 57% for Upper Dempsey water. Measured concentrations of K and Cl were consistently higher in reconstituted waters compared with target waters or nominal reconstituted waters, but the absolute differences in concentrations were generally small (i.e., <5 mg/L).

The charge balance between major cations and major anions was relatively similar (within 2.7%) between target site water quality and measured reconstituted water quality for the Upper Dempsey and Boardtree waters (Supplemental Data, Table S1).

However, for the Winding Shoals water, there was a 6.6% positive charge imbalance in the target site water and a 3.2% positive charge imbalance in the measured reconstituted water quality (Supplemental Data, Table S1). This imbalance indicates that combined concentrations of major anions in the Winding Shoals water were low (either analytical error or an omission of measuring other anions beyond HCO<sub>3</sub>, Cl, or SO<sub>4</sub>). Mean water-quality characteristics for each dilution averaged across all species exposures (Table 1) were similar to mean water-quality characteristics for each dilution for individual species exposures (Supplemental Data, Table S4).

#### Exposures

Control survival of *L. siliquoidea, H. azteca, C. dubia*, and *C. triangulifer* met the test acceptability criteria of >80% mean survival (Table 2). Reproduction of *C. dubia* also met the test acceptability criteria of >15 young/surviving female in the control and >60% of surviving females in controls producing 3 broods.

Dilutions of the Winding Shoals reconstituted waters were toxic to *L. siliquoidea* and *C. triangulifer* at the lowest concentrations tested (i.e., LOEC of 10% Winding Shoals for *L. siliquoidea* survival and LOEC of 33% Winding Shoals for *C. triangulifer* survival and biomass; Table 2). Survival of *L.* 

siliquoidea and biomass of *C. triangulifer* decreased with increasing percentage of Winding Shoals reconstituted water (Figures 1 and 2). The Winding Shoals reconstituted water was also toxic to *H. azteca* (LOEC of 33% Winding Shoals for biomass; Table 2 and Figures 1 and 2). The Winding Shoals

reconstituted water was not toxic to *C. dubia* (LOEC of >100% Winding Shoals; Table 2). By comparison, the USEPA [13] reported a 7-d EC25 for *C. dubia* of 32% Winding Shoals site water. The 100% site water from Winding Shoals had a conductivity of 2147  $\mu$ S/cm [13], which is comparable to the

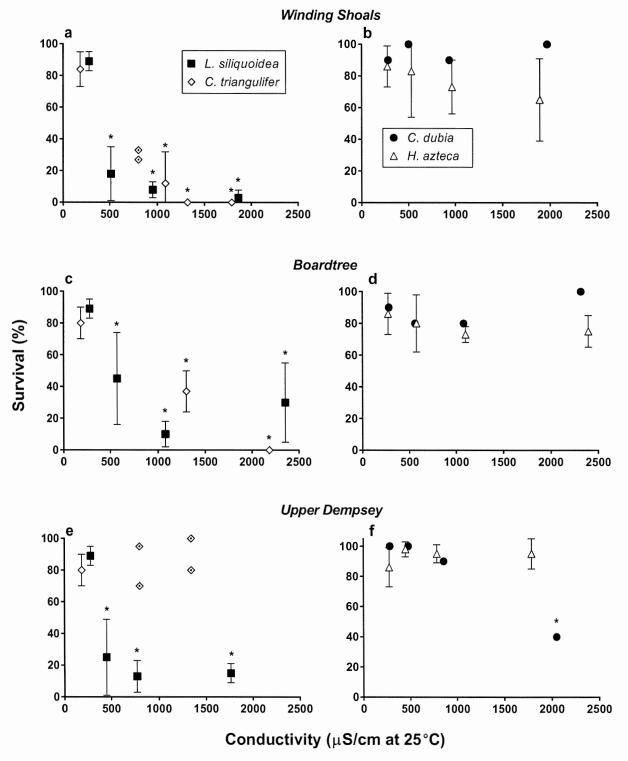


Figure 1. Mean survival of 4 freshwater invertebrates—Lampsilis siliquoidea, Centroptilum triangulifer, Ceriodaphnia dubia, and Hyalella azteca—versus conductivity of reconstituted waters. An asterisk (\*) indicates significant difference compared with control group ( $p \le 0.05$ ). Error bars indicate standard deviation. The *C. dubia* data points do not have error bars, because survival was calculated as the percentage surviving among 10 individually exposed organisms. The *C. triangulifer* data points marked with a dot represent individual replicate responses for situations in which there were <3 replicates and a standard deviation could not be calculated.

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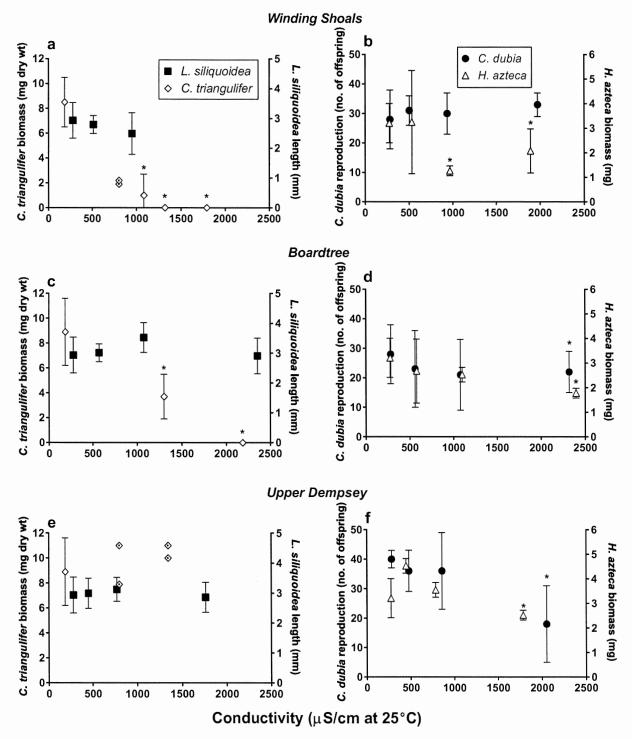


Figure 2. Sublethal response of 4 freshwater invertebrates—Lampsilis siliquoidea, Centroptilum triangulifer, Ceriodaphnia dubia, and Hyalella azteca—versus conductivity of reconstituted waters. An asterisk (\*) indicates significant difference compared with control group ( $p \le 0.05$ ). Data points represent the mean of replicates for a given treatment. Error bars indicate standard deviation. The C. triangulifer data points marked with a dot represent individual replicate responses for situations in which there were <3 replicates and standard deviation could not be calculated.

conductivity of 1906  $\mu$ S/cm in the 100% Winding Shoals reconstituted water (Table 1). Relative to the dilution water control, the 10% dilution of Winding Shoals reconstituted water that was toxic to *L. siliquoidea* had relatively higher proportional concentrations of Mg (3.8 $\times$ ), SO<sub>4</sub> (6.4 $\times$ ), and conductivity (1.8 $\times$ ; Supplemental Data, Table S1).

Dilutions of the Boardtree reconstituted waters were toxic to *L. siliquoidea* and to *C. triangulifer* at the lowest concentrations

tested (LOEC of 10% Boardtree for *L. siliquoidea* survival and LOEC of 50% Boardtree for *C. triangulifer* survival or biomass; Table 2). Survival of *L. siliquoidea* and survival or biomass of *C. triangulifer* decreased with increasing percentage of Boardtree reconstituted water (Figures 1 and 2). The Boardtree reconstituted water was also toxic to *H. azteca* (LOEC of 100% Boardtree for biomass) and *C. dubia* (LOEC of 100% Boardtree for reproduction; Table 2). By comparison, the USEPA [13]

reported a 7-d EC25 for *C. dubia* of 15% Boardtree site water. The 100% Boardtree site water had conductivity of 2582  $\mu$ S/cm [13], which is comparable to the conductivity of 2367  $\mu$ S/cm in the 100% Boardtree reconstituted water (Table 1). Relative to the dilution water control, the 10% dilution of Boardtree reconstituted water that was toxic to *L. siliquoidea* had relatively higher proportional concentrations of Mg (4.1×), SO<sub>4</sub> (8.2×), and conductivity (2.1×; Supplemental Data, Table S1).

The Upper Dempsey reconstituted waters were toxic to L. siliquoidea, H. azteca, and C. dubia (LOEC of 10% Upper Dempsey for L. siliquoidea survival and LOECs of 100% Upper Dempsey for H. azteca biomass and C. dubia survival or reproduction; Table 2). Survival of L. siliquoidea, survival and biomass of H. azteca and reproduction of C. dubia decreased with increasing percentage of Upper Dempsey reconstituted water (Figures 1 and 2). The Upper Dempsey reconstituted water was not toxic to C. triangulifer (LOEC > 100% Upper Dempsey; Table 2). A 7-d EC25 for C. dubia reproduction of about 54% for Upper Dempsey reconstituted water was estimated in the present study. By comparison, the USEPA [13] reported a 7-d EC25 of 25% Upper Dempsey site water for C. dubia. The 100% Upper Dempsey site water had a conductivity of 2143 µS/cm [13], which is comparable to the conductivity of 1813 μS/cm in the 100% Upper Dempsey reconstituted water (Table 1). Relative to the dilution water control, the 10% dilution of Upper Dempsey reconstituted water that was toxic to L. siliquoidea had relatively higher proportional concentrations of Na (4.3 $\times$ ), SO<sub>4</sub> (3.5 $\times$ ), and conductivity (1.6×; Supplemental Data, Table S1).

#### DISCUSSION

Salinization of freshwaters is a growing problem on a global scale [1]. With 4 major cations (Na, K, Ca, and Mg) and 3 major anions (Cl, HCO<sub>3</sub>, and SO<sub>4</sub>) all potentially contributing to the ionic matrix of a high-TDS water, the composition of any one site can vary dramatically from the next, particularly across geographic regions. For example, freshwater streams impacted by marine encroachment display elevated Na and Cl [33], whereas some streams affected by uranium mining display elevated Mg and SO<sub>4</sub> [34]. The ionic composition of streams in Kentucky, Virginia, and West Virginia, USA, impacted by mountaintop removal associated with coal mining are characterized by mixtures of elevated Ca, Mg, K, HCO3, and SO4 (as represented by Winding Shoals and Boardtree waters), although Na is not typically elevated compared with reference streams [10,35]. Elevated Na in mine drainage (as represented by the Upper Dempsey water) could be due to the following: 1) treating mine effluents with sodium hydroxide (NaOH) or sodium carbonate (soda ash, Na<sub>2</sub>CO<sub>3</sub>) to reduce Mn; 2) cation exchange processes in overburden that exchange Ca for Na; or 3) the presence of sodium-rich shale layers in the overburden (M. Passmore, USEPA, Wheeling, WV, USA, personal communication).

The reconstituted waters in the present study were generated to closely match site waters from the Winding Shoals, Boardtree, and Upper Dempsey Branches in West Virginia, with the exception of HCO<sub>3</sub>, which was below the field target concentrations by 38 to 72% (Supplemental Data, Table S1). This is important because bicarbonate can contribute to toxicity to *C. dubia* [7]. Research is ongoing to develop methods for preparing reconstituted waters with varying concentrations of HCO<sub>3</sub> using CO<sub>2</sub> gas to equilibrate relatively insoluble salts (CaCO<sub>3</sub> or MgCO<sub>3</sub>; D. Mount, USEPA, Duluth, MN, USA, personal communication). Use of this new approach should

result in developing reconstituted waters that better represent the alkalinity and HCO<sub>3</sub> of site waters of interest.

Sites impacted by elevated TDS may also have elevated concentrations of trace elements such as Se and Mn [10] and As, Co, and Cu [15]. Therefore, results of whole-effluent toxicity testing can be difficult to interpret with regard to which component of the effluent matrix is contributing to the toxicity of elevated TDS mixtures [14]. Investigators concerned with elevated TDS independent of other toxic elements may have to generate reconstituted waters that approximate the ionic matrix of concern as closely as possible. Importantly, the high-TDS reconstituted waters, created with the addition of elevated major ions without the addition of trace elements, were toxic to test organisms in the present study, indicating that trace elements associated with site waters were not likely substantial contributors to the toxicity observed in previous ambient-water exposures to C. dubia [13]. Moreover, ambient water-quality criteria for trace metals were exceeded only sporadically, and these overages were not related to the toxicity of the ambient waters to C. dubia [13].

Previous laboratory toxicity tests with ambient water samples from the 3 sites were identified as toxic to C. dubia [13]. Two of the 3 reconstituted waters tested (Boardtree and Upper Dempsey) in the present study were also identified as toxic to C. dubia (Table 2). Winding Shoals reconstituted water with an ionic composition similar to that of Boardtree reconstituted water was not toxic to C. dubia. In previous study, a 7-d EC20 of 996 mg SO<sub>4</sub>/L was observed for reproduction of C. dubia at a water hardness of about 100 mg/L (as CaCO<sub>3</sub>; N. Wang, unpublished data). Perhaps the slightly higher concentration of SO<sub>4</sub> in the 100% Boardtree reconstituted water (1450 mg/L) contributed to the reproductive toxicity observed in C. dubia compared with the concentration of SO<sub>4</sub> in the 100% Winding Shoals reconstituted water (1108 mg/L) that was not toxic to C. dubia, given that all other major ions were relatively similar between the Winding Shoals and Boardtree reconstituted waters (Table 1). Sulfate might have contributed to the toxicity but likely was not the sole cause of the toxicity observed to L. siliquoidea in the 10% dilutions of the reconstituted water samples (ranging from 144 to 172 mg SO<sub>4</sub>/L in the 10% dilution waters; Table 1 and Supplemental Data, Table S4). A 28-d EC20 of 696 mg SO<sub>4</sub>/L (as Na<sub>2</sub>SO<sub>4</sub>) was observed for biomass of the pink mucket (L. abrupta) at a water hardness of about 100 mg/L (as CaCO<sub>3</sub>; N. Wang, unpublished data). Potassium might also have contributed to the toxicity but likely was not the sole cause of the toxicity observed to L. siliquoidea in the 10% to 100% dilution waters (ranging up to 21 mg/L; Table 1 and Supplemental Data, Table S4). A 28-d LC20 of >25 mg K/L was observed for survival of the L. siliquoidea at a water hardness of about 100 mg/L (N. Wang, unpublished data). Additional testing of reconstituted waters with varying proportions of major ions is needed to identify the toxicity thresholds for individual ions or to determine whether major ions such as Cl, Ca, or K might ameliorate the toxicity of major ions such as SO<sub>4</sub> [5,36].

Toxicity tests have rarely been conducted with taxa resident to naturally dilute, freshwater Appalachian streams such as mayflies, stoneflies, and caddisflies (i.e., Ephemeroptera, Plecoptera, Trichoptera taxa) or with freshwater mussels. One study investigated the toxicity of high-TDS water on *C. dubia* and a mayfly (*Isonychia bicolor*) and found that *I. bicolor* was substantially more sensitive than *C. dubia* [37] in whole-effluent toxicity testing with ambient waters. Similarly, *C. triangulifer* and the *L. siliquoidea* in the present study were more responsive

to the Winding Shoals and Boardtree reconstituted waters compared with the *C. dubia* and the *H. azteca*. Furthermore, all 3 waters were toxic to *L. siliquoidea* and *H. azteca*, whereas *C. triangulifer* was unaffected by exposure to Upper Dempsey water, and *C. dubia* was unaffected by exposure to Winding Shoals water. Therefore, toxicity testing conducted with test organisms such as *C. dubia* may underrepresent the sensitivity of species present in the Appalachian streams of interest. This highlights the need for developing standard methods for testing of multiple taxa in laboratory toxicity tests that represent the sensitivity of resident taxa. Standard methods have been developed for all of the organisms tested in the present study, except for *C. triangulifer*.

Natural streams in the central Appalachian region are dilute, with conductivity generally between 40 and 100 μS/cm; however, mining-impacted streams display conductivities of about 1000 to 2500 µS/cm [10,35]. Because many macroinvertebrates have evolved in and are adapted to dilute streams in this region, overall ionic strength is a potential physiological mechanism of impairment deserving additional study. Field studies have reported declines in abundance and diversity of aquatic insects (particularly mayflies) in mountaintop miningimpacted streams across Appalachia associated with elevated TDS [9,10,35]. Based on the distributions of benthic invertebrates along conductivity gradients in sites within the Appalachian region, the USEPA [35] proposed a regional conductivity benchmark of 300 µS/cm for protection of aquatic life. The benchmark of 300 μS/cm was designed to protect 95% of the 162 genera that were used to develop the benchmark. The benchmark for conductivity applies to waters in the central Appalachians and Allegheny Plateau with circumneutral to mildly alkaline pH where the elevated conductivity is dominated by HCO<sub>3</sub> and SO<sub>4</sub>. However, no laboratory toxicity studies have investigated the potential effects of elevated conductivity associated with mountaintop mining on resident taxa (e.g., aquatic insects). The Winding Shoals and Boardtree reconstituted waters were toxic to C. triangulifer at a conductivity of about 800 to 1300  $\mu\text{S/cm}$  (Supplemental Data, Table S4) with elevated concentrations of Mg, Ca, Na, K, SO<sub>4</sub>, or HCO<sub>3</sub>. It is interesting to note that the regional 95% extirpation concentration (XC95) based on conductivity for the genus Centroptilum in the benthic community field surveys was determined to be 1092 μS/cm [35]. However, the genus Centroptilum was not among the more sensitive taxa used to derive the regional benchmark (i.e., 56 of the native taxa had XC95 values <1092 µS/cm [35]).

The Winding Shoals and Boardtree reconstituted waters were toxic to L. siliquoidea, H. azteca, or C. dubia at conductivities ranging from about 500 to 2400 µS/cm (Table 1). The Upper Dempsey reconstituted water at conductivities ranging from about 500 to 1800 μS/cm with elevated concentrations of Na. K. SO<sub>4</sub>, and HCO<sub>3</sub> was toxic to L. siliquoidea, H. azteca, and C. dubia but was not toxic to C. triangulifer at conductivity as high as about 1800 μS/cm (Supplemental Data, Table S1). Hence, it might be informative to establish toxicity thresholds based on single major ions or mixtures of major ions to individual taxa rather than to depend solely on the use of surrogate regional benchmarks such as TDS or conductivity. On the other hand, conductivity benchmarks developed within a region for specific ion matrices where natural background conductivity levels are comparable can be useful screening tools for measuring potential toxicity in the field.

Currently, the toxic mechanism of high-TDS waters is largely unknown. It has been speculated that osmoregulatory stress is potentially responsible for adverse effects in sensitive taxa as well as disruption of hemolymph pH or osmolarity [38]. Many surficial ionoregulatory pumps on aquatic insects use adenosine triphosphate to move ions against a concentration gradient [39]. It is possible that exposure to high TDS incurs bioenergetic costs as manifested by developmental delays observed in mayflies (D. Funk, Stroud Water Research Center, Avondale, PA, USA, personal communication). Soucek [40] found that *C. dubia* displayed reduced feeding and oxygen consumption when exposed to elevated concentrations of sodium sulfate. Given the large number of ions, the even larger number of ion combinations, and wide range of invertebrate physiologies, there is much to be learned about why ionic stress is toxic to aquatic organisms.

In the present study, 2 central ideas in the assessment of toxicity associated with major ions were reinforced: 1) specific ionic composition of the water is critical, and 2) selection of laboratory test species is also critical for relating major ion toxicity to field data. For example, survival of C. triangulifer and L. siliquoidea was reduced in all dilutions of Boardtree and Winding Shoals reconstituted waters (with elevated Mg, Ca, K, SO<sub>4</sub>, and HCO<sub>3</sub>), yet C. triangulifer did not exhibit adverse effects with exposure to Upper Dempsey reconstituted water (with elevated in Na, K, SO<sub>4</sub>, and HCO<sub>3</sub>) at conductivity comparable to the toxic dilutions of Boardtree and Winding Shoals reconstituted waters. Moreover, effects were observed on both L. siliquoidea and H. azteca with exposure to all 3 reconstituted waters. Conversely, C. dubia displayed no adverse effects of exposure to Winding Shoals reconstituted water. The ionic composition of the Winding Shoals and Boardtree reconstituted waters are characteristic of mountaintop-miningimpacted streams, and C. triangulifer is representative of native Appalachian taxa, albeit more tolerant to elevated conductivity based on benthic community survey data [35]. Future studies should focus on identifying the primary toxic ions or, conversely, determine whether a characteristic ionic matrix is necessary to produce toxicity. Additionally, future studies should focus on conducting toxicity tests with environmentally relevant and sensitive species, including mayflies and mussels, in order to make stronger links to benthic community survey data.

#### SUPPLEMENTAL DATA

# Tables S1-S8. (115 KB PDF).

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# OBSERVED AND PREDICTED REPRODUCTION OF *CERIODAPHNIA DUBIA* EXPOSED TO CHLORIDE, SULFATE, AND BICARBONATE

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Abstract—Chronic toxicities of Cl<sup>-</sup>, SO<sub>4</sub><sup>2</sup>-, and HCO<sub>3</sub><sup>-</sup> to *Ceriodaphnia dubia* were evaluated in low- and moderate-hardness waters using a three-brood reproduction test method. Toxicity tests of anion mixtures were used to determine interaction effects and to produce models predicting *C. dubia* reproduction. Effluents diluted with low- and moderate-hardness waters were tested with animals acclimated to low- and moderate-hardness conditions to evaluate the models and to assess the effects of hardness and acclimation. Sulfate was significantly less toxic than Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> in both types of water. Chloride and HCO<sub>3</sub><sup>-</sup> toxicities were similar in low-hardness water, but HCO<sub>3</sub><sup>-</sup> was the most toxic in moderate-hardness water. Low acute-to-chronic ratios indicate that toxicities of these anions will decrease quickly with dilution. Hardness significantly reduced Cl<sup>-</sup> and SO<sub>4</sub><sup>2</sup><sup>-</sup> toxicity but had little effect on HCO<sub>3</sub><sup>-</sup>. Chloride toxicity decreased with an increase in Na<sup>+</sup> concentration, and HCO<sub>3</sub><sup>-</sup> toxicity may have been reduced by the dissolved organic carbon in effluent. Multivariate models using measured anion concentrations in effluents with low to moderate hardness levels provided fairly accurate predictions of reproduction. Determinations of toxicity for several effluents differed significantly depending on the hardness of the dilution water and the hardness of the water used to culture test animals. These results can be used to predict the contribution of elevated anion concentrations to the chronic toxicity of effluents; to identify effluents that are toxic due to contaminants other than Cl<sup>-</sup>, SO<sub>4</sub><sup>2</sup>-, and HCO<sub>3</sub><sup>-</sup>; and to provide a basis for chemical substitutions in manufacturing processes. Environ, Toxicol. Chem. 2010;29:347–358. Published 2009 SETAC

Keywords—Ceriodaphnia dubia Anions Hardness Acclimation Effluents

#### INTRODUCTION

Interpretations of toxicity elicited from municipal and industrial wastewaters can be confounded by the presence of common ions (Na<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, K<sup>+</sup>, Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, HCO<sub>3</sub><sup>-</sup>) at elevated concentrations. Slightly saline effluents characterized by high levels of dissolved salts are produced by many manufacturing processes and potentially cause toxic responses in required evaluations [1]. Salinity and total dissolved solids (TDS) are inclusive metrics describing the total mass of ions in solution and are often cited as the most probable source of chronic toxicity in effluents [1-3]; however, common ions do not generally pose a risk to human health or to the environment [1]. Based on acute toxicities and typical effluent concentrations, the anions are often likely to be responsible for this type of toxicity. A salinity-toxicity relationship established for the acute toxicity of these ions indicated that Na+ and Ca2+ were not toxic to aquatic animals and determined the relative toxicity of the others to be  $K^{+} > HCO_{3}^{-} \sim Mg^{2+} > Cl^{-} > SO_{4}^{2-}$  [4]. Sodium is often the dominant cation in wastewaters, followed by Ca2+ and lesser concentrations of K+ and Mg2+, but concentrations of one or more of the dominant anions (Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, HCO<sub>3</sub><sup>-</sup>) are often equal to or greater than that of Na<sup>+</sup> and vary substantially among effluents [5–7]. However,

Ceriodaphnia dubia (Cladocera) is a standard test organism used for assessments of effluents discharged into freshwater systems, and reproduction is the typical endpoint used to determine chronic toxicity [13]. Although this animal is generally sensitive to elevated TDS (>1,000 mg/L), often significant differences in the toxicities of effluents with similar TDS concentrations suggest that toxicity is dependent on specific ions in solution [1,14]. At the same time, C. dubia can also be sensitive to ionic conditions that are lower than those used for their culture [1], and water hardness is one of those parameters [11,12]. Hardness can play a significant role in ameliorating anion toxicity to C. dubia in toxicity tests, and reproduction of C. dubia acclimated to moderately hard water can be significantly reduced when exposed to low-hardness water [12]. The same effects could be expected in effluent assessments. Toxicities elicited may reflect gradients in hardness, as well as effluent concentration within the dilutions, and could be quite different depending on the dilution water used. A problematic

within the discharge permits of many wastewaters, chronic toxicity is the criterion that is monitored. To improve effluent-toxicity interpretations, information concerning the toxicities of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> is required [8]. Some information on the chronic toxicity of Cl<sup>-</sup> to *Ceriodaphnia dubia* is available [9–11], and *C. dubia* have been exposed to Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> to examine effects of water quality and acclimation, but estimates of their toxicities were not made [12]. Additional information is needed regarding the toxicities of these anions, including their interactions with one another and with other water-quality parameters such as hardness.

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situation can occur when *C. dubia* is used to assess the toxicity of wastewaters with elevated TDS and low hardness. Some effluents, such as those produced in the processes of manufacturing textiles, often have these characteristics.

Wastewaters generated during the production of textiles frequently have high TDS content because of the use of Na+ salts and alkaline solutions in many dyeing and finishing operations. The resulting high-pH and -alkalinity conditions require acidification prior to discharge, resulting in even greater TDS. Meanwhile, Ca<sup>2+</sup> and Mg<sup>2+</sup> (primary hardness components) can have adverse effects during textile production, such as scaling and chemical interference, and are usually removed from process waters prior to use [15,16]. In addition, lowhardness effluents are typical of regions such as the southeastern United States, where many ground waters and surface waters are naturally low in Ca<sup>2+</sup> and Mg<sup>2+</sup> [11,17]. The appropriateness of using C. dubia to evaluate the toxicity of effluents with low hardness has been questioned because of its preference for moderately hard water [11]. Use of moderately hard water for the cultures of C. dubia and dilution of low-hardness effluents may result in test solutions that vary in effluent content and hardness concentration, potentially leading to a more toxic response because of the reduced protective effect of hardness and increased negative effect of exposure to lower hardness conditions.

To address these issues common to a wide range of effluents, the present study was designed to determine the chronic toxicities of Cl $^-$ , SO $_4$  $^2-$ , and HCO $_3$  $^-$ , identify interaction effects among the three anions, develop models to predict their chronic toxicities, evaluate the effectiveness of the models in predicting effluent toxicity, and assess the influence of hardness in culture and dilution water on effluent toxicity estimates. The results provide a better understanding of the impact of slightly saline effluents and insight into reducing the toxicity of some wastewaters through substitution with less toxic components.

#### MATERIALS AND METHODS

The toxicities of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> were originally determined in water with characteristics similar to those of many effluents produced by textile manufacturers in the southeastern United States (low hardness, moderate alkalinity). However, a negative effect was apparent when C. dubia cultured in water with moderate hardness and moderate alkalinity (M water) were exposed to low-hardness control waters. To remove the influence of the change in hardness, cultures of C. dubia were initiated in low-hardness/moderate-alkalinity water (T water). These cultures were propagated for 10 months and then used to assess anion toxicities under hardness and alkalinity conditions similar to those of textile effluents. Additional cultures were later started in low-hardness/low-alkalinity water (L water) similar to many surface waters in the Southeast and used to demonstrate the significant effects of sudden decreases in hardness on C. dubia reproduction [12]. To provide a more complete assessment of anion toxicities in low- to moderate-hardness waters, toxicity tests of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> were subsequently conducted with animals from the L and M cultures approximately 12 months after the tests using animals from the T cultures. As a result, anion toxicities were determined in three types of formulated water with animals that had been acclimated to the hardness and alkalinity concentrations within the test solutions. Responses were evaluated for each type of water as well as combined to provide information for low to moderate hardness and alkalinity conditions.

Study design

The reproductive responses of C. dubia to increasing anion concentrations in three different water formulations were used to generate predictive models of chronic anion toxicity in lowto moderate-hardness water. Water formulations included M, which contained 100 mg/L hardness and 70 mg/L alkalinity and is suggested for culturing C. dubia to be used for whole-effluent toxicity testing [13]; L, which contained 40 mg/L hardness and 40 mg/L alkalinity; and T, which contained 40 mg/L hardness and 100 mg/L alkalinity. Initial toxicity assessments focused on single-anion toxicity tests used to establish low-level inhibition concentrations (ICs) for the individual anions that represented a reduction in reproduction compared with the control of 6 (IC6), 12 (IC12), 25 (IC25), and 50 (IC50). These single-anion toxicity tests were repeated twice in the T water and three times in the L and M waters. Mean IC values for Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> were used in a three-factor experiment with four levels to create a matrix of treatments which was used to evaluate anion interactions and to model anion-mixture effects. The four concentrations of each factor (Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup>) included their IC6, IC12, and IC25 values and their concentration in the control water. In this design, 64 treatments were evaluated within each type of water, and evaluations were repeated three times. The size of this design required partitioning the treatments into three groups for the T water or four groups for the L and M waters. To provide comparable responses, a separate control treatment was evaluated for each test and used to normalize results among the tests by dividing the treatment reproduction by the control reproduction. Responses from anion-mixture exposures were used to produce two multivariate models that predict anion toxicity, one based on anion concentrations alone and one that also incorporates the effects of low to moderate hardness conditions.

The final phase of the study assessed the models by determining the toxicities of actual effluents and comparing them with predicted responses. Two separate dilution series were prepared for each effluent using either L or M water to assess the effects of low and moderate hardness conditions on effluent toxicities. In effluent assessments, animals exposed in a dilution series were cultured in the water formulation used as the diluent. In addition, animals acclimated to L or M water were also used to test the dilution series produced with M or L water, respectively, to examine the influence of acclimation.

Toxicity testing

Waters used for cultures and testing were reconstituted from well-aerated deionized water and reagent-grade chemicals. The L water contained 28 mg/L CaSO<sub>4</sub>  $\cdot$  2H<sub>2</sub>O, 28 mg/L MgSO<sub>4</sub>, and 61 mg/L NaHCO<sub>3</sub>; M water contained 60 mg/L CaSO<sub>4</sub>  $\cdot$  2H<sub>2</sub>O, 60 mg/L MgSO<sub>4</sub>, and 96 mg/L NaHCO<sub>3</sub>; T water contained 28 mg/L CaSO<sub>4</sub>  $\cdot$  2H<sub>2</sub>O, 28 mg/L MgSO<sub>4</sub>, and 158 mg/L NaHCO<sub>3</sub>. All three types of water contained 4 mg/L KCl and 5  $\mu$ g/L Na<sub>2</sub>SeO<sub>4</sub>.

Toxicity testing of anion and effluent solutions followed procedures recommended for assessing the chronic toxicity of effluents with C. dubia [13]. Exposures were conducted in 30-ml plastic cups containing 15 ml of test solution, one animal, and 0.1 ml of Pseudokirchneriella subcapitata and 0.1 ml of a yeast, alfalfa, Tetrafin® (Tetra Werke, Melle, Germany) food preparation [13]. Ten replicates of each treatment were randomly assigned a position within holding boards and placed in an environmental chamber that maintained temperature at 25°C and a 16:8-h light:dark photoperiod. Neonates (<24 h old and produced within an 8-h period) used in testing were produced in individual cultures according to recommended procedures using the L, M, or T water. Test animals were transferred daily into prepared cups with reproduction and survival being recorded. Exposures lasted for 6 or 7 d, and total reproduction was the test endpoint with only the first three broods included. Tests were considered acceptable if control survival was greater than 80%; at least 60% of the control animals produced three broods, and mean control reproduction was above 15 neonates per adult. In all tests, the survival rate of control animals was ≥90%; at least 80% of the control replicates had three broods within the test period; and control reproduction was always well above the minimum. Mean numbers of neonates produced in the controls of each test were 26 (coefficient of variation [CV] = 9%, n = 11), 25 (CV = 11%, n = 19), and 25 (CV = 14%, n = 19) in the T, L, and M waters, respectively.

Additions of NaCl, Na<sub>2</sub>SO<sub>4</sub>, or NaHCO<sub>3</sub> provided elevated anion concentrations in the test solutions, which were prepared 24 h in advance of testing and kept under gentle aeration at 25°C until use. To determine single-anion toxicities, five concentrations of each anion were used as treatment levels in addition to the control and consisted of serial dilutions of stock solutions prepared in L, T, or M water. Anion-mixture treatments represented a completely crossed factorial design within each type of water. Four levels of each anion were tested including Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> concentrations in the L, M, or T waters as the lowest concentration of each anion with three additional levels (IC6, IC12, IC25) determined in the single-anion toxicity tests. Anion-mixture solutions were prepared with weighed additions of the sodium salts to 2-L aliquots of L, T, and M water. Nominal concentrations in solutions prepared with L water were 2, 39, 76, 147 mg/L Cl<sup>-</sup>; 44, 161, 278, 430 mg/L  $HCO_3^-$ ; and 38, 254, 469, 625 mg/L  $SO_4^{2-}$ . Nominal concentrations in solutions prepared with M water were 2, 303, 350, 456 mg/L Cl<sup>-</sup>; 85, 139, 209, 379 mg/L HCO<sub>3</sub><sup>-</sup>; and 81, 754, 841, 1,060 mg/L SO<sub>4</sub><sup>2-</sup>. Nominal concentrations in solutions prepared with T water were 2, 193, 243, 340 mg/L Cl<sup>-</sup>; 115, 271, 352, 476 mg/L HCO<sub>3</sub><sup>-</sup>; and 38, 308, 367, 496 mg/L SO<sub>4</sub><sup>2-</sup>.

Nominal concentrations were used for calculations involving single-anion and anion-mixture toxicities because of the large number of tested solutions (714). However, subsets of samples were collected for quality assurance and analyzed by contract laboratory using an ion chromatograph for  $\rm Cl^-$  and  $\rm SO_4^{2-}$  concentrations and a carbon analyzer to determine total and inorganic carbon ( $\rm HCO_3^-$ ). Analytical samples were collected from all treatments in the first set of single-anion tests prepared in the T, L, and M waters and from one set of anion-mixture treatments prepared in each water (n=92 for  $\rm Cl^-$  and  $\rm SO_4^{2-}$  and n=76 for  $\rm HCO_3^-$ ). Samples were collected at test initiation along with measurements of basic water chemistry. Analytical determinations for  $\rm Cl^-$  and  $\rm SO_4^{2-}$  varied

above and below their nominal values, but analytical measurements of HCO<sub>3</sub> were consistently at or below its nominal values. The difference between measured and nominal HCO<sub>3</sub> concentrations tended to be greatest in the upper-concentration treatments prepared in M water. Mean differences from nominal concentrations were 7% (CV = 27) for Cl<sup>-</sup>, 8% (CV = 40) for  $SO_4^{2-}$ , and 6% (CV=9) for  $HCO_3^{-}$ . For single-anion and anion-mixture tests conducted in L and M waters, conductivity was measured again at the conclusion to provide an estimate of the loss of ions due to precipitation during the testing period. Few comparisons of initial and final conductivities indicated a substantial loss of dissolved ions. Nine of 498 solutions had conductivities that decreased by greater than 1.5% (3.9% maximum) during the tests. Most of these were treatments prepared in M water that contained the upper concentrations of HCO<sub>3</sub><sup>-</sup> and SO<sub>4</sub><sup>2-</sup>, but losses were not consistent among repeated tests.

#### Effluent testing

Eleven municipal and industrial effluents with some history of chronic toxicity attributed to TDS were sampled and assessed with the C. dubia reproduction test to evaluate the efficacy of the aniontoxicity models. Each effluent was represented by a grab sample collected 48 h prior to testing. The effluent was passed through a 163-µm mesh cloth to remove large particles and refrigerated and aerated overnight. Effluents were brought to 25°C and prepared for testing on the following day. The toxicity and chemical constituents of each effluent were initially unknown. Therefore, conductivity was used to provide an estimate of an appropriate range of dilutions [1]. If the conductivity of the raw effluent was above 2,000 microsiemens/cm (µS/cm), it was diluted with M water to produce a dilution series with conductivities of 2,000, 1,500, 1,000, and 500 µS/cm. However, if effluent conductivity was below 2,000 µS/cm, a dilution series of 100, 50, 25, and 12.5% was employed. Those dilution proportions were then matched with dilutions prepared with L water. Effluent dilutions were prepared in 2-L glass flasks, covered, and refrigerated until use. All effluent dilutions were measured for concentrations of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, HCO<sub>3</sub><sup>-</sup>, NO<sub>3</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, and total and inorganic C and five trace metals (Cr, Cu, Ni, Pb, Zn). Samples for analytical analyses were passed through a 0.2-µm nylon filter and split into subsamples for anions and metals. Samples for metal analyses were acidified with 1% nitric acid (v/v). Cation concentrations were measured by contract laboratory using inductively coupled plasma mass spectroscopy. Anions were determined as described above, and dissolved organic carbon (DOC) was calculated by subtracting inorganic carbon from total carbon. Hardness concentrations of effluent dilutions were calculated using the measured concentrations of Ca<sup>2+</sup> and Mg<sup>2+</sup> [18]. Mean recoveries of ions from spiked effluents were as follows: Cl<sup>-</sup> 97% (SD = 6, n = 16),  $SO_4^{2-}$  104% (SD = 9, n = 16),  $HCO_3^{-}$  92% (SD = 7, n=8), Na<sup>+</sup> 97% (SD=4, n=6), Ca<sup>2+</sup> 93% (SD=8, n=6), and  $Mg^{2+}$  91% (SD = 10, n = 6).

#### Water chemistry

Basic water chemistries of the single-anion solutions, anion-mixture solutions, and effluent dilutions measured in house included conductivity, pH, dissolved oxygen, hardness (effluents excluded), alkalinity, and total ammonia (effluents only) and were determined at test initiation. Hardness and

alkalinity were determined by titration [19] and are reported in units of mg/L as CaCO<sub>3</sub>. Conductivity, pH, dissolved oxygen, and total ammonia were determined using appropriate meters (Orion<sup>TM</sup> models 1214000, 720 A with triode electrode, 1113000, 720 A with ammonia gas electrode, respectively; Thermo Fisher Scientific). Values of TDS and ionic strength were based on nominal concentrations in anion solutions and measured concentrations in effluent dilutions. Ionic strength was calculated according to the formula described by Stumm and Morgan [20].

The formulated waters used for dilutions provided several comparisons based on their chemistries. Calcium, Mg<sup>2+</sup>, and  $SO_4^{2-}$  concentrations in the L and T waters were less than half those in the M water, and concentrations of Na+ and HCO3 were greatest in the T water, followed by the M and L waters. The T and M waters were similar in TDS (210 and 208) mg/L, respectively) and contained almost twice as much TDS as L water (115 mg/L). Mean pH ranged from 8.0 to 8.3, and mean hardness concentrations were 44 (CV = 4), 44 (CV = 6), and 93 (CV=4) mg/L; mean alkalinities were 45 (CV=7), 101 (CV=6), and 66 (CV=4) mg/L; and conductivities averaged 178 (CV = 4), 306 (CV = 4), and 313 (CV = 6)  $\mu$ S/cm for the L (n=14), T (n=11), and M (n=14) waters, respectively. Dissolved oxygen was near saturation in all test solutions. Additions of Cl and SO42 had little effect on solution pH, but the pH of solutions containing elevated concentrations of HCO<sub>3</sub> was between 8.2 and 8.7.

#### Data analyses

A linear interpolation method [21] was employed to determine single-anion toxicities in the form of point estimates (inhibition concentration; IC) that reduced reproduction by 6, 12, 25, and 50% compared with the control treatment. These IC values were averaged for the repeated tests (n=2 or 3), compared among anions and waters using the Tukey and Tukey-Kramer means comparison methods [22], and incorporated in the formulations of anion-mixture solutions to be assessed. Tukey's method was also used to assess differences in reproduction among control treatments. Significant differences within statistical comparisons were based on an alpha value  $\leq 0.05$ . The distribution and variance homogeneity of reproduction data was assessed by using the Shapiro-Wilkes test and Bartlett's test, respectively, and no-observable-effect concentrations and lowest-observable-effect concentrations were determined with Dunnett's test or Steel's many-one rank test, depending on whether the distribution was normal or nonnormal [13].

Anion-mixture toxicities were evaluated and modeled using statistical procedures provided by SAS® software (SAS Institute). Data sets representing each water type contained the means of 30 replicates (10 replicates from three tests) for each of the 64 mixture solutions. Analysis of variance determined the significance of anion interactions, and the general linear models procedure modeled *C. dubia* reproduction using anion concentrations and water hardness. Fixed concentrations of hardness were assigned to each data set; 40 mg/L for L and T waters and 100 mg/L for M water. Reproduction data from anion-mixture and effluent toxicity assessments were divided by the control reproduction for that water and test prior to use in calculations. Model assumptions were evaluated by plotting calculated

residuals against predicted values and the independent variables of the model and by creating normal probability plots of the residuals. Model validity was assessed with Pearson's coefficient of regression between observed and predicted reproduction and by evaluating F statistics for the model components. The goodness of fit provided by predictive models was assessed using Akaike's information criterion (AIC) [23] and  $R^2$  values.

Inhibition concentrations of effluents (point estimates as a percentage of effluent) that reduced *C. dubia* reproduction by 25% and 50% were also calculated by linear interpolation [21]. Significant differences between effluent IC values were based on the lack of overlap between 95% confidence intervals. Predicted point estimates for each effluent were calculated by regression analysis using the predicted response for each dilution with the percentage of effluent in the dilution. Predicted IC values were compared with the 95% confidence intervals of IC values determined from observed responses as a means of judging model accuracy.

#### RESULTS

Single-anion toxicity

Toxicity tests, conducted in three types of formulated water, provided information describing C. dubia reproduction under low to moderate hardness and alkalinity conditions. The toxicities of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> relative to one another varied depending on the unit of measurement (mg/L, mM/L, meq/L) used for anion concentration (Table 1). Toxicities based on mass or equivalents were fairly consistent across the three types of water, with  $SO_4^{2-}$  being the least toxic. With concentrations based on moles,  $SO_4^{2-}$  was often the most toxic of the three. Although mass is often used to describe the toxicity of a contaminant, the use of molar concentrations has been promoted because of the intrinsic relationship between toxicity and number of toxicant molecules [24]. However, the use of mea/L incorporates both the number of moles and the ionic charge and is therefore more appropriate for comparing anion toxicities than using mass or molar concentrations. Based on equivalents, the relative toxicities of theses anions in moderate-hardness water were  $HCO_3^- > CI^- > SO_4^{2-}$ .

Response curves illustrate differences among the toxicities of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> when assessed under different water quality conditions (Fig. 1). Sulfate and CI were significantly less toxic in moderate-hardness water than in low-hardness water, but the toxicity of HCO<sub>3</sub><sup>-</sup> was relatively unaffected by water hardness. Sulfate toxicity was unaffected by increasing alkalinity, but Cl toxicity was reduced in water with moderate alkalinity (T) compared with water with low alkalinity (L). For the L, T, and M waters, respectively, the mean no-observable-effect concentrations/lowest-observable- effects concentrations for Cl<sup>-</sup> were 101/250, 250/500, and 500/650 mg/ L; those for  $SO_4^{2-}$  were 500/750, 250/500, and 1,000/1,250; and those for HCO<sub>3</sub><sup>-</sup> were 350/500, 300/500, and 243/450. Variability among repeated tests was greatest for Cland  $SO_4^{2-}$  in low-hardness water (Table 1). However, in exposures to HCO<sub>3</sub><sup>-</sup>, toxicity was consistent in tests using low-hardness waters but variable in moderate-hardness water.

When toxicity was determined with concentrations of TDS, ionic strength, or Na<sup>+</sup> in the test solutions, inconsistencies among IC values determined in the Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> solutions indicated that neither TDS, nor ionic strength, nor Na<sup>+</sup>

Table 1. Mean inhibition concentrations (IC with coefficient of variation) for *Ceriodaphnia dubia* reproduction based on three units of measurement for Cl<sup>-</sup>,  $SO_4^{2-}$ , and  $HCO_3^{-}$  in low-hardness/low-alkalinity (L, n = 3), moderate-hardness/moderate-alkalinity (M, n = 3), and low-hardness/moderate-alkalinity (T, n = 2) waters<sup>a</sup>

Water	Unit		IC50			IC25	
L (44 mg/	L hardness / 45	mg/L alkalinity)					
	mg/L	SO <sub>4</sub> <sup>2-</sup> 766 (13) A	HCO <sub>3</sub> <sup>-</sup> 587 (1) B	Cl <sup>-</sup> 342 (17) C	SO <sub>4</sub> <sup>2-</sup> 625 (14) A	HCO <sub>3</sub> <sup>-</sup> 430 (13) B	CI <sup>-</sup> 147 (20) C
	mM/L	Cl <sup>-</sup> 9.7 A	HCO <sub>3</sub> <sup>-</sup> 9.6 A	SO <sub>4</sub> <sup>2-</sup> 8.0 A	HCO <sub>3</sub> <sup>-</sup> 7.0 A	SO <sub>4</sub> <sup>2-</sup> 6.5 A	Cl <sup>-</sup> 4.2 B
	meq/L	SO <sub>4</sub> <sup>2-</sup> 15.9 A	Cl <sup>-</sup> 9.7 B	HCO <sub>3</sub> <sup>-</sup> 9.6 B	SO <sub>4</sub> <sup>2-</sup> 13.0 A	HCO <sub>3</sub> <sup>-</sup> 7.0 B	Cl <sup>-</sup> 4.2 B
M (93 mg	L hardness / 66	6 mg/L alkalinity)					
	mg/L	SO <sub>4</sub> <sup>2-</sup> 1252 (5) A	HCO <sub>3</sub> <sup>-</sup> 725 (14) B	Cl <sup>-</sup> 653 (4) B	SO <sub>4</sub> <sup>2-</sup> 1060 (4) A	Cl <sup>-</sup> 456 (18) B	HCO <sub>3</sub> <sup>-</sup> 379 (46) B
	mM/L	Cl <sup>-</sup> 18.4 A	SO <sub>4</sub> <sup>2-</sup> 13.1 B	HCO <sub>3</sub> <sup>-</sup> 11.9 B	Cl <sup>-</sup> 12.9 A	$SO_4^{2-}$ 11.0 AB	HCO <sub>3</sub> <sup>-</sup> 6.2 B
	meq/L	SO <sub>4</sub> <sup>2-</sup> 26.1A	Cl <sup>-</sup> 18.4 B	HCO <sub>3</sub> - 11.9 C	$SO_4^{2-}$ 22.0 A	CI <sup>-</sup> 12.9 B	HCO <sub>3</sub> - 6.2 C
T (44 mg/	L. hardness / 10	1 mg/L alkalinity)	10.7 <b>D</b>	11.7 C	22.0 A	12.7 B	0.2 C
. (	mg/L	SO <sub>4</sub> <sup>2-</sup> 715 (6) A	HCO <sub>3</sub> <sup>-</sup> 651 (2) AB	Cl <sup>-</sup> 563 (6) B	SO <sub>4</sub> <sup>2-</sup> 496 (8) A	HCO <sub>3</sub> <sup>-</sup> 476 (9) A	Cl <sup>-</sup> 340 (16) A
	mM/L	Cl <sup>-</sup> 15.9 A	HCO <sub>3</sub> - 10.7 B	SO <sub>4</sub> <sup>2-</sup> 7.5 C	CI 9.6 A	$HCO_3^-$ 7.8 AB	SO <sub>4</sub> <sup>2-</sup> 5.2 B
	meq/L	Cl <sup>-</sup> 15.9 A	SO <sub>4</sub> <sup>2-</sup> 14.9 A	HCO <sub>3</sub> <sup>-</sup> 10.7 B	SO <sub>4</sub> <sup>2-</sup> 10.5A	7.6 AB CI <sup>-</sup> 9.6 A	HCO <sub>3</sub> <sup>-</sup> 7.8 A

<sup>&</sup>lt;sup>a</sup> IC50 and IC25 values in a row with the same letter are not significantly different ( $\alpha = 0.05$ ).

was responsible for the observed toxicity (Table 2). Their relative toxicities varied considerably based on the predominant anion and the type of water used to prepare the solutions. Total dissolved solids and ionic strength were significantly less toxic in  $SO_4^{2-}$  solutions than in  $HCO_3^-$  solutions and most of the  $Cl^-$  solutions, and  $Na^+$  was almost twice as toxic in  $HCO_3^-$  solutions compared with  $SO_4^{2-}$  solutions. Inhibition concentrations based on these parameters simply describe their status at the anion-based point estimate of toxicity.

## Anion-mixture toxicity

Analysis of variance conducted with each data set (L, M, or T, n = 64) consistently identified Cl<sup>-</sup>,  $SO_4^{2-}$ , and  $HCO_3^{-}$  and the interaction between Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> as highly significant predictors ( $\alpha < 0.0001$ ) of *C. dubia* reproduction. Interaction between  $SO_4^{\ 2-}$  and  $HCO_3^{\ -}$  was significant in solutions prepared in the L water, but not in M or T water, and interaction between Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> was not significant. Regressions incorporating the SO<sub>4</sub><sup>2-</sup>:Cl<sup>-</sup> ratio provided better fits to the data (based on  $R^2$ ) than those using the Cl<sup>-</sup>:SO<sub>4</sub><sup>2-</sup> ratio or their product, indicating that mixtures containing lower proportions of Cl<sup>-</sup> (compared with SO<sub>4</sub><sup>2-</sup>) were less toxic. Models generated with the combined data set (n = 192) also demonstrated that all three anions contributed significantly to the toxicity of mixture solutions, that there was a significant interaction effect between SO<sub>4</sub><sup>2-</sup> and Cl<sup>-</sup>, and that hardness significantly reduced toxicity (Table 3). A five-variable model (Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, HCO<sub>3</sub><sup>-</sup>, SO<sub>4</sub><sup>2-</sup>:Cl<sup>-</sup>, hardness) provided the best combination of precision and complexity, but a model with four variables using anion concentrations and the SO<sub>4</sub><sup>2-</sup>:Cl<sup>-</sup> ratio also represented the data well. Use of only individual anion concentrations provided an  $R^2$  of 0.75 and an AIC of 128. Including the  $SO_4^{2-}$ :Cl<sup>-</sup> ratio into this model increased the  $R^2$  to 0.84 and decreased the AIC to 47; adding the hardness concentration as an additional variable in the model further increased the  $R^2$  to 0.89 and brought the AIC to 0. Model coefficients were calculated using units of mass (mg/L) because of its use in reporting analytical data. However, molar or equivalent units can be substituted by dividing the coefficients by the molecular or equivalent weights of their anions.

Assumptions of uncorrelated and normally distributed errors used in statistical analyses of regression data were validated with several criteria. Highly significant F statistics for all model components and Pearson correlation coefficients close to 1.0 (0.94-0.97) in the L, T, M, and combined data sets demonstrated strong relationships between independent variables and C. dubia reproduction. Model residuals were ordered along straight lines in normal probability plots, and, with one exception, plots of residuals versus predicted responses and independent variables contained no patterns. Plots comparing residuals with predicted responses in M water revealed two distinct groups of observations that roughly corresponded to levels of reproduction (above 60% or below 30%). This pattern appeared to correspond to a gap among anion concentrations in test solutions (depicted as TDS in Fig. 2) that was due to the combinations of greater IC values established for the individual anions in moderate-hardness water (Table 1).

Total dissolved solids is an easily used term to describe mixture solutions including effluents. However, the correspondence between TDS and toxicity in these effluents was not as consistent as predictions of the anion-concentration models, although it was slightly better than that between toxicity and ionic strength or conductivity. A regression using TDS and reproduction in the combined data set provided an  $R^2$  of 0.70. Regressions comparing ionic strength or conductivity with reproduction resulted in  $R^2$  values of 0.62 and 0.68, respectively. *Ceriodaphnia dubia* that had been cultured and tested in M water were more tolerant of greater TDS concentrations than those produced and exposed to either L or T water (Fig. 2). Inhibition concentrations (IC50) based on TDS were 1,139, 1,081, 1,569, and 1,248 mg/L for the L, T, M, and combined data sets, respectively. With similar TDS concentrations in the

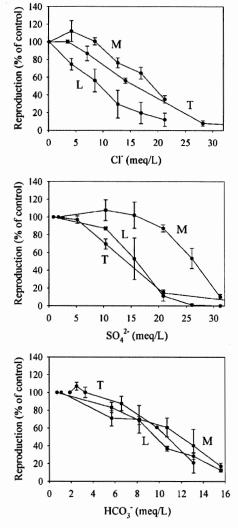


Fig. 1. Mean reproduction of *Ceriodaphnia dubia* ( $\pm 1$  SD) in single-anion toxicity tests of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> using low-hardness/low-alkalinity water (L; n=3), low-hardness/moderate-alkalinity water (T; n=2), and moderate-hardness/moderate-alkalinity water (M; n=3).

Table 2. Mean inhibition concentrations (IC50 with coefficient of variation) that reduce *Ceriodaphnia dubia* reproduction by 50% for total dissolved solids (TDS; mg/L), ionic strength (IS), and Na<sup>+</sup> (mg/L) determined from assessments of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> in low-hardness/low-alkalinity (L, n = 3), low-hardness/moderate-alkalinity (T, n = 2), and moderate-hardness/moderate-alkalinity (M, n = 3) waters<sup>a</sup>

			IC50	
Water		Cl-	HCO <sub>3</sub>	SO <sub>4</sub> <sup>2-</sup>
L	TDS	689 (12) B	901 (3) B	1,206 (12) A
	IS	0.012 (12) B	0.012 (3) B	0.025 (13) A
	Na <sup>+</sup>	244 (13) B	232 (3) B	370 (13) A
T	TDS	1,506 (5) A	989 (4) B	1,517 (2) A
	IS	0.026 (5) B	0.013 (4) C	0.031 (2) A
	Na <sup>+</sup>	558 (6) A	256 (4) C	466 (2) B
M	TDS	1,318 (2) B	1,034 (8) C	1,900 (4) A
	IS	0.024 (2) B	0.014 (8) C	0.040 (4) A
	Na <sup>+</sup>	468 (2) B	252 (9) C	574 (4) A

<sup>&</sup>lt;sup>a</sup> Concentrations in a row with the same letter are not significantly different ( $\alpha = 0.05$ ).

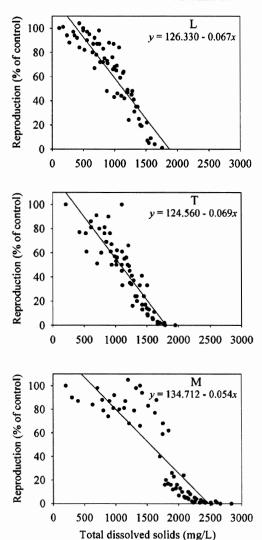


Fig. 2. Mean reproduction of *Ceriodaphnia dubia* plotted against total dissolved solids when exposed to anion-mixture solutions prepared in low-hardness/low-alkalinity water (L), low-hardness/moderate-alkalinity water (T), and moderate-hardness/moderate-alkalinity water (M).

Table 3. Predictive models of chronic toxicity that use concentrations of anions (mg/L) and hardness (mg/L as  $CaCO_3$ ) as independent variables to estimate *Ceriodaphnia dubia* reproduction as a percentage of the control reproduction (n = 192)

				Standard	
	$R^2$	Variable	Coefficient	error	α
Model 1	0.84	Intercept	111.516	2.8458	< 0.0001
		$SO_4^{2-}$	-0.079	0.0038	< 0.0001
		Cl-	-0.049	0.0088	< 0.0001
		$HCO_3^-$	-0.105	0.0073	< 0.0001
		SO <sub>4</sub> <sup>2-</sup> /Cl <sup>-</sup>	0.110	0.0106	< 0.0001
Model 2	0.89	Intercept	92.182	3.2874	< 0.0001
		$SO_4^{2-1}$	-0.087	0.0034	< 0.0001
		Cl	-0.091	0.0089	< 0.0001
		HCO <sub>3</sub>	-0.093	0.0063	< 0.0001
		$SO_4^{2-}/C1^-$	0.078	0.0097	< 0.0001
		Hardness	0.553	0.0639	< 0.0001

Table 4. Inorganic components and total dissolved solids (TDS) of effluents used to assess anion-toxicity models

Effluent	Cl (mg/L)	SO <sub>4</sub> (mg/L)	HCO <sub>3</sub> (mg/L)	Ca (mg/L)	Mg (mg/L)	Na (mg/L)	TDS (mg/L)	Cr (µg/L)	Cu (µg/L)	Ni (μg/L)	Pb (µg/L)	Zn (µg/L)
1	411	728	199	31	6	419	1,796	6	16	4	2	22
2	51	53	117	19	10	48	303	4	12	1	0	15
3	390	715	1,940	7	4	1,210	4,416	8	13	6	2	27
4	3	51	274	2	1	125	501	8	5	0	0	97
5	76	59	224	5	1	159	534	9	3	1	0	18
6	79	17	241	16	2	175	602	6	5	18	1	241
7	50	410	107	14	1	265	871	35	9	3	3	242
8	14	20	148	5	2	60	254	0	2	1	0	32
9	91	118	1,065	9	12	608	1,972	9	2	2	0	24
10	612	373	479	8	3	616	2,146	0	19	0	0	34
11	263	1,740	719	30	10	1,060	3,893	0	4	5	0	25

T and M waters used for *C. dubia* cultures, the difference in the TDS IC values for these waters suggests that exposure in moderate hardness conditions was responsible for the decreased toxicity.

#### Effluent testing/model prediction

Effluents collected from various textile manufacturers and municipal waste-treatment plants around the southeastern United States provided realistic examples and were comprised primarily of Na<sup>+</sup> and a mixture of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> (Table 4). Calcium and Mg<sup>2+</sup> concentrations indicated soft to moderately hard conditions, with hardness concentrations ranging from 9 to 115 mg/L. Effluent alkalinities varied from 58 to 860 mg/L, with pH values of 8.2 to 9.1; DOC concentrations ranged from 1 to 108 mg/L; total ammonia was present in measurable amounts only in effluent 10 (0.2 mg/L). Chronic toxicities of seven effluents were predicted reasonably well by both models. However, only four effluents (3, 9, 10, 11) provided dilutions in which toxicity appeared to be caused primarily by anion concentrations, although the potential influence of other measured or unmeasured components could not be ruled out. Three effluents (2, 6, 8) contained relatively low anion concentrations and exhibited no chronic toxicity as was predicted by the models. On the other hand, four effluents (1, 4, 5, 7) also had relatively low anion concentrations but elicited much more toxicity than predicted. These results presumably were due to toxic levels of trace metals or other undetermined components. Copper and Ni exceeded concentrations causing chronic toxicity in the upper dilutions of effluent 1, and Cu and Zn exceeded these concentrations in effluent 7 (http://epa.gov/ waterscience/criteria/nrwqc-2006.pdf), but causes for toxicities of effluents 4 and 5 were unknown. Only test results from effluents 3, 9, 10, and 11 were considered further. In treatments with the greatest proportion of these effluents, K<sup>+</sup> ranged from 11 to 47 mg/L and  $NO_3^-$  ranged from 0 to 9 mg/L, and  $PO_4^{3-}$ and DOC concentrations were low as well (0-2 mg/L and 1-7 mg/L, respectively). Concentrations of K<sup>+</sup> and NO<sub>3</sub><sup>-</sup> were below reported levels of chronic toxicity [25,26].

Effluent toxicity differed significantly with the type of dilution water used in two of the four effluent assessments (Table 5, Fig. 3). Toxicity tests of effluents 9 and 10 provided essentially the same responses regardless of the type of dilution water used, but effluent 3 was significantly more toxic when diluted with moderate-hardness water, whereas effluent 11 was significantly more toxic when diluted with low-hardness water. Both models produced fairly accurate estimates of reproduction, which were used to approximate effluent toxicity

(Fig. 3). Predictions with model 1 were generally closer to observed responses for effluents 3 and 10 (hardness concentrations < 40 mg/L), whereas predictions with model 2, which included hardness, were more accurate for effluents 9 and 11 (hardness concentrations > 70 mg/L). However, the observed responses might also reflect influences of other effluent components that can often confound assessments of toxicity, such as DOC and undetermined contaminants.

There was a significant difference in the toxicity of effluent 3 depending on the type of dilution water used, with increased toxicity in moderately hard water. Both models adequately predicted the toxicity of effluent 3 when it was diluted with L water, but model 2 substantially underestimated its toxicity when diluted with M water (Table 5, Fig. 3). Predicted point estimates for effluent 3 were within confidence intervals of those determined from the observed responses in low-hardness dilutions, as was the IC50 estimate of model 1 in moderatehardness dilutions. Predicted reproduction was often within 1 SD of the observed responses. In dilutions prepared with low-hardness water, predictions by both models were virtually the same, because the hardness concentration in all four dilutions was close to 40 mg/L (Table 6), which was the low concentration used to establish model 2. The greater toxicity in moderate-hardness dilutions was unexpected because of the small differences in anion concentrations and suggested a negative effect because of the acclimation of test animals to moderate-hardness conditions followed by exposure to lowerhardness test solutions. Chemistries of the low- and moderate-

Table 5. Observed and predicted inhibition concentrations (% effluent) that reduce *Ceriodaphnia dubia* reproduction by 25% and 50% for effluents diluted with low-hardness (L) or moderate-hardness (M) waters<sup>a</sup>

				Pı	redicted	respon	se	
			response CI)	Mod	lel 1	Model 2		
Effluent	Water	IC25	IC50	IC25	IC50	IC25	IC50	
3	L	15 (10–18)	22 (20–25)	11	20	11	20	
	M	3 (2-7)	13 (9-18)	9	18	15	23	
9	L	43 (36-49)	61 (55-63)	20	38	26	49	
	M	41 (29-53)	60 (51-65)	17	35	37	56	
10	L	29 (26-31)	48 ND	30	55	22	42	
	M	36 (19-38)	46 (43-48)	27	51	30	48	
11	L	4 (3–11)	23 (16–25)	14	25	14	26	
	M	30 ND	NT	13	24	20	33	

<sup>&</sup>lt;sup>a</sup> ND = confidence interval (CI) could not be determined; NT = reproduction in greatest effluent concentration >50% of control reproduction.

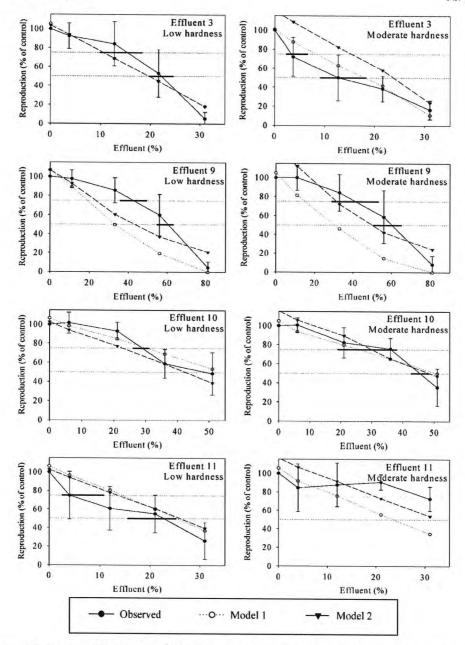


Fig. 3. Mean reproduction of  $Ceriodaphnia\ dubia\ (\pm 1\ SD; n=10)$  in effluents diluted with low- and moderate-hardness waters compared with predicted estimates from two anion-toxicity models. Model 1 incorporates concentrations of  $Cl^-$ ,  $SO_4^{\ 2^-}$ , and  $HCO_3^-$ , and model 2 uses hardness as an additional variable. Confidence intervals (95%) calculated for 50 and 25% inhibition concentrations are denoted by thick lines along the 50 and 75% reproduction levels.

hardness dilutions were similar and varied primarily as a result of differences in the dilution waters; hardness concentrations declined with increasing effluent concentrations in moderate-hardness dilutions while remaining constant in low-hardness dilutions (Table 6). Most of the toxicity observed in effluent 3 appeared to be due to HCO<sub>3</sub><sup>-</sup> and Cl<sup>-</sup>. The IC25 and IC50 values determined for low-hardness dilutions of effluent 3 were 15% and 22%, respectively. In the actual 13% and 22% dilutions of effluent 3 (Table 6), concentrations of HCO<sub>3</sub><sup>-</sup> were between the IC25 and the IC50 values established in low-hardness solutions, and effluent Cl<sup>-</sup> concentrations were between the IC12 and IC25 values (Table 1).

Effluent 9 elicited the same pattern of toxicity regardless of the dilution water used (Fig. 3). Its toxicity was substantially overestimated in low-hardness dilutions by both models, and model 1 overestimated its toxicity in moderate-hardness dilutions. However, point estimates and predicted responses calculated with model 2 for dilutions prepared with moderate-hardness water were reasonably accurate (Table 5). Effluent 9 contained moderate hardness that remained relatively constant among M water dilutions and increased with effluent concentration in L water dilutions. It also contained the highest concentration of DOC (7 mg/L) of the four effluents. The toxicity of effluent 9 appeared to be due primarily to HCO<sub>3</sub> (Table 6). The IC25 and IC50 values determined for the effluent in both low- and moderate-hardness waters were approximately 42% and 60%, respectively. Treatments within this range (33 and 56% effluent) contained concentrations of

Table 6. Chemical concentrations (mg/L) measured in effluent dilutions and used in model predictions

			Low-hardn	ess dilution water	r		Moderate-har	dness dilution w	rater
Effluent	Percentage	CI <sup>-</sup>	SO <sub>4</sub> <sup>2-</sup>	HCO <sub>3</sub>	Hardness	CI-	SO <sub>4</sub> <sup>2-</sup>	HCO <sub>3</sub>	Hardness
3	4	14	59	125	35	13	102	154	72
	13	52	123	296	35	53	160	322	70
	22	84	180	465	34	108	259	484	65
	31	139	260	631	35	152	317	653	61
9	11	7	32	192	39	9	65	244	78
	33	29	54	540	46	29	78	555	74
	56	48	75	802	55	48	92	835	72
	81	70	95	971	60	63	91	994	69
10	6	26	44	81	28	26	75	107	58
	21	99	89	143	27	100	119	171	59
	36	194	142	209	28	189	168	228	47
	51	292	194	272	28	291	211	296	49
11	4	9	88	84	31	9	119	111	63
	12	26	201	141	33	26	229	166	65
	21	50	360	204	41	48	389	228	72
	31	74	546	275	48	72	551	293	77

 $\mathrm{HCO_3}^-$  above its IC25 and IC50 values, whereas the concentrations of  $\mathrm{Cl}^-$  and  $\mathrm{SO_4}^{2-}$  in effluent 9 would not have contributed much toxicity given the level of hardness present (Tables 1 and 6).

The toxicity of effluent 10 was also not significantly affected by the difference in dilution water (Table 5). However, in contrast to effluent 9, effluent 10 contained less hardness and HCO<sub>3</sub><sup>-</sup> and more Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup>, and both models provided fairly accurate estimates of reproduction and effluent IC values (Tables 5 and 6). Chloride appeared to be the most toxic component, with concentrations that ranged between its IC25 and IC50 values (Table 1). Bicarbonate concentrations were between its IC6 and IC12 values, and  $SO_4^{2-}$  was slightly below its IC6 value. Concentrations of hardness in the low-hardness dilutions of effluent 10 were well below 40 mg/L (Table 6), causing model 2 to predict lower reproduction than model 1 and to overestimate toxicity. A confidence interval could not be calculated for the IC50 determined for the low-hardness dilutions because of a lack of sufficient range in effluent concentration.

The type of dilution water had a significant effect on the toxicity of effluent 11, with a toxic response in dilutions using low-hardness water and a nontoxic response in dilutions using moderate-hardness water (Table 5, Fig. 3). Both models underestimated the toxicity of effluent 11 in lowhardness dilutions and overestimated its toxicity in moderatehardness dilutions. However, predicted reproduction was within 1 SD of the observed responses in the low-hardness dilutions and the lower two dilutions prepared with moderatehardness water. Hardness concentrations in effluent dilutions increased with effluent concentrations, causing a reduction in the slope of toxicity midway through the dilutions (Table 6 and Fig. 3). Chloride,  $SO_4^{2-}$ , and  $HCO_3^{-}$  all contributed to the toxicity of the low-hardness dilutions (Table 1), but only HCO<sub>3</sub> was present in toxic concentrations (between IC12 and IC25) in the moderate-hardness dilutions. Confidence intervals and the IC50 value could not be determined for moderate-hardness dilutions of effluent 11 as a result of mean reproduction greater than 70% of the control in all dilutions.

The effects of acclimation to hardness conditions were also noticeable in the effluent assessments. Exposing *C. dubia* cul-

tured at one hardness concentration to test solutions with lesser or greater concentrations as a result of effluent dilutions resulted in substantially different levels of reproduction, particularly in dilutions eliciting responses between the 75 and 50% reproduction levels used to determine IC25 and IC50 values (Fig. 4). In the 13% dilutions of effluent 3, animals cultured in lowhardness water had greater reproduction than those cultured in moderate-hardness water. The 56, 36, and 22% dilutions of effluents 9, 10, and 11, respectively, indicated less reproduction from animals cultured in moderate-hardness water and exposed to low-hardness dilutions and more reproduction from animals cultured in low-hardness water and exposed to moderate-hardness dilutions. Reproduction of animals in control treatments also reflected this pattern with consistently lower reproduction from animals cultured in moderate-hardness water and tested in low-hardness water (Table 7). These results highlight the potential variability in toxicity determinations when using C. dubia reproduction because of differences in hardness concentrations within the effluent, dilution water, and culture water.

# DISCUSSION

The chronic toxicities of Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> determined in the present study were similar to those established in other research. but information regarding the chronic toxicity of HCO<sub>3</sub><sup>-</sup> was not available for comparison. Chloride IC50 values for C. dubia were similar to those determined elsewhere in low-hardness water [10] and in moderate-hardness water [9,11]. The IC50 value for  $SO_4^{2-}$  determined in moderate-hardness water was close to that determined by Soucek [27], who also tested Na<sub>2</sub>SO<sub>4</sub> in a similar water formulation. With the toxicity of  $SO_4^{2-}$  measured as TDS, the IC25 (1,518 mg TDS/L) determined in moderate-hardness water was comparable to the IC25 (1,443 mg/L) determined in water with a slightly lower hardness [14]. Hardness significantly reduced the chronic toxicities of Cl<sup>-</sup> and  $SO_4^{2-}$  and has been observed to reduce the acute toxicity of  $SO_4^{2-}$  as well [7,8,14,28]. In using acutetoxicity values for 24-h tests [7] and IC25 values from the present study, acute-to-chronic ratios (in moderate-hardness water) for Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> were low (4.5, 2.3, and

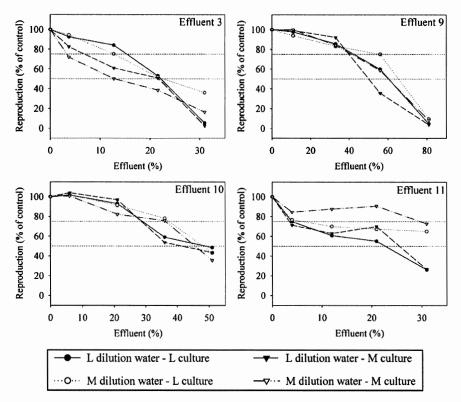


Fig. 4. Mean reproduction of Ceriodaphnia dubia in effluent assessments using animals cultured in low-hardness (L) or moderate-hardness (M) water and exposed to effluent dilutions prepared with low- and moderate-hardness waters.

2.7, respectively), demonstrating that dilution quickly reduces their toxicity [24]. These ratios were similar to those found in surveys of effluents from chemical manufacturers and publicly owned wastewater treatment facilities [29], supporting the assertion that the major anions may contribute significantly to the chronic toxicity of many effluents.

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The relatively high IC values of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> compared with other contaminants may indicate impairment of the animal's osmoregulatory functions or a disruption of its acid-base balance. Freshwater invertebrates concentrate Cl<sup>-</sup> against a chemical gradient by using Na<sup>+</sup>/Cl<sup>-</sup>/K<sup>+</sup>/adenosine triphosphate transport mechanisms and excrete SO<sub>4</sub><sup>2-</sup> and HCO<sub>3</sub><sup>-</sup> as byproducts of respiration [30,31]. Chloride is directly related to the active uptake of Na<sup>+</sup> and elimination of HCO<sub>3</sub><sup>-</sup>, and all three anions contribute to the acid-base balance [31]. The importance of the ionic charge to these processes justified using equivalent units to compare

Table 7. Mean reproduction (with coefficient of variation, n=10) of Ceriodaphnia dubia from low-hardness (L) and moderate-hardness (M) cultures when exposed to low- and moderate-hardness control treatments during effluent assessments<sup>a</sup>

	L c	ulture	М сі	M water 31.1 (7.3) A 29.8 (11.8) A		
Effluent	L water	M water	L water	M water		
3 9 10 11	24.1 (10.8) B 30.7 (6.9) A 28.8 (9.8) A 31.4 (7.7) AB	25.5 (11.6) B 29.7 (7.8) A 28.0 (7.1) A 30.0 (14.2) AB	23.4 (12.1) B 24.4 (13.0) B 25.7 (21.3) A 27.2 (23.8) B			

<sup>&</sup>lt;sup>a</sup> Values in a row with the same letter are not significantly different  $(\alpha = 0.05)$ .

common ion toxicities. Based on equivalents,  $HCO_3^-$  is the most acutely toxic of the anions, followed by  $Cl^-$  and  $SO_4^{2-}$  [4]. The same pattern was evident in chronic exposures from the present study conducted in moderate-hardness water  $(HCO_3^- > Cl^- > SO_4^{2-})$ .

Substantial differences were observed in the toxicities of Cl and  $SO_4^{2-}$  because of characteristics of the waters in which they were tested. However, the toxicity of HCO<sub>3</sub><sup>-</sup> remained fairly consistent across the three types of water. Hardness reduced the toxicities of SO<sub>4</sub><sup>2-</sup> and Cl<sup>-</sup>, and Cl<sup>-</sup> toxicity was also reduced in water with moderate alkalinity compared with low-alkalinity water. However, the reduction in Cl<sup>-</sup> toxicity most likely was due to the increase of Na<sup>+</sup> rather than the increase of alkalinity. Alkalinity in these solutions was provided by additions of NaHCO<sub>3</sub>, and after disassociation the additional HCO<sub>3</sub><sup>-</sup> should add toxicity. However, ion regulation in freshwater animals is dependent on cellular interactions involving Na<sup>+</sup> [30,31] and the additional Na<sup>+</sup> in the moderate-alkalinity water might have mitigated the impacts of the Cl and HCO3. The toxicity of Cl<sup>-</sup> relative to SO<sub>4</sub><sup>2-</sup> and HCO<sub>3</sub><sup>-</sup> was dependent on the type of water in which it was tested. Variability in Cl toxicity resulting from changes in hardness and Na<sup>+</sup> resulted in its toxicity being similar to that of  $SO_4^{\ 2-}$  and significantly less than that of HCO<sub>3</sub><sup>-</sup> in water with more Na<sup>+</sup> and low hardness, similar to  $HCO_3^-$  and significantly greater than  $SO_4^{2-}$  in waters containing less Na+ and low hardness, and more toxic than  $SO_4^{2-}$  but less toxic than  $HCO_3^-$  in water with more Na<sup>+</sup> and moderate hardness.

The toxicities of anion mixtures generally represent the additive effects of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup>, with significant effects contributed by hardness and an interaction between Cl<sup>-</sup> and  $SO_4^{2-}$ . Although measurements such as TDS, conductivity, and ionic strength can provide rough estimates of toxicity, the inherent variability in these measurements precludes their use as reliable predictors. An example can be seen in a comparison of effluent toxicities based on conductivity. Dilutions of the four effluents (3, 9, 10, 11) all represented the same sequence of conductivity (500, 1,000, 1,500, 2,000 µS/cm), but the variability in reproduction among dilutions with the same conductivity was often substantial (Fig. 3). Reproduction in effluent dilutions with conductivities of 1,000 μS/cm ranged from 50% to 93%; reproduction was between 39 and 91% in dilutions with conductivities of 1,500 µS/cm and between 5 and 73% in dilutions with conductivities of 2,000 µS/cm. The apparent reduction of TDS toxicity in moderate-hardness water (Fig. 2) most likely was due to the collective effect of hardness on the anion toxicities. Even though there appeared to be an upper limit of TDS/conductivity/ionic strength with regard to reproduction, the effects almost certainly were due to the cumulative anion toxicities.

Models created from anion-toxicity data predicted the chronic toxicity of effluents relatively well when toxicity was related to concentrations of common ions. Model 2, which includes hardness as a variable, provided better estimates of reproduction than model 1 when effluents contained moderate hardness but became less effective when concentrations of hardness dropped below 40 mg/L. The models can be used to estimate the proportion of effluent toxicity expected to be contributed by the major anions, to indicate when the presence of other toxicants should be investigated, and to suggest changes in chemical components that may result in lower toxicity. Assessments of effluent 9, which was dominated by HCO<sub>3</sub> and contained moderate hardness, were virtually identical with low- and moderate-hardness waters, supporting single-anion toxicity results indicating that hardness does not significantly affect HCO3 toxicity, although the DOC content of effluent 9 may have reduced its toxicity below levels predicted by the models. This would suggest that DOC has an antagonistic effect on the toxicity of HCO<sub>3</sub><sup>-</sup> similar to its effect on the toxicity of other inorganic and organic contaminants [32–34]. However, the possibility of an undetermined effluent component causing the reduction in toxicity also exists. The beneficial effect of hardness was most pronounced in assessments of effluent 11. Low-hardness dilutions contained toxic concentrations of Cl and SO<sub>4</sub><sup>2-</sup>, but, in moderatehardness dilutions, similar concentrations of Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> were not toxic. Sulfate and Cl<sup>-</sup> should be favored over HCO<sub>3</sub><sup>-</sup> in processes that produce wastewaters because of their lower chronic toxicity and because their toxicities can be significantly reduced by relatively small amounts of hardness.

The concentration of hardness in dilution and culture waters had a significant effect on reproduction in some situations but not others. Similarly to previous research [12], acclimation of test organisms to low and moderate hardness conditions had significant effects on reproduction in effluent treatments as well as control treatments. These results support other standard procedures [35] suggesting that *C. dubia* be reared in water with a hardness similar to the hardness of solutions intended for testing. Test animals often produced fewer neonates when exposed to an effluent dilution with a lower hardness concentration than their culture water. Effluents with low concentrations of hardness assessed with animals cultured in moderate-

hardness water may elicit a false-positive response simply because of the change in hardness. Given the possibility of these effects, acclimation of *C. dubia* to the hardness concentration to be tested is a logical solution that requires relatively little effort.

#### CONCLUSIONS

The chronic toxicity observed in evaluations of many municipal and industrial effluents may often be due to concentrations of Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup> that exceed the tolerance of *C. dubia*. Toxicities of these anions decrease quickly with dilution and so present the greatest risk to small receiving streams in which the effluent is the major source of water. For the purposes of permitting effluents, an effort should be made to separate the effects of these anions from those of more persistent and toxic compounds also present in order to provide a more realistic assessment of environmental impact. Information developed in the present study can help to facilitate this distinction. Differences in hardness and conditions of acclimation affected determinations of chronic toxicity in effluents, with relatively small additions of hardness often reducing effluent toxicity. By using measured anion concentrations and the predictive models of chronic anion toxicity, the proportion of effluent toxicity attributable to the major anions can be estimated and potential substitutions with less toxic components suggested.

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# Produced Water Series

# STATISTICAL MODELS TO PREDICT THE TOXICITY OF MAJOR IONS TO CERIODAPHNIA DUBIA, DAPHNIA MAGNA AND PIMEPHALES PROMELAS (FATHEAD MINNOWS)

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Abstract—Toxicity of fresh waters with high total dissolved solids has been shown to be dependent on the specific ionic composition of the water. To provide a predictive tool to assess toxicity attributable to major ions, we tested the toxicity of over 2,900 ion solutions using the daphnids, *Ceriodaphnia dubia* and *Daphnia magna*, and fathead minnows (*Pimephales promelas*). Multiple logistic regression was used to relate ion composition to survival for each of the three test species. In general, relative ion toxicity was  $K^+ > HCO_3^- \approx Mg^{2+} > Cl^- > SO_4^{2-}$ ;  $Na^+$  and  $Ca^{2+}$  were not significant variables in the regressions, suggesting that the toxicity of  $Na^+$  and  $Ca^{2+}$  salts was primarily attributable to the corresponding anion. For *C. dubia* and *D. magna*, toxicity of  $Cl^-$ ,  $SO_4^{2-}$ , and  $K^+$  was reduced in solutions enriched with more than one cation. Final regression models showed a good quality of fit to the data ( $R^2 = 0.767-0.861$ ). Preliminary applications of these models to field-collected samples indicated a high degree of accuracy for the *C. dubia* model, while the *D. magna* and fathead minnow models tended to overpredict ion toxicity.

Keywords---Ions

Total dissolved solids

Salinity

Toxicity

Ceriodaphnia dubia

#### INTRODUCTION

Natural fresh waters contain several ionic constituents at greater than trace levels. Indeed, ions such as Na<sup>+</sup>, Ca<sup>2+</sup>, Cl<sup>-</sup>, and others are required at a minimum level to support aquatic life, and these major ions are components of most formulas for "reconstituted" water used in aquatic toxicity testing [1,2]. However, many natural and anthropogenic sources can increase ion concentrations to levels toxic to aquatic life. Studies of oil and gas produced waters [3–5], irrigation drain waters [6,7], shale oil leachates [8], sediment pore waters [9,10], and industrial process waters [11,12] have shown toxicity caused by elevated concentrations of common ions.

Typically, integrative parameters such as conductivity, total dissolved solids (TDS), or salinity are used as a measure of the concentrations of common ions in fresh waters. While for a given ionic composition there is undoubtedly a correlation between increasing conductivity or TDS and increasing toxicity, these parameters are not robust predictors of toxicity for a range of water qualities. For example, Burnham and Peterka [13] noted that fathead minnows could tolerate TDS concentrations up to 15,000 mg/L in Saskatchewan lakes dominated by Na<sup>+</sup> and SO<sub>4</sub><sup>2-</sup>, but populations did not persist above 2,000 mg/L in Na+/K+/HCO3-dominated lakes of Nebraska. In studies of irrigation drain waters, Dickerson et al. [7] found Ceriodaphnia dubia 50% lethal concentration (LC50) values corresponding to approximate conductivities of 3,500 to 4,000 μS/cm (calculated), while Jop and Askew [11] showed major ion toxicity to C. dubia in an industrial process water with a conductivity of only 1,800 µS/cm (K.M. Jop, personal communication). Studies by Dwyer et al. [14] demonstrated that the toxicity of high TDS waters to *Daphnia magna* and striped bass *Morone saxatilis* was dependent on the specific ionic composition of those waters.

Given the substantial differences in toxicity among major ion salts [15], these differing responses in waters with different ionic compositions are to be expected. Still, they emphasize the inadequacy of generic measures for assessing the potential toxicity of major ions and the need for a broader understanding of major ion toxicity. This paper presents research to develop more comprehensive tools for assessing major ion toxicity. Acute toxicity tests using three freshwater organisms were conducted on solutions enriched with varying combinations of major ions. Results of these tests were incorporated into multivariate logistic regression models that predict survival of the three test species based on major ion concentrations.

#### MATERIALS AND METHODS

Test organisms

All organisms used in testing were obtained from in-house cultures (ENSR, Fort Collins, CO, USA); daphnids were less than 24 h old at test initiation, while fathead minnows were 1 to 7 d old. *Ceriodaphnia dubia* were cultured in either moderately hard reconstituted water (MHRW) or 20% mineral water [1] at 25°C, while *D. magna* were cultured in hard reconstituted water [1] at 20°C. Fathead minnow brood stock were cultured at 20 to 25°C in tap water that was pretreated with activated carbon. Eggs and larva were held in MHRW; larva were fed brine shrimp nauplii (*Artemia* sp.) twice daily until they were used in testing.

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# Test procedures

Toxicity tests followed the general guidance of the U.S. Environmental Protection Agency (USEPA) [1,16] for conducting acute whole effluent toxicity tests. All tests were conducted in 30-ml plastic beakers containing 10 ml of test solution and five organisms per chamber. Tests were conducted under a 16-h:8-h light: dark photoperiod; *C. dubia* and fathead minnows were tested at 25°C, while *D. magna* were tested at 20°C. Dilution/control water for all tests was MHRW. Exposure periods were 48 h for *C. dubia* and *D. magna* and 96 h for fathead minnows, with daily observations of mortality. The criteria for death were no visible movement and no response to prodding.

Standard guidance for acute effluent toxicity testing [1] is to withhold food during testing of daphnids, presumably because of concerns that the addition of food might alter the toxicity of the sample. However, in water devoid of food (e.g., reconstituted laboratory water), withholding food likely places some stress on the test organisms. Moreover, effluents and ambient waters, to which the results of these experiments apply, can be expected to contain bacteria, algae, and other sources of food. Hence, addition of daphnid food (yeast/cerophyl/trout chow [YCT] and algae [2]) to clean laboratory water might better simulate the characteristics of field-collected samples. To assess the potential effect of feeding on major ion toxicity, initial tests using C. dubia were conducted both with and without feeding. Analysis of these initial experiments (see Results) showed that the addition of food represented only a small influence on C. dubia survival. Because the effect of feeding was small and its inclusion was believed to provide a more representative test matrix, remaining C. dubia tests included feeding, as did all D. magna and fathead minnow tests. For daphnid tests, 100 µl of a 1:1 mix of YCT and algal suspension was added to each test chamber at test initiation. For fathead minnow tests, 100 µl of concentrated brine shrimp nauplii was added after 48 h of exposure, though solutions were not subsequently renewed as recommended by the USEPA [1].

Because toxicity testing of salt solutions was to be completed over several months, we recognized the possibility that systematic drift in test organism sensitivity could bias the results of toxicity tests conducted at different times. In an attempt to account for this potential variability, each set of toxicity tests included a reference toxicant test using NaCl. LC50 values were computed for each of these tests and were included in the statistical modeling as another independent variable. Thus, if drifts in organism sensitivity did occur and were reflected in the response to NaCl, they could be accounted for in the regression modeling.

# Chemical measurements

Concentrations of major ions were determined analytically in all stock solutions used in testing.  $Ca^{2+}$ ,  $Na^+$ ,  $Mg^{2+}$ , and  $K^+$  were determined using inductively coupled plasma emission spectroscopy (ICP) according to USEPA method 200.7 [17];  $CI^-$  and  $SO_4^{2-}$  concentrations were determined by anion chromatography [18]; and  $HCO_3^-$  concentrations were determined indirectly by the measurement of phenolphthalein alkalinity [19]. As  $HCO_3^-$  is the predominate carbonate species present in the pH range of interest (pH 6.5–9.0), alkalinity equivalents were converted directly to  $HCO_3^-$  concentration.

Dissolved oxygen (DO) and pH were measured in selected test solutions during actual toxicity testing, primarily on solutions near the threshold for acute toxicity. DO was measured with a Yellow Springs Instrument model 54 DO meter (Yellow Springs, OH, USA) while pH was measured with a Orion pH meter model SA250 (Boston, MA, USA). Measured DO concentrations were always within an acceptable range (>40% saturation) [1]. Measured pH varied according to the components of the solution but was generally between pH 7.5 and 9.0.

#### Preparation of test solutions

Test solutions were prepared by dissolving individual ion salts in MHRW. Salts used in testing were NaCl, Na<sub>2</sub>SO<sub>4</sub>, NaHCO<sub>3</sub>, KCl, K<sub>2</sub>SO<sub>4</sub>, KHCO<sub>3</sub>, CaCl<sub>2</sub>, CaSO<sub>4</sub>, MgCl<sub>2</sub>, MgSO<sub>4</sub>, CaCO<sub>3</sub>, and MgCO<sub>3</sub>; all were of reagent grade or better (Sigma Chemical Company, St. Louis, MO, USA). Stock solutions were prepared from these salts by dissolving 10,000 mg/L of a salt in MHRW. CaSO<sub>4</sub> was not fully soluble at 10,000 mg/L; for this reason, CaSO<sub>4</sub> solutions were filtered through a 1-µm glass fiber filter prior to testing and ion concentrations were measured in filtered solutions. Test solutions using CaCO<sub>3</sub> and MgCO<sub>3</sub> had pH in excess of 10 and were acidified with HCl or H<sub>2</sub>SO<sub>4</sub> until pH stabilized at approximately 8.5.

For tests evaluating only one salt (one cation and one anion), test solutions were prepared by serially diluting the 10,000-mg/L stock solutions with MHRW to develop a series of test concentrations spaced on a 0.5 × dilution factor (i.e., 10,000, 5,000, 2,500, 1,250 mg/L). For tests involving two salts, solutions were prepared by combining equal volumes of the two stock solutions, then diluting as necessary. As testing proceeded and effect thresholds were determined, test concentrations were often spaced much more closely (e.g., 2,500, 2,000, 1,500, 1,000, 500 mg/L) to better define responses near the effect threshold.

All ion concentrations measured in the stock solutions were compared to nominal values. If the measured concentrations differed from the nominal value by more than 20%, the actual measured concentrations were substituted for the nominal concentrations. Aside from CaSO<sub>4</sub>, which did not completely dissolve, substantial discrepancies between nominal and measured concentrations occurred in two instances, once for a MgCl<sub>2</sub> stock solution and once for a CaCl<sub>2</sub> stock solution. In some analyses, the measured concentrations of cations and anions (expressed as milliequivalents or meg) in a salt solution were not similar. Because charge balance is a physical/chemical requirement, such solutions were further evaluated to determine which concentration (cation or anion) was closer to the nominal value. In all cases, the cation concentration was closer to the nominal value; based on this, the anion concentration in the stock solution was changed to the concentration (in meq) of the corresponding cation.

To calculate ion concentrations in actual test solutions, the concentrations in the applicable stock solutions were multiplied by the relative proportion of each solution in the test solution. Because the dilution water (MHRW) also contained small concentrations of each ion, these background concentrations were then added to the calculated contributions from the stock solutions.

In cases where an SO<sub>4</sub><sup>2</sup> salt (e.g., Na<sub>2</sub>SO<sub>4</sub>) was combined with a Ca<sup>2+</sup> salt (e.g., CaCl<sub>2</sub>), the potential existed for supersaturation of test solutions with respect to CaSO<sub>4</sub>. This potential was confirmed by the appearance of white precipitates in some test solutions. Because precipitation would affect the dissolved ion concentrations in the test solutions, all ion com-

binations tested were checked for  $CaSO_4$  supersaturation by comparing the nominal test concentrations of  $Ca^{2+}$  and  $SO_4^{2-}$  with the solubility product for  $CaSO_4$  (226.5) calculated from measured concentrations of  $Ca^{2+}$  and  $SO_4^{2-}$  in a saturated  $CaSO_4$  solution. If a particular solution was supersaturated with respect to  $CaSO_4$ ,  $Ca^{2+}$  and  $SO_4^{2-}$  concentrations were reduced on an equimolar basis until the concentrations reached the calculated saturation point. These corrected concentrations were then used for data analyses.

#### Replication

To incorporate intertest variability into the data set, emphasis was placed on replication between batches of tests conducted through time rather than on having replicate chambers tested simultaneously. Accordingly, most ion combinations evaluated were tested on at least two and as many as five different occasions (see results). The exception was for two cation/one anion solutions tested with *D. magna* and fathead minnows, and two cation/two anion solutions tested with *C. dubia*; for these tests, duplicate chambers (10 animals total) were tested simultaneously. When calculating LC50 values, replicate tests conducted on different days were analyzed separately, but duplicate chambers tested simultaneously were combined into one analysis.

# Data collection, management, and analysis

Data generated by all toxicity tests were entered into a database using Paradox 3.1 software (Borland International, Scotts Valley, CA, USA). Regression modeling was based on individual ion concentrations rather than salt concentrations. By converting salts to ion concentrations, we were able to separate out the effects of individual cations and anions instead of the effects of cation–anion pairs. Statistical modeling of the toxicity data consisted of stepwise logistic multiple regression using the LR program within BMDP statistical software [20].

Logistic regression relates binary observations (e.g., alive or dead) to one or more independent variables (in this case, ion concentrations). The completed regression predicts a probability of survival based on concentrations of ions showing relationships to survival. The linear logistic regression model used is of the form

logit(P) = ln[P/(1 - P)]  
= 
$$\beta_0 + \beta_1 X_1 + \beta_2 X_2 + ... + \beta_n X_n$$
 (1)

where P = proportion surviving,  $\beta =$  regression coefficient, X = ion concentration, and n = total number of significant terms in the model.

During development of the final models, various data transformations (e.g., log) and independent variable interactions (e.g.,  $Cl \times SO_4$  interaction) were considered. Each potential model was evaluated using the following criteria: (1) each independent variable in the model must significantly improve the fit of the model to the data ( $\alpha = 0.05$ ); (2) the model should maximize  $R^2$  (maximize the amount of variance in the data that is explained by the model) and minimize the number of independent variables; and (3) the model should provide reasonable predictions even when extrapolating outside the limits of the data used to generate the model.

Data collection and model development were iterative processes in which a series of statistical models (regressions) were developed followed by supplemental data collection. To begin,

data were generated for single ion pairs or salts (e.g., Na2SO4, CaCl<sub>2</sub>). Based on these data, an initial regression equation was developed  $(F_1)$ . Next, additional toxicity data were generated using combinations of two cations and one anion (e.g., Na+, Ca2+, and SO2-) and one cation and two anions (e.g., Na+,  $Cl^-$ , and  $SO_4^{2-}$ ). The  $F_1$  equation was then used to predict survival for these additional data. In addition, a second regression equation  $(F_2)$  was then developed using all data generated to date. The predictive abilities of both models were then compared by examining the relationship between predicted and observed survival for all of the ion combinations tested. If  $F_2$  had notably better predictive ability than  $F_1$ , we concluded that important relationships in the data were not accounted for in the  $F_1$  equation. The process was repeated by testing more complex ion solutions and developing additional regression equations, until the incorporation of additional data did not substantially alter the basic equation. This iterative process of data generation, model development, and additional data generation continued throughout model development.

As part of this iterative process, characteristics of specific points that had poor correlation between predicted and observed survival were considered. In some cases, it was found that such data points had poor agreement between replicate tests of the same ion combination, hence it was impossible for the regression equation to fit both responses. In these instances, additional toxicity tests were conducted using that particular combination of ions to better characterize the response. Of 2,904 total data points, 59 were discarded as spurious; of these, 46 were for C. dubia, 5 for D. magna, and 8 for fathead minnows. Thirty-eight of the 59 discarded points were cases where mortality (typically one or two dead out of five organisms) was observed two or more concentrations below the primary concentration response, suggesting that ion toxicity may not have been the cause of mortality. Though these points may represent innate variability in the survival of test organisms, our intent was to represent mortality due to ion stress; random mortalities at low ion concentrations tended to decrease the slope of the regression model and obscure the response threshold. Of the remaining discarded points, 10 were discarded because the CaSO<sub>4</sub> solution was not filtered prior to testing (C. dubia); 10 were from a K2SO4 dilution series in which there was erratic and substantial mortality without evidence of a concentration response (C. dubia); and one was from a test chamber that was spilled after the 24-h observation (P. promelas).

In other cases, it was found that outlier points tended to share certain characteristics. For example, it was noted that for *C. dubia*, early regressions showed poor predictive ability for ion combinations containing Cl<sup>-</sup> opposed by two cations (e.g., Na<sup>+</sup> and Ca<sup>2+</sup> with Cl<sup>-</sup>); these solutions showed lower toxicity than those with just one Cl<sup>-</sup> salt (e.g., NaCl). Further testing with these ion combinations showed that this response was reproducible. To account for this phenomenon, a new variable called NumCat was created. The value of NumCat is equal to the number of cations representing at least 10% of the total molar concentration of cations and present at greater than 100 mg/L. The development and implications of the NumCat variable are discussed in detail in the Results.

In addition to the more rigorous statistical modeling described above, LC50 concentrations were also calculated using a computer program following the trimmed Spearman–Karber method [21]. Independent LC50 values were calculated for each unique (i.e., nonsimultaneous) test of ion toxicity. For

Table 1. Number of ion solutions tested for toxicity<sup>a</sup>

		Numl	per of ca	ıtions/an	ions <sup>b</sup>		_	Reference toxicant and				
Species	1/1	1/2	2/1	2/2	3/1	4/1	Subtotal	controls	Total			
Ceriodaphnia dubia	464	449	438	401	108	20	1,887	232	2,119			
Daphnia magna	354	147	65	0	0	0	566	122	688			
Fathead minnows	242	142	59	0	0	0	451	56	499			

a Replicate analyses counted separately.

ion combinations that were tested repeatedly, average LC50s were calculated as the arithmetic mean of the values. In some cases, tests did not capture the effect threshold and an LC50 could only be expressed as a range (e.g., LC50 < 625 mg/L). Where this range did not conflict with the other calculated values, the indefinite value was dropped and the mean was calculated from the remaining values (e.g., 500, 700, and <625 would average to 600 with n=2). If the indefinite value represented an extreme value, the mean was calculated as an inequality relative to the mean of the numerical values (e.g., 775, 700, and <625 would average to <700 with n=3).

#### RESULTS AND DISCUSSION

In total, survival data were collected for 2,904 ion solutions, excluding reference toxicant tests and controls (Table 1). Data collection and modeling were conducted first for *C. dubia*, and the resulting data set encompasses both greater replication and a greater variety of ion combinations. The full data sets are too extensive to provide here but are provided in print in Mount and Gulley [22].

To present the data in a more condensed form, LC50 values were calculated for all ion solutions tested (Tables 2 and 3). Coefficients of variation for LC50 values for individual ion combinations were typical for acute toxicity tests [1], with means of 17% for *C. dubia* (SD = 14; range 0.0–61), 17% for *D. magna* (SD = 7.5; range 4.8–31), and 24% for fathead minnows (SD = 15; range 1.4–62).

The effect of feeding on the response of *C. dubia* was assessed during the first three sets of tests conducted. In each of these, toxicity of each single salt solution was tested both with and without the addition of food. Average LC50 values for tests with and without feeding were similar (Fig. 1), although there was a tendency for tests without feeding to have slightly lower LC50 values. Logistic regression modeling of these data confirmed this trend; feeding was judged a significant variable by the regression algorithm, with a positive coefficient indicating that feeding did increase overall survival. However, the influence of feeding in the model was quite small, explaining less than 1% of the overall variance. Because we believed that the addition of food might provide a more natural test matrix, all remaining tests were conducted with feeding.

To determine whether the results of reference toxicant tests related to the responses observed in the concurrent exposures to ion combinations, LC50 values were calculated for the reference toxicant tests from the first 11 test groups with *C. dubia* (total of 1,045 ion solutions tested). During this period, 48-h LC50 values for NaCl averaged 1,042 mg/L as Cl<sup>-</sup> with a coefficient of variation equal to 24%. The LC50 value from the concurrent reference toxicant test was included as an independent variable for each ion solution and thus considered by the stepwise logistic regression. In this analysis, the ref-

erence toxicant variable was not selected as being statistically significant, explaining only 0.12% of the overall variance. From this, we surmised that there was no consistent relationship between the sensitivity of the test organisms (as measured by the reference toxicant test) and the responses of organisms in the concurrent ion exposures. For this reason, the reference toxicant test results were not considered further in subsequent regressions.

As described previously, the development of the final predictive models was an iterative process in which a series of regression models was developed. Initial regressions were developed based on more limited data sets (e.g., results from toxicity tests using single salts only); as data collection proceeded to more complicated solutions (enrichment with three and four ions), these equations were refined. Throughout the project, 74 distinct models were developed and considered. The majority of these models were discarded, either because they were superseded by later models that incorporated larger data sets, or were found to have undesirable characteristics (e.g., poor predictive ability). Several of these analyses involved experimentation with alternative variables or data transformations. To illustrate the model development process, we selected three intermediate models that demonstrate major advances in the model development, including the creation of a new variable, referred to as NumCat. The three example models are referred to as the single salt, double salt, and double salt with NumCat models and are based on 48-h survival data for C. dubia.

The single salt model was developed relatively early in the data collection process using 362 data points involving single salt solutions only (i.e., enriched with one cation and one anion; Fig. 2). This regression equation fit the observed survival values very well, with an  $R^2$  value of 0.950. Significant variables in this equation were the concentrations of  $K^+$ ,  $Mg^{2+}$ ,  $HCO_3^-$ ,  $Cl^-$ , and  $SO_4^{2-}$ ;  $Na^+$  and  $Ca^{2+}$  were not significant variables indicating that the toxicity of  $Na^+$  and  $Ca^{2+}$  salts could be accounted for primarily by the toxicity of the co-occurring anion. No first-order interaction terms (e.g.,  $K \times Cl$ ) were selected as significant.

Data collection was then expanded to include solutions with one cation and two anions and two cations and one anion. When the single salt model was used to predict survival for this expanded data set (1,045 data points) it showed considerably less predictive ability than it had for the smaller initial data set. Accordingly, a new model was developed using data from all test solutions. This double salt model had the same significant variables as did the single salt model but did a better job of predicting survival for the entire data set than did the single salt model. Although it did have better predictive ability for the combined data set, the  $R^2$  value of 0.837 indi-

<sup>&</sup>lt;sup>b</sup> Number of ions enriched above background concentrations.

Table 2. Mean 24-h (upper right) and 48-h (lower left) LC50 values for salt combinations tested with Ceriodaphnia\*

	) <sub>4</sub> 24-h	[1] NaCl	[3] $Na_2SO_4$ 820)	[1] NaHCO <sub>3</sub>	[2] KCI 260)	[4] K <sub>2</sub> SO4 770)	[1] KHCO <sub>3</sub>	[2] CaCl <sub>2</sub> 3,700)	[3] $CaSO_4$ 5,610)	[3] MgCl <sub>2</sub>	3] MgSO <sub>4</sub>			3]	
	${ m MgSO_4}$	] 3,400 [1]	] 3,480 [3] 0) (3,080–3,820)		] 1,070 [2] 0) (880–1,260)	1,510 [4] (1,340–1,770)	] 940 [1]	) >3,690 [2] 0) (3,670->3,700)	>5,610 [3] (>2,610->5,610)	] 1,560 [3] 0) (1,360–1,770)	1,770[3] (1,770–1,770)			$ \begin{array}{ccc} & 1,770 [3] \\ (1,770-1,770) \end{array} $	${ m MgSO_4}$
eriodapinia"	$MgCl_2$	3,230 [4] (3,080–3,460)	3,100 [2] 0) (2,750–3,460)	1,800 [1]	$\begin{array}{c} 1,400  [2] \\ (1,030-1,770) \end{array}$	>1,550 [1]	860 [1]	3,500 [3] ) (3,420–3,540)	>2,760 [1]	1,270[3] $(880-1,770)$		(6	(880–880)	1,490 [3] (1,360–1,560)	$MgCl_2$
Table 2: Mical 24-11 (upper right) and 46-11 (rower reft) LCOO values for collibriations tested with Certoadprinta	CaSO <sub>4</sub>	>2,430 [1]	>4,940 [2] (4,170->5,700)	>1,040 [1]	1,580 [1]	1,140 [3] 480–1,870	1,560 [1]	3,880 [2] (3,660–4,100)	>1,940 [4] (>1,940->1,990)			>1,910 [4] (1,910->1,970)	<2,370 [1]	>5,610 [2] (>5,610->5,610)	CaSO <sub>4</sub>
ioi sait comoniat	CaCl <sub>2</sub>	3,340 [3] (2,960–3,540)	4,120 [2] (3,800–4,150)	2,680 [2] 0) (2,320–3,080)	1,740 [3] (1,690–1,770)	2,250 [1]	1,910[1]	2,260 [3] (1,770–2,680)			1,830 [4] (1,770–2,030)	3,050 [2] (2,450–3,660)	2,600 [3] (2,430–2,680)	>3,690 [2] (3,670->3,700)	CaCl <sub>2</sub>
COO values	KHCO <sub>3</sub>	1,360 [1]	1,300 [1]	920 [3] (880–1,000)	550 [3] (290–770)	390 [3] (290–440)	630 [2] (580–670)			630 [2] (580–670)	1,810[1]	1,560 [1]	860 [1]	940 [1]	KHCO <sub>3</sub>
יס-וו (וסשכו וכוו) ד	$K_2SO_4$	>1,800 [1]	1,390 [2] (1,020–1,770)	1,110 [1]	620 [3] (250–880)	770 [3] (770–780)			<680 [3] (<620-710)	390 [3] (290–440)	1,820 [1]	1,130 [3] (480–1,830)	1,040 [1]	1,480 [4] (1,340–1,770)	$K_2SO_4$
uppei rigili) alla 4	KCI	1,650 [2] ) (1,540–1,770)	1,730 [1]	1,200[1]	630 [3] (580–630)			630 [3] (580–670)	480 [3] (250–670)	480 [3] (290–580)	1,730[3] $(1,640-1,770)$	1,580 [1]	1,270[3] $(1,000-1,770)$	1,060 [2] (880–1,220)	KCl
C 2. IVICAII 24-II (I	NaHCO <sub>3</sub>	2,200 [4] (1,770–2,680)	2,800 [5] (2,220–3,540)	$\begin{array}{c} 1,420  [4] \\ (1,240-1,770) \end{array}$			1,020 [4] (880–1,170)	1,140 [1]	<1,000 [1]	800 [3] (580–950)	<2,640 [2] (<2,250–3,030)	>1,040 [1]	1,510[1]	1,670 [1]	NaHCO <sub>3</sub>
1401	Na <sub>2</sub> SO <sub>4</sub>	3,320 [4] (3,110–3,540)	3,590 [4] (3,540–3,740)			3,080 [4] (1,770–3,540)	2,630 [4] (1,880–3,540)	1,730 [1]	1,590 [3] (1,020–2,000)	1,300 [1]	>3,940 [2] (3,800->4,080)	>4,940 [2] (4,170->5,700)	<2,520 [2] (<2,320–2,720)	3,190 [3] (2,680–3,540)	Na <sub>2</sub> SO <sub>4</sub>
	NaCl	3,380 [3] (3,080–3,540)			1,960 [3] (1,770–2,330)	3,070 [4] (2,530–3,540)	1,890 [3] (1,770–2,030)	1,560[3] $(1,540-1,600)$	1,660 [1]	1,360 [1]	3,030 [4] (2,240–3,540)	>2,430 [1]	2,380 [4] (1,770–2,730) (	3,250 [1]	NaCl
					NaCl	$\mathrm{Na_2SO_4}$	NaHCO <sub>3</sub>	KCl	$K_2SO_4$	KHCO,	$CaCl_2$	CaSO <sub>4</sub>	MgCl <sub>2</sub>	MgSO <sub>4</sub>	48-h

<sup>a</sup> Values are arithmetic means [n] (range) expressed as total ion concentrations added in mg/L. Tests with two salts involved 1:1 combinations of stock solutions containing 10,000 mg/L, except CaSO<sub>4</sub> (1,970 mg/L).

Table 3. Mean LC50 values for salt combinations tested with Daphnia magna and fathead minnows<sup>a</sup>

	Daphnic	a magna		Fathead minnow	
Salt	24-h	48-h	24-h	48-h	96-h
NaCl	6,380 [2]	4,770 [2]	8,280 [3]	6,510 [3]	6,390 [3]
	(6,160–6,600)	(3,790–5,740)	(7,240–10,000)	(6,090–7,070)	(6,020–7,070)
Na <sub>2</sub> SO <sub>4</sub>	6,290 [4]	4,580 [4]	>8,080 [3]	>7,960 [3]	7,960 [3]
	(5,790–7,070)	(4,060-5,360)	(7,070->10,000)	(6,800->10,000)	(6,800–10,000)
NaHCO <sub>3</sub>	2,380 [4]	1,640 [4]	4,850 [2]	2,500 [2]	<850 [3]
	(1,900–2,870)	(1,170–2,030)	(3,540–6,160)	(950–4,060)	(<310-1,220)
KCl	740 [5]	660 [5]	950 [3]	910 [3]	880 [3]
	(580–880)	(440–880)	(750–1,090)	(750–1,090)	(750–1,020)
K <sub>2</sub> SO <sub>4</sub>	850 [4]	720 [4]	990 [4]	860 [4]	680 [4]
	(670–1,170)	(580–880)	(770–1,170)	(580–1,170)	(510–880)
KHCO <sub>3</sub>	670 [4]	650 [4]	940 [4]	820 [4]	<510 [4]
	(440–880)	(380–820)	(750–1,340)	(750–880)	(<310-750)
CaCl <sub>2</sub>	3,250 [4]	2,770 [4]	>6,660 [3]	>6,560 [3]	4,630 [3]
	(2,680–4,010)	(2,330–3,230)	(4,700->10,000)	(4,390->10,000)	(3,930–5,360)
CaSO <sub>4</sub>	>1,970 [3]	>1,970 [3]	>1,970 [2]	>1,970 [2]	>1,970 [2]
	(>1,970->1,970)	(>1,970->1,970)	(>1,970->1,970)	(>1,970->1,970)	(>1,970->1,970)
$MgCl_2$	1,560 [4]	1,330 [4]	3,520 [3]	2,840 [3]	2,120 [3]
	(1,250–1,810)	(1,170–1,580)	(2,520–4,490)	(1,970–3,880)	(1,580-2,740)
$MgSO_4$	2,360 [4]	1,820 [4]	4,630 [3]	3,510 [3]	2,820 [3]
	(2,180–2,500)	(1,540–2,330)	(3,180-7,070)	(3,000-4,350)	(2,610-3,080)
NaCl/Na <sub>2</sub> SO <sub>4</sub>	6,140 [2] (5,360–6,930)	5,700 [2] (5,360–6,030)	>9,040 [2] (8,080->10,000)	>8,460 [2] (6,930->10,000)	6,090 [2] (6,030–6,160)
NaCl/NaHCO <sub>3</sub>	4,440 [2]	2,950 [2]	4,580 [2]	3,790 [2]	2,540 [2]
	(3,520–5,360)	(2,830–3,080)	(3,540–5,630)	(2,330–5,250)	(2,330–2,750)
Na <sub>2</sub> SO <sub>4</sub> /NaHCO <sub>3</sub>	4,480 [2]	3,180 [2]	5,350 [2]	5,050 [2]	4,060 [2]
	(4,060–4,900)	(2,830–3,540)	(4,660–6,030)	(4,060-6,030)	(3,080–5,040)
KCl/K <sub>2</sub> SO <sub>4</sub>	740 [2]	740 [2]	900 [2]	760 [2]	760 [2]
	(600-880)	(600–880)	(790–1,020)	(630–880)	(630–880)
KCl/KHCO5 <sub>3</sub>	740 [2]	740 [2]	800 [2]	770 [2]	770 [2]
	(640–830)	(640–830)	(770–830)	(700–830)	(700–830)
K <sub>2</sub> SO <sub>4</sub> /KHCO <sub>3</sub>	630 [2]	630 [2]	1,060 [2]	720 [2]	720 [2]
	(540–720)	(540–720)	(1,030–1,090)	(610–830)	(610–830)
CaCl <sub>2</sub> /CaSO <sub>4</sub>	3,250 [2] (3,140–3,360)	2,950 [2] (2,760–3,150)	>5,510[1]	>5,510 [1]	>5,510 [1]
$MgCl_{2}/MgSO_{4}$	2,110 [2]	1,510 [2]	3,830 [2]	3,330 [2]	2,800 [2]
	(1,940–2,280)	(1,340–1,680)	(3,790–3,870)	(3,300-3,370)	(2,240–3,370)
NaCl/KCl	3,930 [1]	3,930 [1]	1,410[1]	1,410[1]	1,410 [1]
NaCl/CaCl <sub>2</sub>	5,250 [1]	5,250 [1]	8,410 [1]	8,080[1]	6,460 [1]
NaCl/MgCl <sub>2</sub>	3,820 [1]	3,070 [1]	5,250 [1]	3,520 [1]	3,160 [1]
KCl/CaCl <sub>2</sub>	2,620 [1]	2,450 [1]	2,810 [1]	2,810[1]	2,810 [1]
KCl/MgCl <sub>2</sub>	2,280 [1]	2,020 [1]	1,580 [1]	1,410 [1]	1,410 [1]
CaCl <sub>2</sub> /MgCl <sub>2</sub>	4,850 [1]	4,390 [1]	5,630 [1]	5,250 [1]	5,250 [1]
Na <sub>2</sub> SO <sub>4</sub> /K <sub>2</sub> SO <sub>4</sub>	4,800 [1]	4,610 [1]	1,580 [1]	1,580 [1]	1,580 [1]
Na <sub>2</sub> SO <sub>4</sub> /MgSO <sub>4</sub>	8,400 [1]	7,980 [1]	8,840 [1]	5,740 [1]	4,800 [1]
K <sub>2</sub> SO <sub>4</sub> /CaSO <sub>4</sub>	1,160 [1]	1,200 [1]	1,980 [1]	1,720 [1]	1,720 [1]
K <sub>2</sub> SO <sub>4</sub> /MgSO <sub>4</sub>	2,760 [1]	2,210 [1]	1,380 [1]	1,290 [1]	1,290 [1]
CaSO <sub>4</sub> /MgSO <sub>4</sub>	>6,470 [1]	>6,470 [1]	NT <sup>b</sup>	NT	NT
NaHCO <sub>3</sub> /KHCO <sub>3</sub>	1,220 [1]	1,040 [1]	1,140 [1]	820 [1]	740 [1]

<sup>&</sup>lt;sup>a</sup> Values are arithmetic means [n] (range) expressed as total ion concentrations added in mg/L. Tests with two salts involved 1:1 combinations of stock solutions containing 10,000 mg/L, except for CaSO<sub>4</sub> (1,970 mg/L), MgCl<sub>2</sub> (5,480 mg/L), and CaCl<sub>2</sub> (7,480 mg/L).

<sup>b</sup> Not tested.

cated a lower quality of fit than was observed for the single salt model fit to the initial, less complex data set.

There were two basic explanations for the decreased quality of fit observed with the double salt model: (1) the larger data set contained greater inherent variability (measurement error) and hence it was not possible to achieve as high an  $R^2$  value; or (2) there were important toxic interactions represented in the three ion solutions that were not represented in the solutions containing only a single salt (although the regression

algorithm had not selected any interaction terms as being significant). When the ion combinations for which the model made poor predictions were analyzed, some patterns were apparent. In particular, it appeared that the model was overpredicting toxicity for solutions containing two Cl<sup>-</sup> salts.

This phenomenon is perhaps best illustrated by data collected for solutions of NaCl and  $CaCl_2$  tested both alone and in combination. As explained above, the single salt model indicated that the toxicity of Na<sup>+</sup> and Ca<sup>2+</sup> salts could be

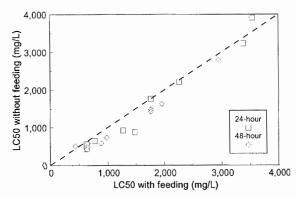


Fig. 1. Average LC50 values for *Ceriodaphnia dubia* exposed to single salts with and without feeding.

adequately explained on the basis of the anion concentration alone; in other words, NaCl and CaCl<sub>2</sub> had approximately the same toxicity when expressed on the basis of Cl<sup>-</sup>. A plot of these data (Fig. 2) supports this conclusion and also shows a good fit of the single salt model to these data. However, when NaCl and CaCl<sub>2</sub> were tested in combination, the resulting solution was less toxic (on the basis of Cl<sup>-</sup> concentration) than either of the solutions tested singly. The single salt model was unable to account for this decreased toxicity and, consequently, made poor predictions for the combined NaCl/CaCl<sub>2</sub> solutions (Fig. 2). The same trend toward lower toxicity of Cl<sup>-</sup> in the presence of two cations was also evident for solutions containing K<sup>+</sup> or Mg<sup>2+</sup>.

The double salt model compensated for the lower toxicity of two cation solutions but only partially. The double salt model simply fit a shallow response curve between the single cation and two cation data, predicting a "mean" probability of survival somewhere between the observed single salt and two salt survival values. While this compromise provided a better overall fit to the data than did the single salt model, it was clearly not a good representation of the response. Given that the regression algorithm did not find any interaction terms to be significant, it appeared that a new variable was required to provide a better fit to the data.

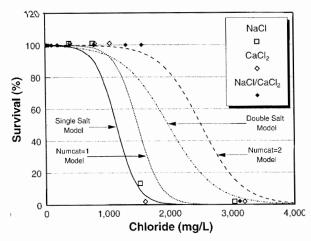


Fig. 2. The 48-h survival of *Ceriodaphnia dubia* exposed to solutions enriched with NaCl, CaCl<sub>2</sub>, or a 1:1 combination of NaCl and CaCl<sub>2</sub>, normalized to Cl<sup>-</sup> concentration. Curves represent regression model predictions for the single salt, double salt, and double salt with NumCat models. Values at 0% and 100% offset slightly for clarity.

We attempted without success to derive a continuous variable that would respond appropriately to the relative concentration of cations in solution and thus identify the two cation solutions as different than solutions with a single cation. After our lack of success with continuous variables, we created a categorical variable called NumCat. The NumCat variable was intended to simply represent the number of major cations in the solution. For the initial modeling trials, the NumCat variable was arbitrarily defined as the number of cations in the solution that represented at least 10% of the total molar cation concentration and that were also present at a concentration greater than 100 mg/L. Our expectation was that the NumCat variable would show a significant interaction with Cl- and any other ion whose toxicity was influenced by the number of cations present. The resulting model, called the "double salt with NumCat" model, showed a markedly improved fit  $(R^2 =$ 0.899); significant terms were the original five ions in the single and double salt models, plus NumCat and the Num-Cat  $\times$  Cl, NumCat  $\times$  SO<sub>4</sub>, and NumCat  $\times$  K interaction terms. The NumCat × Cl term allowed the model to better represent the toxicity of NaCl, CaCl<sub>2</sub>, and NaCl + CaCl<sub>2</sub> solutions shown in Figure 2. NumCat also showed significant (positive) interactions with SO<sub>4</sub><sup>2-</sup> and K<sup>+</sup>, suggesting that the presence of two cations (or one additional cation in the case of K+) ameliorated the toxicity of these ions as well.

After subsequent data collection and analysis, two additional steps were taken to optimize the NumCat variable. First, we conducted supplemental testing of *C. dubia* exposed to mixtures of three and four Cl<sup>-</sup> salts (data not shown). Modeling of these data (NumCat = 3 or 4) yielded a substantial underprediction of toxicity. Direct inspection of these data confirmed that the protective effect observed with two cations did not seem to increase with the addition of three or four cations. Accordingly, we chose to limit the NumCat variable to values of 0, 1, or 2; for solutions where the >10% and >100-mg/L criterion yielded values of 3 or 4, these values were reset to 2.

The second step involved rigorously evaluating the definition criteria for the NumCat variable. Although the NumCat variable was clearly effective at increasing the predictive capability of the model, its original definition had been arbitrary. To provide a stronger technical basis for defining NumCat, we conducted a sensitivity analysis by varying the two components of the NumCat definition, the relative molar concentration (originally >10%), and the absolute concentration (originally 100 mg/L). A complete matrix of relative concentration (0, 5, 10, 15, 20, and 25%) and absolute concentration (0, 100, 200, and 300 mg/L) was modeled using 48-h C. dubia data. The resultant models were evaluated based on their  $R^2$  values (Fig. 3). The NumCat criteria that produced the model with the highest  $R^2$  (best fit of the model to the observed data) were the 15% with >100 mg/L ( $R^2 = 0.8559$ ) and the 10% with >100-mg/L ( $R^2 = 0.8553$ ) criteria. Given that the difference in  $R^2$  was only 0.0006 (0.06% of the variance) and that we had already worked extensively with the 10% and >100-mg/L criteria, we elected to continue using these criteria in finalizing the model equations.

After completion of data collection, final regression equations were developed to predict *C. dubia* survival after 24 and 48 h of exposure. Through the course of these analyses, several additional variables and data transformations were evaluated and discarded. Aside from the feeding and reference toxicant variables discussed previously, we evaluated the sum of all ions, the sum of all cations, the sum of all anions, and NumAn

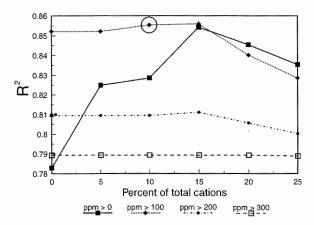


Fig. 3. Effect of varying criteria for the definition of the NumCat variable. Circled point represents the criteria selected initially and maintained for final derivation of the regression equations.

(the anion equivalent of NumCat). First-order interactions of these variables and ion concentrations were also evaluated. None of these variables was selected as significant by the regression algorithm. Models based on log-transformed ion concentrations consistently showed lower  $R^2$  values than those based on untransformed data.

The final 24- and 48-h equations for C. dubia had  $K^+$ ,  $HCO_3^-$ ,  $Mg^{2+}$ ,  $Cl^-$ , and  $SO_4^{2-}$  as significant variables (Table 4). Additionally, NumCat and the interaction terms NumCat  $\times$  Cl, NumCat  $\times$  SO<sub>4</sub>, and NumCat  $\times$  K were found to be significant. As had been the case since early in the modeling process, Na<sup>+</sup> and Ca<sup>2+</sup> concentrations were not significant variables except as they affected the calculation of NumCat.  $R^2$  for the final regressions were 0.861 and 0.842 for the 24-h and 48-h equations.

Model development for D. magna proceeded along the same lines as those described for C. dubia. The initial model developed using only single salt data fit those data very well  $(R^2=0.97)$  but was not as good at predicting survival for more complex ion mixtures. As was observed for C. dubia, solutions with multiple cations tended to be less toxic than comparable solutions with only one cation. As a result, when all D. magna data were analyzed, NumCat was again selected as a significant variable, both by itself and through its interactions with  $Cl^-$ ,  $SO_4^{2-}$ , and  $K^+$  (Table 4). In fact, all significant terms in the C. dubia double salt model with NumCat were

also significant for *D. magna*. Quality of fit for the *D. magna* models was slightly lower than for the *C. dubia* models, though still quite good (0.812 and 0.799).

As for the daphnids, modeling of the fathead minnow data indicated that toxicity was a function of  $K^+$ ,  $Mg^{2+}$ ,  $HCO_3^-$ ,  $Cl^-$ , and  $SO_4^{2-}$  concentrations, as neither  $Na^+$  nor  $Ca^{2+}$  were selected as significant variables (Table 4). The primary difference in the fathead minnow equations was that NumCat was not a significant variable either by itself or in interaction with other terms.  $R^2$  values for the three regression equations were generally comparable to those for the other models, ranging from 0.767 to 0.832.

Because of the large number of independent variables, the actual response surface of the regression models cannot be easily visualized. Nonetheless, marginal plots of the regression equations can be used to illustrate the relative sensitivity of each species to the various ions (Fig. 4). These plots show that *C. dubia* are, in general, the most sensitive of the three species to major ion toxicity, while fathead minnows are the least sensitive. K<sup>+</sup> was the most toxic ion to all species and SO<sub>4</sub><sup>2-</sup> the least. The only inconsistency between species was that Mg<sup>2+</sup> was more toxic than HCO<sub>3</sub><sup>-</sup> for *D. magna* and fathead minnows, but the reverse was true for *C. dubia*.

As a means to visually evaluate the fit of the data sets to the regression equations, each regression equation was used to predict the ion concentrations producing 50% survival for each of the ion combinations tested during data collection. These values were then plotted against the average observed LC50 values from Tables 2 and 3 (Fig. 5). These plots indicate good overall agreement between the calculated and predicted LC50 values for all three species. Note, however, that this analysis is not a direct evaluation of quality of fit for the models because it actually compares a point estimate derived from individual logistic regression equations with the arithmetic mean of multiple point estimates for specific ion combinations derived by a different method (trimmed Spearman-Karber LC50 estimation [21]); it is not a plot of raw data versus model predictions. There are other biases in this comparison as well, such as different weighting of observations. Nevertheless, the concordance between the two methods does provide some assurance that the single multiple regression models provide a reasonable representation of the responses to a broad range of ion combinations.

The absence of interaction terms in the final regression equations, aside from those involving NumCat, suggests that

Table 4. Regression coefficients for final regression equations<sup>a</sup>

	Ceriodaph	nia dubia	Daphnia	magna		Fathead minnow		
	24-h	48-h	48-h         24-h         48-h         24-h           8.83         5.91         5.83         5.69           -0.0299         -0.0200         -0.0185         -0.0108           -0.00668         -0.00450         -0.00510         -0.00225           -0.00813         -0.00330         -0.00395         -0.00117           -0.00439         -0.00204         -0.00255         -0.000728           -0.00775         -0.00276         -0.00397         -0.00200           -0.446         -0.410         -0.511         NS <sup>b</sup> 0.00870         0.00778         0.00677         NS           0.00248         0.00110         0.00146         NS	48-h	96-h			
Constant	9.11	8.83	5.91	5.83	5.69	5.51	4.70	
K+	-0.0320	-0.0299	-0.0200	-0.0185	-0.0108	-0.0113	-0.00987	
$Mg^{2+}$	-0.00594	-0.00668	-0.00450	-0.00510	-0.00225	-0.00316	-0.00327	
Cl-	-0.00706	-0.00813	-0.00330	-0.00395	-0.00117	-0.00125	-0.00120	
$SO_4^{2-}$	-0.00424	-0.00439	-0.00204	-0.00255	-0.000728	-0.000750	-0.000750	
HCO <sub>3</sub>	-0.00745	-0.00775	-0.00276	-0.00397	-0.00200	-0.00274	-0.00443	
NumCat	0.0332	-0.446	-0.410	-0.511	$NS^b$	NS	NS	
NumCat*K+	0.00888	0.00870	0.00778	0.00677	NS	NS	NS	
NumCat*Cl-	0.00196	0.00248	0.00110	0.00146	NS	NS	NS	
NumCat*SO <sub>4</sub> -	0.00121	0.00140	0.000998	0.00132	NS	NS	NS	
Model R2	0.861	0.842	0.812	0.799	0.832	0.828	0.767	

<sup>&</sup>lt;sup>a</sup> Units for ion variables are mg/L.

b NS indicates that this particular variable was not significant and was excluded from the model.

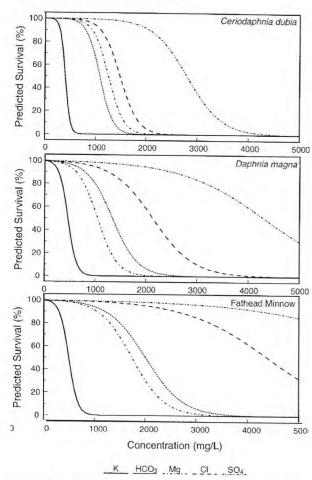


Fig. 4. Marginal plots of regression equations for each of the ions selected as significant. For  $Ceriodaphnia\ dubia$  and  $Daphnia\ magna\ models$ , NumCat=1.

assuming additivity among individual ion toxicities is sufficient to describe the toxicities of the ion mixtures, at least from an empirical standpoint. The apparent amelioration of  $CI^-$ ,  $SO_4^{2^-}$ , and  $K^+$  toxicity by multiple cations could be construed as less than additivity. Alternatively, given that  $Na^+$  and  $Ca^{2^+}$  were not clearly identified as toxic by themselves, it might be more appropriate to consider those ions as water quality variables influencing toxicity, rather than as components of a toxic mixture.

We had little precognition of the important role that the NumCat variable would play in representing the combined toxicity of major ions. In a study of high-TDS irrigation return waters, Dwyer et al. [14] demonstrated that increasing the hardness (Mg2+ and Ca2+) of an NaCl-dominated water decreased toxicity to D. magna and striped bass. For D. magna, this decreased toxicity would be predicted based on the current research, as the addition of hardness to these waters would have increased the value of the NumCat variable, thereby increasing predicted survival. However, results of our study also show that the effect of multiple cations is not an effect of hardness per se. For example, the C. dubia 48-h LC50 values for NaCl and CaCl2 were almost identical when expressed on the basis of Cl- concentration (1,187 and 1,172 mg/L, respectively; Table 2), even though the solutions had greatly different hardness. Moreover, the addition of NaCl to KCl increased the K+ concentration at the C. dubia 48-h LC50

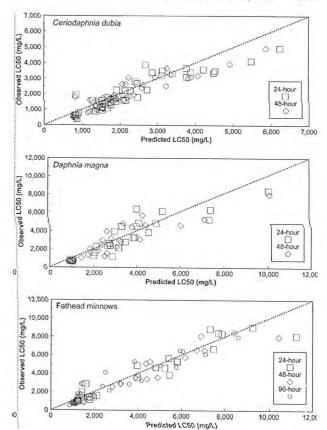


Fig. 5. Relationship between the ion concentrations predicted to cause 50% mortality and the average of LC50 values for individual salts and salt combinations (Tables 2 and 3). Line of unity (slope = 1) added for reference.

from 329 mg K/L for KCl to 458 mg K/L for an NaCl + KCl mix (Table 2), even though hardness was the same in both solutions.

Despite its importance in modeling the response of *C. dubia* and *D. magna*, the NumCat variable was not selected as significant for fathead minnows. Given that the addition of Mg<sup>2+</sup>/Ca<sup>2+</sup> improved survival of striped bass in high TDS solutions tested by Dwyer et al. [14], it seems that the protective effect in multiple cations is not restricted to cladocerans. It is worth noting that combinations of two cations and one anion were only tested once (in duplicate) for fathead minnows. If by chance those test results had a systematic bias, it might mask the presence of a cation-related effect for fathead minnows; coefficients of variation for fathead minnow LC50s were higher than for the other two species. More testing would be required to confirm or deny this possibility.

Though the effect of multiple cations is quite consistent both within the *C. dubia* and *D. magna* data sets and with other research [14], it must be emphasized that its identification and quantification through our modeling is empirical. Notwithstanding the effectiveness of the categorical NumCat variable in modeling our data set, it seems reasonable from a physiological standpoint to assume that the effect is in reality some type of continuous function, rather than the step function represented by our >10% and >100-mg/L criteria. In our modeling efforts, we were unable to devise a continuous variable that corresponded to the observed influence of multiple cations. Nonetheless, with continued research it seems likely that

such a relationship could be uncovered and, if so, might provide a more rigorous representation of the actual relationship than that provided by NumCat as currently defined. A better understanding of the mechanisms of major ion toxicity would likely enhance this effort.

As a related matter, even though we conducted a sensitivity analysis to determine the optimum criteria for the NumCat variable, this analysis was subject to bias from the structure of our data set. Specifically, we tested binary combinations of salt solutions in 1:1 ratios only. As such, only certain areas within the total sampling space (all possible ion combinations) were represented in the data set. Thus, there is no assurance that the ion combinations tested were near critical points in the response surface that might alter the apparent thresholds for response. While we believe the NumCat variable is a significant advance in understanding the response of cladocerans to high TDS solutions, it is probably a somewhat crude representation of the actual physiological response.

Because most chemical reactions are related to molar concentrations, an argument could be made for modeling survival on the basis of molar concentrations rather than mass-based concentration. In retrospect, it seems this would have made little difference in the outcome of the modeling. As the equations are based on first-order concentrations of single ions only, transformation between mass-based and molar concentrations is a simple algebraic manipulation and does not affect the nature of the response surface. In fact, the equations in Table 4 can be converted to a molar basis by simply dividing each coefficient by the molecular weight of each ion. Conversion to chemical activity, however, would be much more involved.

Ultimately, the test of the toxicity models we have generated lies in their ability to make accurate predictions for samples outside those used to generate the original data set. Thus far, the equations have performed well in predicting major ion toxicity in field-collected samples, particularly so for the C. dubia equations. For example, Mount et al. [4] showed a strong correlation ( $R^2 = 0.95$ ) between predicted and observed survival of C. dubia exposed to ambient samples from a watershed receiving oil field-produced waters. The C. dubia regression model was a better predictor of survival than any individual ion concentration, illustrating the ability of the model to predict the combined toxic effects of multiple ions. In a separate analysis, Mount et al. [15] showed a strong relationship between predicted and observed survival of C. dubia exposed to six produced waters collected from coalbed methane operations in Alabama. Obviously, these comparisons assume that major ions were the primary cause of toxicity in the field-collected samples.

Another application of the ion toxicity models that may prove equally or even more valuable lies in using model predictions to determine whether the presence of toxicants other than major ions is indicated. Research by Tietge et al. [5] both demonstrates this application and provides a rigorous evaluation of the predictive capability of the regression models. Six produced waters from various fossil fuel production sites were tested for toxicity and analyzed for major ion concentrations. The ion toxicity models presented here were used to predict survival of *C. dubia, D. magna,* and fathead minnows based on major ion concentrations. Differences between observed and predicted toxicity were used to make inferences as to whether the observed toxicity could be wholly explained by the major ion concentrations alone, or if the presence of other toxicants was indicated. The accuracy of these inferences was

then evaluated by conducting Phase I TIE manipulations [16] and by testing the toxicity of laboratory waters reconstituted to the same major ion concentrations. This study indicated that the *C. dubia* model provided highly accurate predictions, while the fathead minnow and *D. magna* models tended to overpredict ion toxicity. The tendency of the *D. magna* and fathead minnow models to overpredict toxicity in field-collected samples was also noted in comparisons made by Mount et al. [4].

Dickerson et al. [7] used the *C. dubia* and fathead minnow models to evaluate toxicity in surface waters influenced by irrigation drain water. Although independent tests were not performed to confirm model predictions, it appeared that predictions by the *C. dubia* model correlated well with observed toxicity. As in the study by Tietge et al. [5], however, the fathead minnow model seemed to overpredict toxicity; several sites had higher observed survival than was predicted by the fathead minnow model.

In summary, applications of the C. dubia models and, to a lesser extent the D. magna and fathead minnow models, have proven them to be highly effective and comprehensive tools for evaluating major ion toxicity. To date, they have been successfully applied to studies of ambient waters [15], produced waters [4,5], irrigation drain waters [7], water purification byproducts [23], municipal effluents, and effluents from pulp and paper, refining, and manufacturing industries (J.R. Hockett, unpublished data). Because the models represent the combined toxicity of all seven ions, they have much broader application than ion toxicity studies based on generic measures like conductivity or TDS, or studies focusing on certain waters or ion combinations. Application of these models can reduce the need for extensive characterization and fractionation manipulations during TIE studies of high TDS waters [11]. They can also be used to project changes in toxicity resulting from modifications in industrial processes, effluent treatment, or other remedial measures.

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# Environmental Toxicology

# THE ACUTE TOXICITY OF MAJOR ION SALTS TO *CERIODAPHNIA DUBIA*: I. INFLUENCE OF BACKGROUND WATER CHEMISTRY

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Abstract: The ions Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup>/CO<sub>3</sub><sup>2-</sup> (referred to in the present study as "major ions") are present in all freshwaters and physiologically required by aquatic organisms but can increase to harmful levels from a variety of anthropogenic activities. It is also known that the toxicities of major ion salts can vary depending on the concentrations of other ions, and understanding these relationships is key to establishing appropriate environmental limits. The authors present a series of experiments with *Ceriodaphnia dubia* to evaluate the acute toxicity of 12 major ion salts and to determine how toxicity of these salts varies as a function of background water chemistry. All salts except CaSO<sub>4</sub> and CaCO<sub>3</sub> were acutely toxic below saturation, with the lowest median lethal concentrations found for K salts. All 10 salts that showed toxicity also showed some degree of reduced toxicity as the ionic content of the background water increased. Experiments that independently varied Ca:Mg ratio, Na:K ratio, Cl:SO<sub>4</sub> ratio, and alkalinity/pH demonstrated that Ca concentration was the primary factor influencing the toxicities of Na and Mg salts, whereas the toxicities of K salts were primarily influenced by the concentration of Na. These experiments also indicated multiple mechanisms of toxicity and suggested important aspects of dosimetry; the toxicities of K, Mg, and Ca salts were best related to the chemical activity of the cation, whereas the toxicities of Na salts also reflected an influence of the anions and were well correlated with osmolarity. Understanding these relationships between major ion toxicity and background water chemistry should aid in the development of sensible risk-assessments and regulatory standards. *Environ Toxicol Chem* 2016;9999:1–19. Published 2016 Wiley Periodicals Inc. on behalf of SETAC. This article is a US government work and, as such, is in the public domain in the United States of America.

Keywords: Aquatic toxicology Major ions Ceriodaphnia dubia Toxicity mechanism Dose-response modeling

# INTRODUCTION

Inorganic ions generally present at the highest concentrations in freshwaters are Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HCO<sub>3</sub><sup>-</sup>/CO<sub>3</sub><sup>2-</sup> (referred to as "major ions" herein) and are used to describe the basic chemistry of natural waters [1]. All have physiological roles and are actively regulated by aquatic organisms [2] but can also cause toxicity when present in sufficient excess [3]. Concentrations in natural waters are governed by a variety of atmospheric, geochemical, and biological processes [1], but these natural concentrations can be greatly increased by a wide variety of anthropogenic influences, such as mineral mining, oil and gas extraction, irrigation, road deicing, water softening, and wastewaters from various industrial processes.

A variety of studies have shown or implicated major ions as causes of aquatic toxicity in surface waters, with sources such as oil and/or gas production [4,5], irrigation return flows [6,7], mining [8,9], road salt [10], and industrial wastewater [11]. In fact, toxicity identification studies on industrial and municipal effluents have shown major ions to be among the more common causes of effluent toxicity [12]. Field studies in Appalachian streams have also found associations between changes in macrobenthic communities and increased major ion concentrations from mining activities [13–15].

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Understanding the aquatic hazards posed by increased major ion concentrations presents a number of challenges. First, concentrations of major ions cannot be manipulated individually because charge balance demands that increased concentrations of any ion be offset by equal and opposite charge from other ions, making it more difficult to infer the effects of individual ions. Second, the relative concentrations of major ions vary widely across watersheds and anthropogenic inputs, and such differences are known to influence aquatic toxicity. For example, based on total salt concentration, a 1:1 mixture (by mass) of NaCl and CaCl<sub>2</sub> has substantially lower acute toxicity to Ceriodaphnia dubia than either salt alone [3], indicating that toxicity of this salt mixture is not simply additive. Third, the toxicity of a single salt can vary based on the characteristics of the water to which it is added, such as water hardness [16–20] and, more specifically, Ca [21]. Although relationships between water hardness and the toxicity of various other chemicals are often attributed, explicitly or implicitly, to the Ca and Mg ions that comprise most hardness, more detailed studies sometimes show that the concentrations of other ions covarying with hardness are playing important roles. For example, though "hardness" was long reported to influence toxicity of metals such as copper, later research demonstrated more detailed roles of specific ions; this enhanced understanding was incorporated into a more refined toxicity model, the biotic ligand model [22].

In previous work, Mount et al. [3] approached the toxicity of major ion mixtures by developing a multivariate regression model based on a large number of acute toxicity tests conducted with many different combinations of major ion salts. The resulting models predict the survival of 3 test species,

cladocerans *Ceriodaphnia dubia* and *Daphnia magna*, and the fathead minnow (*Pimephales promelas*) based on concentrations of the 7 major ions. Although this model represented a step forward in addressing the complexities of evaluating ion mixtures and showed effectiveness as a predictive tool [5,23], there are important aspects of major ion toxicity that were not fully addressed. Notable among these was that all of the ion solutions tested were created by adding ions to a single base water. This issue underlies the failure of the model to represent some influences of background water chemistry on ion toxicity, such as the reduction of NaCl and Na<sub>2</sub>SO<sub>4</sub> toxicity afforded by adding hardness within ranges common to natural waters [16–21]. Other aspects of the interactions among the ions (e.g., independent vs additive toxicity) also are incompletely addressed by this regression model.

The present study is the first of 3 articles that establish a better foundation for predicting the acute toxicity of elevated major ion concentrations to *C. dubia*. The present study describes a comprehensive study of the influence of background water chemistry that extends earlier work on hardness effects [16–21] to more water chemistry factors and to more major ion salts. Toxicities of major ion salts were evaluated using a wide range of dilution waters; some mimicked natural waters, while others were designed specifically to isolate different components of background water chemistry to better understand their roles in influencing major ion toxicity and thereby determine what is important to risk assessment of ions in natural systems.

These experiments also allowed preliminary consideration of exposure metrics that more effectively describe major ion toxicity to *C. dubia* than total salt concentration. For example, are there different toxicity mechanisms among the salts that need to be addressed? Should the toxicity of a salt be related to an individual ion or both ions, and how should concentrations of multiple ions be combined? Should reductions in toxicity attributable to the formation of complexes between ions and to the general effects of high ion concentrations on chemical reactivity be addressed?

The second article will present results of mixture tests with pairs of salts to more rigorously address the preceding questions. The third will address how the information from the other articles can be incorporated into a mathematical model applicable to any ion mixture and will test the predictions of that model for more complex mixtures relevant to field exposures.

The acute toxicity to *C. dubia* was selected as the endpoint for these efforts because this is a widely distributed organism with considerable sensitivity to ions and for which it was practical to conduct the large number of tests needed to adequately address the multiple factors and interactions of interest. The knowledge gained from the present study with *C. dubia* supports more informed testing and model development for other endpoints and species, which are now under way and will be the subject of additional publications. When combined with the efforts of other investigators, this body of information will support better assessment of the risks of major ions to aquatic communities.

#### MATERIALS AND METHODS

Test water composition and study design

Twenty-six experiments were conducted on the acute toxicity of individual major ion salts to *C. dubia*. Each experiment consisted of 3 to 8 simultaneous toxicity tests with different combinations of dilution waters and toxicants, for a total of 149 median lethal concentration (LC50) determinations. Test waters were developed from deionized water, sand-filtered and ultraviolet light-treated Lake Superior water (LSW), or a combination of both. Deionized water was produced from a Millipore Super-Q system configured as specified by the US Environmental Protection Agency (USEPA) [24]. Lake Superior water was obtained from an intake located offshore from our laboratory at 46.840°N, 92.004°W; typical hardness and alkalinity are 47 mg/L and 43 mg/L as CaCO<sub>3</sub>, respectively, with conductivity of 104  $\mu$ S/cm and pH approximately 7.5. The full ionic composition of LSW is provided in Table 1.

Table 1. Composition of dilution waters

Dilution water description	Abbreviation	Base water	Na (mg/L)	K (mg/L)	Ca (mg/L)	Mg (mg/L)	Cl (mg/L)	SO <sub>4</sub> (mg/L)	Alkalinity (mg CaCO <sub>3</sub> /L)	Hardness (mg CaCO <sub>3</sub> /L
Lake Superior water	LSW		1.62	0.60	14.0	2.92	1.50	3.40	43.0	47.0
Amended Lake Superior water	ALSW	LSW	6.48	1.51	14.6	4.09	7.66	14.9	43.0	53.3
1/3× strength ALSW	$1/3 \times ALSW$	$1/3 \times LSW$	2.16	0.50	4.87	1.36	2.55	4.97	14.3	17.8
			$(4.77)^{a}$	$(1.18)^{a}$	$(4.87)^{a}$	$(1.36)^{a}$	$(5.04)^{a}$	$(7.88)^{a}$	$(14.4)^{a}$	
3× strength ALSW	$3 \times ALSW$	LSW	19.4	4.52	43.8	12.3	23.0	44.7	129	160
			$(16.7)^{a}$	$(2.74)^{a}$	$(43.9)^{a}$	$(12.3)^{a}$	$(19.0)^a$	$(46.0)^a$	$(125)^{a}$	
Moderately hard reconstituted water	MHRW	Deionized water	26.3	2.10	14.0	12.0	1.90	81.0	57.2	84.4
One-third strength MHRW	$1/3 \times MHRW$	Deionized water	8.75	0.70	4.65	4.01	0.63	27.0	19.1	28.1
One-eighth strength MHRW	$1/8 \times MHRW$	Deionized water	3.28	0.26	1.75	1.50	0.24	10.1	7.2	10.6
ALSW with high Ca:Mg ratio	High Ca:Mg	$0.214 \times LSW$	6.48	1.51	19.0	1.40	7.66	14.9	43.0	53.3
ALSW with low Ca:Mg ratio	Low Ca:Mg	0.214× LSW	6.48	1.51	3.00	11.1	7.66	14.9	43.0	53.3
ALSW with high Cl:SO <sub>4</sub> ratio	High Cl:SO <sub>4</sub>	LSW	6.48	1.51	14.6	4.09	14.3	3.40	43.0	53.3
ALSW with low Cl:SO <sub>4</sub> ratio	Low Cl:SO <sub>4</sub>	LSW	6.48	1.51	14.6	4.09	1.50	20.7	43.0	53.3
ALSW with high Na:K ratio	High Na:K	0.333× LSW	7.65	0.20	14.6	4.09	7.66	14.9	43.0	53.3
ALSW with low Na;K ratio	Low Na:K	0.333× LSW	1.89	10.0	14.6	4.09	7.66	14.9	43.0	53.3
ALSW with high alkalinity	High Alk	0.233× LSW	28.3	1.51	14.6	4.09	7.66	14.9	90.0	53.3
ALSW with low alkalinity	Low Alk	0.233× LSW	28.3	1.51	14.6	4.09	7.66	14.9	10.0	53.3
ALSW with varying Na		LSW	1.62	1.51	14.6	4.09	1.50	14.9	43.0	53.3
			3.00				3.62			22.2
			10.0				14.4			
			30.0				45.3			
			100				153			
			300				460			

<sup>&</sup>lt;sup>a</sup>Parentheses denote estimated geometric average ion concentrations at designated hardness for selected U.S. waters. ALSW = amended Lake Superior water; LSW = Lake Superior water; MHRW = moderately hard reconstituted water.

Table 2. Major salts tested within each set of experiments

Experimental Set Description	cription	NaCl	NaCl Na <sub>2</sub> SO <sub>4</sub> NaH	NaHCO <sub>3</sub>	KCI	K2SO4	K2SO4 KHCO3	$MgCl_2$	$MgSO_4$	MgCO <sub>3</sub>	CaCl2	MgCl <sub>2</sub> MgSO <sub>4</sub> MgCO <sub>3</sub> CaCl <sub>2</sub> Na gluconate		Mg gluconate Ca gluconate Mannito	Manni
1/3× ALSW culture vs 1× ALSW culture	s 1× AI SW culture	кX						>							
tested in 1/3× ALSW	W	4						<							
1/3× MHRW culture v	1/3× MHRW culture vs 1× MHRW culture,														
tested in 1/3× MHRW	W														
1/8× MHRW culture vs 1× MHRW	vs 1× MHRW	Xp	×	×	×			×	×		>				
culture, tested in 1/8× MHRW	8× MHRW								*		4				
MHRW vs ALSW		X	×	×	X	×	×	×	×	^	>				
$1/3 \times ALSW \text{ vs } 3 \times ALSW$	LSW	X	Xe	×	:×	:×	***	×	<×	< ×	<>				
Low Ca:Mg vs high Ca:Mg	a:Mg	×	X	×	×			×	:×	4	ζ.				
Low CI:SO <sub>4</sub> vs high CI:SO <sub>4</sub>	1:SO <sub>4</sub>	×	×	×		×		4	<×	×					
Low Na:K vs high Na:K	:K	X	×	X				×	×	4	>				
Low vs high alkalinity		X	X		×			×	:×		4				
ALSW with low vs ambient vs high pH	abient vs high pH	×			i i			×	**						
ALSW with varying Na (0.07-13 mM)	Ia (0.07-13 mM)				×			47							
CaCO, precipitation effects, aged dilution	fects, aged dilution			X						×					
water vs not aged, 1	water vs not aged, $1 \times ALSW$ vs $3 \times ALSW$									17					
Organic salts and mannitol, 1x ALSW only	nitol, 1× ALSW only	X										>	>	>	>

 $^{\rm u}$ Organisms from both amended ALSW cultures also tested in  $1\times$  ALSW.  $^{\rm b}$ Organisms from both cultures also tested in  $1\times$  MHRW.

<sup>c</sup>Duplicate tests conducted which also included  $1 \times$  ALSW. ALSW = amended Lake Superior water, MHRW = moderately hard reconstituted water.

Some test waters were based on the commonly used reconstituted water formulas given by the USEPA [24] and originally proposed by Marking and Dawson [25], specifically the formula for "moderately hard reconstituted water" (MHRW), which was also used by Mount et al. [3]. While commonly used, MHRW (and other reconstituted waters based on this formula) has a chemistry (Table 1) that is atypical for surface waters of the United States: the Ca:Mg ratio is low, the Cl:SO<sub>4</sub> ratio is very low, and the Na concentration is extremely high relative to hardness. The high Na concentration is a consequence of using NaHCO3 to impart alkalinity; in nature, alkalinity generally comes from dissolution of carbonate minerals of Ca and Mg, but these have solubilities and dissolution rates that are inconveniently low for water preparation in the laboratory. The low Ca:Mg and Cl:SO4 ratios in MHRW are not a practical requirement, and the reason for their original selection is not clear.

To design a dilution water with chemistry more like natural waters, we analyzed data from the US Geological Survey national stream water-quality monitoring networks [26]. Data records were obtained for 425 sites in operation from 1983 to 1992. Records (n = 32895) were used that had 1) measurements for all major ions, 2) a charge imbalance of no more than 10%, 3) a hardness  $\geq$ 10 and  $\leq$ 400 mg/L as CaCO<sub>3</sub>, and 4) pH no lower than 6.0. These data were subjected to regression analyses using Sigmaplot (v11.0; Systat Software) to establish relationships between log(hardness) and the logarithms of various other ion concentrations and ratios. A formula was then developed for an "amended Lake Superior water" (ALSW; Table 1) consistent with these relationships. Salts were added to LSW to increase hardness to the estimated geometric average (53.3 mg CaCO<sub>3</sub>/ L) in the US Geological Survey data for the alkalinity of LSW (43.0 mg CaCO<sub>3</sub>/L) and to increase other ions to average values for this hardness. Benefits of modifying LSW to create this test dilution water, rather than creating a completely synthetic water from deionized water, are that it provides alkalinity without having to dissolve carbonate salts or add excessive Na and that it will contain trace constituents from the source water that make the water more realistic and possibly beneficial to the test organisms.

The present study was organized into 12 sets of 1 to 4 experiments addressing different aspects of dilution water chemistry or experimental procedure. These sets of experiments and the toxicants that were tested are summarized in Table 2, and the chemistries of the various dilution waters are provided in Table 1. The purpose and design of each experimental set were as follows.

Sets 1 and 2: Effects of acclimation to dilution water. Some previous studies of the effect of dilution water on major ion toxicity to C. dubia used organisms that were cultured in the dilution water prior to testing [16-20]. This was not done routinely in the present study because the logistics involved in culturing organisms in so many different dilution waters (Table 1) were prohibitive. However, because a preliminary comparison of NaCl toxicity in MHRW and MHRW diluted by a factor of 3 (1/3× MHRW) showed much less of an impact of this dilution than the  $1\times$ ,  $1/2\times$ ,  $1/4\times$ , and  $1/8\times$  MHRW series tested by Elphick et al. [19], we conducted experiments to determine whether observed salt toxicity in waters more dilute than MHRW or ALSW varied significantly depending on whether the organisms were cultured in these dilute test waters or in the standard-strength water formulations. Initially, organisms from our primary culture (in 1× MHRW) were used to create secondary cultures in 1× MHRW, 1/3× MHRW,  $1\times$  ALSW, and  $1/3\times$  ALSW, which were maintained for a minimum of 2 generations before being used in tests. The toxicities of NaCl and MgCl<sub>2</sub> in  $1/3\times$  MHRW and  $1/3\times$  ALSW were simultaneously tested using the cultures from both the dilute water and the corresponding full-strength water (Table 2, set 1). For NaCl, tests were also conducted in 1x ALSW using both the  $1\times$  ALSW and  $1/3\times$  ALSW cultures. To further explore this issue, we conducted additional acclimation studies (Table 2, set 2) comparing the toxicity of several salts in  $1/8\times$  MHRW using *C. dubia* cultured in MHRW and in  $1/8\times$  MHRW (the same composition as the 10 and 80 hardness waters in the Elphick et al. [19] study). For NaCl, tests were also conducted in  $1\times$  MHRW using both cultures.

Set 3: Salt toxicity in MHRW versus ALSW. Because ALSW was developed in response to concerns about the composition of MHRW and because much existing ion toxicity information was conducted in MHRW or similar waters, comparative toxicities in these 2 waters were of interest. Therefore, a set of experiments (Table 2, set 3) compared the toxicity of all major ion salts in MHRW and ALSW, except for CaSO<sub>4</sub> and CaCO<sub>3</sub>, for which preliminary experiments showed no toxicity at saturation in these waters.

Set 4: Salt toxicity in different strengths of ALSW. Natural waters vary widely in total ion concentration, the 10th and 90th percentiles in our analysis of US Geological Survey data being approximately 1.5 mEq/L and 16 mEq/L for the sum of major cations and anions. To determine the effect of varying overall ion concentrations, another set of experiments (Table 2, set 4) compared toxicities of 9 salts in dilution waters that had onethird and 3 times the ion concentrations of ALSW ( $1/3 \times$  ALSW and 3× ALSW; Table 1). We note that, in natural waters, average concentrations of the various ions do not vary exactly in proportion with total ion concentrations, so  $1/3 \times$  ALSW and  $3 \times$ ALSW do not match our estimated field water averages at the same hardnesses, which are also provided in Table 1 for comparison purposes. We elected not to match the field chemistry so that all ions would be in the same proportions across the  $1/3\times$ ,  $1\times$ , and  $3\times$  ALSW series but the ion ratios would still be well within the ranges observed in the US Geological Survey data.

The results of these tests were combined with separate data for  $1 \times ALSW$  (from set 3) to compare across a  $1/3 \times$ ,  $1 \times$ , and  $3 \times ALSW$  dilution water series; for NaCl and Na<sub>2</sub>SO<sub>4</sub> an additional experiment simultaneously tested this entire series to verify results. Preparation of  $1/3 \times ALSW$  was simply by dilution of ALSW with deionized water. Creating the  $3 \times ALSW$  chemistry required dissolution of CaCO<sub>3</sub> to provide higher alkalinity while maintaining the desired cation ratios, in addition to adding other more soluble salts. This was accomplished by adding the appropriate amounts of the various salts to LSW in a large graduated cylinder and bubbling CO<sub>2</sub> gas through the solution until pH stabilized near 5.0 and all salts were dissolved. The solution was then aerated with ambient air until pH was approximately 7.8.

Sets 5–7: Effects of modifying specific ion ratios. Most previous work on the effects of dilution water chemistry on major ion toxicity to daphnids has focused on water hardness [16–20] as a generalized parameter without parsing the influence of other ions that covaried with the hardness, although Davies and Hall [21] did identify Ca as being more important than Mg for the effects of hardness on Na<sub>2</sub>SO<sub>4</sub> toxicity. Regarding other ions in dilution water, Soucek [17] reported an ameliorative effect of low concentrations of added NaCl on the toxicity of Na<sub>2</sub>SO<sub>4</sub> to Hyalella azteca (attributed to

the chloride requirements for this species) but no such effect on *C. dubia*. At higher NaCl concentrations, there was an exacerbation of Na<sub>2</sub>SO<sub>4</sub> toxicity for both species, attributed to additive toxicity of the 2 salts. Soucek et al. [18] also reported a small increase in apparent NaCl toxicity when Na<sub>2</sub>SO<sub>4</sub> was added, again attributed to additive toxicity of the 2 salts.

To more completely evaluate the influence of particular dilution water characteristics on the toxicity of major ion salts, we tested selected salts in ALSW modified to provide different ratios of certain ion pairs, based on the tails of the distributions of these ratios in our analysis of US Geological Survey data. Waters with high Ca:Mg (13.6 by mass) and low Ca:Mg (0.27) ratios (Table 2, set 5) were created by changing the salts added to LSW to achieve the target ratios without altering concentrations of any other ions in the water (Table 1). Similar approaches were used to create waters with high and low Cl:SO<sub>4</sub> ratios (4.2 and 0.072 by mass; Table 2, set 6) and high and low Na:K ratios (38 and 0.19 by mass; Table 2, set 7). Some of these waters required that LSW be initially diluted with deionized water to lower the concentrations of a target ion to its lowest value, followed by addition of salts to match the ALSW chemistry except for the ion pair being manipulated (Table 1). Where required, the CO<sub>2</sub>-aided dissolution of CaCO<sub>3</sub> was used to restore alkalinity to waters based on diluted LSW.

Sets 8 and 9: Effects of manipulating alkalinity and pH. The effects of pH and alkalinity were evaluated in 2 ways. First, the alkalinity of ALSW was reduced to 10 mg CaCO<sub>3</sub>/L and raised to 90 mg CaCO<sub>3</sub>/L (resulting, respectively, in pHs of 7.3–7.5 and 8.1–8.3; Table 2, set 8). Alkalinity was raised simply by adding NaHCO<sub>3</sub>, whereas it was lowered by diluting LSW and adding salts to restore all cations and Cl to their same values as in the high-alkalinity water, with SO<sub>4</sub> replacing the alkalinity (Table 1). Second, pH was reduced to approximately 6.8 and raised to approximately 8.5 (Table 2, set 9) without altering alkalinity or other ions by bubbling the ALSW used in test solution preparation with air containing 1% CO<sub>2</sub> and 0% CO<sub>2</sub>, respectively, and enclosing test vessels in sealed chambers containing these same CO<sub>2</sub> concentrations [27].

Set 10: Effect of sodium on potassium toxicity. Based on the results of the tests contrasting MHRW and ALSW (set 3), we hypothesized that the toxicity of K salts was dependent on the Na concentration in the dilution water. To evaluate this hypothesis, the toxicity of KCl was tested in 6 different dilution waters with Na ranging from 1.6 mg/L to 300 mg/L (Table 2, set 10). Test waters were prepared by modifying the formulation of ALSW to have no added Na and Cl, then adding NaCl to achieve the desired levels of Na (Table 1).

Set 11: Effects of time-dependent calcium precipitation. Testing of the toxicity of NaHCO3 and MgCO3 resulted in oversaturation of CaCO<sub>3</sub> and possible loss of Ca from solution, which would affect toxicity given the established ameliorative effect of Ca. Although Ca was monitored in these tests, the sampling was too limited to precisely determine the Ca concentration to associate with the LC50. Therefore, an additional experiment was conducted to more thoroughly characterize the dependence of any Ca loss on time and test salt concentration. This was a  $2 \times 2 \times 2$  factorial experiment, the 3 factors being test salt (NaHCO<sub>3</sub>, MgCO<sub>3</sub>), dilution water  $(1 \times ALSW, 3 \times ALSW)$ , and solution age (freshly prepared vs aged for 48 h before introducing test organisms; Table 2, set 11). Extra replicate test cups were prepared to allow sampling of Ca at multiple times and treatment concentrations. This experiment provided not only NaHCO3 and MgCO3 LC50s from tests that were monitored better but also data with which to better estimate Ca concentrations at the LC50 in other CO<sub>3</sub>/HCO<sub>3</sub> salt tests.

Set 12: Toxicities of gluconate salts and mannitol. Charge balance requires including both cations and anions in any testing of ion toxicity, making it more difficult to infer the relative toxicities of the individual ions. In addition, results of other experiments in the present study led us to hypothesize that the toxicity of some salts might be related to their effect on the osmolarity of the test solution and thus on the osmotic gradient the organism experiences. To provide more information on the role of the individual cations, we tested the toxicity of the gluconate salts of Na, Ca, and Mg (Table 2, set 12). Gluconate is an organic ion that is not expected to be absorbed appreciably and, thus, should act only through its effect on osmotic potential or on charge gradients, providing an informative contrast to Cl, SO<sub>4</sub>, and HCO<sub>3</sub>/CO<sub>3</sub>. This experiment also evaluated the toxicity of mannitol, a sugar alcohol also not expected to be appreciably absorbed, enabling us to manipulate the external osmolarity without adding any ions. NaCl was also included as a reference major ion salt.

# Toxicity test procedures

Salts for all 12 combinations of the 4 major cations and 3 major anions; gluconate salts of Na, Ca, and Mg; and mannitol were obtained from Sigma-Aldrich or Fisher Scientific. All chemicals were American Chemical Society reagent grade or better, with the exception of MgCO<sub>3</sub> which was specified as meeting US Pharmacopeia requirements. The certificate of analysis for this salt was used to determine the ratio of MgCO<sub>3</sub> to total salt weight for computing nominal MgCO<sub>3</sub> concentrations. This certificate of analysis also specified the Ca content to be 0.73% of the Mg content, enough to appreciably affect the background Ca; this was used to adjust the nominal Ca background concentration for tests on the toxicity of MgCO<sub>3</sub>.

Test organisms were <24-h-old *C. dubia* obtained from inhouse cultures. Most test organisms were cultured in MHRW, but organisms for testing the effects of acclimation were also cultured in the dilution waters for the tests (for a least 2 wk prior to the tests); and some tests late in the present study were conducted with organisms cultured in ALSW after a switch of our culture to this water. The culture water for each toxicity test is indicated in Table 3. General culture procedures followed those described by the USEPA [24].

Static 48-h toxicity tests were conducted in 30-mL plastic cups (Berry Plastics) filled with 10 mL test solution and held in polystyrene boards with holes sized to the cups. These boards were floated in a temperature-controlled water bath with a glass sheet covering all test cups. Because preliminary studies indicated that the response curves for major ion salts were quite steep, we used closely spaced exposure concentrations, with each test concentration being approximately 80% of the next higher. Test solutions were prepared by combining the applicable salt and dilution water to achieve the highest test concentration (100%). From this, 2 additional solutions, 80% and 64%, were prepared by dilution of the first; then, each of these 3 solutions was serially diluted by  $0.5 \times$  to produce a total of 9 to 12 exposure concentrations, plus a dilution water control. The highest salt concentration in each test varied according to its expected toxicity (based on a combination of preliminary testing, any preceding experiments, and literature values), but all tests used the same relative dilution spacing. Most experiments were structured to compare toxicity under 2 related conditions (e.g., high and low Ca:Mg); tests were assigned to experiments such that both conditions would be tested simultaneously for a particular salt, using the same preparation of dilution water and the same cohort of test organisms.

All salts except CaCO<sub>3</sub>, MgCO<sub>3</sub>, and CaSO<sub>4</sub> could be easily dissolved at concentrations high enough to cause mortality. For CaCO<sub>3</sub> and MgCO<sub>3</sub>, the CO<sub>2</sub> procedure in the description of set 4 was used to dissolve the salts. After adjusting to pH 7.8, MgCO<sub>3</sub> solutions so prepared were stable for 48 h at lethal concentrations, thereby allowing for testing as for other salts. However, a 15 mM (1500 mg/L) CaCO<sub>3</sub> solution dissolved in ALSW using CO<sub>2</sub> showed substantial precipitation and settling once the pH was raised, with the total concentration being only 6.5 mM at test start and dropping to 2.4 mM (2.2 mM dissolved) after 24 h and to 1.2 mM after 48 h, with no mortality observed. For CaSO<sub>4</sub>, consistent with the findings of Mount et al. [3], a saturated solution at 16.2 mM (2200 mg CaSO<sub>4</sub>/L) was not acutely toxic to *C. dubia* in MHRW or ALSW. Based on these results, no further testing was conducted with CaSO<sub>4</sub> or CaCO<sub>3</sub>.

Test cups were usually prepared in duplicate, but in a few tests with  $CO_3/HCO_3$  salts more replicates were used to provide test solution for more monitoring of alkalinity or Ca than in other tests. Each cup received food in the form of  $100\,\mu\text{L}$  of a 50:50 mixture of yeast, cereal leaves, and trout chow [24] and algae (*Pseudokirchneriella subcapitata* at  $3.5\times10^7$  cells/mL). Food was added based on previous work indicating that it had minimal effect on toxicity of major ions [3] and to avoid possible stress. Test temperature was  $25\pm1\,^{\circ}\text{C}$  with fluorescent lighting on a 16:8-h light: dark photoperiod. Five *C. dubia* neonates were added to each cup, and survival was determined after 24 h and 48 h of exposure; death was defined as no visible movement after gentle prodding and at least 10 s of observation.

#### Exposure monitoring

Temperature was monitored daily in test cups from each simultaneous test and continuously in the surrounding water bath. Hardness and alkalinity of dilution waters in every test were measured by titration [28]. For tests with MgCO<sub>3</sub> and NaHCO<sub>3</sub>, alkalinity was also measured in the highest test concentration. Conductivity was used to verify that dilution of the salt solutions was done properly and was measured (model 2052; Amber Science) in each treatment at the start of the exposure, in every treatment that had 100% mortality after 24 h, and in all remaining treatments at 48 h. The pH was measured (PH150 with A57198 probe; Beckman Coulter) at the start of each test in the highest concentration and control (in all concentrations for tests with CO<sub>3</sub>/HCO<sub>3</sub> salts), in every concentration that had 100% mortality at 24 h, and in all remaining treatments at 48 h. Dissolved oxygen was measured (model 58; Yellow Springs Instruments) in the highest concentration and control at the start of the test, in the highest and lowest concentrations with 100% mortality at 24 h, and in 2 arbitrarily selected treatments at 48 h.

Because of the large number of exposure treatments (~1500), it was not practical to measure major ion concentrations in every treatment. Instead, the analytical sampling program was structured with 2 primary purposes: 1) to verify that the dilution waters and highest concentrations were prepared properly and 2) to verify the exposure concentrations near the LC50. Accordingly, analytical samples were collected at the start of exposure from each dilution water and from the highest concentration of each salt in each dilution water. In addition, analytical samples were collected from each exposure series after 24 h in the lowest concentration with 100% mortality and at 48 h from the concentration nearest the LC50. Beyond this routine sampling program, some tests included additional

Table 3. Results of tests on the acute toxicity of major ion salts, gluconate salts, and mannitol to Ceriodaphnia dubia in various tests waters<sup>a</sup>

arion	m CI	31.8 35.4 0.02 0.05	28.9 26.8 30.3 33.4 31.0 28.1	26.8 25.4 11.2 14.1 6.42 6.57	15.5 31.0 17.2 29.4 1.46 2.44	0.01 0.01 0.01 0.01 2.38 3.01	0.01 0.01 30.2 30.0	4.92 6.28 34.5 36.0 36.0 18.3	0.22 0.05 0.22 0.05	
Mola III v	Mg mM	0.16 0.50 0.16 0.50	0.17 0.06 0.06 0.17 0.50 0.16	0.16 0.16 5.61 7.06 3.37 3.44	0.06 0.50 0.06 0.50 0.79 1.28	0.06 0.06 1.39 2.48 0.06	0.06 0.06 0.06 0.06	0.17 0.50 0.17 0.50 9.21 8.19	0.17 0.50 0.17 0.50	
and mainment to certoaupinia audia in various tests waters	Ca mM	0.116 0.348 0.116 0.348	0.364 0.121 0.121 0.364 0.348 0.116	0.116 0.116 0.121 0.121 0.116	0.044 0.348 0.044 0.348 0.044 0.044	0.044 0.044 0.044 0.044 0.044 0.044	0.044 0.044 15.1 15.0	0.364 0.348 17.5 18.3 0.364 0.348	0.364 0.348 0.364 0.364	
T to Cert	мm	0.02 0.05 0.02 0.05	0.04 0.01 0.04 0.05 0.05	0.02 0.02 0.01 0.01 0.02	0.01 0.05 0.01 0.05 0.01 0.01	0.01 0.01 0.01 2.38 3.01	0.01 0.01 0.01	4.75 6.28 0.04 0.05 0.05 0.05	4.19 7.01 0.04 0.05	
manning	Na mM	32.2 36.5 26.6 44.2	28.9 26.8 30.3 33.5 32.1 28.4	27.1 25.7 0.09 0.09 0.38 0.38	15.7 32.0 17.3 30.4 0.14 0.14	9.59 13.0 0.14 0.14 0.14 0.14	12.9 12.0 0.14 0.14	0.28 1.14 0.28 1.14 0.28 1.14	0.28 1.14 48.3 47.0	
giuconate sans,	LC50 mM (>95% CL)	31.8 (29.8–34.0) 35.3 (34.5–42.6) 13.1 (12.2–14.2) 21.5 (19.8–23.6)	28.7 (26.8–30.7) 26.7 (24.4–29.4) 30.2 (27.0–33.9) 33.2 (30.5–36.2) 31.0 (28.8–33.4) 28.1 (26.2–30.1)	26.7 (24.6–29.2) 25.3 (23.0–28.1) 5.56 (4.80–643) 7.01 (5.84–8.51) 3.20 (2.85–3.55) 3.28 (2.02–4.38)	15.5 (14.2–17.0) 30.9 (28.1–34.0) 17.2 (16.1–18.4) 29.3 (27.1–31.7) 0.72 (0.56–0.95) 1.22 (0.99–2.12)	4.72 (4.32–5.17) 6.45 (5.84–7.12) 1.33 (1.14–1.53) 2.42 (2.19–2.67) 2.37 (2.13–2.64) 3.0 (2.74–3.29)	12.7 (11.9–13.6) 11.9 (10.9–13.0) 15.1 (14.2–16.1) 15.0 (14.6–17.5)	4.71 (4.31–5.15) 6.22 (5.30–7.62) 17.2 (15.2–19.5) 18.0 (15.9–20.4) 9.04 (7.97–10.3) 7.70 (6.46–9.40)	2.08 (1.95–2.22) 3.48 (3.20–3.78) 24.0 (22.2–26.0) 22.9 (21.5–24.4)	
only of inagor foll saits,	LC50 mg/L (>95% CL)	1860 (1740–1990) 2060 (2020–2490) 1860 (1730–2010) 3060 (2810–3350)	1680 (1560–1790) 1560 (1420–1720) 1770 (1580–1980) 1940 (1780–2110) 1810 (1680–1950) 1640 (1530–1760)	1560 (1440–1710) 1480 (1340–1640) 529 (457–612) 667 (556–810) 305 (271–338) 312 (192–417)	907 (828–993) 1810 (1640–1990) 1010 (940–1080) 1710 (1580–1850) 69 (53–90) 116 (94–202)	671 (613–734) 916 (829–1012) 160 (137–184) 291 (264–321) 177 (159–197) 224 (204–245)	1070 (1000–1140) 1000 (920–1090) 1670 (1570–1780) 1660 (1620–1940)	351 (321–384) 464 (395–568) 1900 (1690–2170) 1990 (1760–2260) 861 (759–981) 733 (615–895)	362 (339–386) 606 (557–659) 3410 (3150–3700) 3260 (3060–3460)	
lile acute toxi	LC50 Method	Midpoint Probit Midpoint Probit	Midpoint Probit Probit Probit Probit Midpoint	Probit Probit Probit Probit Probit	Midpoint Midpoint Midpoint Probit Probit	Midpoint Midpoint Probit Probit Probit Midpoint	Midpoint Midpoint Midpoint Probit	Probit Probit Probit Probit Probit Probit Probit	Midpoint Midpoint Midpoint Midpoint	
Table 3. Acsults of tests off the acute toxicity of inajor foll saits,	Test Water	0.33× MHRW MHRW 0.33× MHRW MHRW	ALSW 0.33× ALSW 0.33× ALSW ALSW MHRW 0.33× MHRW	0.33× MHRW 0.33× MHRW 0.33× ALSW 0.33× ALSW 0.33× MHRW	0.125× MHRW MHRW 0.125× MHRW MHRW 0.125× MHRW 0.125× MHRW	0.125× MHRW 0.125× MHRW 0.125× MHRW 0.125× MHRW 0.125× MHRW	0.125× MHRW 0.125× MHRW 0.125× MHRW 0.125× MHRW	ALSW MHRW ALSW MHRW ALSW MHRW	ALSW MHRW ALSW MHRW	
Laure	Culture Water	MHRW MHRW MHRW MHRW	ALSW ALSW 0.33× ALSW 0.33× ALSW MHRW 0.33× MHRW	0.33× MHRW MHRW 0.33× ALSW ALSW MHRW 0.33× MHRW	0.125× MHRW 0.125× MHRW MHRW MHRW 0.125× MHRW	0.125× MHRW MHRW 0.125× MHRW MHRW 0.125× MHRW MHRW	0.125× MHRW MHRW 0.125× MHRW MHRW	MHRW MHRW MHRW MHRW MHRW	MHRW MHRW MHRW MHRW	
	Test Chemical	$\begin{array}{c} NaCI \\ NaCI \\ Na_2SO_4 \\ Na_2SO_4 \end{array}$	NaC NaC NaC NaC NaC	NaCI NaCI MgCl <sub>2</sub> MgCl <sub>2</sub> MgCl <sub>2</sub>	NaCI NaCI NaCI NaCI MgCl <sub>2</sub>	N <sub>22</sub> SO <sub>4</sub> N <sub>32</sub> SO <sub>4</sub> MgSO <sub>4</sub> MgSO <sub>4</sub> KCI KCI	NaHCO <sub>3</sub> NaHCO <sub>3</sub> CaCl <sub>2</sub> CaCl <sub>2</sub>	KCI KCI CaCl <sub>2</sub> CaCl <sub>2</sub> MgCl <sub>2</sub>	K <sub>2</sub> SO <sub>4</sub> K <sub>2</sub> SO <sub>4</sub> Na <sub>2</sub> SO <sub>4</sub> Na <sub>2</sub> SO <sub>4</sub>	
	Set #		1			7			c	

Table 3. (Continued)

	Set #					4			5	
	Test Chemical	MgSO <sub>4</sub> MgSO <sub>4</sub>	KHCO3 KHCO3 NaHCO3 NaHCO3	NaCI NaCI MgCO <sub>3</sub> MgCO <sub>3</sub>	NaHCO <sub>3</sub> NaHCO <sub>3</sub> NaCI NaCI Na <sub>2</sub> SO <sub>4</sub> Na <sub>2</sub> SO <sub>4</sub>	KCI KCI MgSO <sub>4</sub> MgCl <sub>2</sub> MgCl <sub>2</sub>	CaCl <sub>2</sub> CaCl <sub>2</sub> K <sub>2</sub> SO <sub>4</sub> K <sub>2</sub> SO <sub>4</sub> MgCO <sub>3</sub>	NaCI NaCI NaCI Na <sub>2</sub> SO <sub>4</sub> Na <sub>2</sub> SO <sub>4</sub>	NaCI NaCI NaHCO <sub>3</sub> NaHCO <sub>3</sub> Na <sub>2</sub> SO <sub>4</sub> Na <sub>2</sub> SO <sub>4</sub>	KCI
	Culture Water	MHRW MHRW	MHRW MHRW MHRW MHRW	MHRW MHRW MHRW MHRW	MHRW MHRW MHRW MHRW MHRW MHRW	MHRW MHRW MHRW MHRW MHRW MHRW	ALSW ALSW ALSW ALSW ALSW ALSW ALSW	ALSW ALSW ALSW ALSW ALSW ALSW ALSW	MHRW MHRW MHRW MHRW MHRW MHRW	MHRW MHRW
	Test Water	ALSW	ALSW MHRW ALSW MHRW	ALSW MHRW ALSW MHRW	0.33× ALSW 3.0× ALSW 0.33× ALSW 3.0× ALSW 0.33× ALSW 3.0× ALSW	0.33 × ALSW 3.0 × ALSW 0.33 × ALSW 3.0 × ALSW 0.33 × ALSW 3.0 × ALSW	0.3× ALSW 3.0× ALSW 0.3× ALSW 3.0× ALSW 0.3× ALSW 3.0× ALSW	0.33 × ALSW ALSW 3.0 × ALSW 0.33 × ALSW ALSW 3.0 × ALSW	Low Ca:Mg HighCa:Mg Low Ca:Mg HighCa:Mg Low Ca:Mg HighCa:Mg	Low Ca:Mg HighCa:Mg
	LC50 Method	Probit Probit	Probit Probit Midpoint Probit	Probit Midpoint Midpoint Midpoint	Midpoint Probit Midpoint Probit Probit Midpoint	Midpoint Probit Midpoint Midpoint Probit Probit	Midpoint Midpoint Probit Midpoint Midpoint Midpoint	Midpoint Probit Midpoint Midpoint Midpoint Probit	Midpoint Midpoint Midpoint Midpoint Probit Midpoint	Midpoint Midpoint
7.0	LC50 mg/L (>95% CL)	2040 (1880–2230) 1990 (1830–2180)	322 (306–340) 454 (407–507) 1980 (1880–2090) 1610 (1440–1700)	1910 (1730–2110) 2000 (1830–2190) 906 (859–956) 892 (848–938)	1950 (1780–2130) 1710 (1560–1890) 1640 (1530–1760) 1980 (1770–2160) 1940 (1770–2100) 3350 (3140–3570)	192 (180–205) 481 (432–534) 1590 (1440–1770) 2860 (2620–3130) 374 (344–409) 1290 (1160–1490)	1600 (1490–1710) 2000 (1870–2130) 234 (215–254) 518 (449–598) 390 (365–417) 977 (916–1041)	1700 (1600–1800) 1910 (1760–2070) 2420 (2210–2660) 1890 (1760–2020) 3440 (3220–3670) 3520 (3200–3880)	1580 (1420–1740) 1920 (1790–2050) 1540 (1440–1640) 2000 (1870–2140) 1430 (1340–1560) 3140 (2940–3360)	245 (229–263) 281 (258–306)
ore 5. (Commuted)	LC50 mM (≥95% CL)	16.9 (15.6–18.5) 16.6 (15.2–18.1)	3.22 (3.06–3.40) 4.53 (4.07–5.06) 23.6 (22.3–24.9) 19.1 (17.2–20.3)	32.7 (29.6–36.1) 34.3 (31.4–37.5) 10.8 (10.2–11.3) 10.6 (10.1–11.1)	23.2 (21.2–25.4) 20.4 (18.6–22.5) 28.1 (26.2–30.0) 33.8 (30.2–36.9) 13.6 (12.5–14.8) 23.6 (22.1–25.2)	2.58 (2.41–2.75) 6.45 (5.79–7.16) 13.2 (11.9–14.7) 23.8 (21.7–26.0) 3.93 (3.61–4.30) 13.6 (12.2–15.7)	14.4 (13.4–15.4) 18.0 (16.9–19.2) 1.34 (1.23–1.46) 2.97 (2.58–3.43) 4.63 (4.33–4.95) 11.6 (10.9–12.4)	29.1 (27.4–30.9) 32.6 (30.1–35.4) 41.5 (37.8–45.5) 13.3 (12.4–14.2) 24.2 (22.6–25.8) 24.8 (22.5–27.3)	27.0 (24.3–29.8) 32.8 (30.7–35.0) 18.3 (17.2–19.5) 23.8 (22.2–25.5) 10.1 (9.41–11.0) 22.1 (20.7–23.6)	3.29 (3.07–3.53)
	Na mM	0.28	0.28 1.14 23.8 20.3	33.0 35.4 0.28 1.14	23.3 21.2 28.2 34.7 27.3 23.8	0.09 0.85 0.09 0.85 0.09 0.85	0.09 0.85 0.09 0.85 0.09	29.2 32.9 42.3 26.7 48.7	27.2 33.1 18.6 24.1 20.4 44.5	0.28
	K mM	0.04	3.25 4.59 0.04 0.05	0.04 0.05 0.04 0.05	0.01 0.12 0.01 0.12 0.01 0.12	2.59 6.57 0.01 0.12 0.01 0.12	0.01 0.12 2.70 6.06 0.01	0.01 0.04 0.12 0.01 0.04 0.12	0.04 0.04 0.04 0.04 0.04	3.32
	Ca mM	0.364	0.364 0.348 0.187 <sup>b</sup> 0.141 <sup>b</sup>	0.364 0.348 0.443 0.426	0.121 0.231 <sup>b</sup> 0.121 1.093 0.121 1.093	0.121 1.093 0.121 1.093 0.121 1.093	14.5 19.1 0.121 1.093 0.155 0.318 <sup>b</sup>	0.121 0.364 1.093 0.121 0.364 1.093	0.075 0.475 0.075 0.200 <sup>b</sup> 0.075 0.475	0.075
	Mg mM	17.1	0.17 0.50 0.17 0.50	0.17 0.50 10.9 11.1	0.06 0.50 0.06 0.50 0.06 0.06	0.06 0.50 13.3 24.3 3.98 14.0	0.06 0.50 0.06 0.50 4.68 12.1	0.06 0.17 0.50 0.06 0.17 0.50	0.46 0.06 0.46 0.06 0.46 0.06	0.46
	CI	0.22 1 0.05	0.22 0.05 0.22 0.05	32.9 (34.4 (0.22 (0.05 (	0.07 (0.65 (28.1 (34.5 (0.07 (0.65 (0.07 (0.05 (0.07 (0.05 (0.07 (0.05 (	2.65 (7.10 (0.07 10.65 27.93 (0.27.8 (0.07 10.65 27.93 (0.07 10.65 27.8 (0.07 10.65 10.65 27.8 (0.07 10.65 1	28.8 (9.007 1 0.05 3 0.07 (9.05 0.05 0.05 0.05 0.05 0.05 0.05 0.05	29.1 (2.42.1 (2.00.07 1) 0.02 2	27.2 C 33.0 C 0.22 C 0.22 C 0.22 1 0.22 2	3.50 03.99 0
	SO4 mM	7.10	0.16 0.84 0.16 0.84	0.16 0.84 0.16 0.84	0.05 0.47 0.05 0.47 13.7	0.05 0.47 13.3 24.2 0.05	0.05 0.47 1.39 3.44 0.05	0.05 0.16 0.47 13.4 24.4 25.3	0.15 0.15 0.15 0.15 10.2 22.3	0.15
	Alk meq/L	0.86	4.08 5.68 24.4 20.3	0.86 1.14 22.4 22.3	23.5 23.0 0.29 2.58 0.29 2.58	0.29 2.58 0.29 2.58 0.29 2.58	0.29 2.58 0.29 2.58 9.54 25.8	0.29 0.86 2.58 0.29 0.86	0.86 0.86 19.2 24.6 0.86	0.86 0.86
	hф	7.85	8.70 8.85 9.35 9.25	8.00 8.10 9.25 9.25	9.30 9.35 7.40 8.30 7.55 8.45	7.65 8.45 7.45 8.35 8.35	7.35 7.95 7.40 8.50 8.95 9.15	7.50 7.90 8.40 7.55 7.95 8.50	7.90 7.90 9.25 9.30 7.95 8.00	7.95
	LC50 µS/cm	2240 2280	487 844 1940 1680	3110 3470 1160 1230	1800 1800 2700 3490 2480 4090	430 1220 1780 2910 910 2760	2870 3510 426 1120 619 1280	2960 3320 4110 2480 4000	2790 3218 1611 1970 2027 3822	626 663
	Osmol mOsM	25.2 26.3	8.0 13.3 41.7 36.6	63.6 68.1 23.2 25.2	40.3 39.1 53.9 69.1 36.5 65.4	5.7 18.1 19.4 36.9 11.6 40.8	38.7 52.4 4.5 13.7 11.8 28.9	55.7 63.5 83.1 35.7 63.8 68.5	53.0 63.8 33.8 41.2 28.6 58.7	8.4

(continued)

Table 3. (Continued)

Table 3. (Continued)

er cnem	listry eff	fects on acute ion t	oxicity to Ceriodaphnia	
Osmol	31.0	5.6 8.0 12.5 17.3 26.4 45.3	35.5 38.2 47.2 51.7 20.9 18.0 23.8 18.4	65.0 75.7 76.1 84.6 50.6
LC50 µS/cm	2057 2257	422 599 872 1193 1656 2616	1670 1793 2087 2284 1043 879 1054	3296 2188 2074 1638
Hd	6.75	7.95 8.00 7.95 7.95 7.95 7.95	9.35 9.35 9.15 9.10 9.30 9.30 9.15	7.85 7.75 7.85 7.80 7.80
Alk meq/L	0.86	0.86 0.86 0.86 0.86 0.86	20.6 22.4 26.4 29.3 19.0 14.6 19.6	0.86 0.86 0.86 0.86 0.86
SO4 mM	0.16	0.16 0.16 0.16 0.16 0.16 0.16	0.16 0.47 0.16 0.47 0.16 0.47 0.16	0.16 0.16 0.16 0.16 0.16
CI	22.0	2.09 3.34 5.68 8.16 13.0 23.1	0.22 0.65 0.22 0.22 0.22 0.65 0.65	33.6 0.22 0.22 0.22 0.22
Mg mM	11.0	0.17 0.17 0.17 0.17 0.17	0.17 0.50 0.17 0.50 9.27 6.52 9.56 8.04	0.17 0.17 0.17 0.17 17.3
Ca mM	0.364	0.364 0.364 0.364 0.364 0.364 0.364	0.134° 0.126° 0.207° 0.139° 0.431 0.443 0.433	0.364 0.364 0.364 33.3 0.364
Ж ММ	0.04	2.09 3.27 5.31 6.92 8.68 10.13	0.04 0.12 0.04 0.12 0.04 0.12 0.04	0.04 0.04 0.04 0.04 0.04
Na mM	0.28	0.07 0.13 0.43 1.30 4.35 13.0	20.0 20.7 25.9 27.5 0.28 0.85 0.85	33.7 0.28 39.5 0.28 0.28
LC50 mM (>95% CL)	10.9 (9.39–13.0) 11.9 (10.9–13.4)	2.05 (1.89-2.24) 3.23 (2.76-3.59) 5.27 (4.83-5.75) 6.88 (6.29-7.51) 8.64 (8.08-9.26) 10.1 (8.60-18.2)	19.7 (18.2–21.3) 19.8 (18.3–21.5) 25.6 (23.7–27.6) 26.7 (24.8–28.8) 9.10 (8.29–10.0) 6.01 (5.54–6.54) 9.39 (8.73–10.1) 7.53 (6.27–8.92)	33.4 (31.5–35.5) 73.7 (66.2–82.1) 39.2 (36.8–42.0) 32.9 (29.9–36.2) 17.1 (14.3–18.7)
LC50 mg/L (≥95% CL)	1040 (894–1230) 1130 (1040–1280)	153 (141–167) 241 (206–268) 393 (360–429) 513 (469–560) 644 (602–690) 752 (641–1360)	1660 (1531–1790) 1670 (1530–1810) 2150 (1990–2320) 2240 (2080–2420) 767 (699–843) 507 (467–551) 792 (736–853) 635 (529–752)	1950 (1840–2070)
LC50 Method	Probit Probit	Probit Probit Midpoint Midpoint Midpoint Probit	Midpoint Midpoint Probit Midpoint Probit Probit Midpoint Probit	Midpoint Probit Midpoint Probit
Test Water	Low pH ALSW High pH ALSW	1.6 Na ALSW 3 Na ALSW 10 Na ALSW 30 Na ALSW 100 Na ALSW 300 Na ALSW	ALSW Aged 3.0× ALSW Aged ALSW Fresh 3.0× ALSW Fresh ALSW Aged 3.0× ALSW Aged ALSW Aged 3.0× ALSW Fresh 3.0× ALSW Fresh	ALSW ALSW ALSW ALSW ALSW
Culture Water	ALSW	MHRW MHRW MHRW MHRW MHRW MHRW	ALSW ALSW ALSW ALSW ALSW ALSW ALSW ALSW	ALSW ALSW ALSW ALSW ALSW
Test Chemical	MgCl <sub>2</sub> MgCl <sub>2</sub>	KCC KCC KCC KCC KCC	NaHCO <sub>3</sub> NaHCO <sub>3</sub> NaHCO <sub>3</sub> NaHCO <sub>3</sub> NaHCO <sub>3</sub> MgCO <sub>3</sub> MgCO <sub>3</sub> MgCO <sub>3</sub>	NaCl Mannitol NaGluconate CaGluconate MgGluconate
Set #		01	11	12

\*See text for designations and formulations for test and culture waters; horizontal gaps delineate experiments and horizontal lines delineate experimental sets. For median lethal concentrations (LC50s), "Probit" denotes geometric mean of confidence limits; parentheses denote confidence limits; see text for methodology. Except where noted, LC50s and component

concentrations are nominal, and expected to be within +/-10% based on analysis of subset of test solutions.

<sup>b</sup>Ca concentrations are an extrapolation of Ca measurements to median lethal concentration based on model for time dependence of Ca as a function of alkalinity; see Supplemental Data for more information.

<sup>c</sup>Ca concentrations are a weighted average of measurements and 24 and 48 hours that bracket the median lethal concentration.

ALSW = amended Lake Superior water; LC50 = median lethal concentration; MHRW = moderately hard reconstituted water.

sampling from intermediate treatments at the beginning and end of exposure to verify that exposure concentrations were stable over the course of the exposure. Every dilution water was analyzed for all 4 cations plus Cl and SO<sub>4</sub>. Because cation analysis was more time-efficient than anion analysis, exposure concentrations of the salt tested were verified by analyzing for its cation. For tests of the toxicity of CO<sub>3</sub>/HCO<sub>3</sub> salts, which caused oversaturation of CaCO<sub>3</sub> even at the low Ca concentrations in dilution water, Ca concentrations were also measured. In the Ca precipitation experiment with NaHCO<sub>3</sub> and MgCO<sub>3</sub> (set 11), extra test chambers were used to monitor Ca at 0 h, 24 h, and 48 h at the 25%, 50%, and 100% treatment concentrations, in addition to the cation samples normally taken.

For cation analysis, filtered (0.45-µm nylon syringe filter; Grainger) and unfiltered samples were collected in early experiments; but these analyses consistently showed no significant difference between the 2, so later cation samples were not filtered, except for tests with NaHCO3 and MgCO3 in which CaCO3 precipitation was a concern. Samples for cation analysis were acidified by adding 0.2% (v/v) concentrated HNO3 and held at room temperature; for tests with NaHCO3 and MgCO3, this amount of acid was increased by an amount calculated to neutralize the extra alkalinity. Cation measurements were made using an Agilent 240 FS flame atomic absorption spectrophotometer (Agilent Technologies), calibrated with a blank and a series of 5 standards, and verified with a quality control standard and an independent calibration standard at both the beginning and the end of the run.

Concentrations of Cl and SO<sub>4</sub> were determined on samples filtered into a polypropylene centrifuge tube, stored under refrigeration, and analyzed within 28 d. Quantifications were made using a Dionex DX600 Ion Chromatograph with an AS50 autosampler, an LC25 chromatography oven, an ED50 electrochemical detector, and a GP50 gradient pump (Thermo Fisher Scientific). A typical instrument run included a blank, a series of 8 standards, a blank spike, and a quality control standard analyzed at both the beginning and the end of the run. Thirteen percent of all ion analyses were run in duplicate; the average relative percentage difference was 1.6% (maximum 8%, standard deviation 1.6%).

Across all test solutions (i.e., samples other than controls) of the 10 major ion salts for which LC50s were determined, comparison of measured and nominal cation concentrations averaged 98.7% with a standard deviation of 4.9% (n = 329). Individual results ranged from 83% to 116% of nominal, and 95% fell between 90% and 110% of nominal. Conductivity measurements on all treatments confirmed that the intended gradient of exposure existed among samples not sampled for cation analysis. Based on these results, and the complexity of estimating concentrations in test solutions that were not directly measured, nominal exposure concentrations were used for calculating LC50 values.

Analysis of dilution waters yielded a similarly high average agreement between the measured cation concentrations and nominal values based on past reported measurements (99.3%, n=313) but a higher standard deviation of 10.2%; 97% of values fell between 80% and 120% of nominal and 79% of samples fell between 90% and 110% of nominal. This higher variability is associated with the much lower ion concentrations in control waters, which can inflate percentage error values; in the more dilute waters, K in particular was at very low concentrations, sometimes lower than can be confidently quantified by the methods used (e.g.,  $<0.5 \,\text{mg/L}$ ). Wherever measured concentrations deviated more than 20% from nominal

(n=8), all data from that test were reviewed; and in each instance, the information suggested the large deviation was likely spurious (e.g., concentrations of all other ions measured in the same water were close to nominal). Accordingly, ion concentrations in dilution water were also assumed to be equal to nominal in the data analyses, except for Ca concentrations in tests on the toxicity of  ${\rm CO_3/HCO_3}$  salts, in which case Ca was based on measured concentrations. Because these Ca concentrations varied with time and added salt concentration, this involved interpolation of Ca measurements, which is further discussed in the Supplemental Data.

#### Data analysis

Concentration–response curves were extremely steep, so that even with the closely spaced exposure concentrations  $(0.8\times)$ , it was common for there to be only 1 (or no) exposure treatment with partial survival less than that attributable to background (control) mortality. For such tests, continuous concentration–effect models cannot be fit to provide point estimates for LC50s. Consequently, a tiered approach was used, based on the type of calculation the data would support. For each test, a background mortality range was defined as all treatments up to the highest concentration at which the fraction of mortality was no greater than at any lower concentration. The number of treatments with partial mortality (i.e., above this background mortality range and below the lowest treatment with complete mortality) was then determined and used to select the type of analysis.

For tests with at least 2 partial mortalities that increased with concentration, a tolerance distribution analysis was conducted, using a 3-parameter model that included a background survival parameter and assumed a log-normal distribution for the lethal concentrations. Parameters were estimated by maximum likelihood analysis using custom software written with Intel Professional Fortran Composer XE 2011. Of the 149 LC50s reported in the present study, 82 supported calculation of such "probit LC50s." The 95% confidence limits were calculated using the likelihood ratio method [29]. Statistical significance of LC50 differences between treatments within an experiment was assessed based on these confidence limits not overlapping, so that such differences have a significance level of at least 95%.

For tests with insufficient partial mortalities for this probit LC50 analysis, the same likelihood ratio method was used to calculate confidence limits for the LC50 (such confidence limits can be assigned even when a unique point estimate for the LC50 cannot be calculated). For these tests, we assigned the geometric mean of these confidence limits to be the point estimate for the LC50 ("midpoint LC50s"). For cases in which there were no partial mortalities, these confidence limits are the bracketing concentrations (the upper end of the background mortality range and the lowest treatment with complete mortality) and the LC50 is equivalent to linear interpolation of survival versus log concentration between these 2 concentrations. To test the performance of this methodology when tolerance distribution assumptions are met, it was applied to simulated data sets generated based on the range of observed parameters, which demonstrated the bias for LC50 estimation to be <1% and the confidence limits to equal or exceed 95%.

These LC50 calculations were based on the nominal concentration of added salt, without consideration of background ion concentrations, because different ion ratios in the dilution water and the added salt make it impossible to express total ion concentrations as an equivalent concentration of salt. Initial comparisons of how dilution water differences affect salt toxicity are also on the basis of added salt for the same reason.

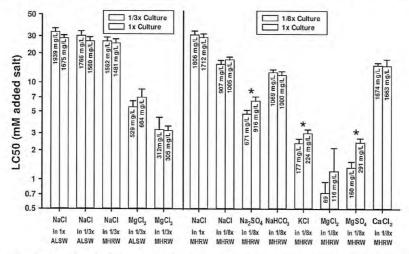


Figure 1. Effects of culturing and testing organisms in different strengths of amended Lake Superior water and moderately hard reconstituted water on median lethal concentrations for selected major ion salts to *Ceriodaphnia dubia*. Error bars denote upper 95% confidence limits, and asterisks denote where confidence limits do not overlap. ALSW = amended Lake Superior water; LC50 = 48-h median lethal concentration; MHRW = moderately hard reconstituted water.

Background concentrations of the component ions of the test salt were <5% in all but 6 tests and never >10%; these had negligible impact on comparisons based on added salt except as noted in *Results and Discussion*. However, for further analyses of the aggregated data, exposure metrics included both background and added ions.

Although test preparation and chemical analysis were based on weight of the salts and the various ions, this is not a good basis for comparing the relative toxicity of the individual salts and ions because the toxic action of these ions should be related to their molarity, not their mass. As such, salt and ion concentrations were converted to molarity, which is the primary basis for toxicity comparisons of the added salts. In addition, evaluation of the dosimetry at these elevated ion concentrations should consider formation of complexes between the various cations and anions, which would affect their chemical reactivities and thus their toxicities. It should also consider that the high ionic strength of these solutions will reduce the reactivity of the ions, as represented in lower activity coefficients, and that exposure metrics thus would be best expressed as chemical activity rather than molarity. To this end, the ion composition at each LC50 was analyzed using the chemical speciation program Visual MINTEQ (Ver 3.0) to estimate chemical speciation and the chemical activity of each chemical species. The osmolarity of each LC50 solution was also calculated based on these MINTEQ activity estimates using the method described by Robinson and Stokes [30].

# RESULTS AND DISCUSSION

For each test (*n* = 149), added-salt LC50 values with confidence limits are provided in Table 3, as both milligrams per liter and millimolarity, along with total ion concentrations and osmolarities at these LC50s. The pH associated with each LC50 at the end of exposure is also provided based on measurements in treatments bracketing the LC50. All reported tests had conductivity measurements that varied across treatments in a manner consistent with the intended exposure concentrations, and LC50s directly based on these conductivities are also provided in Table 3. Dissolved oxygen was always above 7.5 mg/L, and temperature was always within 1 °C of 25 °C. Measurements made at the beginning and end of

experiments indicated that evapoconcentration during the experiment was low (<5%). Over the background mortality range, as defined in *Data analysis*, survival was  $\geq$ 90% in all toxicity tests and averaged 98% overall.

Effects of acclimation to dilution water

Figure 1 compares LC50s for *C. dubia* cultured in dilute solutions to those from our standard culture waters. Culturing organisms in 1/3× ALSW or 1/3× MHRW had no significant effect on the toxicities of NaCl and MgCl<sub>2</sub>, either in test waters at these dilutions or (for NaCl only) in 1× ALSW, the LC50 estimates differing by 20% or less. Culturing organisms in 1/8× MHRW also had no effect on the toxicities of NaCl, NaHCO<sub>3</sub>, and CaCl<sub>2</sub>, the LC50s differing by no more than 10%. However, for Na<sub>2</sub>SO<sub>4</sub>, KCl, MgCl<sub>2</sub>, and MgSO<sub>4</sub>, LC50s for 1/8× MHRW test water are 20% to 45% lower for organisms cultured in 1/8× MHRW than those cultured in 1× MHRW, these differences being statistically significant except for MgCl<sub>2</sub>, for which the LC50 is uncertain because of highly variable responses in this dilute test water.

Thus, "acclimation" to 1/8× MHRW resulted in *C. dubia* being more, rather than less, sensitive to some salts when tested in this dilute solution. Although both we and Elphick et al. [19] established successful cultures in this water, based on our experience with *C. dubia*, such a dilute solution appears to be stressful, an indication of this being the greater variability of organism response to salt toxicity. It is not clear whether this should be considered an artificial stress created by manipulating organisms long adapted to laboratory waters with higher ion concentrations, or a natural consequence for a species not adapted to and not endemic to such dilute waters.

These acclimation experiments were prompted by the work of Elphick et al. [19], who acclimated cultures to dilutions waters (for 2 generations or more) and found that, relative to  $1 \times MHRW$ , the 7-d NaCl LC50 for *C. dubia* decreased by 2.1-fold in  $1/2 \times MHRW$ , 3.6-fold in  $1/4 \times MHRW$ , and 8.4-fold in  $1/8 \times MHRW$ . In contrast, for organisms both cultured and tested in the waters, we found only a 9% decrease in the NaCl LC50 between  $1 \times MHRW$  and  $1/3 \times MHRW$  and only a 1.9-fold decrease between MHRW and  $1/8 \times MHRW$  (Figure 1). The lack of culture water effect on our LC50s for NaCl suggests that these differences between the 2 studies resulted from other

factors, such as the longer duration in Elphick et al. [19] or a variability associated with culturing and/or testing *C. dubia* in such dilute waters. More recent communication with J. Elphick has indicated that they have observed variability in NaCl LC50s for *C. dubia* in very dilute test waters (D.R. Mount, personal communication).

Overall, we concluded that the absence of dilution water-specific culturing was not a substantial factor for most of our experiments but that data for the 1/8× MHRW waters should be interpreted with caution. The results presented hereafter will be only for organisms from our standard culture waters and will consider LC50s for the most dilute test waters to be more useful for qualitative insights into ion toxicity rather than establishing quantitative relationships. Any quantitative uncertainties of LC50s for such dilute waters are probably of limited practical concern because few unimpacted surface waters will likely experience elevations of a single salt to toxic levels while other ions remain at such low levels.

Our culture experiments only addressed more dilute waters than our standard culture water because discrepancies of our results from that of Elphick et al. [19] were noted for those waters. Although we did not evaluate effects of culturing at higher ion concentrations, the relationship of our NaCl LC50s to Ca concentration shows good agreement with that of Soucek et al. [18] and Elphick et al. [19], who did culture organisms in their different test waters. Also, Soucek and Kennedy [16] reported only small and statistically nonsignificant differences among Na<sub>2</sub>SO<sub>4</sub> LC50s in MHRW for *C. dubia* cultured in MHRW versus MHRW with elevated Na<sub>2</sub>SO<sub>4</sub> levels. This suggests that acclimation to higher ion concentrations may not be a significant factor, but more study would be needed to reach a definitive conclusion.

#### Salt toxicity in MHRW versus ALSW

Figure 2 compares LC50s for 10 major ion salts tested in both ALSW and MHRW. For NaCl, MgCl<sub>2</sub>, CaCl<sub>2</sub>, Na<sub>2</sub>SO<sub>4</sub>, MgSO<sub>4</sub>, and MgCO<sub>3</sub>, LC50 differences are <15% and not statistically significant. For NaHCO<sub>3</sub>, the LC50 is 20% lower for MHRW and is statistically significant; however, this test involved significant precipitation of Ca, and this precipitation was more pronounced in the MHRW test (Table 3). Given previous studies on the effect of hardness on salt toxicity [16–21], such a Ca difference would contribute to the

observed LC50 difference. The fact that MHRW provides a higher background Na concentration and alkalinity would also contribute to the difference in LC50s expressed in terms of added NaHCO<sub>3</sub>.

Only for the 3 K salts are there clear, substantial differences between the 2 dilution waters. The LC50s are 32% to 67% higher for MHRW than for ALSW, and these differences were all statistically significant. Notable compositional differences between these waters (Table 1) include 4-fold higher Na in MHRW (because of the use of NaHCO<sub>3</sub> to add alkalinity), 3-fold higher Mg in MHRW (but similar Ca), 5-fold higher SO<sub>4</sub>, and 3-fold lower Cl. The higher Na for MHRW is of particular interest because Na and K are linked physiologically through the central role of Na K adenosine triphosphatases in ion regulation and other key cellular processes [2]. Additional information regarding this issue is provided below (see *Effects of sodium on potassium toxicity*).

Comparing the LC50s in Figure 2 across salts also supports some inferences regarding the contributions of individual ions to toxicity. On a total molarity basis, the Na salts are significantly less toxic than salts of other cations with the same anion. On a total molarity basis NaCl is also significantly less toxic than the other Na salts. For Na<sub>2</sub>SO<sub>4</sub>, the greater toxicity could be caused by it containing 2, instead of 1, Na atoms, in addition to any anion effects. For NaHCO<sub>3</sub>, the greater toxicity should be partly the result of lower Ca concentrations (Table 3) from the CaCO<sub>3</sub> precipitation induced by this salt. These differences among Na salts will be further addressed below (see Effects of calcium on the toxicities of sodium salts).

The K salts are much more toxic than the corresponding salts with other cations, especially Na (Figure 2). The ratio of the Na salt LC50 to the K salt LC50 ranges across the different anions from 4.2 to 6.6 for MHRW and from 5.8 to 9.1 for ALSW, with smaller ratios for MHRW resulting from the aforementioned dilution water effect on K toxicity. These large ratios indicate that K is the principal source of toxicity for these salts because the anion concentrations at the LC50s for K salts are in all cases a small fraction of those present at the LC50s for the Na salts. The LC50 on a molarity basis for K<sub>2</sub>SO<sub>4</sub> is approximately half that of KCl, consistent with toxicity being related to the molarity of K rather than the molarity of the salt. Factors controlling K salt toxicity are explored further below (see *Effects of sodium on potassium toxicity*).

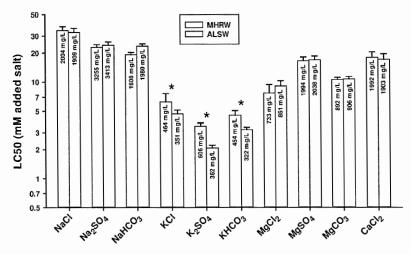


Figure 2. Median lethal concentrations for major ion salts to *Ceriodaphnia dubia* in moderately hard reconstituted water and amended Lake Superior water. Error bars denote upper 95% confidence limits, and asterisks denote where confidence limits do not overlap. ALSW = amended Lake Superior water; LC50 = 48-h median lethal concentration; MHRW = moderately hard reconstituted water.

Although less toxic than the K salts in these test waters, the Mg salts are also significantly more toxic than their corresponding Na salts (ratios of 1.4–4.4; Figure 2), suggesting that, like K, Mg is also more important to the toxicity of these salts than are the anions. On a total molarity basis, MgSO<sub>4</sub> was approximately 2-fold less toxic than MgCl<sub>2</sub>. Given the same Mg stoichiometry in each salt, this suggests that the toxicity of Mg is differentially modified by the anions, perhaps by SO<sub>4</sub> complexing Mg and thus reducing its toxicity. Unlike for NaHCO<sub>3</sub>, there was no Ca precipitation in the tests with MgCO<sub>3</sub> (Table 3; and see Supplemental Data for more information on Ca precipitation relationships), and MgCO<sub>3</sub> was only slightly less toxic than MgCl<sub>2</sub>. The relationship of these toxicity differences among Mg salts to ion speciation and activity is discussed later (see Effects of calcium on the toxicities of magnesium salts).

For Ca, only the Cl salt was acutely toxic below its solubility. On a total molarity basis, CaCl<sub>2</sub> is more toxic (by 1.9 fold) than NaCl (Figure 2); however, because CaCl<sub>2</sub> has twice as many chlorides as NaCl, this result by itself provides no clear evidence regarding relative ion toxicities.

#### Salt toxicity in different strengths of ALSW

Figure 3 shows the effect of varying all ions in dilution water by comparing LC50s for  $1/3 \times$ ,  $1 \times$ , and  $3 \times$  ALSW. These comparisons were developed by combining data from simultaneous tests in  $1/3 \times$  and  $3 \times$  ALSW with the  $1 \times$  ALSW data generated separately (Figure 2). Also, NaCl and Na<sub>2</sub>SO<sub>4</sub> were evaluated in an additional experiment with all 3 waters tested simultaneously (marked "dup" in Figure 3); this was done to determine if combining data across experiments might influence conclusions. For the  $3 \times$  ALSW water, ions in the dilution water equaled between 3% and 8% of the added salt LC50, so that any amelioration of toxicity in this water would be slightly underestimated when based just on the added salt.

For the K salts, there is a 2.3-fold to 2.5-fold increase in LC50s from  $1/3 \times$  to  $3 \times$  ALSW (Figure 3), with the LC50 for  $1 \times$  ALSW being near the midpoint of these ranges. Because all ions varied proportionately between these dilution waters, the specific factors responsible for these differences are not directly identifiable. However, the decreased toxicity from  $1 \times$  to  $3 \times$ 

ALSW (entailing a 3-fold change in both Na and hardness) is similar to the change between ALSW and MHRW (Figure 2), between which Na differs 4-fold but hardness is just 1.5-fold different and Ca is virtually constant, suggesting that Na is more important than other cations to these toxicity differences.

For the Mg salts, there are substantial increases in the LC50s from 1/3× to 3× ALSW, totaling 3.4-fold, 2.5-fold, and 1.8-fold for MgCl<sub>2</sub>, MgCO<sub>3</sub>, and MgSO<sub>4</sub>, respectively (Figure 3). For MgCO<sub>3</sub>, the lack of difference between 1× and 3× ALSW is associated with substantial precipitation of CaCO<sub>3</sub> in 3× ALSW (Table 3). For MgCl<sub>2</sub> and MgSO<sub>4</sub>, the LC50 increases between 1× and 3× ALSW are similar to the corresponding K salts; but because these Mg salts show no LC50 difference between ALSW and MHRW whereas the K salts do, the underlying reasons must be different. These effects of dilution water strength should reflect the effects of the anions on the speciation of Mg and Ca; whether such consideration of speciation accounts well for LC50 differences among dilution waters, and among the salts, is addressed later.

For the Na salts, LC50s show responses to ALSW strength that are different from each other and from the K and Mg salts (Figure 3). The LC50 for Na<sub>2</sub>SO<sub>4</sub> increases by 1.8-fold from  $1/3 \times$  to  $3 \times$  ALSW in both replicate tests, similar to MgSO<sub>4</sub> and  $K_2SO_4$ ; but the increase is all between  $1/3 \times$  to  $1 \times$  ALSW. The LC50 for NaCl increases much less from 1/3× to 3× ALSW (1.2-fold and 1.4-fold in the different replicates). For NaHCO<sub>3</sub>, there is a nonsignificant but slight decrease of the LC50 from  $1/3 \times$  to  $3 \times$  ALSW. This is again associated with precipitation of Ca, this precipitation being more extensive in the higher ALSW strengths, leading to Ca concentrations actually being similar across these waters (Table 3) despite the original formulations being 9-fold different. Possible reasons for these LC50 differences across Na salts, including chemical speciation and the ameliorative effects of Ca, are further addressed below (see Effects of calcium on the toxicities of sodium salts).

For CaCl<sub>2</sub>, there is an increase of only 1.2-fold in the LC50 from  $1/3 \times$  to  $3 \times$  ALSW of marginal statistical significance. This small dependence of Ca toxicity on ALSW strength, compared with the large dependence of Mg toxicity, results in the toxicity of MgCl<sub>2</sub> being very similar to CaCl<sub>2</sub> in  $3 \times$  ALSW (a factor of 1.3 difference) and very different in  $1/3 \times$  ALSW (a

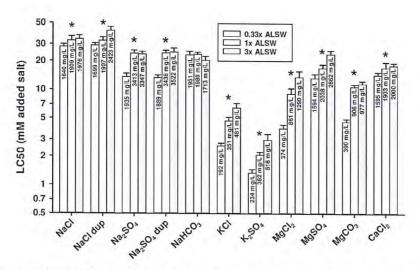


Figure 3. Median lethal concentrations for selected major ion salts to  $Ceriodaphnia\ dubia$  in different strengths of amended Lake Superior water (ALSW). Error bars denote upper 95% confidence limits, and asterisks denote where confidence limits do not overlap for  $0.33 \times$  and  $3 \times$  ALSW. LC50 = 48-h median lethal concentration.

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factor of 3.7 difference). To the extent that the toxicity of MgCl<sub>2</sub> (or another salt) depends on Ca, judging relative potencies is best done at high, but nontoxic, Ca concentrations because any adverse consequences of low Ca cannot, by definition, affect Ca toxicity. Thus, Ca and Mg should be considered to have similar intrinsic toxicities.

Effects of manipulating specific ion ratios

To evaluate the factors behind the responses in the  $1/3 \times$ versus 3× ALSW and MHRW versus ALSW, 3 sets of experiments were conducted in which the Ca:Mg, Cl:SO4, and Na:K ratios were manipulated whereas other ions remained constant. The LC50s for MgCl2, Na2SO4, and MgSO<sub>4</sub> are at least 2-fold greater at high Ca:Mg than low (Figure 4). Because total hardness and all other ion concentrations were the same in high Ca:Mg and low Ca: Mg waters, these differences are presumably attributable to Ca, rather than overall hardness. These same salts have large LC50 differences in the  $1/3 \times$  versus  $3 \times$  ALSW comparisons (Figure 3), for which Ca also changed substantially (in concert with other ions), but small LC50 differences for the MHRW versus ALSW comparisons (Figure 2), for which Ca was essentially the same but other ions varied markedly. Overall, this suggests that Ca is the important factor in differences among these various dilution waters for these salts. The LC50s of NaCl and NaHCO3 had smaller (20-30%) increases at high Ca:Mg (Figure 4), but the magnitude of these effects is also consistent with the results shown in Figures 2 and 3 and their associated Ca concentrations (Table 3). In contrast, although KCl has a higher (15%) LC50 at high Ca:Mg, this is not statistically significant and is much smaller than the LC50 differences for both the MHRW and ALSW comparison (Figure 2) and the  $1/3 \times$  to  $3 \times$  ALSW comparison (Figure 3) for this salt. This reinforces the earlier suggestion of the importance of Na to K salt toxicity (see Salt toxicity in MHRW versus ALSW).

When the  $\text{Cl:SO}_4$  ratio was altered (Figure 4), no significant differences in LC50s (<15%) exist for any of the tested salts (Table 2), indicating that these anions are not important characteristics of dilution water for major ion toxicity to *C. dubia*. When the Na:K ratio was altered (Table 3), there also are no significant effects on the LC50s of the tested salts. No K

salts were included in Na:K ratio studies because the effect of Na on K salt toxicity was evaluated separately.

Effects of manipulating alkalinity and pH

Two sets of toxicity tests evaluated the effects of dilution water alkalinity and pH on salt toxicity (Figure 5). In the first set, alkalinity was decreased (by dilution, then restoring other ions) and increased (by NaHCO3 addition) relative to 1× ALSW water, resulting in pHs of approximately 7.4 and 8.3 in the lowalkalinity and high-alkalinity treatments, respectively, compared with a typical pH of approximately 7.9 in ALSW. The effects of these combined changes in alkalinity and pH on LC50s are <10% and not statistically significant for NaCl, Na<sub>2</sub>SO<sub>4</sub>, and MgSO<sub>4</sub>, but are larger and statistically significant for MgCl2 and KCl. For MgCl2, increasing the alkalinity decreased the LC50 by 1.4-fold. This test was repeated, yielding virtually identical results (Table 3). We initially speculated that complexation by the higher HCO<sub>3</sub>/CO<sub>3</sub> decreased Ca activity and thus increased toxicity. However, to maintain a constant Na concentration in these tests, the low-alkalinity water had higher SO<sub>4</sub>, which also complexes Ca. As a result, the calculated Ca activity in the high-alkalinity treatment is actually higher than that in the low-alkalinity treatment, so it would not explain the observed LC50 shift. We similarly have no explanation for the lower LC50 (20%) for KCl at high alkalinity. One area of needed future work is direct ion activity measurements to verify speciation model calculations before further exploring the cause of these effects.

In the second set of tests, pH was manipulated by CO<sub>2</sub> partial pressure, leaving alkalinity and all other ions constant as pH varied. For this experiment, pH had no effect on NaCl toxicity over the pH range 6.75 to 8.20 or on MgCl<sub>2</sub> toxicity over the pH range 6.75 to 8.50 (Figure 5). (The high pH treatment for the NaCl tests had lower pH than for the MgCl<sub>2</sub> tests because of an air leak in the chamber for the low CO<sub>2</sub> treatment, so that the range of tested pHs was less than intended.) This insensitivity of toxicity to pH manipulation suggests that something other than pH itself is responsible for the effects of altered alkalinity on MgCl<sub>2</sub> toxicity noted in the previous experiment. However, although these tests at different pHs and alkalinities indicate limited or no effect of these factors on the toxicity of SO<sub>4</sub> and Cl salts, they do not

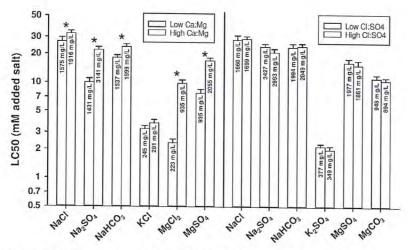


Figure 4. Median lethal concentrations for selected major ion salts to  $Ceriodaphnia\ dubia$  in amended Lake Superior water modified to have different ratios of Ca to Mg and different ratios of Cl to SO<sub>4</sub>. Error bars denote upper 95% confidence limits, and asterisks denote where confidence limits do not overlap. LC50 = 48-h median lethal concentration.

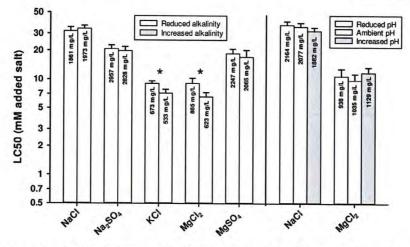


Figure 5. Median lethal concentrations for selected major ion salts to *Ceriodaphnia dubia* in amended Lake Superior water modified to have different alkalinities and pH or different pH with alkalinity unchanged. Error bars denote upper 95% confidence limits, and asterisks denote where confidence limits do not overlap. LC50 = 48-h median lethal concentration.

encompass the much higher pHs and buffering levels of the tests with HCO<sub>3</sub>/CO<sub>3</sub> salts. Therefore, pH might still have a role in the relative toxicity of those salts.

#### Effects of sodium on potassium toxicity

To evaluate the suspected effect of Na on the toxicity of K salts, 6 dilution waters were prepared, each having the composition of ALSW except that the Na $^+$  concentrations were varied to be 0.070 mM (background in LSW), 0.13 mM, 0.43 mM, 1.3 mM, 4.3 mM, and 13 mM (1.62 mg/L, 3 mg/L, 10 mg/L, 30 mg/L, 100 mg/L, and 300 mg/L), added as NaCl so that the anion was the same as for the toxicant, KCl. The LC50 of KCl differs consistently and substantially across these dilution waters (Figure 6), from a low of 2.05 mM (153 mg/L) at the lowest Na to a high of 10.1 mM (752 mg/L) at the highest Na. The magnitudes of these changes are consistent with the behavior observed for KCl in the MHRW versus ALSW and 1/3× versus 3× ALSW experiments (Figures 2 and 3 and included in Figure 6).

Mount et al. [3] and our initial experiments (Figure 2) demonstrated that K is the most toxic of the major ions to C. dubia and, as a result, the effects of K seem to greatly exceed any

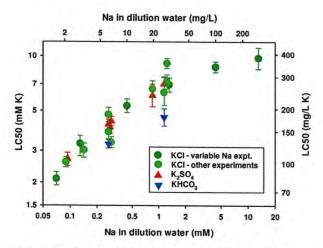


Figure 6. Effect of Na on median lethal concentrations for K salts to *Ceriodaphnia dubia*. Error bars denote 95% confidence limits. LC50 = 48-h median lethal concentration.

effect of the different anions. This is demonstrated in Figure 6, which shows agreement of other K salts and dilution waters with the experiment regarding KCl toxicity across different NaCl levels. Note that by compiling data for tests conducted across a variety of dilution water compositions, the correlation of Na with other major ions that might exist in a single experiment is reduced, strengthening the case for Na being the primary factor influencing toxicity of K salts. There might be some lesser effects of other factors, as evidenced by slightly lower LC50s (expressed as K) for KHCO<sub>3</sub> than for KCl and K<sub>2</sub>SO<sub>4</sub> (Figures 2 and 6). As noted previously, the carbonate tests entailed much higher pH and buffering for which effects are uncertain. However, any such effects appear to be of less consequence than the primary effect of Na on K toxicity.

One implication of the Na dependence of K toxicity demonstrated in the present study is that the high toxicity of K in typical test waters with low Na may be of limited field relevance because K would be unlikely to be present at toxic concentrations when Na is so low. For example, the K-dominated effluent evaluated by Jop and Askew [11] had approximately 1.5 mM Na, which would put the expected acute LC50 for K at 10 mM or higher (Figure 6). This is similar to the toxicity of Mg salts and only 2-fold to 3-fold more toxic than Na salts.

## Effects of calcium on the toxicities of sodium salts

Aside from the effect of Na on the toxicity of K, Ca is the only other ion in the present study which exerted a substantial influence on the acute toxicity of major ion salts to C. dubia. Several studies have shown that hardness influences major ion toxicity to C. dubia [16–20], though the way in which hardness was manipulated differed across studies. Experiments by Soucek et al. [16-18] varied hardness by adding CaCl<sub>2</sub> + MgCl<sub>2</sub> (for NaCl toxicity tests) or CaSO<sub>4</sub>+MgSO<sub>4</sub> (for Na<sub>2</sub>SO<sub>4</sub> toxicity tests) in fixed ratios and keeping the remaining ions constant, whereas Elphick et al. [19,20] varied all ions proportionately (as in the  $1/8\times$ ,  $1/3\times$ , and  $1\times$  MHRW waters used in the present study). In the present experiments, the combination of manipulating all ions simultaneously in some tests and only specific ion pairs in others (at constant hardness) allowed us to conclude that the "hardness effect" on the toxicity of Na salts is primarily an effect of Ca, rather than total hardness or the other ions that covary with hardness. The same conclusion was reached by Davies and Hall [21], who manipulated both

total hardness and the Ca:Mg ratio to show the dominance of Ca in determining toxicity of Na<sub>2</sub>SO<sub>4</sub> to *D. magna*.

Figure 7 shows the relationship of LC50s to Ca for all our Na salt toxicity tests using organisms cultured in  $1\times$  ALSW or MHRW. Each panel considers different exposure metrics for how LC50 and Ca are expressed. As a comparative performance measure for these different metrics, residual standard deviations of the logLC50s (RSD<sub>log</sub>) around the mean trend of logLC50 with Ca are provided. Mean data trends were calculated by least-square regression of logLC50 versus logCa using Sigmaplot with a model for an exponential rise to a maximum. This is not intended to provide a definitive model for the relationship of toxicity to Ca but rather only an empirical description of the mean data trend suitable for calculating the RSD<sub>log</sub>.

In Figure 7A, LC50s are plotted as millimolarity of added salt and Ca is plotted as total concentration (measured Ca for NaHCO<sub>3</sub> tests, nominal for others). Although the 3 inorganic salts have a similar and substantial (2-fold to 3-fold) dependence

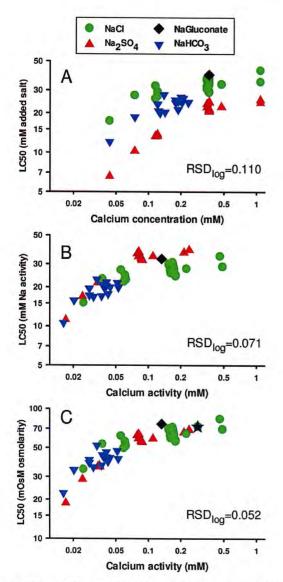


Figure 7. Effect of Ca on 48-h median lethal concentrations (LC50s) for Na salts to *Ceriodaphnia dubia* using different exposure metrics. (A) Metrics are LC50 as concentration of added salt and Ca concentration. (B) Metrics are Na and Ca activities. (C) The LC50 metric is changed to osmolarity. RSD<sub>log</sub> denotes the residual standard deviation of log(LC50)s from their mean trend with Ca.

on Ca, there is an apparent difference in the relative toxicities of the salts, with LC50s being ordered NaCl  $\approx$  Na gluconate > NaHCO<sub>3</sub> > Na<sub>2</sub>SO<sub>4</sub>. The RSD<sub>log</sub> is 0.110, for which  $\pm 2$  SDs corresponds to nearly a 3-fold range in LC50 at a fixed Ca concentration, indicative of the substantial variability among the different salts.

Because chemical reactions are governed by chemical activity rather than concentration, the relative toxicities of salts and their relationships to Ca should be based on chemical activities rather than total concentrations, as in Figure 7A. Activities will differ from total concentrations because of formation of chemical complexes as well as reduced activity coefficients associated with high ion concentrations (especially for the divalent Ca ion), and these factors will differ among the salts and dilution waters. Another issue for comparing toxicities across these salts is that, per mole of salt, Na<sub>2</sub>SO<sub>4</sub> provides twice as many Na ions and 1.5 times as many total ions as do the other salts. To address these issues, in Figure 7B, Ca is plotted as Ca activity and the LC50s are based on the activity of Na. This results in much closer agreement among the salts, with the  $RSD_{log}$  reduced to 0.071 and the  $\pm 2$  SD range for LC50 reduced to less than 2-fold; NaCl and NaHCO3 now show similar toxicity because complexation of Ca by CO<sub>3</sub> and HCO<sub>3</sub> in the NaHCO<sub>3</sub> exposures causes Ca activity to be lower than that in NaCl exposures at the same total Ca concentration. Accounting for Na<sub>2</sub>SO<sub>4</sub> having twice as many Na atoms as other Na salts has also made its LC50s more similar to the other salts; however, Na<sub>2</sub>SO<sub>4</sub> now appears to be less, rather than more, toxic than NaCl, based on the repeated tests with Na<sub>2</sub>SO<sub>4</sub> and NaCl in 1× ALSW and other waters with similar Ca concentration (i.e., the cluster of red triangles near 0.1 mM Ca activity and green circles near 0.2 mM Ca activity). This continued discrepancy between Na<sub>2</sub>SO<sub>4</sub> and NaCl suggests that the anions play some role in determining toxicity beyond their effects on cation speciation.

A simple metric for expressing the aggregate effect of Na and anions is osmolarity, which reflects the combined, unweighted influence of all dissolved species. Figure 7C provides LC50s on the basis of calculated osmolarity, resulting in excellent agreement among all the tests of Na salt toxicity. The LC50s for the different salts now overlap within experimental variability; the RSD<sub>log</sub> is reduced to 0.052 and the  $\pm 2$  SD range to 1.6-fold. Further support for using osmolarity as an exposure metric comes from the test conducted with mannitol (black star in Figure 7C), whose LC50 is not significantly different from those of the Na salts even though its only expected effect would be on osmolarity. While this case for osmolarity is simply correlative and does not establish osmotic potential as the primary stressor, an exposure metric such as osmolarity that includes multiple ions in a nonspecific manner is indicated by these data.

## Effects of calcium on magnesium toxicity

Figure 8 examines the relationship of LC50s to Ca for all our Mg salt toxicity tests using organisms cultured in  $1 \times$  ALSW or MHRW. The panels address different exposure metrics parallel to those examined in Figure 7 for Na salts and include RSD<sub>log</sub> as a performance measure for these different expressions of exposure.

In Figure 8A, LC50s are plotted as millimolarity of added salt and Ca is plotted as total concentration (measured Ca as appropriate for MgCO $_3$  tests, nominal for others). There is an even greater effect of Ca ( $\sim$  10-fold LC50 range over the 0.05–1.0 mM Ca range) for Mg salts than for Na salts (note scale change from Figure 7). On the basis of total added salt

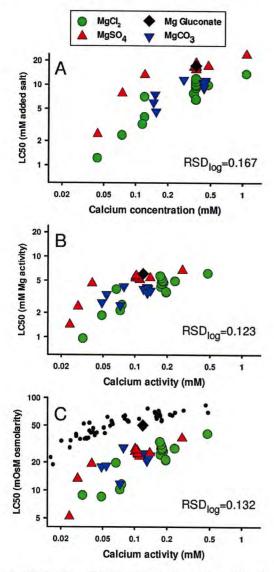


Figure 8. Effect of Ca on 48-h median lethal concentrations (LC50s) for Mg salts to *Ceriodaphnia dubia* using different exposure metrics. (A) Metrics are LC50 as concentration of added salt and Ca concentration. (B) Metrics are Mg and Ca activities. (C) The LC50 metric is changed to osmolarity, and black dots provide comparison to Na salt data from Figure 7C. RSD<sub>log</sub> denotes the residual standard deviation of log(LC50)s from their mean trend with Ca.

concentrations, the salts have different apparent toxicities, with MgSO<sub>4</sub> and Mg gluconate being approximately 2-fold to 3-fold less toxic than MgCl<sub>2</sub> and MgCO<sub>3</sub>. These differences make the RSD<sub>log</sub> (0.167) even larger than for Na salts (Figure 7A), corresponding to a  $\pm 2$  SD range of almost 5-fold at a fixed Ca concentration.

Expressing exposure on the basis of chemical activities of Mg and Ca (Figure 8B) reduces the residual variability and the differences between the salts because of the different degrees of complexation of both Ca and Mg by the different anions. The total range of the LC50s is also less because of a concentration dependence for complexation and activity coefficients. However, the closer agreement among the data is mainly for Ca activity >0.05 mM; at lower Ca activity, there remains considerable disparity between MgSO<sub>4</sub> and MgCl<sub>2</sub>. As noted earlier (see *Effects of acclimation to dilution water*), these tests in very dilute test water raised some concerns about variable response and stresses associated with low ions, which might

involve effects other than from Mg and Ca. Thus, although basing LC50s on Mg activity does provide a better metric than total Mg concentration, it does not account for all factors of significance. For all the data, the RSD $_{\rm log}$  is reduced to 0.123; but if only the data at Ca activity >0.05 mM are considered, the RSD $_{\rm log}$  is 0.102 and the  $\pm 2$  SD range is then 2.5-fold, half of what is was for LC50s based on total added salt concentration.

Although osmolarity provides a possible unifying exposure metric for Na salts (Figure 7C), it is not a useful metric for Mg salt toxicity (Figure 8C). Using osmolarity to express Mg salt toxicity did not improve, but rather increased, the RSD<sub>log</sub> of the data. More importantly, the osmolarities at the LC50 for Mg salts are 2-fold to 4-fold lower than those for Na salts at the same Ca activity. This indicates specific action of Mg and is incompatible with using a nonspecific, total ion metric such as osmolarity for the toxicity of Mg salts.

Our results indicate a strong effect of Ca on MgSO<sub>4</sub> and MgCl<sub>2</sub> toxicity, nearly a 10-fold change in LC50s from the Ca concentration in  $1/8 \times$  MHRW (0.044 mM) to that in  $1 \times$ MHRW (0.35 mM), with Figure 1 indicating even lower LC50s at 1/8× MHRW if organisms are cultured in this water. This is in contrast to results from van Dam et al. [31] for an Australian cladoceran (Moinodaphnia macleayi) collected from and cultured in very low ionic strength water (Ca < 0.02 mM). In 6-d to 7-d reproductions tests, the MgSO<sub>4</sub> concentration reducing reproduction by 50% for M. macleayi was 2.6 mM in the culture water but only 2-fold higher (5.0 mM) when CaSO<sub>4</sub> was added to increase the Ca concentration by roughly 20-fold, to 0.34 mM. While there are several possible explanations for the difference in Ca effect for the 2 species, the van Dam et al. [31] study does reinforce concerns that species like C. dubia tested at very low ionic strength may not respond the same way as species (or strains) naturally adapted to low ionic strength water.

#### The toxicity of calcium

Because of solubility constraints, only limited testing of Ca salts could be conducted; and conclusions about any specific Ca toxicity could not be made based on simple comparisons of the toxicities of CaCl<sub>2</sub> and NaCl in our standard test waters. However, there are some indications that the toxicity of CaCl<sub>2</sub> is dominated by the cation, as are the toxicities of K and Mg salts. First, any comparison of the toxicity of CaCl<sub>2</sub> to NaCl should be made for NaCl tests in the dilution waters with the highest Ca. Otherwise, the comparison is confounded by the exacerbation of Na toxicity by low Ca. For the CaCl2 tests (Table 3), osmolarities at the LC50s average 45 milliosmoles/L (range, 39-52), well below the average osmolarity of 76 milliosmoles/L (range, 69–83) for the NaCl tests in 3× ALSW. Unless the Cl from these 2 salts acts differently, this indicates some Caspecific toxicity beyond the toxicity exerted by Na and the various anions (Figure 7). Second, in ALSW, the calculated Ca activity is 7.5 mM for CaCl2 and 8.1 mM for Ca gluconate; such similarity would not be expected if these disparate anions were contributing to toxicity. However, the conclusion that there is some specific toxicity of Ca is tentative and will be further addressed in the next article in this series.

#### Relationship of toxicity to conductivity

Figure 9 replots the data from Figures 7 and 8 with LC50s based on conductivity. Given the results already presented for the toxicity of different salts, it is not surprising that these conductivity LC50s also show a strong dependence of toxicity on Ca and substantial variation across the different salts. The



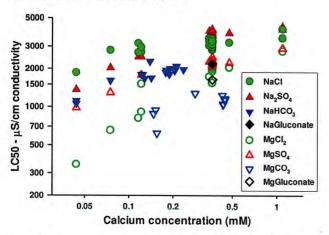


Figure 9. Median lethal concentration of Na and Mg salts expressed as conductivity. LC50 = 48-h median lethal concentration.

total range of conductivity LC50s is approximately 10-fold, with up to a 5-fold range among salts at the same Ca concentration. This indicates that assessment approaches using conductivity as an exposure metric should have a restricted scope of applicability regarding relative ion concentrations, as is the case for the conductivity benchmark of Cormier et al. [14,15].

#### SUMMARY AND IMPLICATIONS

In the present study, the key influences of background water chemistry on the acute toxicity of major ions salts to C. dubia were the effect of Na on toxicity of K salts and the effects of Ca on the toxicity of Na and Mg salts. Although the practical implications of the Na effect on K toxicity are limited because of the unlikelihood of K-dominated exposures, this result does mean that when K exposures are of concern [11] effects concentrations based on typical laboratory tests with low Na concentrations would overestimate risk. Regarding the Ca effects, although a hardness dependence of toxicity was already known for some Na and Mg salts [16-20] and Davies and Hall [21] implicated Ca as the specific factor for Na<sub>2</sub>SO<sub>4</sub> toxicity, our results more thoroughly establish a role of Ca for a variety of salts. Predicting salt toxicity across waters will thus be more accurate if done on the basis of Ca rather than hardness, but the benefits of accuracy must be weighed against the availability of Ca versus hardness data. Uncertainties associated with using hardness as a surrogate for Ca can be addressed based on the regional variability of the Ca:Mg ratio, the resultant uncertainty on Ca, and the consequent uncertainty on toxicity estimated from Ca (e.g., Figures 7 and 8); in many cases, the resultant uncertainty should be limited.

An important tool in addressing the toxicity of ion mixtures has been the model of Mount et al. [3], and part of the purpose of the present study was to address some limitations of that model. The present results do indicate that not addressing the hardness (or Ca specifically) of the background water can introduce uncertainty into this model and raise questions regarding how joint toxicity across the different ions was addressed by Mount et al. [3]. However, any quantitative evaluation of that model relative to our results must wait for the alternative model to be presented in our subsequent article.

In addition to describing the dependence of major ion salt toxicities on background water chemistry, the results of the present study allowed some inferences about mechanisms and some conclusions about how salt toxicity dosimetry should be expressed. At least 3 separate mechanisms were apparent. First, a K-related mechanism is indicated by the high toxicities of K salts, the dependence of this toxicity on Na concentration, and its good correlation to K concentration. Second, a Mg-related mechanism is indicated based on the greater toxicity of Mg salts than Na salts and the correlation of Mg salt toxicity to Mg activity. Third, a mechanism related to multiple ions is indicated, based 1) on Na salts being less toxic than salts of other cations, 2) on anions affecting the toxicity of Na salts, 3) on the good correlation of this toxicity to osmolarity, and 4) on the agreement of toxicity from Na gluconate and mannitol with that of inorganic Na salts when expressed as osmolarity. Calcium salt toxicity also likely represents a mechanism different from that of Na salts because of the greater toxicity of CaCl<sub>2</sub> compared with Na salts tested at high but nontoxic Ca concentrations; however, this might be the same mechanism as Mg, which has similar toxicity to Ca when tested in waters with higher Ca background.

Regarding dosimetry, rigorously understanding and describing salt toxicity requires examining toxicity on the basis of chemical activities and consideration of various ion interactions. The cation-related mechanisms would be expected to be best related to cation activities, and the multiple ion mechanism associated with Na salts requires some measure, such as osmolarity, which addresses multiple chemical species. These conclusions regarding mechanisms and dosimetry are preliminary and will be further addressed in the subsequent articles mentioned in the *Introduction*, but it is evident that regulations based solely on a single ion may lack robustness from failing to address the relative toxicities and interactions of multiple ions.

One limitation of the present results is that they just concern the toxicities of single salts, with other ions being at low to moderate background concentrations reported for US waters. Most field situations would involve enrichment of more ions—to well above ambient levels, if not to toxic levels. For example, it would not be expected that Mg would be enriched to toxic levels while Ca stays at background levels, and the high toxicity of K salts depends on very low Na concentrations, which would be an unusual exposure scenario. The exposure metrics and relationships noted in the present study would be expected to apply to more complex mixtures (e.g., all ions contributing to osmolarity), but there is a need to better define the scope of applicability of these relationships. Their extension to more complex mixtures will be the subject of subsequent articles.

Although a complete model for the acute toxicity of major ions to C. dubia will be forthcoming, Figures 6 through 8 make it clear that this will be complicated, involving chemical speciation calculations, multiple toxicity mechanisms, and interactions among 7 toxicants. Any practical application will require simplifications, but we feel that the detail provided in the present study and in later articles will provide a starting point for determining what simplifications are possible and how they should be structured. Another application issue concerns the relevance of acute ion toxicity for C. dubia to the assessment of an entire aquatic community, especially in low-ion, lotic systems. This toxicity endpoint is the starting point for work that will address more endpoints and species. However, the relationships developed for C. dubia will be directly relevant to many aquatic systems and will also have value in developing and interpreting major ion toxicity data for other species.

Supplemental Data—The Supplemental Data are available on the Wiley Online Library at DOI: 10.1002/etc.3487.

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Disclaimer—The views expressed in the present study are those of the authors and do not necessarily reflect the views or policies of the US Environmental Protection Agency. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.

Data availability—Data are available through the USEPA Environmental Dataset Gateway (https://edg.epa.gov/metadata/catalog/main/home.page) or on request from the corresponding author (erickson.russell@epa.gov).

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## EFFECTS OF HARDNESS, CHLORIDE, AND ACCLIMATION ON THE ACUTE TOXICITY OF SULFATE TO FRESHWATER INVERTEBRATES

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Abstract—The acute toxicity of sulfate to *Ceriodaphnia dubia*, *Chironomus tentans*, *Hyalella azteca*, and *Sphaerium simile* was assessed to support potential updates of Illinois (USA) sulfate criteria for the protection of aquatic life. The mean lethal concentrations to 50% of a sample population (LC50s), expressed as mg SO<sub>4</sub><sup>2-</sup>/L, in moderately hard reconstituted water (MHRW) were as follows: 512 mg/L for *H. azteca*, 2,050 mg/L for *C. dubia*, 2,078 mg/L for *S. simile*, and 14,134 mg/L for *C. tentans*. At constant sulfate (~2,800 mg/L) and hardness (106 mg/L), survival of *H. azteca* was positively correlated with chloride concentration. Hardness also was found to ameliorate sodium sulfate toxicity to *C. dubia* and *H. azteca*, with LC50s for *C. dubia* increasing from 2,050 mg SO<sub>4</sub><sup>2-</sup>/L at hardness = 90 mg/L to 3,516 mg SO<sub>4</sub><sup>2-</sup>/L at hardness = 484 mg/L. Using a reformulated MHRW with a similar hardness but higher chloride concentration and different calcium to magnesium ratio than that in standard MHRW, the mean LC50 for *H. azteca* increased to 2,855 mg/L, and the LC50 for *C. dubia* increased to 2,526 mg/L. Acclimation of *C. dubia* to 500 and 1,000 mg SO<sub>4</sub><sup>2-</sup>/L for several generations nominally increased mean LC50 values compared with those cultured in standard MHRW.

Keywords-Sulfate

Total dissolved solids

Osmoregulation

Hyalella

Toxicity

#### INTRODUCTION

Aquatic ecotoxicological research has primarily focused on the impairment of fauna by contaminants that are toxic at minute concentrations; however, ordinarily benign major ions (e.g., sodium, sulfate) can reach concentrations in wastewater discharges that severely impair sensitive in-stream macroinvertebrates and laboratory test organisms [1-5]. Concentrations of these major ions and therefore, of total dissolved solids (TDS), which is essentially the sum of the concentrations of all common ions (e.g., sodium, potassium, calcium, magnesium, chloride, sulfate, and bicarbonate) in freshwaters, can be elevated by numerous practices, such as reverse osmosis systems, pH modifications, and mining operations [6]; and investigations of major-ion toxicity have involved irrigation drainage water [1,7-9], inundation of freshwater systems by brackish water [3,10], laboratory-formulated salt solutions [11,12], and mining activities [4,5,13].

Coal preparation facilities wash coal to reduce sulfur emissions prior to burning in coal-fired power plants and treat wastewaters for acid-soluble metals. This practice often produces a waste containing sulfuric acid that is usually neutralized by the addition of sodium hydroxide or sometimes quicklime (CaO) prior to release to a receiving system [14]. The result is an effluent containing high concentrations of sulfate, sodium, and/or calcium ions and therefore, TDS. Other ions potentially present at high concentrations because of coal preparation activities include magnesium and chlorides; therefore, the interacting effects of these various ions should be considered. Researchers have found hardness and multiple "nontoxic" cations in solution to ameliorate major-ion toxicity ([8,11,15] (http://scholar.lib.vt.edu/theses/available/etd-051499-130633/), and several

There are no federal water quality criteria for the protection of freshwater life for TDS, sulfate, or sodium [19], but several states, including Illinois, are developing standards for sulfate to protect aquatic life. Although major-ion (i.e., TDS) toxicity is caused by osmoregulatory stress from the combination of all cations and anions, chloride standards currently exist, and Illinois plans to additionally regulate for sulfate in order to address the major non-chloride component of TDS in these waters. Therefore, the objectives of the current study were to generate lethal concentrations to 50% of a sample population (LC50s) and lethal concentrations to 10% of a sample population (LC10s) for sulfate with selected freshwater invertebrates (Ceriodaphnia dubia, Chironomus tentans, Hyalella azteca, and Sphaerium simile) in the U.S. Environmental Protection Agency (U.S. EPA)'s [20] moderately hard reconstituted water (MHRW) and to determine the effects of laboratory water composition, water hardness, and test organism acclimation on the acute toxicity of sulfate. The endpoints generated are described in terms of sulfate concentrations to address regulatory issues; however, it is important to note that in our exposures, sodium was the major cation, and effects observed are probably caused by the combination of all dissolved ions.

#### MATERIALS AND METHODS

Toxicity of sulfate to freshwater invertebrates in MHRW

Four invertebrates were selected for initial testing. Three of these, *C. dubia*, *H. azteca*, and *C. tentans*, are standard U.S. EPA organisms used to test for either water column or sediment toxicity [20,21]. The fourth, *S. simile*, is a fingernail clam (Bivalvia, Sphaeriidae) that was easily obtained from the field and represented the phylum Mollusca. Reliable toxicity data for sodium sulfate have been generated for *C. dubia* [11], so this organism was used in the present study for comparative

studies indicate that calcium is more important than magnesium in this regard [16–18].

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purposes. Additionally, previous studies have found *C. dubia* to be more sensitive to major-ion or TDS toxicity than other U.S. EPA-recommended test species (e.g., *Daphnia magna*, *Pimephales promelas*) [5,9,11].

The cladoceran, C. dubia, was cultured in-house (Soucek Laboratory, Illinois Natural History Survey) according to U.S. EPA methods [20]. The mean LC50 in NaCl reference tests for these C. dubia cultures was 2,030 mg NaCl/L, which was comparable to the value of 1,960 mg/L reported in previous studies [11]. The midge, C. tentans, also was cultured in-house according to U.S. EPA methods [21]. Prior to testing, larvae were fed a diet of ground Tetra Min® (TetraWerke, Melle, Germany) flake food and rabbit pellets (free of antibiotics). Amphipods, H. azteca, were obtained from a commercial source (Aquatic Research Organisms, Hampton, NH, USA) and were acclimated to MHRW at 22°C and a 16:8-h (light: dark) photoperiod for at least 7 d prior to testing. Sphaeriid clams were collected from Spring Creek, near Loda, Illinois, USA, and acclimated to MHRW at 22°C and a 16:8-h (light: dark) photoperiod for 5 to 7 d prior to testing. Clams were identified to species by Gerald Mackie (University of Guelph, Department of Zoology, Guelph, ON, Canada).

For toxicity testing, a pure (99%) grade of anhydrous sodium sulfate ( $Na_2SO_4$ ) (CAS 7757-82-6) was obtained from Fisher Scientific (Pittsburgh, PA, USA) to serve as the source of sulfate. A concentrated solution of this salt (19,040 mg  $SO_4^{2-}/L$ ), as well as a sample of laboratory-deionized water, was acidified to pH <2.0 and analyzed for priority metal concentrations at the Illinois State Water Survey (Champaign, IL, USA) using inductively coupled plasma-atomic emission spectrometry according to U.S. EPA methods [22]. All metals analyzed were below acute standard levels ([19], and R. Mosher, Illinois Environmental Protection Agency, Springfield, IL, USA, personal communication) in the concentrated sulfate sample, and all were below detection limits in the deionized water sample except for iron (37  $\mu$ g/L) and zinc (9  $\mu$ g/L). The actual metal concentrations have already been reported [23].

For definitive static, nonrenewal toxicity tests, conducted according to American Society for Testing and Materials E729-96 methods [24], treatments comprised a 75% dilution series (i.e., the 100% concentration was serially diluted by 25%), rather than the standard 50%, because major-ion toxicity tests often cause 100% mortality in one concentration and 0% mortality in the next highest concentration if the spread is too great. Five to six concentrations were tested using MHRW as both the diluent and control, with four replicates tested per concentration. Tests with C. dubia and C. tentans were conducted for 48 h with a 16:8-h (light:dark) photoperiod, with the C. dubia tests being conducted at 25°C and the C. tentans tests at 22 °C. H. azteca and S. simile were exposed for 96 h at 22°C and a 16:8-h (light:dark) photoperiod. C. dubia, C. tentans, and H. azteca were exposed in 50-ml glass beakers with five organisms per beaker, and for C. tentans and H. azteca, 1 g of quartz sand was added to each beaker to serve as substrate. Clam tests were conducted in 150-ml glass beakers (no substrate) with three to five organisms per replicate, depending on the animal size. All clams used were juveniles. In the first experiment, clams averaged 4.6 mm in length (anterior to posterior margin), whereas in the second and third tests, they averaged 5.4 and 8.3 mm in length, respectively. This slight difference in size for the last test did not substantially affect toxicity. C. dubia used were <24 h old, C. tentans were 10 d old, and H. azteca were approximately third instar (7–14 d old). Percent survival in each replicate was recorded every 24 h and at the end of the exposure period. A dissecting microscope was used to assess survival of *Hyalella* and *Sphaerium*.

Standard water chemistry parameters, including temperature, pH, conductivity, dissolved oxygen, alkalinity, and hardness, were measured at both the beginning and the end of each exposure period. The pH measurements were made using an Accumet® (Fisher Scientific, Pittsburgh, PA, USA) model AB15 pH meter equipped with an Accumet gel-filled combination electrode (accuracy <±0.05 pH at 25°C). Dissolved oxygen was measured using an air-calibrated Yellow Springs Instruments (Yellow Springs, OH, USA) model 58 meter with a self-stirring biochemical oxygen-demand probe. Conductivity measurements were made using a Mettler Toledo® (Fisher Scientific) model MC226 conductivity/TDS meter. Alkalinity and hardness were measured (beginning of tests only) by titration as described in work by the American Public Health Association [25]. Samples from each treatment were analyzed to confirm sulfate concentrations by ion chromatography at the Illinois Natural History Survey Aquatic Chemistry Laboratory (Champaign, IL, USA).

All LC50 values were calculated using either the Spearman-Karber method or probit analysis. To increase confidence in LC50 values, three assays were conducted with each organism, except that only two were conducted for C. tentans because of their relative tolerance and low variation in LC50s for the first two tests. This provided a stronger estimate of the mean LC50 value for each species. Geometric means are reported because they are less affected by extreme values. In addition, LC10 values were calculated for all species. With the exception of those for H. azteca, all LC50 values presented are geometric means of the Spearman-Karber LC50s for a given species, generated from measured sulfate concentrations. The H. azteca data did not permit use of the Spearman-Karber method, so probit analysis was used. The LC10 values presented were generated using probit analysis (the Spearman-Karber program does not calculate LC10s) with the combined data from all tests for a given species.

Influence of dilution water composition on sulfate toxicity

Based on observations of others that H. azteca had much better control survival in water-only whole-effluent toxicity tests using modified laboratory water [26], experiments were conducted to determine sulfate LC50 values for C. dubia and H. azteca using the alternate water type referred to as reformulated moderately hard reconstituted water (RMHRW). Reformulated moderately hard reconstituted water is similar to MHRW with two basic differences: The nominal chloride concentration in RMHRW is nearly 18-fold higher than that in MHRW, and the calcium and magnesium salt concentrations are adjusted so that RMHRW has a Ca:Mg molar ratio of 3.25: 1, whereas MHRW has a Ca:Mg molar ratio of 0.88:1 (Table 1). A minor modification in the present study was that anhydrous CaSO<sub>4</sub> (CAS 7778-18-9) was used for both RMHRW and MHRW. The nominal concentrations shown in Table 1 take this modification into account. Mean LC50s and LC10s were generated for both species in this water using the same laboratory and calculation methods as described above, with the only exception being the changed diluent/control water.

An additional experiment was conducted with *H. azteca* to attempt to isolate the two basic differences between MHRW and RMHRW. In this experiment, only one nominal sulfate

Table 1. Nominal chemical composition of two laboratory waters used in testing with *Hyalella azteca* and *Ceriodaphnia dubia* 

Component (units)	$MHRW^a$	RMHRW <sup>b</sup>	
K+ (mg/L)	2.1	2.1	
Na+ (mg/L)	26.3	26.3	
Ca <sup>2+</sup> (mg/L)	17.6	32.7	
$Mg^{2+}$ ( $mg/L$ )	12.1	6.1	
$SO_4^{2-}$ (mg/L)	90.2	59.2	
Cl- (mg/L)	1.9	33.9	
$HCO_3^-$ (mg/L)	69.7	69.7	
Hardness (mg/L as CaCO <sub>3</sub> )	94	107	
Ca/Mg (molar ratio)	0.88	3.25	
pH <sup>c</sup>	7.9	7.9	
Conductivity (S/cm)d	295	341	

<sup>a</sup> MHRW = moderately hard reconstituted water [20].

<sup>b</sup> RMHRW = reformulated moderately hard reconstituted water [26].

<sup>c</sup> The average pH for all treatments during all tests was  $8.0\pm0.2$  (standard deviation), and dissolved oxygen never dropped below 6.5 mg/L.

<sup>d</sup> Conductivity of samples in MHRW varied depending upon  $SO_4^2$  concentration and followed a linear trend described by the formula: Conductivity (S/cm) =  $1.7111[SO_4^{2-} (mg/L)] + 717.15$ ,  $r^2 = 0.9963$ .

concentration (2,500 mg/L) was tested with various base waters. The first of these was MHRW; the second was RMHRW; the third, called chloride, had the same chloride concentration (33.9 mg/L) as RMHRW (Table 1) but the same Ca:Mg molar ratio (0.88:1) as MHRW; and the final medium, called Ca/Mg, had the same Ca:Mg molar ratio (3.25:1) as RMHRW, but the same chloride concentration (1.9 mg/L) as MHRW. Hyalella was exposed to these four treatments for 96 h at 22°C. Mean percent survivorship values for each treatment were compared using analysis of variance with JMP-IN® software [27].

#### Influence of hardness on the toxicity of sodium sulfate

In these experiments, we tested the toxicity of sulfate (with sodium as the major cation) to C. dubia in six freshwater solutions having nominal hardness values of <100 (standard U.S. EPA MHRW), 200, 300, 400, 500, and 600 mg/L (as CaCO<sub>3</sub>). Hardness was increased by adding enough CaSO<sub>4</sub> (CAS 7778-18-9) and MgSO<sub>4</sub> (CAS 7487-88-9) in the same molar ratio as that in U.S. EPA MHRW (Ca/Mg = 0.88) to achieve the nominal hardness values. Then Na<sub>2</sub>SO<sub>4</sub> was added, as was done with the standard MHRW. Whole carboys were made at each elevated hardness level, and this water was used as both diluent and control; therefore, each concentration within a given test had the same hardness (i.e., [Ca<sup>2+</sup>] and [Mg<sup>2+</sup>] did not change with dilution). The only parameters that varied within a particular test were sodium, sulfate, and conductivity. At least three tests were conducted for each hardness level to provide a mean LC50 value and standard deviation. Exposures were conducted using the same laboratory and calculation methods described above, with the only exception being the hardness of the diluent. An additional assay was conducted with H. azteca at only one sulfate concentration (1,460 mg/ L) and three different hardness levels (90, 200, and 300 mg/ L as CaCO<sub>3</sub>). Hyalella was exposed to sulfate at each of these hardness levels for 96 h, and mean percent survival was compared between treatments using analysis of variance with JMP-IN [27].

#### Influence of chloride on the toxicity of sulfate

In this experiment, we tested the toxicity of sulfate to *H. azteca* in six freshwater solutions having nominal chloride

Table 2. Toxicity of sulfate to freshwater organisms in MHRWa

Species	n	$\begin{array}{c} Mean\ LC50^b \\ (mg\ SO_4^{2-}/L) \end{array}$	Range (mg SO <sub>4</sub> <sup>2-</sup> /L)	LC10 <sup>c</sup> (mg SO <sub>4</sub> <sup>2-</sup> /L)
Ceriodaphnia dubia	3	2,050	1,869-2,270	1,759
Chironomus tentans	2ª	14,134	14,123-14,146	11,682
Sphaerium simile	3	2,078	1,901-2,319	1,502
Hyalella azteca	3	512	431-607	262

<sup>a</sup> MHRW = moderately hard reconstituted water [20].

<sup>b</sup> Lethal concentrations to 50% of a sample population (LC50s) are geometric means of all Spearman-Karber values generated for a given organism using measured sulfate concentrations. Control survival was >90% in all exposures.

c Lethal concentration to 10% of a sample population (LC10) values were generated using probit analysis with the combined data from all tests for a given energies.

all tests for a given species.

d Tests produced similar LC50s and because values were so high, a third test was not conducted.

concentrations of 1.9, 10, 15, 20, 32, and 60 mg/L. Chloride, as NaCl (CAS 7647-14-5, Fisher Scientific AC42429-0010), was added at appropriate concentrations to a solution with a hardness of approximately 106 mg/L (Ca/Mg = 3.25, molar ratio) and a nominal sulfate concentration of 2,800 mg/L. The only parameters that varied between treatments were sodium and chloride. In general, tests were conducted using the same laboratory methods as described above for Hyalella. Sulfate, chloride, and bromide were measured in test solutions by ion chromatography. Hyalella was exposed to sulfate at each of the six chloride levels for 96 h, and mean percent survival was compared between treatments using analysis of variance with JMP-IN[27]. One additional aspect of this experiment that was different from others in this study using Hyalella was that the organisms were cultured in RMHRW and not acclimated to MHRW, as in previous experiments, to potentially improve the health of the test organisms [26]. Finally, two test endpoints were recorded. Tests were checked for survival under a dissecting microscope, and total survival included all living individuals, even if they were lying on the bottom and only legs were twitching. Functional survivors included only those individuals that were active and upright or burrowing.

Influence of acclimation on the toxicity of sulfate to C. dubia

This experiment was designed to determine the effects of acclimation to relatively high sulfate levels on the response of *C. dubia* to sulfate. *C. dubia* were cultured in U.S. EPA MHRW with Na<sub>2</sub>SO<sub>4</sub> added to achieve sulfate concentrations of 500 and 1,000 mg/L. After two to three generations had been cultured in these two sulfate concentrations, acclimated organisms were tested in high sulfate solutions using standard MHRW as a diluent and control as described above. Three replicate tests were conducted for each acclimation level to provide a mean LC50 value and standard deviation.

## RESULTS

Toxicity of sulfate to freshwater invertebrates in MHRW

Of the four species tested in MHRW, the most sensitive was H. azteca, with a mean LC50 of 512 mg  $SO_4^{2-}/L$  (Table 2). C. dubia and the fingernail clam, S. simile, were similar in sensitivity, with mean LC50s of 2,050 and 2,078 mg  $SO_4^{2-}/L$ , respectively. C. tentans was tolerant to sulfate exposure, with a mean LC50 of 14,134 mg  $SO_4^{2-}/L$ . The LC10 values were calculated by analyzing all tests for each species

Table 3. Influence of culture/testing water composition on toxicity of sulfate to Hyalella azteca and Ceriodaphnia dubia

Species	Water type	Mean <sup>a</sup> LC50 <sup>b</sup> (mg $SO_4^{2-}/L$ )	Range	$\frac{LC10^{c}}{(\text{mg SO}_{4}^{2}-/L)}$
H. azteca	MHRWd	512 B	431–607	262
H. azteca	RMHRW <sup>e</sup>	2,855 A	2,835-2,876	2,185
C. dubia	MHRW	2,050 B	1,869-2,270	1,759
C. dubia	RMHRW	2,526 A	2,436–2,607	2,216

- <sup>a</sup> Different capital letters indicate means are significantly different (p < 0.05). Only intraspecific comparisons were tested.
- <sup>b</sup> LC50 = lethal concentration to 50% of a sample population.
- <sup>c</sup> LC10 = lethal concentration to 10% of a sample population.
- <sup>d</sup> MHRW = moderately hard reconstituted water [20].
- <sup>e</sup> RMHRW = reformulated moderately hard reconstituted water [26].

simultaneously, and these ranged from 262 mg  $SO_4^{2-}/L$  for *Hyalella* to 11,682 mg  $SO_4^{2-}/L$  for *C. tentans* (Table 2).

Influence of dilution water composition on sulfate toxicity

Testing *H. azteca* in RMHRW produced a strikingly different response compared to results of tests in MHRW (Table 3). The mean LC50 in RMHRW (2,855 mg  $SO_4^{2-}/L$ ) was more than 5.5-fold higher (p < 0.0001) than that generated using MHRW (512 mg  $SO_4^{2-}/L$ ), with a >8-fold increase in the LC10 value. *C. dubia* also had a significantly different (p = 0.0205), though not as striking, response, with the mean LC50 increasing from 2,050 mg  $SO_4^{2-}/L$  in MHRW to 2,526 mg  $SO_4^{2-}/L$  in RMHRW. The LC10 for *C. dubia* increased from 1,759 mg / L in MHRW to 2,216 mg  $SO_4^{2-}/L$  in RMHRW (Table 3).

In the experiment with *H. azteca* designed to dissect the effects observed in RMHRW, only 45% and 55% of the test organisms exposed to 2,500 mg SO<sub>4</sub><sup>2</sup>/L were alive in the MHRW and Ca/Mg treatments, respectively, after 48 h, whereas 85% and 80% survived in the RMHRW and chloride media, respectively (Fig. 1). After 96 h, all of the organisms had died in MHRW and Ca/Mg, whereas 80% still survived in RMHRW and 25% survived in chloride. These data suggest that chloride played the larger role in protecting *H. azteca* against sulfate toxicity and that the different Ca:Mg ratio played a smaller role.

Influence of hardness on the toxicity of sodium sulfate

Increasing water hardness decreased the toxicity of sodium sulfate to *H. azteca* (Fig. 2). In controls, 90% of test organisms

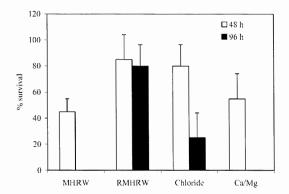


Fig. 1. Effect of various components of reformulated, moderately hard, reconstituted water (RMHRW) on percent survival of *Hyalella azteca* in elevated (2,500 mg SO<sub>4</sub><sup>2</sup>/L) sulfate solutions. The chloride and Ca/Mg treatments consisted of standard moderately hard reconstituted water (MHRW) with chloride or Ca:Mg molar ratio adjusted to match RMHRW.

survived in MHRW (no sulfate added), whereas after 96 h, all organisms were dead in the hardness = 100,  $SO_4^{2-} = 1,460$  mg/L treatment. However, in the hardness = 200 and hardness = 300 mg/L treatments, 15% and 60% of test organisms survived, respectively.

Whereas the mean LC50 for C. dubia in standard MHRW (hardness = 90) was 2,050 mg  $SO_4^{2-}/L$ , the mean LC50s substantially increased at hardness values of 200 and 300 mg/L (Table 4). Mean LC50s were even higher at the higher hardness values of 390, 484, and 578 mg/L, with a maximum of 3,516 mg SO<sub>4</sub><sup>2</sup>-/L at a hardness of 484 (Table 4). The LC10s increased as well, from 1,759 mg SO<sub>4</sub><sup>2</sup>/L at a hardness of 90 mg/L to 2,173 mg SO<sub>4</sub><sup>2</sup>/L, and 2,389 mg SO<sub>4</sub><sup>2</sup>/L at hardness values of 200 and 300 mg/L, respectively. Whereas in the 90 through 500 nominal hardness tests, measured sulfate concentrations were very close to nominal sulfate concentrations, measured sulfate in the 600 nominal hardness tests was somewhat lower than nominal sulfate concentrations, suggesting that some precipitation of CaSO<sub>4</sub> occurred. Therefore, results may be questionable at this hardness level. If the mean LC10 at that hardness is excluded, a linear relationship exists between water hardness and LC10, described by the formula LC10 (mg  $SO_4^{2-}/L$ ) = 2.685(hardness) + 1595.5,  $r^2$  = 0.959. When the LC10 at a hardness of 578 (nominal hardness of 600) is included, the relationship is better described by a logarithmic function with the formula LC10 (mg  $SO_4^{2-}/L$ )  $526.24(\ln[\text{hardness}]) - 574.81 \ (r^2 = 0.8713).$ 

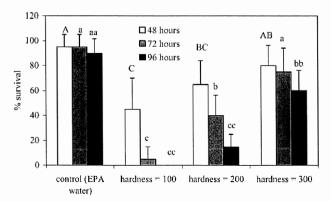


Fig. 2. Effect of hardness on toxicity of elevated sulfate to Hyalella in moderately hard reconstituted water. Average measured sulfate concentration was 1,460 mg/L (standard deviation = 25) for the three treatments excluding the control (106 mg/L sulfate). EPA = Environmental Protection Agency. Different upper- or lower-case letters indicate means are significantly different (p < 0.05).

Table 4. Influence of water hardness on toxicity of sulfate to Ceriodaphnia dubia in MHRW<sup>a</sup>

Hardness, nominal (actual)	n	Mean LC50b (mg SO <sub>4</sub> <sup>2-</sup> /L)	Range (mg SO <sub>4</sub> <sup>2</sup> -/L)	LC10° (mg SO <sub>4</sub> <sup>2-</sup> /L)
90 (89)	3	2,050	1,869-2,270	1,759
200 (194)	3	3,000	2,706-3,265	2,173
300 (288)	4	2,946	2,383-3,361	2,389
400 (390)	3	3,174	3,073-3,369	2,744
500 (484)	3	3,516	3,338-3,716	2,793
600 (578)	3	3,288	2,761-4,220	2,547

- <sup>a</sup> MHRW = moderately hard reconstituted water [20]
- <sup>b</sup> Lethal concentrations to 50% of a sample population (LC50s) are geometric means of all Spearman-Karber values generated for a given organism using measured sulfate concentrations.
- c Lethal concentration to 10% of a sample population (LC10) values were generated using probit analysis with the combined data from all tests for a given treatment.

#### Influence of chloride on the toxicity of sulfate

Sulfate toxicity to *H. azteca* decreased with increased levels of chloride when hardness was held constant (Fig. 3). At the lowest measured chloride concentration tested (5 mg/L), only 20% of the test organisms exposed to 2,846 mg  $SO_4^2$ /L were alive after 96 h, and none of these organisms were functionally alive. At 13 mg Cl<sup>-</sup>/L, both total and functional survival increased nominally, but not significantly (p > 0.05); however, significant increases in total and functional survival were observed at and above 18 mg Cl<sup>-</sup>/L (p < 0.05). Survival was 85% and 100% in the 36 and 67 mg Cl<sup>-</sup>/L treatments, respectively. Bromide concentrations in all treatments were below detection limits (<0.01 mg/L).

## Influence of acclimation on the toxicity of sulfate to C. dubia

In this experiment, with *C. dubia* acclimated for several generations to either 500 or 1,000 mg  $SO_4^{2-}/L$ , nominal increases in mean LC50 values were observed; however, these means were not significantly greater (p < 0.05) than that for organisms cultured in standard MHRW (Fig. 4).

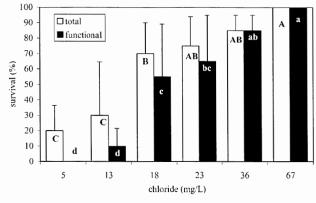


Fig. 3. Effect of increasing chloride concentrations on sulfate toxicity to Hyalella azteca. Mean  $\pm$  standard deviation sulfate concentration for all treatments was 2,846  $\pm$  80 mg SO<sub>4</sub><sup>2</sup>/L, mean hardness was 106  $\pm$  2 mg/L as CaCO<sub>3</sub>, and Ca:Mg was 5.4:1. Different upper- or lower-case letters indicate means are significantly different (p < 0.05). Total = all survivors including those lying on bottom barely moving; functional = survivors that are moving about.

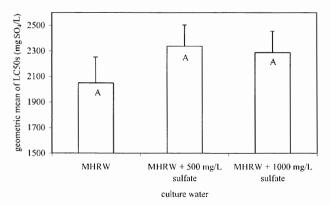


Fig. 4. Effect of acclimation on sulfate toxicity to *Ceriodaphnia dubia*. Organisms were cultured for at least two generations in moderately hard reconstituted water (MHRW), MHRW with 500 mg  $SO_4^{2-}$ , or MHRW with 1,000 mg  $SO_4^{2-}$ . Three tests were conducted with each population of organisms. Treatments with the same upper-case letters indicate that means are not significantly different (p < 0.05). LC50 = median lethal concentration.

#### DISCUSSION

Toxicity of sulfate to freshwater invertebrates in MHRW

The geometric mean for the three tests with C. dubia in this study was 2,050 mg SO<sub>4</sub><sup>2</sup>/L (Table 2), which compares favorably with the value of 3,080 mg Na2SO4/L (equivalent to 2,082 mg SO<sub>4</sub><sup>2</sup>/L) generated in previous studies [11]. Values generated in this study for H. azteca and S. simile were lower than values generated by others for the fathead minnow, P. promelas (5,380 mg/L), and D. magna (3,096 mg/L) [11]. The midge, C. tentans, was relatively insensitive compared with other invertebrates. This finding agrees with the observation of no significant reductions in relative chironomid abundance in waters exceeding 3,000 mg SO<sub>4</sub><sup>2-</sup>/L below a coal processing discharge facility (A.J. Kennedy, unpublished data). The British Columbia Ministry of Environment, Land and Parks (BCMELP) has an online database (wlapwww.gov.bc.ca/wat/ wq/BCguidelines/sulphate/index.html) that includes a variety of sulfate toxicity data for a number species. The values generated by BCMELP for Hyalella were quite variable and not similar to that obtained in this study using MHRW; however, with the exception of hardness estimates, water quality data were not presented in the database, so it is difficult to make comparisons with our study. As will be discussed below, water quality data, including cations and anions present, are critical for predicting the responses of freshwater organisms (especially Hyalella) to elevated sulfate concentrations.

#### Influence of chloride on the toxicity of sulfate

The composition of dilution water used during testing in this study had a dramatic effect on the toxicity of sulfate to *Hyalella*. In fact, the 96-h LC50 in RMHRW was 5.5-fold higher than that generated using MHRW. Both dilution waters were similar in terms of hardness (~90-106 mg/L as CaCO<sub>3</sub>), alkalinity, and pH, but one potential reason for the difference in response is the difference in chloride concentrations between the two media (see Table 1). Freshwater organisms tend to osmoregulate hypertonically with respect to the surrounding medium, achieved by active transport of ions into the hemolymph [28,29]. The principal inorganic anion of crustacean hemolymph is chloride, and it has been suggested that low chloride concentrations may limit the distribution of at least

one euryhaline amphipod (*Corophium curvispinum*) in freshwaters [30]. *H. azteca* is a euryhaline amphipod [7], and perhaps when encountering high ion (Na<sup>+</sup> and SO<sub>4</sub><sup>2-</sup>) concentrations in MHRW, it is not able to osmoregulate because of the relatively low concentration of chloride. This same effect was observed, to a lesser extent, with *C. dubia*.

The experiment with *Hyalella*, in which hardness and sulfate were held constant and chloride was variable (see Fig. 3), supports the hypothesis that chloride has a protective effect against sulfate toxicity, because incremental increases in chloride were associated with incremental increases in survival. Borgmann [31] included bromide as one of the ions required by *Hyalella* for long-term survival, stating that chloride is not required; however, chloride is chemically similar to bromide, and results of this study indicate that if chloride is not indeed required, it does appear to at least provide protection from salt toxicity. Bromide was not present at measurable concentrations (<0.01 mg/L) in our experiments. The results of this study further support the findings of others that MHRW may not be an acceptable medium for water-only testing with *Hyalella* [26].

The fingernail clam, S. simile, had a marginally lower LC10 value for sulfate than that of C. dubia in MHRW, but the former was not tested in RMHRW because of temporal restrictions in its availability. It remains unclear whether or not mollusks will have the same physiological response as two crustaceans to increased chloride in toxicity experiments with sulfate. In a field study, 76% of transplanted Asian clams (Corbicula fluminea) in and below a treated mining discharge survived sulfate levels of approximately 3,600 mg  $SO_4^{2-}/L$  with ~700 mg Cl-/L, although, as will be discussed below, hardness (700-800 mg/L as CaCO<sub>3</sub>) likely played a role in this system [5]. Chloride is a principal anion in the hemolymph of most bivalves [32], but others have found that in the unionoidean Toxolasma texasensis, chloride and bicarbonate are equivalent anionic components [33]. Because bicarbonate is readily available via respiration and metabolism, this mussel may not depend on external chloride concentrations for osmoregulation to the extent that some crustaceans do. If this is the case, the effect of chloride observed for Hyalella and Ceriodaphnia might not be manifested in some unionoidean bivalves, and further work should be done to clarify this.

Influence of hardness on the toxicity of sodium sulfate

Another factor that appears to have a strong effect on the toxicity of sulfate is the presence of other major cations, in this case, calcium and magnesium, measured as hardness. In our sodium-dominated system, increased hardness reduced the toxicity of sulfate to Hyalella (see Fig. 2) and had a dramatic effect on the 48-h LC50 for C. dubia (see Table 4). Mount et al. [11] obtained a similar result in that when using only Na<sub>2</sub>SO<sub>4</sub>, the LC50 for C. dubia was 2,082 mg SO<sub>4</sub><sup>2-</sup>/L, but when using a 1:1 ratio of Na<sub>2</sub>SO<sub>4</sub> and MgSO<sub>4</sub>, the LC50 increased to 2,335 mg SO<sub>4</sub><sup>2</sup>/L. They were careful to point out that the effect was not necessarily caused by hardness, but rather by multiple major cations, citing that the LC50 (expressed as mg Cl<sup>-</sup>/L) for C. dubia in NaCl was nearly identical to that in CaCl<sub>2</sub>, despite the fact that the two solutions had very different hardness values. However, increased calcium is known to decrease the passive permeability of gill epithelia to water and ions in seawater-adapted fish and crabs [34,35]. A similar phenomenon may explain the results of the hardness experiments conducted in this study; i.e., we hypothesize that the increased calcium concentrations at higher hardness levels reduced epithelial permeability, thus reducing passive diffusion and the energy required to osmoregulate and accounting for the decrease in toxicity. In support of this hypothesis, the decreased toxicity of sulfate to *Hyalella* in RMHRW was not entirely explained by the increased chloride concentration (see Fig. 1). The different Ca:Mg ratio also appeared to have an effect, and hardness in RMHRW was similar to that in MHRW (~106 and ~90 mg/L as CaCO<sub>3</sub>, respectively). An alternative hypothesis is that increased calcium is competing for binding sites in a manner similar to that proposed for metals like copper [36]; however, this may be unlikely, because sulfate is an anion and sodium is a monovalent cation. Further experiments are required to test these hypotheses.

Others have observed reduced toxicity of saline solutions because of increased hardness. Dwyer et al. [8] demonstrated that increasing the hardness of an NaCl-dominated irrigation return water reduced its toxicity to striped bass and D. magna. A similar phenomenon was observed with a coal processing discharge in Ohio, USA [5,15]. Although this discharge did include elevated sulfate (3,672 mg/L) and chloride (792 mg/ L) concentrations, the nature of the toxicity was complicated by other ions in solution. The hardness of the field-collected effluent ( $\sim$ 792 ± 43 mg/L as CaCO<sub>3</sub>) and several synthetic solutions of varying hardness appeared to reduce sodium and sulfate-dominated TDS toxicity in a fashion similar to that observed in the current study, on both an acute and chronic scale [5; Alan J. Kennedy, unpublished data]. In addition, the BCMELP database suggests a hardness effect on sulfate toxicity for both D. magna and Hyalella. The present study has shown quantitatively that, in a sodium-dominated system, sulfate toxicity is reduced as hardness progressively increases, although results may require further investigation at the highest hardness tested (578 mg/L). Higher hardness levels should be tested to determine whether the relationship remains linear or is logarithmic and reaches an asymptote.

Influence of acclimation on the toxicity of sulfate to C. dubia

We hypothesized that *C. dubia* acclimated to varying levels of sulfate would be less sensitive to sulfate than naive organisms, as implied in other studies addressing TDS acclimation [1,37] and shock [38]. Although the LC50s for the sulfate-acclimated organisms were nominally higher, the means were not significantly different from those of unacclimated organisms. Perhaps more generations of exposure are required before a significant benefit is seen, and further work should be done in this area.

#### CONCLUSIONS

In conclusion, sulfate toxicity is a complex issue, and a number of factors may interact to determine the responses of various organisms to sodium and sulfate-dominated, saline waters. We have found that in MHRW, *H. azteca* is the most sensitive to sulfate of the four invertebrates tested, followed by *C. dubia* and *S. simile*, then *C. tentans*. Furthermore, we demonstrated that increasing chloride concentration reduces the toxicity of sulfate to *Hyalella*, and increasing water hardness ameliorates sodium sulfate toxicity to *Hyalella* and *C. dubia*. More research is required into the hardness issue to determine whether it was truly calcium that ameliorated sulfate toxicity, because only one Ca:Mg ratio was used in this study when increasing hardness, and other major cations like potas-

sium were not investigated. In addition, the actual mechanism behind the mode of protection from multiple cations should be studied. Finally, the aforementioned issues should be examined at a chronic scale using sublethal and/or multigenerational endpoints as more accurate indicators of population-level effects.

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# INFLUENCE OF WATER HARDNESS AND SULFATE ON THE ACUTE TOXICITY OF CHLORIDE TO SENSITIVE FRESHWATER INVERTEBRATES

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Abstract — Total dissolved solids (TDS) represent the sum of all common ions (e.g., Na, K, Ca, Mg, chloride, sulfate, and bicarbonate) in freshwater. Currently, no federal water quality criteria exist for the protection of aquatic life for TDS, but because the constituents that constitute TDS are variable, the development of aquatic life criteria for specific ions is more practical than development of aquatic life criteria for TDS. Chloride is one such ion for which aquatic life criteria exist; however, the current aquatic life criteria dataset for chloride is more than 20 years old. Therefore, additional toxicity tests were conducted in the current study to confirm the acute toxicity of chloride to several potentially sensitive invertebrates: water flea (*Ceriodaphnia dubia*), fingernail clams (*Sphaerium simile* and *Musculium transversum*), snail (*Gyraulus parvus*), and worm (*Tubifex tubifex*), and determine the extent to which hardness and sulfate modify chloride toxicity. The results indicated a significant ameliorating effect of water hardness (calcium and magnesium) on chloride toxicity for all species tested except the snail; for example, the 48-h chloride median lethal concentration (LC50) for *C. dubia* at 50 mg/L hardness (977 mg Cl<sup>-</sup>/L) was half that at 800 mg/L hardness (1,836 mg Cl<sup>-</sup>/L). Conversely, sulfate over the range of 25 to 600 mg/L exerted a negligible effect on chloride toxicity to *C. dubia*. Rank order of LC50 values for chloride at a given water hardness was in the order (lowest to highest): *S. simile* < *C. dubia* < *M. transversum* < *G. parvus* < *T. tubifex*. Results of the current study support the contention that the specific conductivity or TDS concentration of a water body alone is not a sufficient predictor of acute toxicity and that knowledge of the specific ion composition is critical. Environ. Toxicol. Chem. 2011;30:930–938. © 2010 SETAC

Keywords—Chloride

Sulfate

Acute toxicity

Invertebrates

Water hardness

#### INTRODUCTION

Total dissolved solids (TDS), a water quality parameter that represents the sum of the concentrations of all major ions (e.g., Na, K, Ca, Mg, chloride, sulfate, and bicarbonate) in freshwaters, can reach concentrations in wastewater discharges that severely impair sensitive aquatic species [1-5]. Common causes of elevated TDS include road salting, reverse osmosis systems, pH modifications of wastewater, agricultural runoff, gas and oil production, and coal or metal mining operations [6]. Unfortunately, our understanding of the influence of TDS from such sources on the general health and well-being of aquatic organisms is generally lacking. A number of others have conducted field and laboratory studies to investigate major ion toxicity [3-5,7-17]; however, insufficient data exist from which to derive protective criteria for TDS. In the United States, no Federal water quality criteria exist for the protection of freshwater aquatic life for TDS, sulfate, Na, Ca, Mg, L, or bicarbonate [18], although both acute and chronic criteria currently exist for chloride (860 and 230 mg/L, respectively).

Numerous freshwater species have been tested for sensitivity to sodium chloride, and invertebrates tend to be more sensitive than fish. Within the invertebrates, a wide range of sensitivity can be seen in acute exposures, with mollusks tending to be the most sensitive [16,19–22], followed by crustaceans [11,16,23–25], then insects [16,20,22,26]. The five most

sensitive genera tested thus far are *Sphaerium* [22], *Ceriodaphnia* [11,23,24], *Tubifex* [27], *Daphnia* [11,23,24], and *Gyraulus* [22], with median lethal concentrations (LC50s) ranging from 682 to 1,941 mg Cl<sup>-</sup>/L. The damselfly nymph *Argia sp.* appears to be the least sensitive, with a mean acute value of 14,252 mg Cl<sup>-</sup>/L [16,22].

The fact that TDS toxicity is dependent on the ionic composition of surface water or effluent has been well established [7,9,11,14,15]. Several researchers have observed that hardness or presence of multiple major cations in solution can ameliorate major ion toxicity [7,9,11], and laboratory experiments with synthesized freshwaters have demonstrated that increasing hardness at a constant Ca:Mg ratio results in decreased sodium sulfate toxicity to *C. dubia* and *Hyalella azteca* [14,15]. In addition, chloride concentration has been shown to regulate the toxicity of sodium sulfate to *H. azteca*, and to a lesser extent, *C. dubia* [15]. Although some work has been done to determine how various ions affect the toxicity of sodium sulfate, very little has been done to explore how additional cations or anions impact the toxicity of sodium chloride to freshwater organisms (see Mount et al. [11]).

The purpose of the current study was to generate acute toxicity data for chloride as sodium chloride in support of the development of updated aquatic life criteria for this common component of TDS. In doing so, we had several objectives. Our first aim was to generate new acute chloride toxicity data for several of the most sensitive genera (*Sphaerium*, *Gyraulus*, and *Tubifex*) using standard methods [28,29] and dilution waters with known ionic composition. Second, we sought to characterize the effect of hardness on the toxicity of chloride to

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freshwater invertebrates. This was done primarily with *C. dubia* but also to a lesser extent with the three previously mentioned species. Finally, given that chloride has been shown to regulate sulfate toxicity [15], our third goal was to determine whether varying sulfate concentration has an impact on chloride toxicity to *C. dubia*.

#### MATERIALS AND METHODS

#### General

Acute chloride toxicity tests were performed at the Great Lakes Environmental Center (GLEC; Columbus, OH, USA) and Illinois Natural History Survey (Champaign, IL, USA). The acute toxicity tests for all species followed U.S. Environmental Protection Agency (U.S. EPA) and American Society for Testing and Materials (ASTM) protocols [28,29]. Reconstituted water at different total hardness and sulfate levels was used as dilution water for the chloride toxicity tests for all species. The base water used to prepare the reconstituted dilution water was deionized tap water from the cities of Columbus (OH, USA) and Champaign (IL, USA), respectively. Reagent grade salts were added in the appropriate amounts to deionized water to achieve the desired ionic composition for each test (Table 1).

#### Organism acquisition and acclimation

Ceriodaphnia dubia. Tests were initiated with *C. dubia* from stock cultures maintained at GLEC and Illinois Natural History Survey. Subcultures of *C. dubia* were grown in and acclimated to the above hardness- and sulfate-adjusted reconstituted dilution water mixes. The subcultures were subjected to hardness and sulfate acclimation over two generations. Such an acclimation period allowed the test population sufficient time to adjust to the conditions they encountered during the chloride toxicity testing aside from the elevated sodium chloride concentrations.

Briefly, each subculture was initiated with neonates in reconstituted dilution waters with similar concentrations of

Table 1. Salt concentrations added to deionized water for generation of the acclimation/dilution and test waters used for definitive chloride toxicity testing at different water hardness and sulfate levels (all test organisms)

	Nomi	nal total l	nardness c	of dilution	water (m	ng/L as Ca	aCO <sub>3</sub> )
	25	50	100	200	400	600	800
Salt		Salt (r	ng/L) add	led at eacl	h hardnes	s level	
KCl	4	4	4	4	4	4	4
NaHCO <sub>3</sub>	96	96	96	96	96	96	96
MgCl <sub>2</sub> *	10	20	40	80	160	240	320
CaCl <sub>2</sub> *	16	32	64	130	260	380	510
Na <sub>2</sub> SO <sub>4</sub>	96	96	96	96	96	96	96
	Non	ninal sulf	ate conce	ntration o	f dilution	water (m	g/L)
	25	50	100	200	400	600	
Salt		Salt (mg/	L) added	at each s	ulfate con	centration	1
KCl	4	4	4	4	4	4	
NaHCO <sub>3</sub>	96	96	96	96	96	96	
MgCl <sub>2</sub> *	121	121	121	121	121	121	
CaCl <sub>2</sub> *	192	192	192	192	192	192	
Na <sub>2</sub> SO <sub>4</sub>	37	74	148	296	592	887	
* Anhydrou	e calte ne	ed					

<sup>\*</sup>Anhydrous salts used.

water hardness and sulfate found in the main stock culture water. These neonates were then transferred to fresh reconstituted dilution waters daily. Each daily transfer represented an incremental (50 mg/L) increase or decrease of the hardness or sulfate concentration from the previous day. This daily transfer process was continued until all animals reached their target acclimation concentration. The single exception to this acclimation regimen was for the group of *C. dubia* acclimated, and subsequently tested, at the nominal 800 mg/L total water hardness level. Because of the large increase needed to reach this nominal target total water hardness acclimation level, and because of the short life span of *C. dubia* (14–21 d), acclimation for this group was increased by 50 and 100 mg/L hardness (or sulfate) on alternating days.

After the acclimation period, test organisms were held at that level for 24 to 48 h with no further incremental increases or decreases until their neonates were collected. Second-generation *C. dubia* were cultured at their respective target total water hardness and sulfate levels, and third-generation *C. dubia* were used for testing. Reproduction rates in all test waters were sufficient to provide the required number of neonates for each test.

Aside from the above details, culturing of *C. dubia* followed U.S. EPA methods [28]. *Ceriodaphnia dubia* were fed approximately  $0.3 \,\mathrm{ml}$  of a yeast-trout chow-cereal leaves/ *Pseudokirchneriella subcapitata*  $(3.0 \times 10^7 \,\mathrm{cells/ml})$  mixture  $(1:2, \, \mathrm{v.v})$  daily throughout the entire acclimation process.

Sphaerium simile and Musculium transversum. Sphaerium simile were field-collected from Spring Creek, near Loda, Illinois, in Iroquois County. Musculium transversum were field collected from the Kankakee River near Aroma Park, Illinois, in Kankakee County. This species was identified by its fused siphons and distinctive shell shape. Both clam species were collected as adults, returned to the laboratory (at Illinois Natural History Survey, Champaign, IL, USA) in site water, and shortly thereafter, allowed to release juveniles from their brood chambers in the laboratory. Juveniles were used for testing.

The juvenile clams were gradually acclimated to laboratory conditions for approximately two weeks. Twenty percent of the water was changed daily until the desired nominal hardness level (50-200 mg/L as CaCO<sub>3</sub>) was achieved; afterward, 50% of the water was changed daily. The temperature of the clam holding water was recorded daily and gradually adjusted (1°C/d) from the water temperature at the time of collection  $(24.7^{\circ}\text{C})$  to a test temperature of  $22 \pm 1^{\circ}\text{C}$ . The clams were held in aquaria containing 6 L gently aerated reconstituted water, and maintained in environmental chambers under controlled conditions (photoperiod 16:8 h light:dark). Before testing, clams were fed daily a suspension of a green alga (Ankistrodesmus falcatus) at a rate of 1.25 mg (dry wt, determined by filtering a known volume and drying) per gram of clam (wet wt). Other details of clam holding conditions followed recommendations of ASTM E729 [29]. Musculium transversum was only tested at the 50 mg/L hardness level because of organism availability.

Gyraulus parvus. Snails (mixed ages; 3–5 mm in diameter) were collected from a vernal pool in Big Run Park, southwest Columbus, Ohio, USA (Franklin County). Snails were transported in 2-L, precleaned plastic containers with site water to GLEC, Columbus, Ohio, USA. Snails were held in the GLEC laboratory in site water at the water temperature measured in the field and with field-collected foliage and detritus material. Water quality parameters (water hardness, alkalinity, dissolved oxygen, pH, and specific conductivity) of the site water were measured and recorded at GLEC.

Snails were gradually acclimated to laboratory conditions over a period of approximately five weeks. The temperature of the snail holding water was recorded daily. The snail holding water required only minor temperature adjustment from the water temperature at the time of collection (23°C) to a test temperature of 22 ± 1°C. Snails were cultured in 10-L glass aquaria in their natural source water that was gently aerated and maintained in environmental chambers under controlled conditions (16:8 h light:dark photoperiod). A 90% water replacement occurred four times a week, with the first replacement consisting of a mixture of 80% site water and 20% reconstituted dilution water of designated hardness (i.e., 50 or 200 mg/L as CaCO<sub>3</sub>). Gradually, the percentage of dilution water at the designated hardness was increased by 10% with each transfer until the holding water was 100% test dilution water. Fresh food sources (leaf material with associated detritus) collected from the site along with TetraMin fish food were provided to the snails with each transfer. Other details of acclimation and holding conditions for snails followed the recommendations in ASTM E729 [29].

Tubifex tubifex. Freshwater tubificid worms (mixed ages) were obtained from Aquatic Research Organisms (ARO). The organisms were shipped in moderately hard reconstituted water prepared by ARO. On receipt, temperature, water quality parameters, and the condition of the organisms were recorded. The annelids were then placed inside a 10-L aquarium with crushed TetraMin fish food and a sand substrate provided by ARO, and with GLEC moderately hard reconstituted water. Great Lakes Environmental Center moderately hard reconstituted water was of similar water hardness, but lower total alkalinity; to mimic the reconstituted water from ARO, additional sodium bicarbonate was added.

Temperature of the annelid cultures was recorded daily and gradually adjusted (1°C/d) from the water temperature at the time of receipt (24°C) to a test temperature of  $22 \pm 1$ °C to acclimate the worms for subsequent toxicity testing. Annelids were maintained in environmental chambers under controlled conditions (photoperiod 16:8 h light:dark). Four times a week, a 90% water replacement occurred, with the first replacement consisting of a mixture of 80% moderately hard reconstituted water and 20% reconstituted dilution water with a hardness of 50 or 200 mg/L as CaCO<sub>3</sub>. The percentage of dilution water at a hardness of 50 or 200 mg/L was gradually increased by 10% with each change over until the culture water was 100% test dilution water. Freshly ground TetraMin fish food was provided with each water replacement. Other details of acclimation and holding conditions for the annelids followed the recommendations in ASTM E729 [29].

#### Toxicity testing procedures

Ceriodaphnia dubia. Forty-eight hour static acute toxicity tests were used for the chloride toxicity tests with  $\leq$  24-h-old

C. dubia neonates. After test initiation, beakers were placed into environmental chambers to maintain proper conditions for the duration of the test. Some test conditions, including the number of organisms per chamber (five), number of chambers per treatment (four), lighting regimen (16:8 h light:dark), and endpoint (mortality) were the same for all species. Conditions that varied among species are detailed in Table 2. In addition, tests with C. dubia were fed one drop of a mixture of Pseudokirchneriella subcapitata and Yeast-cereal leaves-trout chow at test initiation and at 24 h. This is a deviation from U.S. EPA [28] methods, but was done to reduce stress to these organisms. Addition of food would be highly unlikely to alter availability of the toxicant. Tests with other species were not fed.

S. simile, M. transversum, G. parvus, and T. tubifex. Ninety-six-hour static acute toxicity tests were used for the chloride/hardness toxicity tests with these four invertebrates. Testing procedures followed those outlined in ASTM E729 [29]. After test initiation, test beakers were placed into environmental chambers to maintain proper test conditions for the duration of the test (Table 2). For S. simile and M. transversum, mortality was determined daily by examination under a dissecting microscope while still in test chambers. Clams that were obviously gaping and not siphoning were gently probed with a blunt object and closely examined for ciliary movement. If no response or movement was observed, clams were considered dead and removed. At the end of the test, clams were placed in control water with food and checked for active siphoning to determine final mortality numbers. For G. parvus and T. tubifex, mortality was determined daily by noting organism color and determining whether they moved over a 2-h period.

### Stock and test solution preparation

Reagent-grade sodium chloride (NaCl, EMD Chemicals, CAS 7647-14-5; Lot 3628B013) was used to make the test solutions for each respective test. The test solutions were prepared by dissolving a specified amount of the reagent-grade salt into the various hardness- and sulfate-adjusted reconstituted dilution water mixes to form the highest test solution (see Table 1). After the reagent-grade salt for the highest test concentration was completely dissolved in solution, the remaining test concentrations were prepared using a 0.65× dilution factor. The range of test concentrations for each water type was determined on the basis of results from 48-h or 96-h static acute range-finding toxicity tests performed 2 to 3 d before definitive testing. Solutions were not renewed in any of the tests because of the known stability of chloride ion in solution and the desire to reduce organism handling stress. Total chloride concentrations comprised chloride contributed from the NaCl stock solution and dilution water. Measured chloride concentrations for each test were within 10% of nominal.

Table 2. Test conditions and specifications for definitive acute chloride toxicity testing with the five invertebrate species<sup>a</sup>

Test condition	C. dubia	S. simile	G. parvus	T. tubifex	M. transversum
Temperature (°C)	$25 \pm 1$	22 ± 1	22±1	22±1	22 ± 1
Beaker size	30 ml/50 ml <sup>b</sup>	250 ml	250 ml	250 ml	250 ml
Solution volume	25 ml/40 ml <sup>b</sup>	200 ml	150 ml	150 ml	150 ml
Test duration	48 h	96 h	96 h	96 h	96 h

<sup>&</sup>lt;sup>a</sup> Only conditions that were different for different species are shown.

b Illinois Natural History Survey, chamber size; Ceriodaphnia dubia, Sphaerium simile, Gyraulus parvus, Tubifex tubifex, Musculium transversum.

#### Different hardness levels

To determine the influence of hardness on the acute toxicity of chloride to C. dubia, organisms were acclimated to and tested in reconstituted dilution water at seven different nominal hardness concentrations: 25, 50, 100, 200, 400, 600, and 800 mg/L as CaCO<sub>3</sub>. For each of these seven different acclimation and test dilution water compositions, the sulfate concentration was held constant at approximately 69 to 74 mg/L through the addition of Na<sub>2</sub>SO<sub>4</sub> (Tables 1 and 3). Chloride concentrations in dilution waters (control treatment—no additional NaCl added) solutions ranged from approximately 20 mg/ L at the lowest total hardness level to approximately 570 mg/L in the highest (based on calculation from the salts added; see Table 1). Chloride concentrations varied in dilution water solutions because reagent-grade MgCl<sub>2</sub> and CaCl<sub>2</sub> (anhydrous) were used to manipulate water hardness to the desired level. These salts were selected over MgSO<sub>4</sub> and CaSO<sub>4</sub> to maintain the sulfate level near 65 mg/L. The Ca:Mg mass ratio was maintained at approximately 2.25 for all hardnesses (Table 3).

For the other three chloride-sensitive invertebrate test species (*S. simile*, *G. parvus*, and *T. tubifex*), the acclimation and dilution water used for acute chloride toxicity testing was reconstituted dilution water mixed with the salts to achieve a nominal total hardness of 50 and 200 mg/L CaCO<sub>3</sub>. As with *C. dubia*, MgCl<sub>2</sub> and CaCl<sub>2</sub> were used to manipulate water hardness to the desired level, and the Ca:Mg mass ratio was maintained at approximately 2.25 for both hardnesses (Table 4). Sulfate was maintained at approximately 60 mg/L in tests with each species (Table 4).

#### Different sulfate levels

To determine the influence of sulfate on the acute toxicity of chloride to *C. dubia* (only species tested), organisms were acclimated to and tested in reconstituted dilution water at one of six different nominal sulfate concentrations: 25, 50, 100, 200, 400, and 600 mg/L (Table 1). Nominal total water hardness was approximately 300 mg/L as CaCO<sub>3</sub> for all test solutions. For each of the six final reconstituted dilution water types, the desired sulfate concentrations were achieved through the addition of Na<sub>2</sub>SO<sub>4</sub>, whereas reagent-grade MgCl<sub>2</sub> and

CaCl<sub>2</sub> were used to achieve the water hardness as described previously. The nominal chloride concentration in base solutions of all six reconstituted dilution water types was approximately 215 mg/L (based on calculation from the salts added per Table 1). As was the case for tests at different hardnesses with *C. dubia* and the other species, one test was conducted by each laboratory for each sulfate concentration.

Water quality measurements and chloride and sulfate analysis

Total hardness and alkalinity of the reconstituted dilution waters were measured before making test solutions. Temperature, pH, dissolved oxygen, and conductivity were measured at test initiation. For the duration of the tests, temperature, pH, and dissolved oxygen were measured every 24 h after test initiation. To reduce the risk of contamination in the test solutions during chemistry measurements, a fifth replicate of each test concentration was used for chemistry measurements. This fifth replicate contained test organisms, but observations for survival and mortality were not performed, and test solution from the replicate was not used for chloride or sulfate determinations.

The methods used for the chemical measurements supporting each toxicity test were as reported by Eaton et al. [30]. The dissolved oxygen and pH meters used for water chemistry measurements to support the tests were calibrated daily according to routine procedures for these instruments.

The quantification of chloride, sulfate, and other select anions (F, Br) and cations (Ca, Mg) in water samples was completed by Stantec Consulting Services (Columbus, OH, USA). Samples were collected from all test concentrations and controls at test initiation and termination and analyzed for chloride and sulfate. All samples were collected in trace clean plastic bottles and stored at 4°C during storage and shipment to the analytical laboratory for analysis. Chloride and sulfate in the various test solutions were quantified using U.S. EPA method 325.2 and 375.4, respectively; Ca and Mg were quantified using U.S. EPA-SW6010B and U.S. EPA-SW3010A (http://water.epa.gov/scitech/drinkingwater/labcert/analyticalmethods.cfm). Duplicate analysis indicated good precision between replicate measurements of

Table 3. Mean 48-h chloride median lethal concentration values (LC50s) for Ceriodaphnia dubia at various water hardness<sup>a,b</sup> and sulfate concentrations<sup>c</sup>

Nominal Hardness	Hardness (range)	Ca (mg/L) (range)	Mg (mg/L) (range)	Nominal SO <sub>4</sub> <sup>2-</sup> (mg/L)	SO <sub>4</sub> <sup>2-</sup> (mg/L) (range)	Mean 48 h LC50 (mg Cl <sup>-</sup> /L)
			Different hardness level	s		
25	28 (25–30)	5.3 (5.1–5.5)	2.3 (2.2–2.4)	65	74 (70–79)	977
50	47 (44-49)	10.8 (10.6–10.9)	4.8 (4.7-4.9)	65	72 (68–76)	861
100	96 (95–96)	20.9 (20.8–20.9)	9.3 (9.3-9.4)	65	72 (70–74)	1,249
200	187 (180-194)	42.0 (41.8-42.2)	18.7 (18.2–19.1)	65	69 (68–70)	1,402
400	388 (375-400)	81.8 (78.2–85.4)	36.9 (36.0–37.8)	65	74 (69–79)	1,589
600	565 (560-570)	123.0 (117.0-129.0)	56.1 (53.9–58.3)	65	72 (68–76)	1,779
800	796 (792-800)	170.5 (163.0-178.0)	77.3 (75.2–79.3)	65	73 (71–76)	1,836
		Di	fferent sulfate concentra	ions		
300	280 (279-280)	66.1 (NM-66.1)	28.1 (NM-28.1)	25	26 (23–28)	1,356
300	278 (276–280)	63.8 (NM-63.8)	27.5 (NM-27.5)	50	55 (50-60)	1,489
300	282 (280-283)	65.4 (63.7–67.1)	28.9 (28.8–28.9)	100	112 (107–117)	1,317
300	281 (280-281)	64.1 (NM-64.1)	27.3 (NM-27.3)	200	234 (229-239)	1,357
300	285 (280–290)	63.8 (62.8–64.8)	28.2 (27.6–28.7)	400	472 (461–482)	1,154
300	279 (278–280)	63.7 (NM-63.7)	27.5 (NM-27.5)	600	712 (694–729)	1,192

a Nominal Ca:Mg mass ratio for all test solutions was 2.25.

<sup>&</sup>lt;sup>b</sup> Unless otherwise noted, all concentrations shown are measured values, and with the exception of LC50s and confidence intervals, are arithmetic averages of measurements in all treatments at test initiation and termination.

<sup>&</sup>lt;sup>c</sup> NM = not measured.

Table 4. Ninety-six-hour chloride median lethal concentration values (LC50s) for four freshwater invertebrates at various water hardness<sup>a,b</sup> and sulfate concentrations<sup>c</sup>

Species Tested	Hardness (mg/L as CaCO3)	Ca (mg/L)	Mg (mg/L)	SO <sub>4</sub> <sup>2-</sup> (mg/L)	96 h LC50 (mg Cl <sup>-</sup> /L)	95% CI (mg Cl <sup>-</sup> /L)
Sphaerium simile	51	NM	NM	59.9	740	678–807
	192	45.3	19.5	61.7	1,100	1.040-1.164
Gyraulus parvus	56	13.5	3.04	60.9	3,078	2,771-3,418
	212	49.9	22.4	59.7	3,009	2,728-3,318
Tubifex tubifex	52	11.5	4.9	57.9	4,278	3,848-4,717
	220	44.9	20.7	58.9	6,008	5,563-6,489
Musculium transversum	48	NM	NM	58.9	1,930	1,655-2,251

<sup>a</sup> Nominal Ca:Mg mass ratio for all test solutions was 2.25.

<sup>c</sup> NM = not measured; CI = confidence intervals.

chloride and sulfate within a sample (mean relative percent difference of 2.2 for chloride [n=56] duplicate samples] and 5.0 for sulfate [n=5] duplicate samples]).

#### Statistical analysis

The LC50 values based on measured concentrations for all acute toxicity tests were calculated using the U.S. EPA Probit Analysis Program, the Spearman-Karber Method, or the Trimmed Spearman-Karber method following U.S. EPA-821-R-02-012 [28]. The mean of the day 0 and day 2 measured concentrations was used to calculate values, because the total chloride concentrations measured at test initiation and at test termination were only minimally different in all tests. All LC50 values are expressed as milligrams chloride per liter based on the measured chloride in the test solutions, which included chloride added in preparation of the reconstituted dilution water and NaCl that was used to prepare the test concentrations. To analyze relationships between chloride LC50s and hardness or sulfate concentration, we generated scatter plots with LC50 as the dependent variable and hardness or sulfate concentration as the dependent variable, and used Sigma Plot version 9.0 to fit various types of lines through the distribution. We used the following fit options in Sigma Plot: linear, sigmoidal (3 parameter), exponential rise to maximum (single 2 parameter and simple exponent 2 parameter), hyperbola (single rectangular), power (2 parameter), and logarithm (2 parameter I). The line that provided the highest  $r^2$  value was considered to be the best fit.

### RESULTS

Although we varied hardness, sulfate, and chloride concentrations of the test solutions according to our experimental design, these variations had virtually no effect on alkalinity, pH, or dissolved oxygen of the solutions. Mean alkalinity values ranged from 56 to 64 mg/L as CaCO<sub>3</sub>, whereas pH and dissolved oxygen ranged from 7.65 to 8.02 and 7.56 to 8.07 mg/L, respectively. Variation was even lower within tests for a particular species. Temperature was maintained to within  $\pm 1^{\circ}\mathrm{C}$  of the target test temperatures of 25°C (*C. dubia*) and 22°C (other test organisms) in both laboratories (data not shown). For all tests, controls met performance criteria of  $\geq 90\%$  survival.

In tests with *C. dubia* at hardnesses ranging from 27.5 to 795.5 mg/L (as  $CaCO_3$ ) and constant sulfate concentration (72.4  $\pm$  1.6 mg  $SO_4^{2-}/L$ ), mean (of single tests from each of the two laboratories) 48-h LC50s ranged from 861 to 1,836 mg  $Cl^-/L$  (Table 3). Plotting individual LC50s from each

laboratory with corresponding hardnesses showed a strong (statistically significant) positive relationship best described (highest  $r^2$ ) by a power function ( $r^2 = 0.8481$ ; Fig. 1).

To explore the relationship between hardness and LC50 in terms of TDS, nominal concentrations of all constituent ions (except  $H^+$  and  $OH^-$ ) were calculated at the concentration associated with the LC50s for *C. dubia* in each test solution type. The total dissolved solids was then calculated as the sum of all nominal constituent ion concentrations. Ions that varied in concentration in these different test solutions included not only Na and chloride but also Ca, Mg, and sulfate because of the increasing hardness. A significant positive relationship between LC50 (mg TDS/L) and hardness was observed, best described by a power function ( $r^2 = 0.9005$ ; Fig. 1).

In tests with *C. dubia* at sulfate concentrations ranging from 26 to 712 mg  $SO_4^{2-}/L$  and constant hardness (280.3  $\pm$  2.3 [standard deviation] mg  $SO_4^{2-}/L$ ), mean (of tests from both laboratories) 48-h LC50s ranged from 1,154 to 1,489 mg Cl<sup>-</sup>/L (Table 3). Plotting individual LC50s from each laboratory with corresponding sulfate concentrations showed a negative but not statistically significant ( $\alpha$  = 0.05) relationship as described by an exponential function ( $r^2$  = 0.3092; Fig. 2).

Calculating LC50s in terms of TDS as described, a significant positive relationship between LC50 (mg TDS/L) and sulfate concentration was observed, best described by an exponential function ( $r^2 = 0.5372$ ; Fig. 2).

Mean chloride 96-h LC50s for S. simile, G. parvus, T. tubifex, and M. transversum at hardnesses of 50 and 200 mg/

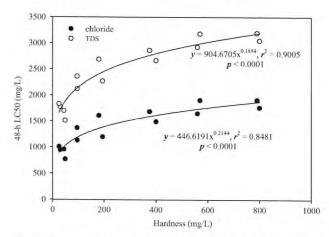


Fig. 1. Relationship between water hardness (mg/L as  $CaCO_3$ ) in diluent and 48-h median lethal concentration (LC50s) values for *Ceriodaphnia dubia* in terms of chloride (mg Cl $^-/L$ ) and total dissolved solids (mg TDS/L).

b All concentrations shown are measured values, and with the exception of LC50s and confidence intervals, are arithmetic averages of measurements in all treatments at test initiation and termination.

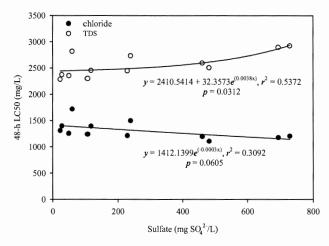


Fig. 2. Relationship between sulfate concentration in diluent and 48-h median lethal concentration (LC50s) values for *Ceriodaphnia dubia* in terms of chloride (mg Cl<sup>-</sup>/L) and total dissolved solids (mg TDS/L).

L ranged from 740 to 6,008 mg Cl<sup>-</sup>/L (Table 4 and Fig. 3). As stated previously, *M. transversum* was only tested at the 50-mg/L hardness level. The most sensitive species of all tested (including *C. dubia*) was *S. simile*, whereas *T. tubifex* was the least sensitive. *Ceriodaphnia dubia* and *S. simile* were relatively similar in sensitivity, however, and *G. parvus* and *T. tubifex* had LC50 values from approximately three-fold to 5.5-fold higher. *Musculium* had an LC50 that was intermediate between the more sensitive and the less sensitive species. For two of the three additional species, increasing hardness resulted in higher LC50 values (740–1,100 mg Cl<sup>-</sup>/L for *S. simile*, and 4,278–6,008 mg Cl<sup>-</sup>/L for *T. tubifex*), whereas for *G. parvus* hardness did not affect the LC50s (3,078 and 3,009 mg Cl<sup>-</sup>/L at 50 and 200 mg/L hardness, respectively).

Comparing the relationship between chloride LC50 and hardness at hardness values of approximately 50 and 200 mg/L, relatively similar slopes were obtained for three of the four species tested (power functions were used in this case to make slopes comparable to those developed for *C. dubia*; Fig. 3). *Tubifex, Sphaerium*, and *Ceriodaphnia* had slopes of 0.2354,

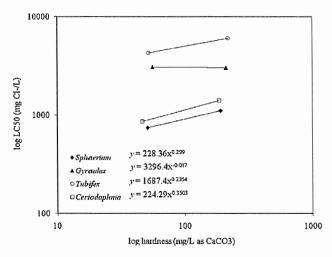


Fig. 3. Relative acute toxicity of chloride (as NaCl) to four freshwater invertebrates at two different hardness levels. Data shown are 96-h log median lethal concentration (LC50s) values for *Gyraulus parvus*, *Sphaerium simile*, and *Tubifex tubifex*, and mean 48-h log LC50s for *Ceriodaphnia dubia*.

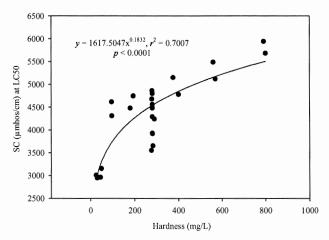


Fig. 4. Relationship between 48-h median lethal concentration (LC50s) values in terms of specific conductivity and dilution water hardness for chloride toxicity tests with *Ceriodaphnia dubia*. SC = specific conductivity.

0.2990, and 0.3503, respectively, whereas *Gyraulus* had a slope of close to zero (-0.017). Note that when including data over the whole range of hardness values tested, the slope for *Ceriodaphnia* was 0.2144.

To evaluate and compare (with chloride concentration) the ability of conductivity to predict TDS toxicity, we plotted the specific conductivity ( $\mu$ mhos/cm) measured in the treatment nearest the LC50 concentration for each chloride toxicity test conducted with *C. dubia* over the range of hardnesses and sulfate concentrations. We observed a significant positive relationship between specific conductivity and hardness that was best described by a power function ( $r^2 = 0.7007$ ; Fig. 4).

#### DISCUSSION

We determined the acute toxicity of chloride to five freshwater invertebrate species: water flea (Ceriodaphnia dubia), fingernail clams (Sphaerium simile and Musculium transversum), planorbid snail (Gyraulus parvus), and tubificid worm (Tubifex tubifex). Rank order of LC50 values for chloride at a given water hardness was in the order (lowest to highest): S. simile < C. dubia < M. transversum < G. parvus < T. tubifex. Tests with C. dubia acclimated and tested under different levels of hardness and sulfate were performed by two different laboratories with comparable results. The 48-h chloride LC50 for C. dubia at 25 to 50 mg/L hardness was approximately half that of C. dubia exposed at 600 to 800 mg/L hardness. Conversely, sulfate over the range of 25 to 600 mg/L exerted a nonsignificant effect on chloride toxicity to C. dubia. The mean 48-h LC50 at 25 mg/L sulfate was approximately 1,356 mg Cl/L, whereas at 600 mg/L sulfate, it was  $1,192 \text{ mg Cl}^-/L$  (reduction of 12%).

Chloride LC50s from the literature for *C. dubia* under various water quality conditions range from 447 to 1,698 mg Cl<sup>-</sup>/L [11,23,24,31], with a geometric mean of 1,172 mg/L. Data reported in the current study span a similar range (861–1,836 mg/L). Wurtz and Bridges [22] generated 96-h chloride LC50s for *Sphaerium tenue* and *Gyraulus circumstriatus* of 682 and 1,941 mg Cl<sup>-</sup>/L, respectively, at a hardness of 100 mg/L. Although we tested related species (*S. sphaerium* and *G. parvus*) at both lower (50 mg/L) and higher (200 mg/L) hardnesses, our LC50 values were all higher than the respective values reported by Wurtz and Bridges [22]. In tests with *Tubifex* 

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*tubifex*, Kangharot [27] generated a 96-h chloride LC50 of 1,203 mg/L at a hardness of 245 mg/L. Again, this value was much lower than ours despite the hardness for both of our tests being lower.

Although the influence of water hardness on chloride toxicity has not been tested for a freshwater fish species, and despite the limited dataset for three of the five species evaluated in the current study, these results indicate that acute chloride toxicity expressed as a function of water hardness is likely appropriate. Given this finding, conducting tests using the K, Ca and magnesium chlorides to determine whether the relationship established here between acute chloride toxicity and water hardness also applies to these other (K, Ca, and Mg) chloride salts as well may be prudent.

The apparent lack of a relationship between acute chloride toxicity and water hardness for *G. parvus* warrants additional research. *Gyraulus parvus* (family Planorbidae) is in the subclass Pulmonata, and although these species are characterized as aquatic snails, they lack gills. Gaseous exchange is via a large pulmonary sac in the mantle cavity. As such, the mechanism of chloride exchange in this species is likely different from that of the other aquatic invertebrate species tested. However, Wurtz and Bridges [22] tested another pulmonate snail (*Physa heterostropha*) at hardnesses of 20 and 100 mg/L and generated 96-h chloride LC50s of 2,487 and 3,444 mg/L, respectively, suggesting that hardness did ameliorate chloride toxicity in that species.

Effect of water hardness on acute chloride toxicity to C. dubia

As indicated, in experiments with C. dubia, increasing hardness reduced the toxicity of chloride. The relationship between hardness and LC50 in terms of milligrams C17/L was best described as a power function, with the steepest part of the line occurring between hardness of 25 and 200 mg/L as CaCO<sub>3</sub> (Fig. 1). A number of other researchers have observed that increased concentrations of Ca, and to a lesser extent Mg, reduce the toxicity of Na-dominated, high TDS solutions to freshwater species [7,9,11,14,15,22], but to our knowledge, the current study is the first to extensively quantify this relationship for sodium chloride. In the current study, the hardness solutions were adjusted using a constant Ca:Mg ratio (2.25 based on mass). Increasing hardness using only Mg or with a different Ca:Mg ratio may produce different results. Furthermore, our data show that increasing Ca and Mg reduced the toxicity of sodium chloride. Our data do not necessarily support the general claim that increasing total hardness reduces the toxicity of chloride, because we did not investigate how additional cations impact the toxicity of CaCl<sub>2</sub>, MgCl<sub>2</sub> chloride, or KCl. Mount et al. [11] provided an excellent illustration of this point by generating LC50s for NaCl, CaCl<sub>2</sub>, and MgCl<sub>2</sub>. Although the Ca and Mg salt solutions would have higher hardnesses than the Na salt solutions, in terms of mg Cl<sup>-</sup>/L, the 48-h LC50s for C. dubia were 1,187, 1,172, and 655 for NaCl, CaCl<sub>2</sub>, and MgCl<sub>2</sub>, respectively [11].

The precise mechanism for the reduction in sodium chloride toxicity to freshwater invertebrates at elevated Ca and Mg concentrations is unknown. Others have shown that Ca reduces membrane permeability to both water and ions. This has been shown in physiological studies with fishes [32–34], and Robertson [35] reviewed studies providing evidence of the same effect in invertebrates nearly 70 years ago. In most cases, these studies dealt with the problem of animals losing ions in freshwater, but in 1932, Pruthi [36] proposed that unusually high Ca concentrations in saline waters of the Salt Range,

Punjab, allowed for their colonization by mayflies and hydrophilid beetles, normally freshwater insects. Calcium and Mg are known to reduce the toxicity of metals by competing for binding sites. These divalent cations could have been competing with Na in the solutions used in the current study, which would explain the reduced toxicity of NaCl at higher hardnesses, but Mount et al. [11] observed that Ca and Na were not significant variables in models developed to predict toxicity attributable to major ions, so they concluded that toxicity attributable to salts of these cations were attributable to the corresponding anion. That suggests that competition at binding sites is not likely the sole mechanism for the reduced toxicity observed at higher hardnesses in the current study. Furthermore, analysis of our nominal ion concentrations using Visual MIN-TEQ (data not shown) suggests that at least 98% of chloride would exist in our solutions as free Cl ion, even at high hardnesses, suggesting that complexation of the toxic ion is not the mechanism either. Therefore, the most likely explanation for this phenomenon is that increased concentrations of Ca, and to a lesser extent, Mg (as shown by Potts and Fleming [34]) tighten cellular junctions, thereby reducing the passive diffusion of chloride ions into the animal (acknowledging that not all movement of chloride into freshwater animals is passive) and the energy required for ionoregulation.

Effect of sulfate on acute chloride toxicity to C. dubia

Although hardness had a strong effect on chloride toxicity to C. dubia and a variety of other invertebrates in the current study (except G. parvus), varying sulfate concentration from 26 to 712 mg/L had a much weaker, statistically insignificant effect in tests with C. dubia. This relationship was investigated because Soucek [15] observed a strong influence of chloride on sodium sulfate toxicity; at chloride concentrations ranging from 5 to 33 mg/L, sulfate toxicity to Hyalella azteca decreased substantially, whereas, for both H. azteca and C. dubia, increasing chloride from approximately 33 to 500 mg/L resulted in lower sulfate LC50s. The trend over the higher chloride range suggested that additive toxicity was observed with chloride concentrations reaching toxic levels, thereby requiring less sulfate to generate an LC50. In the current study, a nominally negative slope in the plot of chloride LC50 versus dilution water sulfate concentration was seen, again suggesting an additive effect. Plotting the same data in terms of TDS resulted in a significant positive slope (Fig. 2), reflecting the relatively constant chloride LC50 at the increasing sulfate concentrations in the dilution water.

Sulfate over a lower concentration range was not expected to affect chloride toxicity as was observed by Soucek [15] in the opposite scenario (increasing chloride from 5 to 33 mg/L decreased sulfate toxicity), because this phenomenon was likely a product of the fact that *H. azteca* simply performs better in water-only toxicity tests when the dilution water has a higher chloride concentration than, for example, is provided in the U.S. EPA's [28] moderately hard reconstituted water has a nominal chloride concentration of 1.9 mg/L, and *H. azteca* performs poorly in toxicity tests using this water ([37]; D. J. Soucek, personal observation). As a result, others have developed reconstituted water recipes with higher chloride concentrations to achieve better test performance with this species [37,38].

Finally, the plot of conductivity at LC50 versus hardness of dilution water (Fig. 4) was included to illustrate the point that when assessing the impacts of elevated major ion concentrations on freshwater organisms, surrogate measures do not

provide adequate information. Others have observed correlations between TDS or conductivity and changes in benthic macroinvertebrate communities, with widely disparate results. For example, Kennedy et al. [5] observed an impairment threshold of 3,700 µmhos/cm, whereas Pond et al. [39] saw effects at approximately 500 µmhos/cm. This difference was attributed to variation in sensitivity of endemic communities [39], but in our experiments with a single species, the conductivity at the LC50 for chloride varied from less than 3,000 to nearly 6,000 µmhos/cm, depending on the hardness of the water. In a similar analysis using data generated in sodium sulfate toxicity tests with C. dubia and H. azteca, conductivities at the respective LC50s ranged from approximately1,000 to more than 8,000 µmhos/cm [15]. These large ranges are a result of the fact that TDS toxicity is highly dependent on the specific ionic composition of the water. Less is known about the toxicity of chloride or sulfate when sodium is not the dominant cation; however, as stated previously, the toxicity of K, Ca, and MgCl<sub>2</sub> is generally more profound. For example, Mount et al. [11] generated a chloride LC50 of 655 mg/L for C. dubia using MgCl<sub>2</sub>. Bicarbonate is more toxic than both sulfate and chloride, with 8-d EC50s as low as 289 mg HCO<sub>3</sub>/L [11,24,25,40]. High TDS solutions dominated by Mg salts or bicarbonates would generate LC50s at much lower TDS concentrations or conductivities than NaCl<sub>2</sub> or sulfate-dominated solutions. Evidence from the current study and the literature clearly support the importance of knowing the composition of water with high conductivity before predicting its toxicity.

#### CONCLUSIONS

Acute toxicity of chloride at different hardness (Ca and Mg) levels was determined for five freshwater invertebrate species with the following rank order of LC50s: S. simile < C. dubia < M. transversum < G. parvus < T. tubifex. Increasing hardness reduced toxicity of chloride to C. dubia by a factor of approximately 2 over the range 25 to 800 mg/L. The relationship was best described by the following power function:

## $LC50 (mgCl^{-}/L) = 446.6191 (hardness in mg/L)^{0.2144}$

The acute toxicity of chloride to two other invertebrates species (*T. tubifex* and *S. simile*) was similarly affected between the range of 50 and 200 mg/L hardness. Conversely, increasing sulfate from 25 to 600 mg/L did not significantly affect acute chloride toxicity to *C. dubia*. Because toxicity in terms of TDS or conductivity can vary widely depending on ion composition, measuring toxicity in terms of specific ions, that is, chloride, sulfate, bicarbonate, Ca, Mg, Na, K, is the most useful and practical mechanism to derive protective criteria for freshwater organisms from the adverse effects of TDS.

Acknowledgement—The authors thank Martin Knapp and Mick Micacchion (Ohio Environmental Protection Agency) for their assistance in locating and identifying *Gyraulus parvus* for organism collection and subsequent toxicity testing. The authors also thank Charles Stephan (U.S. EPA) for comments related to study design and results. This study was funded by the U.S Environmental Protection Agency.

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## **Heather Ferguson**

From: Kesler, Karen [Kesler.Karen@epa.gov] Sent: Friday, November 04, 2016 2:29 PM

To: PJD; 'Heather Ferguson'

Cc: 'Sarah Clem'; 'James Malcolm'; Crocker, Philip

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA

in the Red River in Arkansas

Hi Pat,

Yes, thank you for the clarification and additional papers.

- Karen

From: PJD [mailto:pjd@ftn-assoc.com] Sent: Friday, November 04, 2016 2:27 PM

To: Kesler, Karen < Kesler. Karen@epa.gov>; 'Heather Ferguson' < hlf@ftn-assoc.com>

Cc: 'Sarah Clem' <clem@adeq.state.ar.us>; 'James Malcolm' <jtm@ftn-assoc.com>; Crocker, Philip

<crocker.philip@epa.gov>

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

Karen,

"More than one chemical" refers to a study (maybe 2) that looked at ions plus metals. In those studies the main toxicant was the metal so they were not relevant to this analysis. I have attached several studies that look at combinations of ions.

We hope that this provides sufficient information for you to complete your review.

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Pat Downey

Senior Project Manager FTN Associates, Ltd. 3 Innwood Circle Little Rock, AR 72211

tel 501-225-7779 fax 501-225-6738 cell 501-860-4447 pjd@ftn-assoc.com

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

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Sent: Friday, November 04, 2016 2:03 PM

To: Heather Ferguson

Cc: 'Sarah Clem'; James Malcolm; Patrick Downey; Crocker, Philip

Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

Hi Heather,

Yes, the supplemental information that you have submitted adequately addresses my request for lethal and sublethal minerals tolerance values. I appreciate you putting this together and especially pulling out the associated hardness values with these studies where you could find them.

The one additional question that I have is that I noticed in your filtering of the data that you removed any tests that used more than one chemical. Did any of these tests with two chemicals involve both chlorides and sulfates? If so, those would probably be useful data to see as well.

Thanks, Karen

From: Heather Ferguson [mailto:hlf@ftn-assoc.com]
Sent: Wednesday, November 02, 2016 2:15 PM
To: Kesler, Karen < Kesler.Karen@epa.gov>

Cc: 'Sarah Clem' < <a href="mailto:clem@adeq.state.ar.us">clem@adeq.state.ar.us</a>; James Malcolm < <a href="mailto:jtm@ftn-assoc.com">jtm@ftn-assoc.com</a>; Patrick Downey < <a href="mailto:pid@ftn-assoc.com">pid@ftn-assoc.com</a>> Subject: RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

Hi Karen,

I just wanted to check in with you regarding the supplemental information we provided to you in September in support of the third-party rulemaking for SWEPCO (for dissolved minerals in the lower Red River in Arkansas). We need to proceed with the next steps in the third-party rulemaking and would like to confirm that the supplemental information we submitted adequately addressed your request regarding lethal and sublethal tolerance values for minerals.

Please let us know at your earliest convenience. If you have questions or comments regarding the information, please do not hesitate to call Jim Malcolm, Pat Downey, or me at (501) 225-7779. Thank you!

Kindest regards,



Heather Ferguson FTN Associates, Ltd.

3 Innwood Circle, Suite 220 **№** Little Rock, AR 72211 hlf@ftn-assoc.com

(501) 225-7779 **%** fax (501) 225-6738 http://www.ftn-assoc.com

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov] Sent: Thursday, September 29, 2016 11:34 AM

To: Heather Ferguson

**Subject:** RE: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in Arkansas

Hi Heather,

Thank you for sending this information over. Once I have a chance to review it, I will let you know if I have any questions or comments.

Thanks,

#### Karen

From: Heather Ferguson [mailto:hlf@ftn-assoc.com]

Sent: Tuesday, September 27, 2016 2:19 PM

To: Kesler, Karen < Kesler.Karen@epa.gov >; Crocker, Philip < Crocker.philip@epa.gov >; Nelson, Russell

<nelson.russell@epa.gov>

**Cc:** 'Allan Gates' < <u>AGates@mwlaw.com</u>>; 'Frank Mills' < <u>flmills@aep.com</u>>; Patrick Downey < <u>pjd@ftn-assoc.com</u>>; James

Malcolm < itm@ftn-assoc.com>; 'Sarah Clem' < clem@adeq.state.ar.us>

Subject: Toxicity Endpoints and Hardness Data in support of SWEPCO Dissolved Minerals UAA in the Red River in

Arkansas

Good afternoon Karen,

On behalf of Southwestern Electric Power Company, please find attached the requested supplemental information for dissolved minerals tolerance values.

The attached information includes a memorandum that briefly describes our approach to obtaining the information you requested and a spreadsheet summarizing lethal and sublethal toxicity endpoints and associated hardness values. We believe this information satisfies your request and demonstrates through a weight-of-evidence approach the protection of aquatic life in the downstream reach of the Red River.

If you have any questions or comments regarding the attached information, please do not hesitate to call Jim Malcolm, Pat Downey, or me at (501) 225-7779.

Respectfully submitted,



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## **Heather Ferguson**

From: Sent:

Jim Malcolm [itm@ftn-assoc.com] Friday, November 04, 2016 3:16 PM

To: Cc: 'Kesler, Karen'; 'Clem, Sarah'

Subject:

Patrick J. Downey; "Heather Ferguson"; 'Crocker, Philip'; James Malcolm

SWEPCO Red River SSC for TDS

Hi Karen,

Thanks for the quick reply to Pat. I scanned back through some relevant email threads to review where we are at now regarding SWEPCO's revised proposal for a site specific criterion for TDS in the lower Red River (see below). I believe, from reading your most recent email to Pat (just received), that we have now addressed your request for the supplemental information (see your July 26, 2016 email below). If my interpretation is correct and you have a chance, would you give us an indication affirming that? We'd like to finalize the package of supplemental information to be included in the re-opened rulemaking to give to the AR Pollution Control & Ecology Commission. Of course, you will see all of this again once it winds its way through the rulemaking process here in AR and gets back to EPA for final review.

Thanks, Jim

From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Tuesday, July 26, 2016 11:15 AM

To: Jim Malcolm

Subject: RE: Additional SWEPCO information

Thanks Jim,

I'll let you know if we have any further questions. Again, thank you for putting the additional information together.

#### - Karen

From: Jim Malcolm [mailto:jtm@ftn-assoc.com]

Sent: Tuesday, July 26, 2016 11:13 AM To: Kesler, Karen < Kesler. Karen@epa.gov>

Cc: 'Clem, Sarah' < CLEM@adeq.state.ar.us >; Crocker, Philip < crocker.philip@epa.gov >; 'Philip Massirer' < phm@ftn-

assoc.com>; Patrick J. Downey <pid@ftn-assoc.com>; James Malcolm <jtm@ftn-assoc.com>

Subject: RE: Additional SWEPCO information

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From: Kesler, Karen [mailto:Kesler.Karen@epa.gov]

Sent: Tuesday, July 26, 2016 10:46 AM

To: Jim Malcolm

Cc: Clem, Sarah; Crocker, Philip

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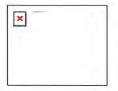
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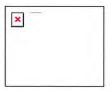
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To:

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Cc:

'Crocker, Philip': 'Sarah Clem': James Malcolm

Subject:

RE: SWEPCO Red River SSC for TDS

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Have a good week, Jim

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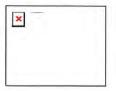
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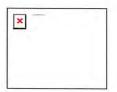
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